

PULMONARY CYTOLOGY

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REVIEW ARTICLE

Pulmonary Cytology

Current Status of Cytologic Typing of Respiratory Tract Tumors

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PREVIOUS ATTEMPTS to classify lung cancer have resulted in conflicting viewpoints on its etiology, diagnosis, treatment, and prognosis.¹ This situation is not surprising, because in a random survey of 57 studies pertaining to human lung cancer, 13 different systems of histologic classification were used.² These classifications characterized the tumors largely according to growth pattern and cell size and shape. Basing classification on such features easily leads to confusion, because the majority of tumors reveal two or more structural patterns on histologic examination of multiple paraffin blocks.³ Therefore, with such systems, tumors that show similar features at the cellular level are placed in different categories, and comparison of data becomes exceedingly complex.

Interobserver variability has been demonstrated in the classification of tumor type from tissue sections, especially for poorly differentiated and undifferentiated carcinomas. Between two separate panels of pathologists, disagreements in typing were observed in 40% or 53% of the poorly differentiated epidermoid carcinomas, 42% or 58% of the poorly differentiated adenocarcinomas, and 23% or 90% of the large cell carcinomas.^{4.5} In addition, individual pathologists from one panel demonstrated intraobserver variability by making different diagnoses between the first and second readings of the same slide in a range of 2% for the most consistent reader to 20% for the least consistent.⁵

World Health Organization Classification

In order to provide for a more accurate and more defined morphologic classification, 19 pathologists

from 17 countries collaborated to produce the WHO manual "Histological Typing of Lung Tumors."¹ Since 1967, over 120 epidemiologic, clinical, and pathologic studies have referred to, or based data on, the WHO lung tumor classification of 1967.⁶ The classification resulted in 13 types, most of which have several subtypes. The purpose of the WHO classification was to define a sufficient number of tumor categories, according to objective criteria, so that only a minimum of tumors remained unclassified.

The majority of lung tumors can be classified within the first six groups of the WHO classification (Table 1). Typing is based largely upon descriptive light-microscopic, cytologic, and histologic criteria, with little consideration given to patterns of differentiation at the cellular level. For example, Group II, small cell anaplastic carcinomas, are subtyped into fusiform, polygonal, lymphocytelike ("oat cell") and others, according to the shape of the cells. However, the various subtypes of tumors within Group II show different features of differentiation at the cellular level.⁷ Some

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Table 1—WHO Classification (1967)

I. Epidermoid	carcinomas
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- II. Small cell anaplastic carcinomas
 - 1. Fusiform cell carcinomas
 - 2. Polygonal cell carcinomas
 - Lymphocytelike ("oat cell") carcinomas
 Others
- III. Adenocarcinomas
- 1. Bronchogenic adenocarcinomas
 - 2. Bronchiolo-alveolar carcinomas
- IV. Large cell carcinomas
 - 1. Solid tumors with mucinlike substances
 - 2. Solid tumors without mucinlike substances
 - 3. Giant cell carcinomas
 - 4. "Clear" cell carcinomas
- V. Combined epidermoid and adenocarcinomas
- VI. Carcinoid tumors

show features characteristic of mucous and/or basal cells, whereas cells of both oat cell carcinomas and carcinoid tumors (Group VI) contain presumptive endocrine granules and may share a common histogenesis.⁸⁻¹⁷ Carcinoid tumors are considered low-malignancy forms, whereas oat cell carcinomas are highly malignant.

Group IV, large cell carcinomas, are divided into the following subtypes: solid tumors with mucinlike substances, solid tumors without mucinlike substances, giant cell carcinomas, and clear cell carcinomas. These represent a collection of dissimilar tumors. Cells constituting solid tumors with mucinlike substances may be highly differentiated for secretion (adenocarcinoma), whereas many clear cell tumors are composed of cells filled with glycogen, showing very poor specialization features. Recently, the WHO classification of lung tumors was updated, and proposed changes have been published.18 The most significant change appears to be that solid carcinomas with mucinlike substances, diagnosed as large cell carcinomas in the 1967 schema, will now be classified as a subtype of adenocarcinomas.

The Importance of Pulmonary Cytology

Cytology has become a reliable tool for the detection of pulmonary carcinoma cells in spontaneous cough and induced sputa, bronchial washings, bronchial brushings, fine needle aspiration biopsies, and pleural and pericardial effusions.

Many studies have provided statistics on the percentage of positive cytologic detection (percent yield) of malignant cells in bronchial secretions, some of which report the percentage of detection by specific cell type.¹⁹⁻²¹ Irrespective of tumor type, the overall percentage of positive preoperative cytologic detection of malignant lung tumors is about 80%,²¹⁻²⁴ with some studies demonstrating greater positive diagnoses of 92%,²⁵ 94%,²¹ and 98%.²⁶

Sputum cytology has detected radiographically undemonstrable primary lung neoplasms;²⁷⁻³⁶ and, in one study, pulmonary cytology yielded the only microscopic diagnoses prior to major surgery in 32% of cases.³⁷

Matching Correlation Between Cytologic and Histologic Diagnoses in Man

Since the first comparison between exfoliated cells and tissue sections was performed for clinical diagnoses of uterine cancer,³⁸ many such correlations have evaluated matching cytologic and histologic diagnoses for pulmonary neoplasia.^{3,21,24,26,39-52} In these studies the histologic diagnosis was accepted as the "true" tumor type.

In general, pulmonary cytology has been very successful in the diagnosis of moderately to well differentiated epidermoid carcinomas and moderately to well differentiated adenocarcinomas. For example, Lange and H ϕ eg⁴⁴ diagnosed 93% (43/46) of well differentiated epidermoid carcinomas and 100% (19/19) of adenocarcinomas by exfoliative cytology in agreement with histologic evaluations. Similarly, Kanhouwa and Matthews⁴⁷ matched 100% (50/50) of moderately to well differentiated epidermoid carcinomas and 100% (14/14) of moderately to well differentiated adenocarcinomas by cytologic and histologic evaluation (Table 2).

However, much confusion and controversy persists in the cytologic classification of poorly differentiated epidermoid carcinomas, poorly differentiated adenocarcinomas, and the so-called undifferentiated carcinomas, such as large cell carcinomas (including giant cell carcinomas and clear cell carcinomas) and oat cell carcinomas. One reason is the lack of rigid cytologic criteria used for the various tumor types. For instance, Johnston and Frable²¹ considered oat cell carcinoma to be a separate entity, whereas Koss,²³ Naib,⁵³ and Frost^{54,55} considered oat cell carcinomas to be poorly differentiated epidermoid carcinomas. Koss²³ classified large cell carcinomas as poorly differentiated epidermoid carcinomas, whereas Frost^{55,56} stated that the cells of adenocarcinomas were basically a large cell undifferentiated type. Although Naib53 categorized giant cell carcinomas as types of epidermoid carcinomas, Koss²³ and Johnston and Frable²¹ classified the basic cellular patterns of most giant cell carcinomas as variants of adenocarcinoma. Furthermore, Takahashi⁵⁷ stated that clear cell carcinoma, a subtype of large cell carcinoma according to the WHO classifica-

Histologic diagnosis	No. cases	Cytologic diagnosis	No. correct*	No. discrepancies	% correct*
Well and moderately differentiated epidermoid carcinoma	50	Epidermoid	50		100
Well and moderately differentiated adenocarcinoma	14	Adenocarcinoma	14		100
Small cell carcinoma	19	Small cell carcinoma Epidermoid	17	2	90
Large cell anaplastic carcinoma	6	Large cell anaplastic Adenocarcinoma	5	1	83.3
Poorly differentiated adenocarcinoma	16	Large cell anaplastic Epidermoid		13 3	0
Poorly differentiated epidermoid carcinoma	6	Adenocarcinoma Large cell anaplastic		1 _. 5	0

Table 2—Comparison of Cytologic and Histologic Typing of Human L	Lung Tumors (Courtesy of Kanhouwa and
Matthews, ⁴⁷ Copyright 1976, International Academy of Cytology)	

* Correct = matching correlations between cytologic and histologic diagnoses.

tion,' was indistinguishable from poorly differentiated epidermoid carcinoma or poorly differentiated adenocarcinoma.

Therefore, it is hardly surprising that previous studies, using light-microscopic criteria based upon growth patterns and cell size and shape, have provided relatively poor matching accuracy of pulmonary cytology versus histology for various forms of poorly differentiated carcinomas. For example, 16 tumors, diagnosed histologically as poorly differentiated adenocarcinomas, shed single cells which were typed as poorly differentiated epidermoid carcinomas using sputum cytology.²⁶ Similarly, 29% (20/68) of adenocarcinomas, diagnosed as such by histology, were interpreted cytologically as epidermoid carcinomas by examination of sputa that sometimes revealed single cells.²⁴

No matching diagnoses were made by Kanhouwa and Matthews⁴⁷ on six cases of poorly differentiated epidermoid carcinomas and 16 cases of poorly differentiated adenocarcinomas, as diagnosed histologically (Table 2). Instead, 18 of these tumors were classified cytologically as large cell anaplastic carcinomas. Evidently, the criteria for cytologic diagnosis of epidermoid carcinomas and adenocarcinomas,^{96,58,59} used by Kanhouwa and Matthews,⁴⁷ were very narrow; so the large cell carcinoma group increased in size.

The comparison of cytologic and histologic typing of human lung tumors between two institutions provided similar results.²¹ The criteria for cytologic diagnosis of large cell carcinomas used by Johnston at Duke University (Table 3) were evidently broad and demonstrated lower numbers of matching correlations for epidermoid carcinomas and adenocarcinomas than the more narrow criteria employed by Frable at the Medical College of Virginia (Table 4). Future epidemiologic or clinical studies using histologic and/or cytologic typing between these two institutions would probably result in the classification of similar tumor types being placed in completely different categories.

Table 3—Comparison of Cytologic and Histologic Typing of Human Lung Tumors: Duke University Results (Courtesy of Johnston and Frable,²¹ Copyright 1976, American Association of Pathologists)

		Су	tologic diagnos	sis		
Histologic diagnosis	Squamous cell carcinoma	Large cell undiffer- entiated carcinoma	Small cell undiffer- entiated carcinoma	Adeno- carcinoma	Muco- epidermoid carcinoma	% corre- lation
Squamous cell carcinoma	86	23	0	4	2	75
Large cell undifferentiated carcinoma	5	24	0	4	2	69
Small cell undifferentiated carcinoma	0	1	28	1	0	93
Adenocarcinoma	0	7	0	35	0	83
Mucoepidermoid carcinoma	0	1	0	0	3	75

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Table 4—Comparison of Cytologic and Histologic Typing of Human Lung Tumors:	Medical College of Virginia Results
(Courtesy of Johnston and Frable, ²¹ Copyright 1976, American Association of Patho	ologists)

		Су	tologic diagnos	sis		
Histologic diagnosis	Squamous cell carcinoma	Large cell undiffer- entiated carcinoma	Small cell undiffer- entiated carcinoma	Adeno- carcinoma	Muco- epidermoid carcinoma	% corre- lation
Squamous cell carcinoma	171	2	3	7	1	93
Large cell undifferentiated carcinoma	5	10	6	3	0	42
Small cell undifferentiated carcinoma	1	1	48	4	0	86
Adenocarcinoma	4	0	0	32	1	86
Mucoepidermoid carcinoma	5	1	0	5	0	0

Matching Correlation Between Cytologic and Histologic Diagnoses in Hamsters

In recent years the Syrian golden hamster has been demonstrated to be a good experimental model for human lung cancer. This animal has a low incidence of spontaneous respiratory tract tumors and offers great resistance to pulmonary infection.⁶⁰ The baseline data of normal hamster tracheobronchial and human bronchial epithelium show many similarities.61.62 With the use of both ultrastructural and histochemical techniques, respiratory tract carcinomas in the hamster, induced by benzo(a)pyrene-ferric oxide (BP-Fe₂O₃), resembled those seen in humans.^{7,63} Using the light-microscopic criteria of the WHO classification, tracheobronchial carcinomas resembling human bronchogenic epidermoid carcinomas, adenocarcinomas, large cell carcinomas, and combined epidermoid and adenocarcinomas were induced in the hamster by intratracheal instillation of BP-Fe₂O₃.^{60,64,65}

Matching accuracy of cytology versus histology for various hamster tumor types was quite similar to that of previously described human studies (Table 5).⁶⁵ Specifically, matching correlations occurred in 95% (18/19) of epidermoid carcinomas and 93% (13/14) of adenocarcinomas. However, cytologic and histologic diagnoses agreed in only 42% (5/12) of large cell anaplastic carcinomas.

Recognition of Cytoplasmic Differentiation Features in Cytologic Specimens

The majority of classifications of human and hamster respiratory tract tumors utilize light-microscopic descriptions based largely on cellular growth pattern, size, and shape, with little reference to differentiation at the cellular level. However, a recent histogenetic classification of respiratory tract carcinomas utilized specific features of cytoplasmic differentiation as observed in ultrastructural and histologic sections^{7.63} and in cytologic preparations⁶⁶ to differentiate and subtype the tumors. This approach demonstrated that various tumors, previously typed in different categories according to conventional criteria, actually share common characteristics at the cell level and thus may be typed in similar groups according to cytoplasmic features of functional differentiation.

In order to classify lung carcinomas according to features of their cytoplasmic differentiation, it is important to consider which epithelial cell types are potentially capable of hyperplastic, metaplastic, dysplastic, and neoplastic changes.⁷ It appears that only five cell types, that is, basal cells, mucous cells, endocrine cells, Clara cells, and possibly Type II alveolar cells can give rise to pulmonary carcinomas in man and hamster.^{7.63} Most respiratory tract carcinomas possess morphologic features of epidermoid differentiation (keratinization) and/or adeno differentiation (secretion) and were thus subdivided into epidermoid carcinomas, combined epidermoid and adenocarcinomas, and adenocarcinomas using a histogenetic classification.^{7.63}

Specific morphologic features of cytoplasmic differentiation can provide for the differential diagnosis of various tumor types, including poorly differentiated epidermoid carcinomas and poorly differentiated adenocarcinomas. On ultrastructural examination, tonofilament bundles represent epidermoid differentiation^{7,67-69} whereas secretory granules,^{7,67} extracellular and/or intracellular alveoli, and/or a welldeveloped Golgi apparatus and endoplasmic reticulum correspond to adeno features,⁷ which can be used as criteria to provide for accurate diagnoses. These features, which can be discerned by light microscopy in Papanicolaou-stained cytologic preparations,66 have often been misinterpreted or even ignored by cytologists in their diagnoses of various lung tumor types.

The transparency afforded by the Papanicolaou procedure, which uses two counterstains in solutions

Table 5—Comparison of Cytologic and Histologic	Typing of Hamster Lung	Tumors (Courtesy of Schreiber et al,65
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			Cytologic type		
Histologic type (WHO Classification in parentheses)	Number of animals	Epidermoid carcinoma	Adeno- carcinoma	Anaplastic carcinoma	Sensitivity† (% correct)
Epidermoid carcinoma (I)	19	18		1	18/19 (95)
Adenocarcinoma (III 1a, 1b)	14		13	1	13/14 (93)
Combined epidermoid-adenocarcinoma (V)	4	4*	4*		
Large cell anaplastic carcinoma (IV 2, 3, 4)	12	6	1	5	5/12 (42)

* Diagnosis of both tumor types was made in these animals.

[†] Sensitivity = matching correlations between cytologic and histologic diagnoses.

of high alcoholic content, offers greater capability of detecting differentiation features at the cellular level in cytologic preparations of lung tumors, as compared with routine histologic stains such as hematoxylin and eosin, applied to tumors in paraffin sections. Thus, the cytoplasmic features of epidermoid differentiation are recognized by a glassy appearance (hyalinization) and/or dense concentric rings or their fragments, corresponding to tonofilament bundles, with Papanicolaou stain.⁶⁶ Adeno characteristics include a foamy appearance in the cytoplasm, which corresponds to secretory granules and a well-developed Golgi apparatus, whereas cytoplasmic basophilia represents well-developed endoplasmic reticulum.66 Extracellular alveoli are sometimes observed between overlapping cells of tight to loose clusters. These are a characteristic of carcinomas that possess a well-differentiated adeno component and retain gross glandular organization.⁶⁶ The overlapping cells are joined at their luminal surfaces by well-developed junctional complexes providing for the presence of the extracellular alveoli. Also, with the use of the Papanicolaou stain, clear vacuoles can sometimes be seen within the cells. These correspond to intracellular alveoli that may be observed in overlapping cells of tight to loose clusters (when gross glandular organization is maintained) as well as in single cells, exfoliated from tumors in which gross glandular organization is lost.66 Since tumors that lack gross glandular organization do not possess well-developed junctional complexes, they appear solid in histologic sections but often possess other well-differentiated adeno features at the cellular level.66

Previously, these cytoplasmic features of epidermoid and adeno differentiation have not been understood, thereby leading to discrepancies in matching correlations between cytologic and histologic diagnoses. Approximately 50% of human and hamster respiratory tract tumors exhibit both epidermoid and adeno differentiation features, to varying degrees, at the cellular level.^{7.63.70} Therefore, large numbers of cells from these combined tumors exhibit mixtures of both epidermoid and adeno cytologic features in Papanicolaou preparations.

Poorly Differentiated Epidermoid and/or Adenocarcinomas

The concept of combined tumors is generally not appreciated and has led to confusion. For example, vacuolated cells shed from tumors classified histologically as poorly differentiated epidermoid carcinomas were typed cytologically with the use of conventional criteria as poorly differentiated adenocarcinomas.^{3,40,43} In contrast, eosinophilic cells exfoliated from tumors diagnosed histologically as adenocarcinomas were suggestive of degenerative changes and consequently were classified by conventional cytology as poorly differentiated epidermoid carcinomas.^{3,23,71} However, these tumors probably exhibited features of both epidermoid and adeno differentiation. Many adenocarcinomas, diagnosed as such by conventional cytologic criteria,55,56 actually possess tonofilament bundles of prekeratin, recognizable as rings in Papanicolaou-stained preparations.66 Similarly, many single cells, with or without hyalinization, shed from tumors diagnosed as epidermoid carcinomas by conventional cytologic criteria,55,59 possess intracellular alveoli, which are often thought to represent degenerative vacuoles by those using conventional cytology. However, these structures were found to be filled with mucosubstances⁶⁶ and therefore represent true secretory vacuoles, indicative of adeno differentiation in tumors in which gross glandular organization is either maintained or lost.66,70 The distinct borders of the intracellular alveoli were similar in various cytologic preparations, regardless of whether the specimen had been prepared fresh (eg, imprint smears, sputa, and bronchial brushings) or prepared by prefixation, ie, fixed in Saccomanno's fluid in the operating room (eg, bronchial washings).66 These tumors are classified histogenetically as combined epidermoid and adenocarcinomas, in which the adeno component predominates over the epidermoid component.^{66,70}

Large Cell Anaplastic Carcinomas and Giant Cell Carcinomas

Large cell anaplastic carcinomas^{54,55} and giant cell carcinomas,^{21,53} diagnosed as such by conventional cytologic criteria, clearly demonstrated specific cytoplasmic features of dual differentiation according to the histogenetic cytologic criteria.^{66,70} Both epidermoid and adeno features can be seen in many of these tumors, often in the same cell. The large number of combined tumors seen cytologically is to be expected, because approximately 50% of respiratory tract tumors present varying degrees of both epidermoid and adeno differentiation features at the cellular level.^{7,63,70}

Obviously the epidermoid and/or adeno features discernible in cytologic preparations of large cell and giant cell tumors reflect the patterns of differentiation seen at the ultrastructural level. Recently, epidermoid and adeno features of differentiation at the cell level were revealed in large cell carcinomas, ie, solid tumors with or without mucinlike substances (WHO Groups IV 1,2), often in the same tumor cell.⁷ Similarly, giant cell carcinomas, diagnosed cytologically by conventional criteria,^{21,53} demonstrated both epidermoid and adeno differentiation features with the use of electron microscopy.⁷² Ultrastructurally, the epidermoid and adeno characteristics of these subtypes were the same as described above.

The concept of dual differentiation in large cell and giant cell carcinomas is generally poorly recognized. Previous cytologic studies^{3,21,49} using conventional criteria have described either epidermoid or adeno characteristics in large cell tumors (diagnosed as such on paraffin sections). Both epidermoid and adeno features were never described in the same large cell carcinoma, let alone in the same tumor cell.^{3,21,49} In one study,⁴⁹ 11 of 14 (79%) large cell carcinomas (diagnosed histologically on paraffin sections) were classified after cytologic examination as either poorly differentiated epidermoid carcinomas or poorly differentiated adenocarcinomas.

As in these cytologic studies, several electron-microscopic studies of large cell and giant cell carcinomas did not recognize the dual differentiation of epidermoid and adeno cytoplasmic features in any one tumor.^{69,73-80} These studies indicated that large cell and giant cell carcinomas are similar to either poorly differentiated epidermoid carcinomas or poorly differentiated adenocarcinomas. For example, four reports of large cell carcinomas demonstrated epidermoid features of bundles of tonofilaments (tonofibrils),^{73,75,79,80} whereas other investigators described large cell carcinomas presenting with adeno features of intra- and/or intercellular alveoli (lumens) and/or mucous granules.^{69,76,80} Similarly, several electron-microscopic studies of giant cell carcinomas have noted features of either epidermoid differentiation⁷⁸ or adeno differentiation,^{74,77} but dual differentiation was never described.

The specific epidermoid and/or adeno characteristics of poorly differentiated, large cell, and giant cell tumors, which are impossible to detect with the use of H&E-stained tissue sections, can be discerned by lightmicroscopic cytology with the Papanicolaou method.⁷⁰ Therefore, Papanicolaou-stained imprint smears of surgical biopsy specimens of lung tumors promise to provide more accurate typing than either H&E-stained frozen or paraffin sections.

Reliability of Revised Cytologic Criteria Based Upon Features of Cytoplasmic Differentiation

The histogenetic cytologic classification is based on objective cytoplasmic features of functional differentiation,⁶⁶ whereas conventional cytologic classifications rely largely on subjective criteria pertaining to cell size, shape, and growth pattern.^{21,23,53-56,59} Using the revised cytologic criteria, the most common type of respiratory tract tumor is the combined epidermoid and adenocarcinoma, which possesses characteristics of mucous and/or basal cells.^{66,70}

This large number of combined tumors is not surprising in light of studies on the histogenesis of epidermoid metaplasia and carcinoma in situ in human and hamster respiratory epithelium.^{81,82} The data have shown that epidermoid metaplasia and carcinoma in situ may have arisen from changes in mucus-secreting cells. The combined epidermoid and adenocarcinomas probably originated from malignantly transformed metaplastic mucous cells and/or from malignantly transformed basal cells that demonstrated mucous cell characteristics. These two cell types can demonstrate both epidermoid and adeno functional differentiation within a single cell. Utilizing ultrastructural and histochemical techniques, Klein-Szanto et al^{83,84} have recently confirmed the presence of both tonofilament bundles (epidermoid differentiation) and various secretory characteristics (adeno differentiation) in metaplastic and dysplastic epithelial cells, carcinoma in situ, and microinvasive carcinomas of the rat trachea.

Utilizing the revised cytologic criteria,⁶⁶ the differentiation features of combined epidermoid and adenocarcinomas are easily recognizable with the Papa-

				Cyte	ologic diagn	osis (imprin	t smears)					
Histogenetic histologic diagnosis (including TEM)	Moderately to well dif- ferentiated epidermoid carcinoma	Poorly dif- ferentiated epidermoid carcinoma	CEACIA	CEAC/B	CEAC/CI	CEAC/CII	CEAC/D	Adenocar- cinoma I	Adenocar- cinoma II	Carcinoid	Oat cell	% correlation
Moderately to well differentiated epidermoid carcinoma	13											(16/17) 94
Poorly differentiated epidermoid carcinoma	2	٣	-									
CEACIA			3									(3/3) 100
CEAC/B	5			9	-							NG (Z1/9)
CEAC/CI CEAC/CII					11 5	5 6		-				} (27/28) 96
CEAC/D					-		ю	۲	-			(3/6) 50
Adenocarcinoma I Adenocarcinoma II					0 0		~	- Q				<pre>{ (8/13) 62</pre>
Carcinoid										-		(1/1) 100
Oat cell											4	(4/4) 100
Atypical endocrine							2					(n (7)n
CEAC = combined epid	ermoid and ader	nocarcinoma. A	= epidermo	oid componer	it, well-differe	ntiated; adeno	o component	, well-differer	tiated. B = e	pidermoid col	mponent, we	ell-differentiated;

Table 6—Reliability of Histogenetic Cytologic Criteria Applied to 86 Resected Human Lung Tumors (Hess et al,⁷⁶ Copyright 1980, International Academy of Cytology)

5 adeno component, poorly differentiated. C = epidermoid component, poorly differentiated; adeno component, well-differentiated. D = epidermoid component, poorly ponent, poorly differentiated. I = gross glandular organization maintained. II = gross glandular organization lost. TEM = transmission electron microscopy.

					Š	ologic diagn	iosis (impr	int smeal	rs)					
	Moderately differen- tiated to well differ- entiated epider-	Poorly differ- entiated epider-	S anaplas	mall cell stic carcin	oma	Moderately differen- tiated to well differ- entiated	Poorly differ- entiated	Larç	ge cell undi carcino	fferentiat oma	R R	Combined epider-		
conventional histologic diagnosis (WHO)	mold car- cinoma	moid car- cinoma	Fusiform I cell	Polygonal cell	Oat- cell	adeno- car- cinoma	adeno- car- cinoma	Solid- mucin	Solid-no mucin	Giant cell	Clear cell	adeno- carcinoma	Carci- noid	% correlation
Moderately to well differentiated epidermoid carcinoma	53	5					5	-	-					(25/30) 83
Poorly differentiated epidermoid carcinoma									-					
Small cell anaplastic carcinoma Fusiform cell Polygonal cell Oat cell	-			-	ę									— (0/1) 0 (3/4) 75
Bronchogenic adeno- carcinoma Moderately to well						ო	Q	ъ С	5	7				131200 46
differentiated Poorly differentiated		-		-			4	-	4					
Bronchioloalveolar carcinoma														I
Large cell carcinoma Solid-mucin	2	5				-	-	2	-	2				(2/11) 18
Solid-no mucin Giant cell	-	- (ო	ო						0 (8/0)
		V												
Combined epidermoid adenocarcinoma														I
Carcinoid tumor					-									(0/1) 0
WHO = World Health	Organization.													

Table 7—Reliability of Conventional Cytologic Criteria Applied to 86 Resected Human Lung Tumors (Hess et al,⁷⁰ Copyright 1980, International Academy of Cytology)

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nicolaou stain⁸⁵ and can be observed in well-preserved cells of imprint smears, obtained from surgically resected tumors, as well as in exfoliated cells from preoperative specimens such as sputa and bronchial washings, that have undergone various degrees of autolysis.⁷⁰ However, combined tumors are diagnosed in different categories as either epidermoid carcinomas, adenocarcinomas, or large cell carcinomas, with the use of conventional cytologic criteria.⁷⁰

The revised cytologic criteria have been demonstrated to be more reliable and accurate in matching the histologic diagnosis, as compared with conventional cytologic criteria.⁷⁰ A greater number of matching correlations was obtained for histogenetic cytodiagnoses versus histogenetic histologic evaluations than were achieved for conventional cytodiagnoses versus WHO histologic evaluations (Tables 6 and 7).⁷⁰ Relatively high matching reliability for combined epidermoid and adenocarcinomas occurred when the revised cytologic criteria were used (Table 6, large box). Different degrees of both epidermoid and adeno differentiation of most of the combined tumors were recognized in the cytologic preparations.

Although the accuracy of the revised cytologic criteria for adenocarcinomas was low (62%) (Table 6), these cytodiagnoses were actually more accurate than the histogenetic histologic diagnoses.⁷⁰ Several of these tumors revealed epidermoid characteristics, with the use of the Papanicolaou stain.^{66.70} Consequently, they were classified cytologically as combined epidermoid and adenocarcinomas with a poorly differentiated epidermoid component. The epidermoid features were not detectable with the use of H&E-stained paraffin sections.

The apparent high number (83%) (Table 7) of matching correlations between conventional cytodiagnoses and WHO histologic diagnoses for epidermoid carcinomas is false, because many of these tumors were actually combined epidermoid and adenocarcinomas, diagnosed as such with the new criteria on the basis of specific cytoplasmic features of functional differentiation revealed by light and electron microscopic examination.⁷⁰

The revised cytologic criteria were derived from a three-year study consisting of consecutive cases of lung cancer patients at four hospitals and consecutive cases of respiratory tract tumors of hamsters from two 1-year studies.⁶⁶ Although the number of cases in some categories is small (Tables 6 and 7),⁷⁰ these studies provided an adequate number of cases to demonstrate convincingly that similar reliability and reproducibility of various tumor types were revealed for varied types of respiratory tract cytologic specimens, such as imprint smears of human and hamster resected

tumors, hamster tracheal washings, and preoperative human specimens of sputa, bronchial washings, and bronchial brushings.

The new cytologic data clarify poorly differentiated epidermoid carcinomas and poorly differentiated adenocarcinomas as well as eliminate vague categories such as large cell carcinomas.⁷⁰ It is hoped that the revised diagnostic criteria will be recognized, understood, and used by cytopathologists, so that comparison of results between various institutions will be more accurate and perhaps more meaningful.

References

- Kreyberg L: Histological typing of lung tumors, International Histological Classification of Tumors. No. 1. World Health Organization, Geneva, 1967, pp 1–28
- Sobin LH: Multiplicity of lung tumor classifications. Rec Results Cancer Res 1972, 39:29-35
- Reid JD, Carr AH: The validity and value of histological and cytological classifications of lung cancer. Cancer 1961, 14:673-698
- Yesner R, Gerstl B, Auerbach O: Application of the World Health Organization classification of lung carcinoma to biopsy material. Ann Thorac Surg 1965, 1:33– 49
- Feinstein AR, Gelfman NA, Yesner R: Observer variability in the histopathologic diagnosis of lung cancer. Am Rev Respir Dis 1970, 101:671-684
- Sobin LH: Standardization and the histopathology of tumors. Histopathology 1977, 1:87–92
- McDowell EM, McLaughlin JS, Merenyi DK, Kieffer RF, Harris CC, Trump BF: The respiratory epithelium: V. Histogenesis of lung carcinomas in the human. J Natl Cancer Inst 1978, 61:587-606
- Bensch KG, Gordon GB, Miller LR: Electron microscopic and biochemical studies on the bronchial carcinoid tumor. Cancer 1965, 18:592-602
- Toker C: Observations on the ultrastructure of a bronchial adenoma (carcinoid-type). Cancer 1966, 19:1943– 1948
- Gmelich JT, Bensch KG, Liebow AA: Cells of Kultschitzky type in bronchioles and their relation to the origin of peripheral carcinoid tumor. Lab Invest 1967, 17: 88-98
- 11. Hattori S, Matsuda M, Tateishi R, Terazawa T: Electron microscopic studies on human lung cancer cells. Gann 1967, 58:283-290
- Pariente R, Even P, Brouet G: Étude ultrastructurale des carcinoïdes bronchiques. Presse Med 1967, 75: 183-188
- 13. Bensch KG, Corrin B, Pariente R, Spencer H: Oat-cell carcinoma of the lung: Its origin and relationship to bronchial carcinoid. Cancer 1968, 22:1163-1172
- Hattori S, Matsuda M, Tateishi R, Tatsumi N, Terazawa T: Oat-cell carcinoma of the lung containing serotonin granules. Gann 1968, 59:123-129
- Hattori S, Matsuda M, Tateishi R, Nishihara H, Horai T: Oat cell carcinoma of the lung: Clinical and morphological studies in relation to its histogenesis. Cancer 1972, 30:1014-1024
- Hage E: Histochemistry and fine structure of bronchial carcinoid tumours. Virchows Arch [Pathol Anat] 1973, 361:121-128
- Fisher ER, Palekar A, Paulson JD: Comparative histopathologic, histochemical, electron microscopic and tis-

sue culture studies of bronchial carcinoids and oat cell carcinomas of lung. Am J Clin Pathol 1978, 69:165-172

- 18. Sobin LH: The WHO histological classification of lung tumors, Lung Cancer: Progress in Therapeutic Re-search. Edited by FM Muggia, M Rozencweig, New York, Raven Press, 1979, pp 83-90
- 19. Grunze H: A critical review and evaluation of cytodiagnosis in chest diseases. Acta Cytol 1960, 4:175-198 20. Russell WO, Neidhardt HW, Mountain CF, Griffith
- KM, Chang JP: Cytodiagnosis of lung cancer: A report of a four-year laboratory, clinical, and statistical study with a review of the literature on lung cancer and pulmonary cytology. Acta Cytol 1963, 7:1-44 21. Johnston WW, Frable WJ: Cytopathology of the respi-
- ratory tract: A review. Am J Pathol 1976, 84:372-424 22. Farber SM, Benioff MA, Frost JK, Rosenthal M, To-
- bias G: Cytologic studies of sputum and bronchial secretions in primary carcinoma of the lung. Dis Chest 1948, 14:633-664
- 23. Koss LG: Cancer of the lung, Diagnostic Cytology and Its Histopathologic Bases. 2nd edition. Philadelphia, J B Lippincott, 1968, pp 338-385
- 24. Oswald NC, Hinson KF, Canti G, Miller AB: Diagnosis of primary lung cancer with special reference to sputum cytology. Thorax 1971, 26:623-631 25. Laurie W, Szaloky LG: Sputum cytology in the diagno-
- sis of bronchial carcinoma. Med J Aust 1971, 1:247-251
- 26. Lukeman JM: Reliability of cytologic diagnosis in cancer of the lung. Cancer Chemother Rep 1973, 4:79–93
- 27. Woolner LB, Andersen HA, Bernatz PE: "Occult" carcinoma of the bronchus: A study of 15 cases of in situ or early invasive bronchogenic carcinoma. Dis Chest 1960, 37:278-288
- 28. Lerner MA, Rosbash H, Frank HA, Fleischner FG: Radiologic localization and management of cytologically discovered bronchial carcinoma. N Engl J Med 1961, 264:480-485
- 29. Melamed MR, Koss LG, Cliffton EE: Roentgenologically occult lung cancer diagnosed by cytology: Report of 12 cases. Cancer 1963, 16:1537-1551
- 30. Holman CW, Okinaka A: Occult carcinoma of the lung. J Thorac Cardiovasc Surg 1964, 47:466-471 31. Pearson FG, Thompson DW, Delarue NC: Experience
- with the cytologic detection, localization, and treatment of radiographically undemonstrable bronchial carci-noma. J Thorac Cardiovasc Surg 1967, 54:371-382
- 32. Meyer JA, Bechtold E, Jones DB: Positive sputum cytologic tests for five years before specific detection of bronchial carcinoma. J Thorac Cardiovasc Surg 1969, 57:318-324
- 33. Grzybowski S, Coy P: Early diagnosis of carcinoma of the lung: Simultaneous screening with chest x-ray and sputum cytology. Cancer 1970, 25:113-120
- 34. Woolner LB, David E, Fontana RS, Andersen HA, Bernatz PE: In situ and early invasive bronchogenic carcinoma: Report of 28 cases with postoperative survival data. J Thorac Cardiovasc Surg 1970, 60:275-290
- 35. Marsh BF, Frost JK, Erozan YS, Carter D: Occult bronchogenic carcinoma: Endoscopic localization and television documentation. Cancer 1972, 30:1348-1352
- 36. Nasiell M, Sinner W, Tornvall G, Roger V, Vogel B, Enstad I: Clinically occult lung cancer with positive sputum cytology and primarily negative radiological findings. Scand J Respir Dis 1977, 58:134-144
- 37. Erozan YS, Frost JK: Cytopathologic diagnosis of cancer in pulmonary material: A critical histopathologic correlation. Acta Cytol 1970, 14:560-565
- 38. Papanicolaou GN, Traut HF: Diagnosis of Uterine Cancer by the Vaginal Smear. New York, Commonwealth Fund, 1943
- 39. Foot NC: The identification of types of pulmonary cancer in cytologic smears. Am J Pathol 1952, 28:963-983

- 40. Spjut HJ, Fier DJ, Ackerman LV: Exfoliative cytology and pulmonary cancer: A histopathologic and cytologic correlation. J Thorac Surg 1955, 30:90-107
- 41. Dahlgren SE: Aspiration biopsy of intrathoracic tumours. Acta Pathol Microbiol Scand 1967, 70:566-576 42. Hastrup J: In Koss, Cancer of the lung,²³ pp 378-379
- 43. Kirsh MM, Orvald T, Naylor B, Kahn DR, Sloan H: Diagnostic accuracy of exfoliative pulmonary cytology. Ann Thorac Surg 1970, 9:335-338
- 44. Lange E, Høeg K: Cytologic typing of lung cancer. Acta
- Cytol 1972, 16:327–330 45. Rosa UW, Prolla JC, Gastal ES: Cytology in diagnosis of cancer affecting the lung: Results in 1,000 consecutive patients. Chest 1973, 63:203-207
- 46. Gagneten CB, Geller CE, Saenz MC: Diagnosis of bronchogenic carcinoma through the cytologic examination of sputum, with special reference to tumor typing. Acta Cytol 1976, 20:530-536
- 47. Kanhouwa SB, Matthews MJ: Reliability of cytologic typing of lung cancer. Acta Cytol 1976, 20:229-232
- 48. Roger V, Nasiell M, Linden M, Enstad I: Cytologic differential diagnosis of bronchiolo-alveolar carcinoma and bronchogenic adenocarcinoma. Acta Cytol 1976, 20:303-307
- 49. Erozan YS, Frost JK: Cytopathologic diagnosis of lung cancer, Lung Cancer: Clinical Diagnosis and Treatment. Edited by MJ Straus. New York, Grune and Stratton, 1977, pp 95-106 50. Augusseau S, Mouriquand J, Brambilla C, Paramelle B:
- Cytological survey of bronchial brushings and aspirations performed during fiberoptic bronchoscopy. Arch Geschwulstforsch 1978, 48:245-249
- 51. Siegismund G, Atay A, Loblich HJ: The diagnostic value of combined bronchus biopsy and contact smear cytology of the biopsy specimen in lung cancer. Cancer Čytol 1978, 18:9–12
- 52. Payne CR, Stovin PGI, Barker V, McVittie S, Stark JE: Diagnostic accuracy of cytology and biopsy in primary bronchial carcinoma. Thorax 1979, 34:294-299
- 53. Naib ZM: The respiratory tract, Exfoliative Cytopathology. 1st edition. Boston, Little, Brown and Co., 1970, pp 186-241
- 54. Frost JK: Cancers without identifiable functional differentiation, The Cell in Health and Disease: An Evaluation of Cellular Morphologic Expression of Biologic Behavior. Monograph in Clinical Cytology. Vol 2. Edited by GL Wied. Baltimore, Williams and Wilkins, 1969, pp 130-136
- 55. Frost JK: Functional differentiation characteristics in cancer, Concepts Basic to General Cytopathology. 4th edition. Baltimore, Johns Hopkins Press, 1972, pp 39-49
- 56. Frost JK: Atypical functional differentiation of adenocarcinoma,⁵⁴ pp 101-111 57. Takahashi M: Respiratory tract, Color Atlas of Cancer
- Cytology. Philadelphia, J. B. Lippincott, 1971, pp 166-209
- 58. Frost JK: Malignant cell morphology: Malignant neoplasia, Concepts Basic to General Cytopathology. 2nd edition. Baltimore, Johns Hopkins Press, 1961, pp 35-49
- 59. Frost JK: Atypical functional differentiation of squamous cell carcinoma,⁵⁴ pp 72–93 60. Saffiotti U, Cefis F, Kolb LH: Method for the experi-
- mental induction of bronchogenic carcinoma. Cancer Res 1968, 28:104–124 61. Becci PJ, McDowell EM, Trump BF: The respiratory
- epithelium: II. Hamster trachea, bronchus, and bron-chioles. J Natl Cancer Inst 1978, 61:551-561
- 62. McDowell EM, Barrett LA, Glavin F, Harris CC, Trump BF: The respiratory epithelium: I. Human bronchus. J Natl Cancer Inst 1978, 61:539-549

- 63. Becci PJ, McDowell EM, Trump BF: The respiratory epithelium: VI. Histogenesis of lung tumors induced by benzo(a)pyrene-ferric oxide in the hamster. J Natl Cancer Inst 1978, 61:607-618
- 64. Stenbäck F: Morphologic characteristics of experimentally induced lung tumors and their precursors in hamsters. Acta Cytol 1973, 17:476-486
- 65. Schreiber H, Saccomanno G, Martin DH, Brennan L: Sequential cytological changes during development of respiratory tract tumors induced in hamsters by benzo-(a)pyrene-ferric oxide. Cancer Res 1974, 34:689-698
- 66. Hess FG, McDowell EM, Trump BF: The respiratory epithelium: VIII. Interpretation of cytologic criteria for human and hamster respiratory tract tumors. Acta Cytol (In press)
- 67. Gyorkey F, Min K-W, Krisko I, Gyorkey P: The usefulness of electron microscopy in the diagnosis of human tumors. Hum Pathol 1975, 6:421-441
- 68. Bonikos DS, Bensch KG, Kempson RL: Contribution of electron microscopy to the differential diagnosis of tumors, Beitr Pathol Bd 1976, 158:417-444
- 69. Mackay B, Osbourne BM, Wilson RA: Ultrastructure of lung neoplasms,⁴⁹ pp 71-84
- Hess FG, McDowell EM, Resau JH, Trump BF: The respiratory epithelium: IX. Validity and reproducibility of revised cytologic criteria of human and hamster respiratory tract tumors. Acta Cytol (In press)
- 71. Smith JH, Frable WJ: Adenocarcinoma of the lung: Cytologic correlation with histologic types. Acta Cytol 1974, 18:316-320
- 72. McDowell EM: Unpublished observations, 1978
- 73. Stoebner P, Cussac Y, Porte A, LeGal Y: Ultrastructure of anaplastic bronchial carcinomas. Cancer 1967, 20: 286-294
- 74. Razzuk MA, Lynn JA, Kingsley WB, Race GJ, Urschel HC Jr, Paulson DL: Giant cell carcinoma of the lung. J Thorac Cardiovasc Surg 1970, 59:574–580
- 75. Razzuk MA, Race GJ, Lynn JA, Martin JA, Urschel

HC Jr, Paulson DL: Observation on ultrastructural morphology of bronchogenic carcinoma. J Thorac Cardiovasc Surg 1970, 59:581–587

- Mollo F, Canese MG, Campobasso O: Human peripheral lung tumours: Light and electron microscopic correlation. Br J Cancer 1973, 27:173–182
- Wilson BO, Battifora HA: Histological classification of malignant lung tumors. Ann Clin Lab Sci 1974, 4:4-12
- Wang NS, Seemayer TA, Ahmed MN, Knaack J: Giant cell carcinoma of the lung: A light and electron microscopic study. Hum Pathol 1976, 7:3-16
- Kandawalla NM, Kasnic G Jr, Azar HA: An ultrastructural study of lung tumors (Abstr). Lab Invest 1977, 36: 342
- 80. Churg A: The fine structure of large cell undifferentiated carcinoma of the lung: Evidence for its relation to squamous cell carcinomas and adenocarcinomas. Hum Pathol 1978, 9:143-156
- 81. Becci PJ, McDowell EM, Trump BF: The respiratory epithelium: IV. Histogenesis of epidermoid metaplasia and carcinoma in situ in the hamster. J Natl Cancer Inst 1978, 61:577-586
- 82. Trump BF, McDowell EM, Glavin F, Barrett LA, Becci PJ, Schürch W, Kaiser HE, Harris CC: The respiratory epithelium: III. Histogenesis of epidermoid metaplasia and carcinoma in situ in the human. J Natl Cancer Inst 1978, 61:563-575
- Klein-Szanto AJP, Topping DC, Heckman CA, Nettesheim P: Ultrastructural characteristics of carcinogeninduced nondysplastic changes in tracheal epithelium. Am J Pathol 1980, 98:61-82
- Klein-Szanto AJP, Topping DC, Heckman CA, Nettesheim P: Ultrastructural characteristics of carcinogeninduced dysplastic changes in tracheal epithelium. Am J Pathol 1980, 98:83-100
- 85. Gill GW, Plowden KAM: Staining, Compendium on Cytopreparatory Techniques. 4th edition. Edited by CM Keebler, JW Reagan, GL Wied. Chicago, Tutorials of Cytology, 1976, pp 16-32