

Letters to the Editor

Activity of Garenoxacin against Macrolide-Susceptible and -Resistant *Mycoplasma pneumoniae*^V

In the treatment of *Mycoplasma pneumoniae* infection, erythromycin and clarithromycin, 14-membered macrolides, and azithromycin, a 15-membered macrolide, are usually considered to be the first-choice drugs (4, 11). Macrolide antibiotics inhibit protein synthesis by binding to domain II and/or domain V of 23S rRNA (1, 13). We found that ca. 20% of *M. pneumoniae* strains isolated from patients in Japan from 2000 to 2003 were macrolide resistant (6, 9). Lucier et al. (5) and Okazaki et al. (7) found that an A-to-G transition or an A-to-C transversion at position 2063 (corresponding to position 2058 in *Escherichia coli*) or 2064 of the 23S rRNA gene results in a high level of resistance to macrolide antibiotics. Given this background, it was considered that an alternative to macrolides in the treatment of pneumonia caused by *M. pneumoniae* is needed.

Garenoxacin, a des-F(6)-quinolone that exhibits excellent activity against respiratory pathogens such as *Streptococcus pneumoniae* and *Chlamydomphila pneumoniae*, is under development for both oral and parenteral administration (10). In the present study, we examined the in vitro activity of garenoxacin against *M. pneumoniae* isolates, including macrolide-resistant strains, and compared it with those of other antibiotics.

The following agents were employed for MIC determinations: garenoxacin (Toyama Chemical Co., Ltd., Tokyo, Japan), gatifloxacin (Kyorin Pharmaceutical Co., Ltd., Tokyo, Japan), levofloxacin and clarithromycin (LKT Laboratories, Inc., Saint Paul, MN), and minocycline (Lederle-Japan Ltd., Tokyo, Japan). The purity of each of these agents was above 99.8%. The following isolates of *M. pneumoniae* were tested: three macrolide-susceptible reference strains, FH, Mac, and M129, and four macrolide-resistant clinical isolates, 1 and 6 (isolates carrying the A-to-G mutation at position 2063 in the 23S rRNA gene for which the MIC of clarithromycin is 32 µg/ml) and 2 and 4 (isolates carrying the A-to-G mutation at position 2064 in the 23S rRNA gene for which the MIC of clarithromycin is 8 µg/ml). The PCR amplification and sequencing of domain II of the 23S rRNA genes were performed by a method reported previously (6).

A broth microdilution method was used to determine the

MICs. Serial twofold dilutions of antibacterial agents prepared in PPLO broth (Difco Inc., Detroit, MI) containing 10⁴ to 10⁵ CFU of *M. pneumoniae*/ml were put into 96-well microplates. The microplates were sealed with adhesive sheets and incubated at 37°C for 4 to 8 days. The MIC was defined as the lowest concentration of a drug at which the metabolism of the organism was inhibited, as evidenced by a lack of color change in the medium at the time when the drug-free control first showed a color change.

The MICs of garenoxacin for the *M. pneumoniae* isolates, including the macrolide-resistant isolates, were 0.016 to 0.031 µg/ml, two- to fourfold lower than those of gatifloxacin and 8- to 32-fold lower than those of levofloxacin and minocycline (Table 1). The results for macrolide-susceptible strains such as *M. pneumoniae* FH, a reference strain, accorded with those given in previous reports (2, 8, 12).

The newer fluoroquinolones such as gatifloxacin and moxifloxacin exhibit potent in vitro activity against a broad spectrum of organisms, including *M. pneumoniae* (3). Garenoxacin showed more-potent in vitro activity against *M. pneumoniae* than gatifloxacin and has attracted interest as a potential therapy for community-acquired pneumonia. These data indicate that garenoxacin may be a viable alternative to macrolides, such as clarithromycin, in the treatment of pneumonia caused by *M. pneumoniae*, including macrolide-resistant strains.

REFERENCES

- Douthwaite, S., L. H. Hansen, and P. Mauvais. 2000. Macrolide-ketolide inhibition of MLS-resistant ribosomes is improved by alternative drug interaction with domain II of 23S rRNA. *Mol. Microbiol.* **36**:183–193.
- Gruson, D., S. Pereyre, H. Renaudin, A. Charron, C. Bebear, and C. M. Bebear. 2005. In vitro development of resistance to six and four fluoroquinolones in *Mycoplasma pneumoniae* and *Mycoplasma hominis*, respectively. *Antimicrob. Agents Chemother.* **49**:1190–1193.
- Kenny, G. E., and F. D. Cartwright. 2001. Susceptibilities of *Mycoplasma hominis*, *M. pneumoniae*, and *Ureaplasma urealyticum* to GAR-936, dalfofpristin, dirithromycin, evernimicin, gatifloxacin, linezolid, moxifloxacin, quinupristin-dalfopristin, and telithromycin compared to their susceptibilities to reference macrolides, tetracyclines, and quinolones. *Antimicrob. Agents Chemother.* **45**:2604–2608.
- Langtry, H. D., and J. A. Balfour. 1998. Azithromycin. A review of its use in pediatric infectious diseases. *Drugs* **56**:273–297.
- Lucier, T. S., K. Heitzman, S. K. Liu, and P. C. Hu. 1995. Transition mutations in the 23S rRNA of erythromycin-resistant isolates of *Mycoplasma pneumoniae*. *Antimicrob. Agents Chemother.* **39**:2770–2773.
- Matsuoka, M., M. Narita, N. Okazaki, H. Ohya, T. Yamazaki, K. Ouchi, I. Suzuki, T. Andoh, T. Kenri, Y. Sasaki, A. Horino, M. Shintani, Y. Arakawa, and T. Sasaki. 2004. Characterization and molecular analysis of macrolide-resistant *Mycoplasma pneumoniae* clinical isolates obtained in Japan. *Antimicrob. Agents Chemother.* **48**:4624–4630.
- Okazaki, N., M. Narita, S. Yamada, K. Izumikawa, M. Umetsu, K. Kenri, Y. Sasaki, Y. Arakawa, and T. Sasaki. 2001. Characteristics of macrolide-resistant *Mycoplasma pneumoniae* strains isolated from patients and induced with erythromycin in vitro. *Microbiol. Immunol.* **45**:617–620.
- Pereyre, S., H. Renaudin, C. Bebear, and C. M. Bebear. 2004. In vitro activities of the newer quinolones garenoxacin, gatifloxacin, and gemifloxacin against human mycoplasmas. *Antimicrob. Agents Chemother.* **48**:3165–3168.
- Suzuki, S., T. Yamazaki, M. Narita, N. Okazaki, I. Suzuki, T. Andoh, M. Matsuoka, T. Kenri, Y. Arakawa, and T. Sasaki. 2006. Clinical evaluation of macrolide-resistant *Mycoplasma pneumoniae*. *Antimicrob. Agents Chemother.* **50**:709–712.
- Takahata, M., J. Mitsuyama, Y. Yamashiro, M. Yonezawa, H. Araki, Y.

TABLE 1. Activities of various antibiotics against *M. pneumoniae* strains

Strain	Gene mutation ^a	MIC (µg/ml) of:				
		Garenoxacin	Gatifloxacin	Levofloxacin	Minocycline	Clarithromycin
FH	None	0.016	0.063	0.50	0.50	0.004
Mac	None	0.016	0.063	0.25	0.25	0.002
M129	None	0.016	0.063	0.25	0.25	0.002
1	A2063G	0.016	0.031	0.25	0.25	32
6	A2063G	0.031	0.063	0.50	0.25	32
2	A2064G	0.016	0.063	0.25	0.25	8
4	A2064G	0.016	0.031	0.25	0.25	8

^a Gene mutation in domain II of 23S rRNA genes.

- Todo, S. Minami, Y. Watanabe, and H. Narita.** 1999. In vitro and in vivo antimicrobial activities of T-3811ME, a novel des-F(6)-quinolone. *Antimicrob. Agents Chemother.* **43**:1077–1084.
11. **Taylor-Robinson, D., and C. J. Bebear.** 1997. Antibiotic susceptibilities of mycoplasmas and treatment of mycoplasmal infections. *J. Antimicrob. Chemother.* **40**:622–630.
12. **Waites, K. B., D. M. Crabb, X. Bing, and L. B. Duffy.** 2003. In vitro susceptibilities to and bactericidal activities of garenoxacin (BMS-284756) and other antimicrobial agents against human mycoplasmas and ureaplasmas. *Antimicrob. Agents Chemother.* **47**:161–165.
13. **Weisblum, B.** 1995. Erythromycin resistance by ribosome modification. *Antimicrob. Agents Chemother.* **39**:577–585.

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