## In Vitro Studies of Activities of Some Antifungal Agents against Candida albicans ATCC <sup>10231</sup> by the Turbidimetric Method

MARIA TERESA BLANCO,\* CIRO PEREZ-GIRALDO, JAVIER BLANCO, FRANCISCO J. MORAN, CIPRIANO HURTADO, AND ANTONIO C. GOMEZ-GARCIA

> Department of Microbiology, Faculty of Medicine, University of Extremadura, Avenida Elvas sn., 06071 Badajoz, Spain

> > Received 11 July 1991/Accepted 6 January 1992

Different criteria (the drug concentration which inhibited 50% of growth  $[IC_{1/2}]$ , the lowest drug concentration at which growth was just less than 30% of that in a positive control well  $[IC_{30}]$ , the visual inhibitory concentration  $[IC_v]$ , and the minimum fungicidal concentration  $[MFC]$ ) were applied to study the effects of some antifungal agents against Candida albicans. Amphotericin B, flucytosine, and bifonazole produced total growth inhibition. Clotrimazole, itraconazole, ketoconazole, and miconazole produced partial growth inhibition. The values of  $IC_{1/2}$  and  $IC_{30}$  were similar for all agents and avoided the problems of partial inhibition; the values of IC<sub>v</sub> and MFC were higher than those of IC<sub>1/2</sub> and IC<sub>30</sub>.

The principal difficulty in interpreting susceptibility test results for antifungal agents is the partial growth inhibition produced by imidazole derivatives (10, 13). There is no unanimous agreement as to the best methodology for interpreting susceptibility test results (5, 7, 8, 19, 21). By the microdilution method, the inhibitory concentration (IC) is the lowest drug concentration that is capable of inhibiting a determined percentage of growth (10, 12, 13, 16, 24-26), and an objective value for assessing the final point of inhibition is achieved. Turbidimetric methods can determine the minimum drug concentration which inhibits 50% of growth  $(IC_{1/2})$   $(1, 6, 10, 18)$  or the lowest drug concentration at which growth is just less than 30% of that in the positive control well  $(IC_{30})$   $(12, 13)$ .

In this study, the effects of increasing concentrations of seven antifungal agents on the growth of Candida albicans ATCC <sup>10231</sup> were studied. The ICs were also evaluated by comparing the criteria mentioned above  $(IC_{1/2}$  and  $IC_{30})$ , the study of the visual inhibitory concentration  $(IC_v)$ , and the minimum fungicidal concentration (MFC).

The antifungal agents used in this study were amphotericin B (Squibb), clotrimazole and bifonazole (Bayer), miconazole (Isdin), itraconazole (Janssen) solubilized in dimethyl sulfoxide, flucytosine (Roche) resuspended in water, and ketoconazole (Janssen) solubilized in HCI (0.2 N). The concentrations of antifungal agents ranged from  $0.01$  to  $40 \mu g/ml$  and were obtained by dilution in 0.01 M phosphate-buffered saline (PBS) at pH 7.

C. albicans ATCC <sup>10231</sup> was grown in yeast nitrogen base medium (Difco) supplemented with 0.5% glucose in 0.01 M PBS (pH 7) (bYNBG).

Growth kinetics, IC, and MFC studies were carried out in microtiter plates with a final volume of 200  $\mu$ l. Each row of the microtiter plates contained the different concentrations of the antifungal agents and the strain tested. The final inoculum in each well was approximately  $10<sup>5</sup>$  organisms/ml. Controls of sterility and growth without antifungal agents were carried out. Plates were incubated at 37°C in a humid atmosphere. At 0, 4, 12, 24, 48, and 72 h of incubation, the optical density (OD) at 492 nm was read in <sup>a</sup> Titertek

Multiskan MC photometer. The data obtained after 24, 48, and 72 h were used to establish the  $IC_{1/2}$  and the  $IC_{30}$ . To determine the MFC, 20  $\mu$ l was taken from the wells where no visible growth appeared, inoculated into medium without antifungal agent, and examined after 48 h of incubation. The criteria for the MFC corresponded to the lowest concentration that yielded no colonies. All of the experiments were performed five times.

The effects of increasing concentrations of the antifungal agents on the growth of C. albicans ATCC <sup>10231</sup> are shown in Fig. 1, 2, and 3. The ODs obtained for the growth controls demonstrated that the culture reached the stationary growth phase after 24 h. The presence of concentrations greater than 1.25  $\mu$ g of amphotericin B per ml (IC<sub>1/2</sub> and IC<sub>30</sub>) brought about sudden growth inhibition. All of the imidazole derivatives except bifonazole showed partial growth inhibition from 24 h of culture for six consecutive concentrations (lower than the  $IC_{1/2}$  and the  $IC_{30}$ ). At 48 and 72 h, inhibition was generally presented as more sudden growth decreases, and the values of both parameters  $(IC_{1/2}$  and  $IC_{30})$  remained constant. The C. albicans strain studied (ATCC 10231) showed great susceptibility to flucytosine, although it was subsequently shown, by use of subcultures, that in order for flucytosine to act as a fungicide, much greater concentrations are required.

The values of  $IC_{1/2}$  and  $IC_{30}$  coincided for all of the antifungal agents studied except bifonazole after 24, 48, and 72 h; bifonazole presented a variation of 1 dilution after 24 h (Table 1). This was to be expected, since theoretical application of the formulae provided endpoints, with variations of ODs never being greater than 0.003 units. The ICs obtained after 48 and 72 h of incubation were similar and were greater than those obtained after 24 h.

After <sup>24</sup> h, the MFCs of all antifungal agents except itraconazole were always greater than the  $IC_{1/2}$  and  $IC_{30}$ , principally for flucytosine.

The IC, showed higher values and greater variability; included in Table 1 are the values which were repeated at least three times. Visual reading gave less objective data, and these data were often in disagreement with the spectrophotometric readings. In a study performed by Lefler and Stevens (14),  $IC_{\nu}$ s showed much variation, whereas the  $IC<sub>1/2</sub>$ s were the same for all strains.

<sup>\*</sup> Corresponding author.





FIG. 1. Effects of increasing concentrations of amphotericin B and flucytosine on the growth of C. albicans ATCC 10231. The concentrations (in micrograms per milliliter) were  $0$  ( $\bullet$ ),  $0.03$  ( $\blacksquare$ ),  $0.07$  ( $\square$ ),  $0.15$  ( $\blacktriangle$ ),  $0.3$  ( $\square$ ),  $0.6$  ( $\blacklozenge$ ),  $1.25$  ( $\square$ ),  $5$  ( $\blacksquare$ ),  $10$  ( $\square$ ),  $20$  ( $\boxplus$ ), and  $40$  ( $\boxtimes$ ).

bYNBG medium was chosen for this study because, although unanimity does not exist, it has been used by various investigators in studies of all antifungal agents and because it has been specified that significant variations do not exist with respect to other media (9, 11, 17, 22-24). bYNBG medium was buffered to avoid an acidic pH, which might adversely affect ketoconazole and amphotericin B (4). PBS was used in this study since it is inexpensive and gives ICs similar to those obtained with morpholinepropanesulfonic acid buffer (15). Some investigators have indicated that bYNBG medium produces poorer growth than those produced by other media (9) and that when agar dilution methods are applied, inhibition zones are not well defined (25). In microdilution tests, by using an IC that produces a



FIG. 2. Effects of increasing concentrations of clotrimazole, ketoconazole, and miconazole on the growth of C. albicans ATCC 10231. The concentrations (in micrograms per milliliter) were  $0$  ( $\bullet$ ), 0.01 (O), 0.03 ( $\blacksquare$ ), 0.07 ( $\square$ ), 0.15 ( $\blacktriangle$ ), 0.3 ( $\triangle$ ), 0.6 ( $\blacklozenge$ ), 1.25 ( $\diamond$ ), 5  $(\Box)$ , 10  $(\Box)$ , 20  $(\boxplus)$ , and 40  $(\boxtimes)$ .



FIG. 3. Effects of increasing concentrations of bifonazole and itraconazole on the growth of C. albicans ATCC 10231. The concentrations (in micrograms per milliliter) were  $0$  ( $\bullet$ ),  $0.01$  ( $\odot$ ),  $(0.03 \, (\blacksquare), \, 0.07 \, (\square), \, 0.15 \, (\triangle), \, 0.3 \, (\triangle), \, 0.6 \, (\blacklozenge), \, 1.25 \, (\diamond), \, 5 \, (\blacksquare), \, 10 \, (\square),$ 20 ( $\boxplus$ ), and 40 ( $\boxtimes$ ).

determined percentage of inhibition of growth with respect to the control, the problem of inhibition zones that are not well defined no longer exists.

The length of incubation has a pronounced effect on antifungal agents; MICs tend to increase with prolonged incubation times (3, 20). Pfaller et al. (23) have pointed out that after 24 h, optimum growth is produced and that the existing inhibition reflects the degree of susceptibility of the yeasts to the antifungal agent. On the other hand, Doern et al. (2) have stated that the reproducibility of the results is greater after 48 h, although they also found that MICs determined after incubation for 48 h were higher.

The partial inhibition produced by the majority of the imidazole derivatives (10, 12, 13) confirms the importance of

TABLE 1. In vitro activities of different antifungal agents against C. albicans ATCC <sup>10231</sup> by incubation time and reading criteria

Antifungal agent	Incubation time (h)	IC $(\mu g/ml)$			
		$IC_{1/2}$	$IC_{30}$	$IC_{\rm V}$	<b>MFC</b>
Amphotericin B	24	1.25	1.25	5	5
	48	5	5	10	10
	72	5	5	10	10
Flucytosine	24	0.07	0.07	0.15	40
	48	0.3	0.3	0.6	40
	72	0.3	0.3	5	40
<b>Bifonazole</b>	24	10	5	10	20
	48	10	10	10	20
	72	10	10	10	20
Clotrimazole	24	0.6	0.6	5	10
	48		$\frac{5}{5}$	5	20
	72	$\frac{5}{5}$		5	20
Itraconazole	24	5	5	5	5
	48	5	$\frac{5}{5}$	5	20
	72	5		10	40
Ketoconazole	24	0.6	0.6	5	20
	48	5	5	$\frac{5}{5}$	20
	72	5	5		20
Miconazole	24	0.07	0.07	5	20
	48	5	5	5	20
	72	5	5	5	20

using a growth control and the necessity of carrying out the reading of the test of susceptibility to antifungal agents on the basis of a percentage of growth inhibition. From the results of this study, we found that to evaluate the activity of an antifungal agent, it is advisable to use turbidimetric methods. Thus, it is possible to avoid the problems brought about by the partial inhibition produced by some antifungal agents. Turbidimetric methods also permit the application of objective criteria for the determination of ICs. By this methodology, the  $IC_{30}$  and the  $IC_{1/2}$  criteria are equally valid.

This work was supported in part by <sup>a</sup> grant (89/0860) from FISS (Spain).

We thank G. Gomez-Landero for excellent technical assistance and J. McCue for help in the preparation of the manuscript.

## **REFERENCES**

- 1. Calhoun, D. L., and J. N. Galgiani. 1984. Analysis of pH and buffer effects on flucytosine activity in broth dilution susceptibility testing of Candida albicans in two synthetic media. Antimicrob. Agents Chemother. 26:364-367.
- 2. Doern, G. V., T. A. Tubert, K. Chapin, and M. G. Rinaldi. 1986. Effect of medium composition on results of macrobroth dilution antifungal susceptibility testing of yeasts. J. Clin. Microbiol. 24:507-511.
- 3. Drutz, D. J. 1987. In vitro antifungal susceptibility testing and measurement of levels of antifungal agents in body fluids. Rev. Infect. Dis. 9:392-397.
- 4. Fromtling, R. A. 1988. Overview of medically important antifungal azole derivatives. Clin. Microbiol. Rev. 1:187-217.
- 5. Galgiani, J. N. 1987. Antifungal susceptibility tests. Antimicrob. Agents Chemother. 31:1867-1870.
- 6. Galgiani, J. N., and D. A. Stevens. 1976. Antimicrobial susceptibility testing of yeasts: a turbidimetric technique independent of inoculum size. Antimicrob. Agents Chemother. 10:721-726.
- 7. Gordon, M. A., E. W. Lapa, and P. G. Passero. 1988. Improved method for azole antifungal susceptibility testing. J. Clin. Microbiol. 26:1874-1877.
- 8. Guinet, R., D. Nerson, F. D. Closets, J. Dupouy-Camet, L. Kures, M. Marjollet, J. L. Poirot, A. Ros, J. Texier-Mauguein, and P. J. Volle. 1988. Collaborative evaluation in seven laboratories of a standardized micromethod for yeast susceptibility testing. J. Clin. Microbiol. 26:2307-2312.
- 9. Hoeprich, P. D., and J. M. Merry. 1986. Influence of culture medium on susceptibility testing with BAY <sup>n</sup> <sup>7133</sup> and ketoconazole. J. Clin. Microbiol. 24:269-271.
- 10. Hughes, C. E., R. L. Bennett, and W. H. Beggs. 1987. Broth dilution testing of *Candida albicans* susceptibility to ketoconazole. Antimicrob. Agents Chemother. 31:643-646.
- 11. Hughes, C. E., R. L. Bennett, I. C. Tuna, and W. H. Beggs. 1988. Activities of fluconazole (UK 49,858) and ketoconazole against ketoconazole-susceptible and -resistant Candida albicans. Antimicrob. Agents Chemother. 32:209-212.
- 12. Johnson, E. M., M. D. Richardson, and D. W. Warnock. 1984. In-vitro resistance to imidazole antifungals in Candida albicans. J. Antimicrob. Chemother. 13:547-558.
- 13. Korting, H. C., M. Ollert, A. Georgii, and M. Froschl. 1988. In vitro susceptibilities and biotypes of Candida albicans isolates from the oral cavities of patients infected with human immunodeficiency virus. J. Clin. Microbiol. 26:2626-2631.
- 14. Lefler, E., and D. A. Stevens. 1984. Inhibition and killing of Candida albicans in vitro by five imidazoles in clinical use. Antimicrob. Agents Chemother. 25:450-454.
- 15. MacKerrow, S. D., J. M. Merry, and P. D. Hoeprich. 1987. Effect of buffers on testing of Candida species susceptibility to flucytosine. J. Clin. Microbiol. 25:885-888.
- 16. Mazens, M. F., G. P. Andrews, and R. C. Bartlett. 1979. Comparison of microdilution and broth dilution techniques for the susceptibility of yeasts to 5-fluorocytosine and amphotericin B. Antimicrob. Agents Chemother. 15:475-477.
- 17. McGinnis, M. R., and M. G. Rinaldi. 1986. Antifungal drugs: mechanisms of action, drug resistance, susceptibility testing, and assays of activity in biological fluids, p. 223-281. In V.

Lorian (ed.), Antibiotics in laboratory medicine. The Williams & Wilkins Co., Baltimore.

- 18. McIntyre, K. A., and J. N. Galgiani. 1989. In vitro susceptibilities of yeasts to <sup>a</sup> new antifungal triazole, SCH 39304: effects of test conditions and relation to in vivo efficacy. Antimicrob. Agents Chemother. 33:1095-1100.
- 19. National Committee for Clinical Laboratory Standards. 1985. Antifungal susceptibility testing: committee report, vol. 5, no. 17. National Committee for Clinical Laboratory Standards, Villanova, Pa.
- 20. Odds, F. C. 1980. Laboratory evaluation of antifungal agents: a comparative study of five imidazole derivatives of clinical importance. J. Antimicrob. Chemother. 6:749-761.
- 21. Odds, F. C., and A. B. Abbott. 1984. Relative inhibition factors-a novel approach to the assessment of antifungal antibiotics in vitro. J. Antimicrob. Chemother. 13:31-43.
- 22. Pfaller, M. A., T. Gerardem, M. Yu, and R. P. Wenzel. 1989. Influence of in vitro susceptibility testing conditions on the anti-candidal activity of LY121019. Diagn. Microbiol. Infect. Dis. 11:1-9.
- 23. Pfaller, M. A., M. G. Rinaldi, J. N. Galgiani, M. S. Bartlett, B. A. Body, A. Espinel-Ingroff, R. A. Fromtling, G. S. Hall, C. E. Hughes, F. C. Odds, and A. Sugar. 1990. Collaborative investigation of variables in susceptibility testing of yeasts. Antimicrob. Agents Chemother. 34:1648-1654.
- 24. Pfaller, M. A., S. Wey, T. Gerarden, A. Houston, and R. P. Wenzel. 1989. Susceptibility of nosocomial isolates of Candida species to LY121019 and other antifungal agents. Diagn. Microbiol. Infect. Dis. 12:1-4.
- 25. Radetsky, M., R. C. Wheeler, M. H. Roe, and J. K. Todd. 1986. Microtiter broth dilution method for yeast susceptibility testing with validation by clinical outcome. J. Clin. Microbiol. 24:600- 606.
- 26. Spitzer, E. D., S. J. Travis, and G. S. Kobayashi. 1988. Comparative in vitro activity of LY121019 and amphotericin B against clinical isolates of Candida species. Eur. J. Clin. Microbiol. Infect. Dis. 7:80-82.