Role of IL-15 in HIV-1-associated hypergammaglobulinaemia

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SUMMARY

IL-15 is a novel cytokine, produced by monocytes/macrophages, with biological activities similar to IL-2 but with no significant sequence homology. IL-15 also stimulates human B cells to proliferation and immunoglobulin secretion. We measured serum levels of IL-15 in 84 HIV-1-infected individuals at different stages of disease in reference to 41 healthy blood donors. Our results show a marked elevation of IL-15 serum levels during HIV-1 infection. Moreover, we found that this increase correlated with serum levels of IgG (r = 0.376, P < 0.0001), and partly with serum IgM (r = 0.265, P = 0.015). A significant increase of IL-15 production by cultured peripheral blood mononuclear cells (PBMC) and purified monocytes in the presence of HIV-1 virus suggests that monocytes/macrophages may be a source of higher IL-15 serum levels in HIV-1-infected individuals. These findings indicate a participation of IL-15 in the hypergammaglobulinaemia frequently associated with HIV-1 infection.

Keywords hypergammaglobulinaemia IL-15 serum IgG serum IgM HIV-1

INTRODUCTION

Infection with HIV-1 is known to cause an intense polyclonal activation of B cells, as manifested by hypergammaglobulinaemia, elevated serum levels of immune complexes and autoantibodies, increased numbers of spontaneous immunoglobulin-secreting cells, and an elevated frequency of B cell lymphomas [1–3]. The HIV-1 envelope glycoprotein gp120 has been identified to bind and induce immunoglobulin secretion via superantigen interaction by a subpopulation of B cells, expressing the V_H3 family of immunoglobulin on their surface [4]. Recently, peptide epitopes that block binding of gp120 to immunoglobulin have been determined [5].

The induction of B cell differentiation is a complex process that is regulated by T lymphocytes and cytokines. Several mechanisms for the excessive B cell activation in HIV-induced hypergammaglobulinaemia have been suggested, including T cell contactdependent interactions [6,7], IL-6 and tumour necrosis factor-alpha (TNF- α) stimulation by monocytes [3,8–10] and IL-10 secretion by B cells [11].

To shed more light on HIV-induced hypergammaglobulinaemia, we investigated the role of a novel cytokine, IL-15, which shares many of the stimulatory activities associated with IL-2 [12,13].

IL-15 is a 14–15-kD protein that stimulates proliferation of the CTLL cell line, phytohaemagglutinin (PHA)-activated blasts [12], activation of natural killer (NK) cells [14] and secretion of IgM, IgG and IgA by B cells [15] to the same extent and with similar potency as IL-2. Expression of IL-15 mRNA has been detected in

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several human tissues such as heart, lung, liver, kidney, placenta, skeletal muscle and epithelial cell lines, but the highest level of secretion has been seen in monocytes/macrophages [12,16]. Although it has no sequence homology with IL-2, IL-15 uses IL-2 receptor (IL-2R) β - and γ -chains for binding and signal transduction to the cell [14,17,18].

The primary purpose for our investigations was to find an explanation for hypergammaglobulinaemia in late stages of HIV infection, when CD4⁺ T cells are diminished and no longer able to provide a stimulatory signal for broad B cell activation. In our study we measured IL-15 serum levels in 84 HIV-1⁺ individuals at different stages of the disease (WR I to WR VI) in comparison with 41 HIV-1⁻ blood donors. Our results show a correlation between IL-15 and immunoglobulin levels in serum of HIV-1⁺ patients. Moreover, we observed an increase in IL-15 secretion from monocytes cultivated in the presence of HIV-1_{IIIB} virus isolate (HIV-1_{IIIB}).

MATERIALS AND METHODS

Samples

Serum samples were kindly provided by Dr R. Zangerle (Universitäts-Klinik für Dermatologie, Innsbruck, Austria). The study population comprised 84 HIV-1⁺ individuals and 41 healthy blood donors (HIV-1⁻). HIV-1⁺ subjects were classified according to the Walter Reed (WR) staging classification with 38 in WR I or II, 16 in WR III or IV and 30 in WR V or VI. Serum samples were stored at -70° C.

Detection of IL-15 in serum samples

Various dilutions (undiluted samples and samples diluted 1:1,

Table 1. Determination of IL-15 serum level in $HIV-1^+$ and $HIV-1^-$ individuals

HIV-1 infection stage	IL-15 serum level	
	Detectable/not detectable	%
WR I–II $(n = 38)$	15/23	39
WR III–IV ($n = 16$)	8/8	50
WR V–VI ($n = 30$)	15/15	50
Healthy blood donors $(n = 41)$	12/29	29

1:2, 1:4) of serum samples were assayed by an ELISA kit for the specific quantitative determination of human IL-15 (Genzyme, Cambridge, MA) according to the manufacturer's instructions. The kit is specific for native or recombinant human IL-15 (detection limit 10 pg/ml) with no detectable cross-reaction with other cytokines and serum proteins.

Detection of IgM and IgG

IgM and IgG levels in serum were measured by routine sandwich ELISA using polyclonal rabbit anti-IgM or anti-IgG as capture antibody and polyclonal goat anti-IgM or anti-IgG antibodies, conjugated to horseradish peroxidase (HRP; Dako A/S, Glostrup, Denmark) for antigen detection.

Mononuclear cells and monocytes

Peripheral blood mononuclear cells (PBMC) were obtained from heparinized peripheral blood of healthy donors by centrifugation on a Ficoll–Hypaque density gradient. Monocytes were separated from PBMC by adherence on gelatin-coated Petri dishes as described previously [19]. Briefly, 10 ml of PBMC suspension $(3-6 \times 10^6 \text{ cells/ml})$ in RPMI 1640 + 10% fetal calf serum (FCS) +3% autologous human serum were incubated on gelatin-coated (2% gelatin in water) Petri dishes for 40 min at 37°C. Non-adherent cells were aspirated and dishes washed three times with prewarmed RPMI 1640 + 10% FCS medium. Adherent cells (92–97% CD14⁺ monocytes) were incubated for 10 min in RPMI 1640 + 10% FCS medium with 5 mM ethylenediaminotetraacetate (EDTA). Detached monocytes were aspirated, washed and resuspended in RPMI 1640 + 10% FCS medium.

Cocultivation of PBMC and monocytes with HIV-1_{IIIB} isolate

HIV-1_{IIIB} virus isolate was used in all experiments. Virus stock was prepared by propagation in PBMC from healthy HIV-1⁻ donors. Virus was concentrated by ultracentrifugation and quantified by capture ELISA for HIV-1 p24 antigen. Heat-inactivated (56°C for 30 min) HIV-1_{IIIB} isolate (iHIV-1_{IIIB}) was used in some experiments. PBMC and monocytes (1×10^5 cells/well, $100 \,\mu$ l) were incubated in RPMI 1640 + 10% FCS medium with different HIV-1_{IIIB} concentrations for 7 days. One hundred microlitres of 1 : 1 diluted supernatants were used immediately for IL-15 determination as described above.

Statistical analysis

Results are expressed as mean \pm s.e.m. of the data obtained from two independent measurements of each serum sample performed in duplicate. Non-parametric Mann–Whitney test for unpaired data was carried out for statistical comparison between HIV-1⁺ and

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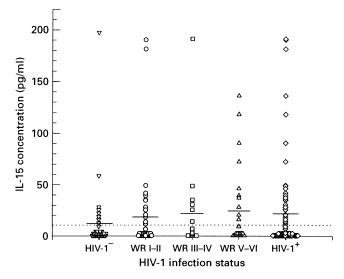


Fig. 1. Determination of IL-15 serum levels in 41 healthy donors and 84 $HIV-1^+$ individuals. $HIV-1^+$ individuals were divided in three groups according to the Walter Reed Classification (WR I–II, WR III–IV, WR V–VI). $HIV-1^-$, healthy donors; horizontal bars, mean values of each group of subjects; dotted line, detection limit.

HIV-1⁻ serum samples. Spearman's non-parametric test was used for correlations between IL-15 and IgG or IgM serum levels. Twotailed Student's *t*-test for unpaired data was applied for comparison of IL-15 levels in supernatants from PBMC and monocytes.

RESULTS

Serum levels of IL-15 in HIV-1-infected subjects

We were able to detect serum levels of IL-15 only in parts of HIV-1⁺ and HIV-1⁻ subjects (Table 1). Only 29% of HIV-1⁻ serum samples contained measurable levels of IL-15. With progression of disease the number of IL-15-positive individuals increased to 50% (WR III–VI).

The mean values of serum IL-15 showed a significant (Mann–Whitney test; P = 0.0034) increase in 84 HIV-1⁺ individuals compared with 41 HIV-1⁻ donors (20.83 ± 4.47 pg/ml *versus* 10.78 ± 5.01 pg/ml) (Fig. 1). Among HIV-1⁺ individuals, IL-15 serum levels were higher in subjects at later stages (WR III–IV 21.99 ± 11.91 pg/ml and WR V–VI 22.65 ± 6.65 pg/ml) than in HIV-1⁺ individuals at earlier stages (WR II–IV stages, the IL-15 serum level in ARC/AIDS patients (WR V–VI) remained stable. In all groups, no significant difference was found for serum IL-15 (P > 0.05).

Correlation between serum levels of IL-15 and gammaglobulins

The hypergammaglobulinaemia in HIV-1⁺ subjects was mainly due to an increase in IgG, although IgM were also elevated. Our results show that, among HIV-1⁺ carriers, the mean IgM and IgG level is higher in patients with detectable serum levels of IL-15 than in those with undetectable levels of IL-15 (for IgM $403\cdot2 \pm 69\cdot4$ mg/100 ml *versus* $247\cdot3 \pm 30\cdot6$ mg/100 ml; for IgG $2191\cdot4 \pm 79\cdot1$ mg/100 ml *versus* $1811\cdot1 \pm 34\cdot1$ mg/100 ml) (Fig. 2a,b). In addition, the difference in serum levels of IgG was statistically significant (for IgG P < 0.0001, for IgM P = 0.1351).

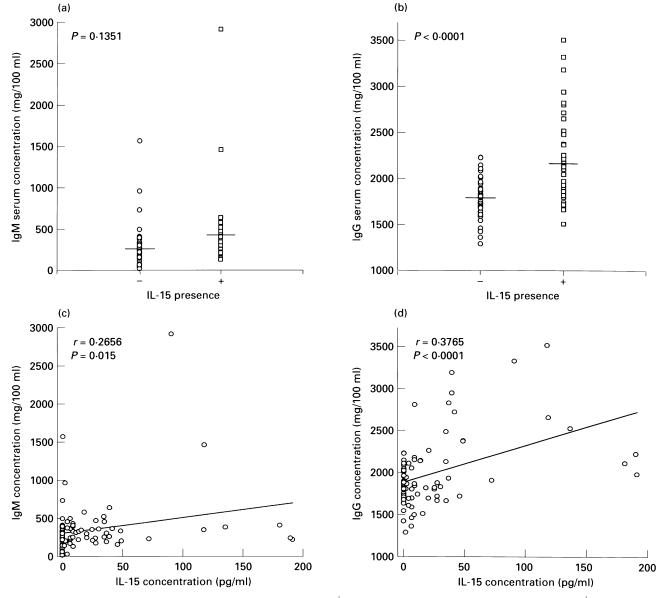


Fig. 2. Correlation between IL-15 and immunoglobulin serum levels in $HIV-1^+$ individuals. IgM (a) and IgG (b) serum levels in $HIV-1^+$ individuals with undetectable (\bigcirc) or detectable (\bigcirc) levels of serum IL-15. Horizontal bars, mean values of each group of subjects; *P* value, non-parametric Mann–Whitney test for unpaired data. Correlations of IgM (c) and IgG (d) levels in sera with serum IL-15. *r* and *P* value, non-parametric Spearman's test for correlation.

A moderate, but significant correlation was observed between IL-15 and IgM serum levels (r = 0.2656; P = 0.015) in HIV-1⁺ individuals (Fig. 2c). Moreover, we actually found a significant correlation between serum levels of IL-15 and IgG (r = 0.3765; P < 0.0001) (Fig. 2d).

IL-15 production by monocytes and PBMC

The production of IL-15 by cultured monocytes and PBMC was examined to prove relevance of increased IL-15 serum levels in HIV-1⁺ individuals (Fig. 3). Monocytes and PBMC obtained from healthy donors showed a spontaneous production of IL-15 which was increased after stimulation with HIV-1_{IIIB} or iHIV-1_{IIIB} isolate. We detected maximum two-fold increase against mock control in IL-15 production by stimulation with 10 ng/ml of HIV-1 p24 antigen (monocytes + iHIV-1_{IIIB} 139·54 pg/ml *versus* 69·24 pg/ml, monocytes + HIV-1_{IIIB} 154·34 pg/ml *versus*

68.33 pg/ml; PBMC + iHIV-1_{IIIB} 76.31 pg/ml versus 40.17 pg/ml, PBMC + HIV-1_{IIIB} 88.93 pg/ml versus 39.74 pg/ml). In addition, a significant increase of produced IL-15 was observed by cultivation of monocytes and PBMC with HIV-1_{IIIB} isolate at the concentration 1 ng/ml and 10 ng/ml of HIV-1 p24 antigen (P < 0.01; Student's *t*-test). IL-15 production from purified monocytes was much higher than that observed from PBMC, thus confirming that monocytes are the main cell type involved in IL-15 production.

DISCUSSION

B lymphocyte dysfunctions, frequently observed during HIV infection, include hypergammaglobulinaemia, circulating activated B cells, spontaneous immunoglobulin secretion and the presence of autoantibodies [1,20]. Several mechanisms have been postulated to explain this abnormal B cell function. Some

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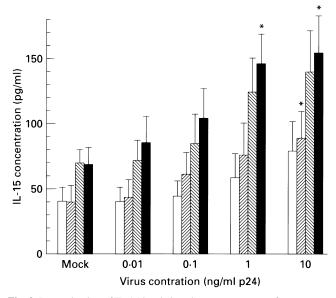


Fig. 3. Determination of IL-15 levels in culture supernatants of monocytes and peripheral blood mononuclear cells (PBMC) $(1 \times 10^6 \text{ cells/ml})$ after 7 days incubation in presence or absence of HIV-1_{IIIB} and iHIV-1_{IIIB}. Data are the mean \pm s.e.m. of six separate experiments performed in duplicate. □, PBMC + iHIV-1; □, PBMC + HIV-1; □, monocytes + iHIV-1; ■, monocytes + HIV-1. **P* < 0.01.

groups [6,7] described T cell contact-dependent activation of B cells by HIV infection. Nevertheless, none of these mechanisms clearly explained hypergammaglobulinaemia at all stages of HIV infection, even in ARC/AIDS patients that exhibit CD4⁺ T cell dysfunction. Unlike CD4⁺ T cells, which are rapidly depleted in HIV-1⁺ individuals, monocytes/macrophages withstand prolonged periods of infection by HIV with significantly less cytopathology and cell death, show altered cytokine production and have an activated phenotype [21]. Therefore, we propose that a monocytes/macrophages-mediated cosignalling pathway rather than T cell contact-dependent interactions is involved in HIV-1-associated hypergammaglobulinaemia.

In the present study we investigated serum levels of IL-15 in 84 $\rm HIV$ -1⁺ individuals at the different stages of disease. Fifty-five percent of $\rm HIV$ -1⁺ and 71% of $\rm HIV$ -1⁻ subjects in our study showed undetectable levels of serum IL-15. However, similar detectability was previously described for IL-6 serum levels in $\rm HIV$ -1⁻ and $\rm HIV$ -1⁺ individuals [8].

We observed a significant increase in serum levels of IL-15 in HIV-1⁺ carriers compared with healthy controls. This elevation was found to correlate with disease progression. In particular, the IL-15 level rose during WR III–IV stages, but stabilized and remained unchanged in ARC/AIDS patients (WR V–VI). A similar tendency has been previously described for IgG serum levels [8]. Therefore, we analysed serum levels of IgG and IgM in HIV-1⁺ individuals in a further set of experiments. We observed significant increases of IgG levels in the group of HIV-1⁺ individuals with detectable levels of serum IL-15. Although the difference in concentrations appears to be of no clinical importance, we conclude that IL-15 may play a role in inducing hypergammaglobulinaemia associated with HIV-1 infection.

Numerous cytokines are able to support proliferation and/or

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differentiation of activated B cells. Among them, IL-6, IL-4 and TNF- α were considered to induce B cell hyperactivity [8,22,23]. Some authors have ascribed the hypergammaglobulinaemia of HIV-1-infected individuals to an increased production of IL-6 by cells of the monocyte-macrophage system as a consequence of HIV-1 infection [22,24]. Nevertheless, hypergammaglobulinaemia in HIV-1-infected persons does not clearly correlate with plasma levels of IL-6 [8]. We demonstrate that IL-15 levels correlate with serum IgG in HIV-1⁺ individuals (r = 0.376; P < 0.0001), but do not clearly correlate with serum IgM (r = 0.265; P = 0.015). This is the first report of serological analysis that describes a correlation between HIV-1-associated hypergammaglobulinaemia and immunological factors.

Since increased production of IL-15 by lipopolysaccharide (LPS)-activated murine macrophages has been reported [16], in further experiments we analysed IL-15 production by purified monocytes and PBMC from healthy donors. We detected a two-fold, significant increase in the amount of IL-15 released from monocytes and PBMC stimulated with HIV-1_{IIIB} virus isolate. These results suggest that HIV-1 virus exerts direct influence on IL-15 production in monocytes and therefore also on IL-15 serum levels.

In conclusion, we determined serum levels of IL-15 in HIV-1⁺ persons at different stages of disease in comparison with healthy controls. We demonstrate that IL-15 levels are significantly elevated in HIV-1-infected individuals. Moreover, as investigated in the present study, we have found correlation between IL-15 and immunoglobulin serum levels, indicating that IL-15 may contribute to the pathogenesis of HIV-1-associated hypergamma-globulinaemia, even in the later stages of infection. IL-15, secreted by activated monocytes/macrophages, costimulates proliferation and differentiation of activated B cells in a manner similar to IL-2 [15]. Because of its broad stimulatory activities, we suggest that this interleukin is a useful parameter for HIV-disease and can provide more precise assessment of the immune status.

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REFERENCES

- 1 Schnittman SM, Lane HC, Higgins SE *et al.* Direct polyclonal activation of human B lymphocytes by the acquired immune deficiency syndrom virus. Science 1986; 233:1084–6.
- 2 Fauci AS. The human immunodeficiency virus: infectivity and mechanisms of immunopathogenesis. Science 1988; 239:617–22.
- 3 Oyaizu N, Chirmule N, Ohnishi Y *et al*. Human immunodeficiency virus type 1 envelope glycoproteins gp120 and gp160 induce interleukin-6 production in CD4⁺ T-cell clones. J Virol 1991; **65**:6277– 82.
- 4 Berberian L, Goodglick L, Kipps TJ *et al*. Immunoglobulin V_H3 gene products: natural ligands for HIV gp120. Science 1993; 261:1588–91.
- 5 Goodglick L, Zevit N, Neshat MS *et al.* Mapping the superantigenbinding site of HIV-1 gp120. J Immunol 1995; **155**:5151–9.
- 6 Chirmule N, Kalyanaraman VS, Lederman S et al. HIV-gp160-induced T cell-dependent B cell differentiation. J Immunol 1993; 150:2478–86.
- 7 Macchia D, Almerigogna F, Parronchi P *et al*. Membrane tumour necrosis factor-α is involved in the polyclonal B-cell activation induced by HIV-infected human T-cells. Nature 1993; **363**:464–6.

- 8 Re MC, Zauli G, Furlini G *et al.* Hypergammaglobulinemia in HIV-1 infected individuals does not clearly correlate with plasma levels of IL-6. AIDS Res Hum Retrovir 1992; 8:1289–95.
- 9 Amadori A, Zamarchi R, Veronese ML et al. B cell activation during HIV-1 infection. J Immunol 1991; 146:57–62.
- 10 Roux-Lombard P, Modoux C, Crushaud A *et al.* Purified blood monocytes from HIV-1-infected patients produce high levels of TNF- α and IL-1. Clin Immunol Immunopathol 1989; **50**:374–84.
- 11 Benjamin D, Knobloch TJ, Dayton MA. Human B-cell interleukin-10: B-cell lines from patients with acquired immunodeficiency syndrome and Burkitt's lymphoma constitutively secrete large quantities of interleukin-10. Blood 1992; 80:1289–98.
- 12 Grabstein KH, Eisenman J, Shanebeck K *et al.* Cloning of a T cell growth factor that interacts with the β chain of the interleukin-2 receptor. Science 1994; **264**:965–8.
- 13 Carson WE, Ross ME, Baiocchi RA *et al.* Endogenous production of interleukin 15 by activated human monocytes is critical for optimal production of interferon-γ by natural killer cells *in vitro*. J Clin Invest 1995; **96**:2578–82.
- 14 Carson WE, Giri JG, Lindemann MJ *et al.* Interleukin (IL) 15 is a novel cytokine that activates human natural killer cells via components of the IL-2 receptor. J Exp Med 1994; 180:1395–403.
- 15 Armitage RJ, Macduff BM, Eisenman J, Paxton R, Grabstein KH. IL-15 has stimulatory activity for the induction of B cell proliferation and differentiation. J Immunol 1995; 154:483–90.

- 16 Doherty TM, Seder RA, Sher A. Induction and regulation of IL-15 expression in murine macrophages. J Immunol 1996; 156:735–41.
- 17 Giri JG, Ahdieh M, Eisenman J *et al.* Utilization of the β and γ chains of the IL-2 receptor by the novel cytokine IL-15. EMBO 1994; **13**:2822–30.
- 18 Balasubramanian S, Chernov-Rogan T, Davis AM *et al.* Ligand binding kinetics of IL-2 and IL-15 to heteromers formed by extracellular domains of the three IL-2 receptor subunits. Int Immunol 1995; 7:1839–49.
- 19 Freundlich B, Avdalovic N. Use of gelatin/plasma coated flasks for isolating human peripheral blood monocytes. J Immunol Methods 1983; 62:31–37.
- 20 Amadori A, Chieco-Bianci L. B-cell activation and HIV-1 infection: deed and misdeeds. Immunol Today 1990; **11**:374–9.
- 21 Mosier D Sieburg H. Macrophage-tropic HIV: critical for AIDS pathogenesis? Immunol Today 1994; 15:332–9.
- 22 Breen EC, Rezai AR, Nakajima K *et al.* Infection with HIV is associated with elevated IL-6 levels and production. J Immunol 1990; **144**:480–4.
- 23 Boue F, Wallon C, Goujard C *et al.* HIV induces IL-6 production by human B lymphocytes. J Immunol 1992; **148**:3761–7.
- 24 Birx DL, Redfield DD, Tencer K *et al.* Induction of interleukin-6 during human immunodeficiency virus infection. Blood 1990; **70**: 2303–10.