

Rearrangement of the T-cell Receptor Delta Chain Gene in T-cell Lymphomas with a Mature Phenotype

J. H. J. M. van Krieken,* Lori Elwood,*
Rafael E. Andrade,* Elaine S. Jaffe,* Jeffrey
Cossman,† and L. Jeffrey Medeiros*

From the Laboratory of Pathology,* Section of
Hematopathology, National Cancer Institute, Bethesda,
Maryland; and the Department of Pathology,† Georgetown
University Medical Center, Washington, DC

The configuration of the T-cell receptor (TCR) delta chain gene was assessed using restriction fragment analysis and the Southern blot technique in 39 T-cell lymphomas with a mature immunophenotype. The TCR δ gene was rearranged in four lymphomas although the γ/δ TCR was not expressed in two cases studied. The TCR δ gene was the only TCR gene rearranged in two cases. Each lymphoma with TCR δ gene rearrangement had an aberrant T-cell immunophenotype and three cases were of the large cell anaplastic type. The TCR δ gene was deleted in 22 cases and was in the germline configuration in 13 lymphomas. Deletion of the TCR δ gene was characteristic of mycosis fungoides, adult T-cell leukemia/lymphoma (human T cell leukemia-lymphoma virus positive), and Lennert's lymphoma, and was not identified in angiocentric lymphomas. In eight cases with TCR δ deletion, however, a large number of polyclonal (presumably reactive) T cells were present and, in these lymphomas, the authors could not determine if TCR δ gene deletion occurred in the polyclonal T cells, the neoplastic cells, or both cell populations. The authors conclude that the TCR δ gene is usually deleted in mature T-cell lymphomas, as would be expected in α/β TCR T cells. However, TCR δ gene rearrangement is detectable in approximately 10% of cases. Analysis of this locus may be useful diagnostically, as it occasionally may be the only molecular marker of clonality in mature T-cell lymphomas. T-cell receptor delta chain gene rearrangement also is found most often in lymphomas of the large cell anaplastic type. (Am J Pathol 1991, 139:161-168)

Mature T-cell lymphomas form a heterogeneous group of malignant neoplasms that may be difficult to diagnose using morphologic and immunophenotypic criteria.^{1,2} Histologically a spectrum of cell sizes and types are typically found in T-cell lymphomas, including many non-neoplastic cells, making differentiation of T-cell lymphomas from reactive lesions problematic. Immunophenotypic analysis also may not be definitive because T cells do not express a clonal marker analogous to immunoglobulin light chain expression by B cells.^{1,2}

Gene rearrangement analysis of the T-cell receptor (TCR) genes has been shown to be a useful method for distinguishing reactive from malignant T-cell infiltrates.²⁻⁴ There are two TCRs, α/β and γ/δ , encoded by four genes that each rearrange during T-cell development before expression of the polypeptide chains that constitute the TCR.²⁻⁴ Studies of the α -chain locus have been limited because of the large size of this gene.^{2,4} Both the TCR β and γ genes are often rearranged in T-cell leukemias and lymphomas.⁵⁻¹⁰ Up to 20% of T-cell lymphomas, however, may be germline for the TCR β gene.⁹ In contrast, the genomic configuration of the recently described TCR δ gene, although well described in T-cell lymphoblastic lymphomas and leukemias,¹¹⁻¹⁵ is less understood in mature T-cell lymphomas.^{16,17}

In this study, we analyzed the TCR δ gene in 39 mature T-cell lymphomas, diagnosed by conventional histologic and immunophenotypic criteria, to assess its possible value in the diagnosis of these neoplasms.

Materials and Methods

Thirty-nine malignant lymphomas with a mature T-cell immunophenotype were analyzed. Each neoplasm was accessioned in the Hematopathology Laboratory of the Na-

Dr. van Krieken was a visiting fellow from the Department of Pathology, University of Leiden, the Netherlands, supported by the Giesela Thier fund at the time this work was done.

Accepted for publication February 21, 1991.

Address reprint requests to L. Jeffrey Medeiros, MD, Building 10, Room 2N108, National Cancer Institute, Bethesda, MD 20892.

tional Cancer Institute and was selected on the basis of frozen tissue or cells being available for DNA extraction and Southern blot analysis. All cases were diagnosed using histopathologic and immunophenotypic criteria. According to the Working Formulation,¹⁸ every case was diffuse: 13 cases were mixed small and large cell type, 23 lymphomas were large cell immunoblastic, two lesions were mycosis fungoides (MF) and one neoplasm was diffuse but not further classified. In addition, 25 lymphomas were further specified in the following manner: 4 cases were angiocentric lymphomas (ie, angiocentric immunoproliferative lesions, grade III),¹⁹ 5 lesions had the morphologic features of Lennert's lymphoma,²⁰ 4 lymphomas occurred in the setting of human T-cell leukemia-lymphoma virus infection,²¹ and 12 cases were of the large cell anaplastic type.²²

Immunophenotypically, all neoplasms had a mature T-cell immunophenotype as defined by the expression of one or more pan-T-cell antigens (CD2, CD3, CD5, or CD7) and the absence of TdT or the CD1 antigen, immunoglobulin heavy and light chains, and pan-B-cell antigens. Twenty-eight tumors (72%) did not express one or more pan-T-cell antigens as described by others.¹ Immunophenotyping was performed using either cell suspensions and flow cytometric analysis or frozen section immunohistochemistry as previously described.²³ The core panel of monoclonal antibodies used included: Leu-1 (CD5), Leu-4 (CD3), Leu-3a (CD4), Leu-2a (CD8) (Becton-Dickinson, Sunnyvale, CA), T11 (CD2), T6 (CD1), B1 (CD20), B4 (CD19), (Coulter Monoclonal Antibodies, Hialeah, FL), 3A1 (CD7) (courtesy of B.F. Haynes, MD), and immunoglobulin light and heavy chains (kappa, lambda, mu, gamma, alpha) (Bethesda Research Labs, Bethesda, MD). The CD30 antigen (using either the Ki-1 or BerH2 antibodies; DAKO, Santa Barbara, CA) was assessed in 33 cases and was positive in 13 cases, including all 10 large cell anaplastic lymphomas studied. In addition, in 10 cases (including two with TCR δ gene rearrangement), expression of the γ/δ TCR was assessed using the TCR δ -1 and TCS-1 antibodies (T Cell Sciences, Cambridge, MA) that react with V δ 1 and a TCR δ framework antigen, respectively.¹⁵ These cases were also stained with the BF-1 antibody (T Cell Sciences), which reacts with a framework antigen of the α/β TCR.

Southern Blot Analysis

High-molecular-weight DNA was extracted from frozen tissue blocks or cell suspensions using a standard proteinase K phenol/chloroform technique. After purification, 10 to 15 μ g of DNA were completely digested using the *Eco*R1, *Hind*III, and *Bam*H1 restriction enzymes. The digested DNA was electrophoresed in 0.7% agarose gels, deperinated in 0.25 N HCl, and transferred in 0.4 N

NaOH²⁴ to nylon membranes (Gene Screen Plus, New England Nuclear Research Products, Boston, MA) The membranes were sequentially hybridized at high stringency in 50% formamide at 42°C with DNA probes radiolabeled with P32 using the random primer method.²⁵ The probes used in this study have been described previously. The TCR β gene was assessed with a cDNA probe of the constant region (C β).²⁶ The JH,²⁶ J δ 1¹⁵ and J γ 1.3²⁷ probes were genomic. Because J δ 1 is the predominantly used J region of the TCR δ locus,^{17,28,29} a probe of this region would be expected to detect most productive rearrangements as well as deletions of this region. Deletion of the TCR δ gene was determined by visual comparison of the intensity of the germline J δ 1 band with the germlines obtained with the other probes on the same blots.

Results

The clinicopathologic and genotypic data are listed in Tables 1 and 2.

TCR δ Gene Rearrangements

The TCR δ gene was rearranged in four lymphomas (10%), and the rearranged restriction fragments were variously sized in each case (Figure 1). In three lymphomas, a single rearranged band and a germline band were identified. In one neoplasm, two rearranged bands and a faint germline band were identified. In two cases, both the TCR β and TCR γ chain genes were also rearranged, whereas in the other two lymphomas, the TCR β and TCR γ genes were germline.

Three lymphomas with TCR δ gene rearrangements were classified histologically as large cell immunoblastic in the Working Formulation and were also of the large cell anaplastic type. Immunophenotypically all cases with TCR δ rearrangement had an abnormal T-cell phenotype. Each neoplasm was CD5 antigen negative. In addition, one lymphoma also lacked the CD3 antigen, in one lymphoma the CD2 and CD7 antigens were also negative, and in one lymphoma the CD3 and CD7 antigens as well CD5 antigen were absent. Each case was strongly CD30 antigen positive.

TCR δ Gene Deletion

In 22 lymphomas, the TCR δ gene (more precisely, the J δ 1 region) was deleted. In 19 of these cases, the TCR β gene was rearranged and 16 of these lymphomas also had rearrangement of the TCR γ gene. In three T-cell lymphomas, deletion of the TCR δ gene was the only geno-

Table 1. Summary of Clinicopathologic, Immunophenotypic, and Genotypic Data

Case	Biopsy site	Histologic diagnosis*	Additional features	Immunophenotype			
				CD2	CD3	CD5	CD7
1742	Paraspinal mass	LCIB	LCAL	-	+	-	-
1807	LN	LCIB	LCAL	+	+	-	+
2005	Skin	LCIB	LCAL	+	-	-	-
2014	Nasopharynx	DM	AL	+	-	-	+
2073	Skin	MF		+	+	+	-
2094	LN	MF		+	+	+	-
856	Skin	DM	LL	+	+	ND	ND
933	LN	LCIB	LL	+	+	+	+
1315	LN	DM	LL	+	-	+	+
2239	Skin	DM	LL	+	+	-	+
862	LN	LCIB	ATL	+	+	+	ND
1148	LN	DM	ATL	+	+	+	-
1372	LN	DM	ATL	+	+	+	-
1894	LN	LCIB	ATL	+	+	+	-
757	LN	LCIB		+	-	ND	ND
1436	LN	DM		+	+	-	-
1485	LN	DM		+	-	+	-
1666	LN	LCIB		ND	+	-	-
2063	LN	LCIB		+	+	-	+
2119	LN	LCIB		+	+	+	+
1092	Skin	LCIB	LCAL	+	+	+	-
1254	LN	LCIB	LCAL	+	+	+	+
1685	Skin	LCIB	LCAL	+	+	-	-
1891	LN	LCIB	LCAL	+	+	-	-
1893	Flank mass	LCIB	LCAL	+	+	-	+
2139	LN	LCIB	LCAL	ND	+	-	+
1595	Nasopharynx	DU	AL	+	-	-	+
1887	Liver	DM	AL	+	+	+	+
2400	Lung	LCIB	AL	+	+	+	+
1378	LN	DM	LL	+	+	+	ND
1000	LN	LCIB	LCAL	+	+	+	+
1222	LN	LCIB	LCAL	+	-	+	-
1658	LN	LCIB	LCAL	-	-	-	+
291	LN	DM		+	+	ND	ND
739	LN	LCIB		+	-	-	-
834	LN	DM		+	-	ND	ND
1565	Skin	LCIB		+	-	-	+
2194	Skin	DM		+	+	+	+
2345	Skin	LCIB		+	+	-	-

* Working formulation. + TCR δ expression.

NC, neoplastic cells; BTC, benign T cells (% positive); LN, lymph node; MF, mycosis fungoides; DM, diffuse mixed small- and large-cell lymphoma; AL, angiocentric lymphoma (angiocentric immunoproliferative lesion, grade III); LL, Lennert's lymphoma; ATL, adult T-cell leukemia/lymphoma (HTLV-I+); LCIB, large-cell immunoblastic lymphoma; LCAL, large-cell anaplastic lymphoma; DU, diffuse unclassified lymphoma; D, deletion; R, rearrangement; P, TCR γ gene polyclonal T cell pattern; R**, TCR γ gene rearrangement and polyclonal T-cell pattern; G, germline; ND, not done.

mic change detected with these probes. In addition to the presence or absence of TCR γ gene rearrangement, analysis with the J γ probe demonstrated up to eight bands in eight of these lymphomas, indicating the presence of a substantial number of polyclonal (non-neoplastic) T cells, constituting at least 10% of the cell population in these cases.²⁷ Because it is difficult to determine the percentage of admixed non-neoplastic T cells, either by morphologic or immunophenotyping methods, it is uncertain whether the TCR δ gene deletion occurred in the neoplastic cells, the polyclonal cells, or both cell populations.

The lymphomas with TCR δ gene deletion were classified in the working formulation as follows: seven diffuse mixed small and large cell, 13 large cell immunoblastic,

and two MF. T-cell receptor delta chain gene deletion was identified in all cases of MF and adult T-cell leukemia/lymphoma and in four of five Lennert's lymphomas. Six of twelve large cell anaplastic lymphomas and 6 of 13 cases of T-cell lymphomas not further characterized as a clinicopathologic entity also had deletion of the TCR δ gene. There was no correlation between immunophenotype and TCR δ gene deletion. The incidence of pan-T-cell antigen absence was similar in cases with and without deletion of the TCR δ gene.

TCR δ Gene Germline

In 13 T-cell lymphomas, the TCR δ gene was in the germline configuration. In five of these tumors, the TCR β

Table 2. Summary of Clinicopathologic, Immunophenotypic, and Genotypic Data

Case	Immunophenotype			TCR δ +		Genotype			
	CD4	CD8	Ig/pan-B	NC	BTC	J δ	C β	J γ	J μ
1742	+	-	-			R	R/R	R	G
1807	-	-	-	-	<5%	R	R/G	R	G
2005	+	-	-			R	G/G	G	G
2014	-	-	-	-	<5%	R	G/G	G	G
2073	+	-	-			D	R/G	R	G
2094	+	-	-			D	R/G	R	G
856	+	-	-			D	R/G	R	G
933	+	-	-	-	<5%	D	R/R	P	G
1315	+	-	-	-	<5%	D	G/G	P	G
2239	+	-	-			D	R/G	R	G
862	+	-	-			D	R/G	R	G
1148	-	-	-			D	R/R	R	G
1372	+	-	-			D	R/R	R	G
1894	+	-	-			D	R/R	R**	G
757	-	+	-			D	R/R	R	G
1436	+	-	-			D	R/R	R**	G
1485	+	-	-	-	<5%	D	R/R	R**	G
1666	+	-	-			D	R/G	R	G
2063	-	-	-	-	<5%	D	R/R	R	G
2119	-	+	-			D	R/R	G	G
1092	+	-	-			D	R/R	R	G
1254	+	-	-	-	<5%	D	G/G	P	G
1685	+	-	-			D	G/G	P	G
1891	+	-	-	-	<5%	D	R/R	P	G
1893	+	-	-			D	R/R	R	G
2139	+	-	-			D	R/G	R	G
1595	-	-	-			G	G/G	G	G
1887	+	-	-			G	G/G	P	G
2400	+	-	-			G	G/G	P	G
1378	ND	ND	-			G	R/R	P	G
1000	+	-	-			G	R/G	P	G
1222	+	-	-			G	G/G	P	G
1658	+	-	-			G	G/G	G	G
291	ND	ND	-			G	R/G	R	G
739	+	ND	-	-	<5%	G	R/G	R	G
834	+	-	-			G	R/R	P	G
1565	+	ND	-	-	<5%	G	G/G	G	G
2194	+	-	-			G	G/G	P	G
2345	-	+	-			G	G/G	P	G

Abbreviations can be found in Table 1.

gene was rearranged (two that also had TCR γ gene rearrangements). Analysis with the J γ probe also demonstrated numerous bands indicating the presence of numerous polyclonal T cells in eight cases.

In eight T-cell lymphomas, all TCR genes were in the germline configuration. These cases included three angiocentric lymphomas and two large cell anaplastic lymphomas.

TCR δ Expression

Expression of the TCR δ protein (and presumably the γ/δ TCR) was assessed in 10 lymphomas in which frozen tissue was available for additional immunohistochemical staining (Figure 2). Each case had been previously stained with an anti-CD3 antibody. The cases studied included two lymphomas with TCR δ gene rearrangement (cases 1807 and 2014; both CD30+, 1 CD3+), six lymphomas

with TCR δ gene deletion, five of which had >10% polyclonal T cells (cases 933, 1254, 1315, 1485, 1891, and 2063; five of six CD3+), and two lymphomas with the TCR δ gene in the germline configuration (cases 739, 1565; both CD3-). In all cases (including the two lymphomas with TCR δ gene rearrangement), the neoplastic cells did not react with either the TCR δ -1 or TCS-1 antibodies. The neoplastic cells in three lymphomas with TCR δ gene deletion were stained by BF-1 (cases 933, 1315, and 2063; all CD3+) indicating α/β TCR lineage. The tumor cells were BF-1 negative in seven cases (739, 1254, 1485, 1565, 1807, 1891, and 2014; three of seven CD3+), including both neoplasms with TCR δ gene rearrangement (cases 1807 and 2014). The majority of the non-neoplastic T cells in each biopsy were stained by the BF-1 antibody, whereas less than 5% (visual estimate) of these cells reacted with the TCR δ -1 and TCS-1 antibodies.

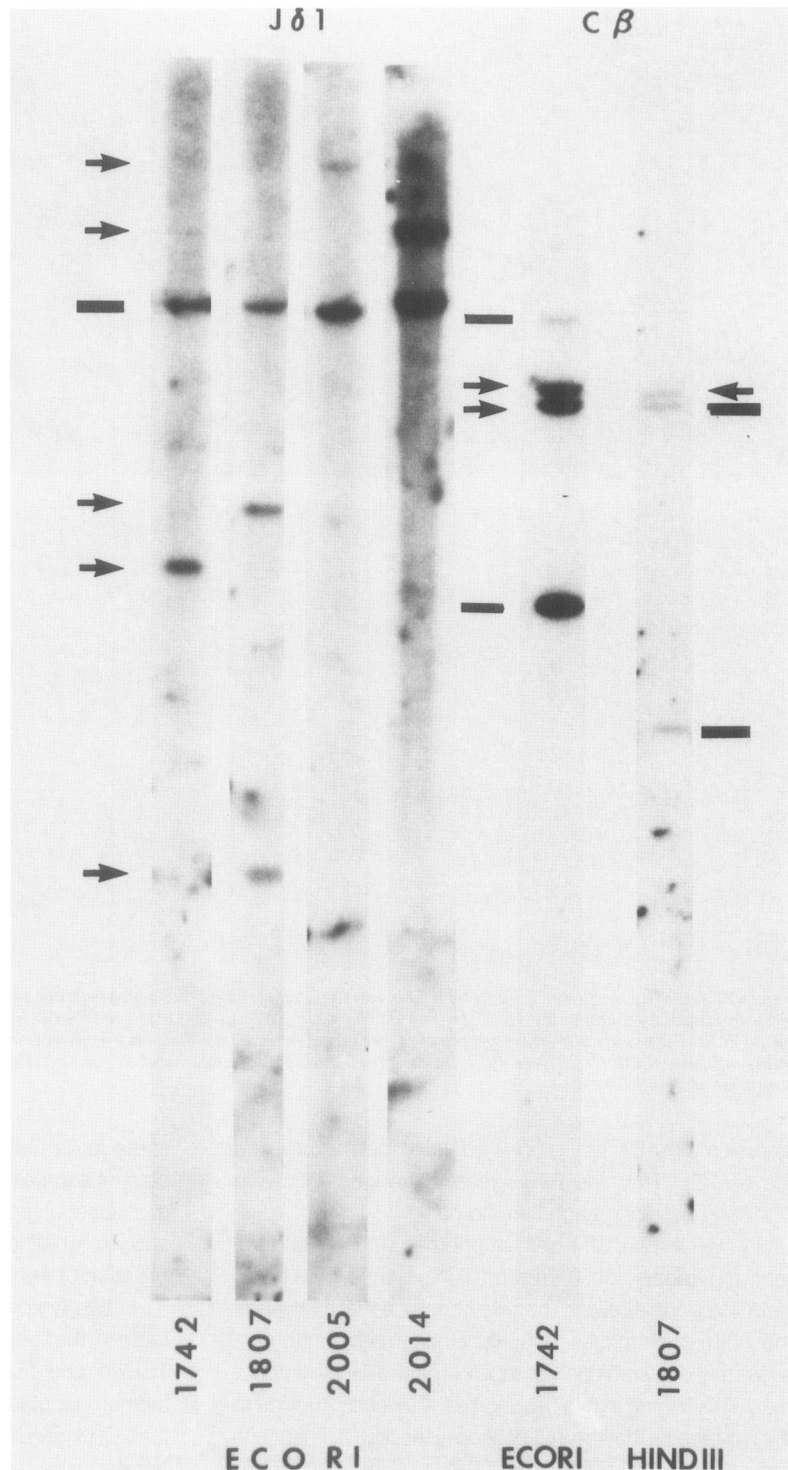


Figure 1. Four of thirty-nine (10%) mature T-cell lymphomas (cases 1742, 1807, 2005, and 2014) had rearrangements of the TCR δ gene. The rearrangements were of different sizes in each case. Case 1807 had two rearrangements. Cases 1742 and 1807 also had rearrangements of the TCR β gene.

Discussion

In this study, we have identified rearrangements of the TCR δ gene in 4 of 39 (10%) T-cell lymphomas with a mature immunophenotype. Although we have not shown directly that the TCR δ gene rearrangements occurred in the neoplastic and not the non-neoplastic (presumably

reactive) T cells, we believe that the rearrangements are likely to have occurred in the neoplastic cell population for the following reasons: First in each case with TCR δ gene rearrangement the neoplastic cells were the predominant cell population (more than 50% of all cells were neoplastic by visual estimate). Second in two cases stained with the TCR δ -1 and TCS-1 antibodies, less than

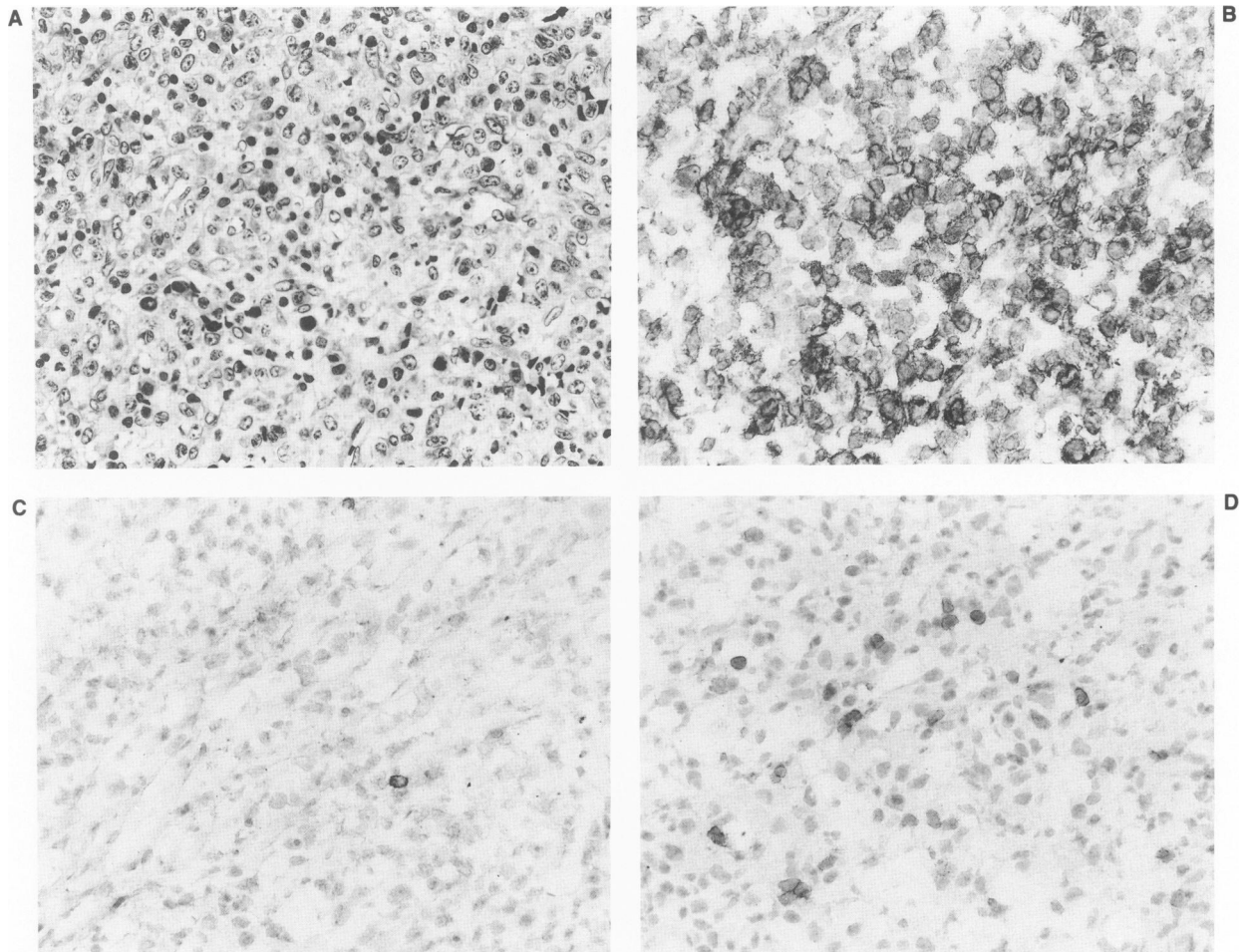


Figure 2. A CD30+ large-cell anaplastic lymphoma (case 1807) with TCR δ gene rearrangement. **A:** The neoplastic cells have bizarre nuclear features and a high mitotic rate. (H&E, $\times 400$). **B:** The tumor cells strongly expressed the CD30 antigen. **C:** The neoplastic cells were unstained by the TCR δ -1 antibody. Rare non-neoplastic T cells were positive. **D:** The tumor cells also were not stained by the BF-1 antibody, although many non-neoplastic cells were BF-1 positive and thus, presumably reactive $\alpha\beta$ T cells. (B–D, immunoperoxidase with hematoxylin counterstain, $\times 400$).

approximately 5% of the non-neoplastic cells expressed the γ/δ TCR. Third the TCR δ gene rearrangements were of different sizes in each lymphoma.

In two of the four T-cell lymphomas with TCR δ gene rearrangements, the TCR β and TCR γ genes were in the germline configuration. Thus we conclude that analysis of the TCR δ gene locus is a useful clonal marker in the assessment of a small percentage of mature T-cell lymphomas. This finding in two lymphomas also lends support to the hypothesis that the TCR δ gene is the first TCR gene to rearrange, followed by the TCR γ , TCR β , and TCR α genes.^{13–15,17}

The cases with TCR δ gene rearrangements in this study had certain features in common. All four lymphomas were characterized by bizarre histologic features; three cases were classified as large cell immunoblastic in the Working Formulation and had histologic features typical of the entity large cell anaplastic lymphoma.²² TCR δ gene rearrangements have been shown previously in

some large cell anaplastic lymphomas.¹⁶ All four neoplasms expressed CD30 and had an aberrant T-cell immunophenotype characterized by preservation of CD2 antigen, absence of the CD5 antigen, and variable expression of the CD3 and CD7 antigens.

In two lymphomas with TCR δ gene rearrangements tested, the neoplastic cells were negative with the TCR δ -1 and TCS-1 antibodies whereas less than 5% of the non-neoplastic cells in the biopsy were positive. Thus the TCR δ gene rearrangements in these two lymphomas appear to be nonproductive. In view of the propensity of the TCR δ locus to translocate,³ it is interesting to speculate that some TCR δ gene rearrangements in mature T-cell lymphomas may be a consequence of chromosomal translocation.

The TCR δ chain gene was deleted in 22 of 39 (56%) mature T-cell lymphomas. In three cases, the TCR β and TCR γ genes were in the germline configuration. In these cases, we can not exclude the possibility that only the J δ 1

region was deleted and that rearrangement into the J δ 2 or J δ 3 regions may have occurred as has been rarely reported.^{17,28,29} The J δ 1 probe we used would not address this possibility. In the remaining 19 cases with TCR δ deletion, the TCR β chain gene was also rearranged, implying that the entire TCR δ gene was deleted, as would be expected on the basis of what is known about the δ/α locus in normal T cells.^{15-17,30} The TCR δ gene is located within the TCR α gene locus and is deleted as a prelude to TCR α gene rearrangement in cells that differentiate into α/β T cells. In eight of these lymphomas, however, a substantial number (more than 10%) of polyclonal T cells was also detected with the J γ probe.²⁷ Six of these lymphomas with TCR δ gene deletion, five of which had many polyclonal T cells, were stained with the TCR δ -1 and TCS-1 antibodies. The neoplastic cells did not express TCR δ . In each case, the majority of the non-neoplastic T cells also did not express the γ/δ TCR and were α/β (BF-1+) cells. Thus we cannot determine whether the TCR δ gene deletion seen in these cases occurred in the neoplastic cells, the non-neoplastic α/β T cells, or in both cell populations. Interestingly TCR δ gene deletion may correlate with specific clinicopathologic entities within the spectrum of mature T-cell lymphoma. For example, all cases of adult T-cell leukemia/lymphoma showed deletion of the TCR δ gene, as has been reported by others.¹⁷ Similarly all cases of MF and four of five Lennert's lymphomas had TCR δ gene deletion. Presumably these are neoplasms of α/β T cells.

In 8 of the 39 (20%) cases in this study, all TCR genes were in the germline configuration. Four of these lymphomas did not express CD3. Our results as well as those of others would suggest that some of these cases arise from lymphoid precursor cells, before the stage of gene rearrangement, that subsequently acquire surface markers suggestive of a more mature phenotype.³¹ Alternatively these cases may be oligoclonal or polyclonal T-cell lymphoproliferations.

Four of the lymphomas without gene rearrangements were CD3+. Perhaps CD3 and TCR expression were discordant, as is seen in lymphoblastic neoplasms.³² A second possibility may be that the lesions were not truly malignant lymphoma. These cases were clearly malignant using histologic criteria, however, and one lymphoma had an abnormal T-cell immunophenotype, as is typically associated with malignant lymphoma.¹ Another possible explanation is that the tissue analyzed for rearrangements was not representative of the neoplasm. All four biopsies were taken from extranodal sites (skin, liver, lung). Historically in our laboratory DNA extracted from biopsies from extranodal sites is less often representative of the lesion than DNA extracted from lymph node biopsies.

In summary, in this study the TCR δ gene was deleted

in the majority of mature T-cell lymphomas and rearranged in a small number of cases, approximately 10%. T-cell receptor delta chain gene rearrangement was the only molecular marker of clonality in two cases and thus analysis of this gene may be useful in the assessment of clonality. In addition, TCR δ gene rearrangement was found most often in large cell anaplastic lymphomas.

Acknowledgment

The authors thank Ms. Kim Condron for her help with the immunohistochemical staining.

References

1. Picker LJ, Weiss LM, Medeiros LJ, Wood GS, Warnke RA: Immunophenotypic criteria for the diagnosis of non-Hodgkin's lymphoma. *Am J Pathol* 1987, 128:181-201
2. Knowles DM: Immunophenotypic and antigen receptor gene rearrangement analysis in T cell neoplasia. *Am J Pathol* 1989, 134:761-785
3. Griesser H, Tkachuk D, Reis MD, Mak TW: Gene rearrangements and translocations in lymphoproliferative diseases. *Blood* 1989, 73:1402-1415
4. Cossman J, Uppenkamp M: T-cell gene rearrangements and the diagnosis of T-cell neoplasms. *Clin Lab Med* 1988, 8:31-44
5. Flug F, Pelicci P-G, Bonetti F, Knowles DM, Dalla-Favera R: T-cell receptor gene rearrangements as markers of lineage and clonality in T-cell neoplasms. *Proc Natl Acad Sci USA* 1985, 82:3460-3464
6. Waldmann TA, Davis MM, Bongiovanni KF, Korsmeyer SJ: Rearrangements of genes for the antigen receptor on T cells as markers of lineage and clonality in human lymphoid neoplasms. *N Engl J Med* 1985, 313:776-783
7. Bertness V, Kirsch I, Hollis G, Johnson B, Bunn PA: T-cell receptor gene rearrangements as clinical markers of human T-cell lymphomas. *N Engl J Med* 1985, 313:534-538
8. O'Connor NTJ, Weatherall DJ, Feller AC, Jones D, Pallesen G, Stein H, Wainscoat JS, Gatter KC, Isaacson P, Lennert K, Ramsey A, Wright DH, Mason DY: Rearrangement of the T-cell-receptor β -chain gene in the diagnosis of lymphoproliferative disorders. *Lancet* 1985, i:1295-1297
9. Smith JL, Haegert DG, Hodges E, Stacey GN, Howell WM, Wright DH, Jones DB: Phenotypic and genotypic heterogeneity of peripheral T-cell lymphoma. *Br J Cancer* 1988, 58:723-729
10. Asou N, Matsuoka M, Hattori T, Kawano F, Maeda S, Shimada K, Takasaki K: T cell γ gene rearrangements in hematologic neoplasms. *Blood* 1987, 69:968-970
11. Griesinger F, Greenberg JM, Kersey JH: T cell receptor gamma and delta rearrangements in hematologic malignancies: Relationship to lymphoid differentiation. *J Clin Invest* 1989, 84:506-516
12. Hara J, Benedict SH, Champagne E, Takihara Y, Mak TW,

- Minden M, Gelfand EW: T cell receptor δ gene rearrangements in acute lymphoblastic leukemia. *J Clin Invest* 1988, 82:1974–1982
13. Foroni L, Laffan M, Boehm T, Rabbitts TH, Catovsky D, Luzzatto L: Rearrangement of the T-cell receptor δ genes in human T-cell leukemias. *Blood* 1989, 73:559–565
 14. Dyer MJS: T-cell receptor δ/γ rearrangements in lymphoid neoplasms. *Blood* 1989, 74:1073–1083
 15. deVillartay J-P, Pullman AB, Andrade R, Tschachler E, Colamenici O, Neckers L, Cohen DI, Cossman J: γ/δ lineage relationship within a consecutive series of human precursor T-cell neoplasms. *Blood* 1989, 74:2508–2518
 16. Tkachuk DC, Griesser H, Takihara Y, Champagne E, Minden M, Feller AC, Lennert K, Mak TW: Rearrangement of T-cell δ locus in lymphoproliferative disorders. *Blood* 1988, 72:353–357
 17. Kimura N, Takihara Y, Akiyoshi T, Yoshida T, Ohshima K, Kawara T, Hisano S, Kozuru M, Okumura M, Kikuchi M: Rearrangement of T-cell receptor δ chain gene as a marker of lineage and clonality in T-cell lymphoproliferative disorders. *Cancer Res* 1989, 49:4488–4492
 18. National Cancer Institute sponsored study of classifications of non-Hodgkin's lymphomas: Summary and description of a working formulation for clinical usage. *Cancer* 1982, 49:2112–2135
 19. Medeiros LJ, Peiper SC, Elwood L, Yano T, Raffeld M, Jaffe ES: Angiocentric immunoproliferative lesions: A molecular analysis of 8 cases. *Hum Pathol* (In press)
 20. Patsouris E, Noël H, Lennert K: Histological and immunohistological findings in lymphoepithelioid cell lymphoma (Lennert's lymphoma). *Am J Surg Pathol* 1988, 12:341–350
 21. Jaffe ES, Blattner WA, Blayney DW, Cossman J, Robert-Guroff M, Gallo RC, Bunn PA: The pathologic spectrum of adult T-cell leukemia/lymphoma in the United States. Human T-cell leukemia/lymphoma virus-associated lymphoid malignancies. *Am J Surg Pathol* 1984, 8:263–275
 22. Stein H, Mason DY, Gerdes J, O'Connor N, Wainscoat J, Pallesen G, Gatter K, Falini B, Delsol G, Lemke H, Schwartz R, Lennert K: The expression of the Hodgkin's disease associated antigen Ki-1 in reactive and neoplastic lymphoid tissue: Evidence that Reed-Sternberg cells and histiocytic malignancies are derived from activated lymphoid cells. *Blood* 1985, 66:848–858
 23. Medeiros LJ, Herrington RD, Gonzalez CL, Jaffe ES, Cossman J: My4 antibody staining of non-Hodgkin's lymphomas. *Am J Clin Pathol* 1991, 95:363–368
 24. Reed KC, Mann DA: Rapid transfer of DNA from agarose gels to nylon membranes. *Nucleic Acids Res* 1985, 13:7207–7221
 25. Feinberg AP, Vogelstein B: A technique for radiolabeling DNA restriction endonuclease fragments to high specific activity. *Anal Biochem* 1983, 132:6–13
 26. Lipford EH, Smith HR, Pittaluga S, Jaffe ES, Steinberg AD, Cossman J: Clonality of angioimmunoblastic lymphadenopathy and implications for its evolution to malignant lymphoma. *J Clin Invest* 1987, 79:637–642
 27. Uppenkamp M, Andrade R, Sundeen J, Raffeld M, Coup-land R, Cossman J: Diagnostic interpretation of T γ gene rearrangement: Effect of polyclonal T cells. *Hematol Pathol* 1988, 2:15–24
 28. Takihara Y, Reimann J, Michalopoulos E, Ciccone E, Mor-etta L, Mak TW: Diversity and structure of human T cell receptor δ chain genes in peripheral blood γ/δ -bearing T lymphocytes. *J Exp Med* 1989, 169:393–405
 29. Lafaille JJ, DeCloux A, Bonneville M, Takagaki Y, Tonegawa S: Junctional sequences of T cell receptor γ/δ genes: Implications for γ/δ T cell lineages and for a novel intermediate of V-(D)-J joining. *Cell* 1989, 59:859–870
 30. deVillartay J-P, Hockett RD, Coran D, Korsmeyer SJ, Cohen DI: Deletion of the human T-cell receptor δ -gene by a site-specific recombination. *Nature* 1988, 335:170–174
 31. Herbst H, Tippelmann G, Anagnostopoulos I, Gerdes J, Schwartz R, Boehm T, Pileri S, Jones DB, Stein H: Immunoglobulin and T-cell receptor gene rearrangements in Hodgkin's disease and Ki-1-positive anaplastic large cell lymphoma: Dissociation between phenotype and genotype. *Leukemia Res* 1989, 13:103–116
 32. Pittaluga S, Raffeld M, Lipford EH, Cossman J: 3A1 (CD7) expression precedes T β gene rearrangements in precursor T (lymphoblastic) neoplasms. *Blood* 1986, 68:134–139