REVIEW

Eotaxin and asthma: some answers, more questions

C. J. CORRIGAN Department of Respiratory Medicine, Imperial College School of Medicine, Charing Cross Campus, London, UK

(Accepted for publication 2 September 1998)

Cytokines and chemokines are characterized by their pleiotropicity and redundancy of function. Furthermore, any one cytokine will generally induce the release of one or more others. Consequently, in any inflammatory process it is usually straightforward to establish excessive production of a wide range of these inflammatory mediators. Discovery of a new cytokine or chemokine is typically followed by a flurry of scientific activity as researchers seek to establish its presence and possible role in their own particular disease. Such searches are seldom disappointing. What is more difficult in these situations is to identify the 'key players', so that a cogent and ultimately successful therapeutic approach can be formulated. The chemokine eotaxin has been suggested to be such a key player in the pathogenesis of asthma. It is the purpose of this brief review to examine the evidence for this proposition.

Eotaxin was first identified as an activity appearing in bronchoalveolar lavage fluid of guinea pigs in the course of antigeninduced pulmonary eosinophil infiltration which caused specific accumulation of indium-labelled eosinophils when injected into guinea pig skin [1,2]. The cDNA encoding the purified eotaxin protein was subsequently cloned [3]. Soon afterwards the human homologous eotaxin cDNA and genomic sequences were identified [4-6]. The human gene is located on chromosome 17 and comprises of three exons and two introns. The 5'-flanking region of the gene contains a number of consensus regulatory elements including binding sites for AP-1, NF-kB, NFIL-6, interferon-gamma (IFN- γ) response element and the glucocorticold receptor, suggesting possible regulation of expression of the gene by cytokines as well as glucocorticoids. Human eotaxin appears to be constitutively expressed, at least at the level of mRNA, at detectable concentrations in a variety of human tissues, and at relatively high concentrations in the small intestine and colon. The significance of this observation is not clear. Another recently described chemokine has rather confusingly been named eotaxin-2, not because of any close structural homology with eotaxin (in fact the molecules share only 39% of identical amino acids) but because it appears so far to have a spectrum of activities identical to those of eotaxin, although it is somewhat less potent [7,8].

Eotaxin is a member of the CC family of chemokines, so called because of their conserved two N-terminal cysteine residues (although natural eotaxin preparations seem to contain N-terminally truncated forms missing two or three amino acids [9]). The genes encoding these chemokines are clustered on chromosome 17. The hallmark of the CC chemokines in general is their ability to chemoattract and activate inflammatory leucocytes, particularly lymphocytes, monocytes, eosinophils and basophils, as well as some stromal cells such as endothelial and smooth muscle cells [10]. An important, and probably related function is their ability to induce diapedesis of cells across vascular endothelium, a function of obvious relevance to the propagation of tissue inflammation. A final notable effect of some of the CC chemokines, including eotaxin, is their ability to cause IgE-independent degranulation of basophils [11]. These activities of CC chemokines are mediated through a family of receptors belonging to the 7 transmembranespanning (serpentine) superfamily. At present, nine CC chemokine receptors (CCR1-9) have been described, and it seems likely that further receptors (as well as further CC chemokines) will be discovered, especially using techniques such as expression sequence tag screening [8]. In general, any particular CC chemokine can bind to several CC receptors. Eotaxin (and eotaxin-2) are exceptional in this regard, since they bind only to the CCR3 receptor. Thus far, only a limited range of cells has been found to express CCR3: eosinophils, basophils, 'Th2-type' T cells and microglial cells in the central nervous system [11-15]. Thus, only eotaxin and eotaxin-2 can chemoattract and activate these target cells specifically. While other CC chemokines (MCP-2, MCP-3, MCP-4 and RANTES) may exert a similar range of effects on these target cells by binding to CCR3 [12], these chemokines are less specific in the sense that they also bind to CC chemokine receptors other than CCR3 on a wider range of target cells.

Asthma is associated with chronic, T cell-mediated inflammation of the bronchial mucosa characterized by a striking and relatively specific influx of eosinophils [16]. Eosinophil products (granule basic proteins and membrane-derived lipid mediators) are believed to cause much of the bronchial mucosal damage which is thought ultimately to give rise to clinical symptoms. Release of mast cell and basophil products, IgE-mediated or otherwise, is additionally thought to cause acute exacerbation of asthma symptoms, e.g. after acute exposure to allergens or after exercise. One fundamental aspect of asthma research has been to determine the mechanism of this specific eosinophil influx. Local release of cytokines from 'Th2-type' activated T cells (particularly IL-5, which acts exclusively on eosinophils and their committed precursors, but also IL-3 and granulocyte-macrophage colonystimulating factor (GM-CSF) which act on eosinophils but less specifically) is thought to be pivotal to this process, since these cytokines (and IL-5 specifically) can promote maturation of eosinophils in the bone marrow, increased adherence of eosinophils to vascular endothelium, and priming, activation and prolonged survival of these cells in the tissues [16]. One function not

Correspondence: Dr C. J. Corrigan, Department of Respiratory Medicine, Imperial College School of Medicine, Charing Cross Campus, Fulham Palace Road, London W6 8RF, UK.

clearly attributable to these cytokines is the induction of diapedesis of cells across vascular endothelium. This role is potentially ideally filled by the CC chemokines. In particular, eotaxin and eotaxin-2 may attract eosinophils, basophils and 'Th2-type' T cells into tissues, and specifically so via interaction with the CCR3 receptor. It is little wonder, then, that these molecules have become one of the current icons of the asthma researcher.

There exists considerable evidence in support of this cooperative scenario between eotaxin and IL-5 in the mechanism of specific eosinophil influx in the tissues from studies on animals. Thus, in guinea pigs accumulation of eosinophils in the skin following subcutaneous injection of eotaxin was time-dependently augmented by concomitant i.v. infusion of IL-5, which released eosinophils from the bone marrow into the circulation [17]. IL-5 alone, when given subcutaneously, did not cause local eosinophil accumulation. Eotaxin, when injected directly into the arterial supply of the guinea pig femoral bone marrow, caused a rapid and selective release of eosinophils into the draining vein [18]. Eosinophil progenitor cells, which have recently also been shown to express CCR3 [19], were also released, an effect not observed with IL-5. Eotaxin administered subcutaneously to IL-5 gene-deleted mice failed to evoke local eosinophil accumulation unless exogenous IL-5 was administered concurrently [20]. The mechanisms by which eotaxin evokes this 'two-way traffic' of eosinophils (into tissues but out of the bone marrow) are as yet poorly defined, but these may at least partly involve interactions of β_1 - and β_2 -integrin adhesion molecules (CD11b/CD18, VLA-4) on eosinophils with counter-receptors (intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1)) on local endotlielial cells [21-24]. Interestingly, IL-4 may also attract eosinophils through secondary eotaxin release [9,24].

What of human asthma? Two recent studies [25,26] have demonstrated elevated numbers of cells expressing eotaxin mRNA and protein in the bronchial mucosa of atopic asthmatics compared with controls. Eotaxin mRNA expression correlated positively with local eosinophil numbers and inversely with disease severity. One of these studies [25] in addition showed CCR3 expression on local infiltrating eosinophils. These studies are consistent with the hypothesis that eotaxin, through its specific action on CCR3, plays a role in the specific recruitment of eosinophils to the asthmatic bronchial mucosa, a phenomenon which in turn regulates asthma severity. The potential cellular sources of eotaxin in inflammatory reactions are beginning to be delineated. One study referred to above [25] clearly identified both epithelial and endothelial cells as the principal cells expressing eotaxin mRNA in the asthmatic bronchial mucosa. Similarly, human epithelial cell lines have been shown to release eotaxin in *vitro* after cytokine (tumour necrosis factor-alpha (TNF- α), IL-1 β , IFN- γ) stimuli [27]. Human [26] as well as animal [28] studies implicate T cells as a probable potential source, while eosinophils themselves have been shown to secrete the eotaxin in response to IL-3 stimulation [29]. Skin fibroblasts secrete eotaxin in response to TNF- α and IL-4 stimulation [9].

To return to the question posed at the beginning of this article, it seems reasonable from these data to suggest that eotaxin may play a key role in asthma pathogenesis. Reservations remain, however. First, knowledge of the entire repertoire of CC chemokines and their receptors is probably incomplete. It is always possible that further CC chemokines with a spectrum of activities similar to eotaxin will be discovered (in this regard, eotaxin-2 is already with us). Second, little or nothing is known about the molecular mechanisms which result in eotaxin release in inflammatory processes in general and asthma in particular. It is possible that a knowledge of these mechanisms will be more revealing than simply a knowledge of the existence of eotaxin itself. Finally, owing to the redundancy of action of chemokines, it is always possible that targeting of one particular chemokine in a disease process will not be particularly effective since other chemokines may assume similar roles. This may explain why, in a model of antigen-induced pulmonary eosinophil accumulation in mice, targeted deletion of the eotaxin gene reduced, but did not abolish, pulmonary cosinophil infiltration [30]. Having said this, targeting of the CCR3 receptor would seem to be a much more promising therapeutic proposition in asthma, since this would block the effects not only of eotaxin and eotaxin-2, but also of those other CC chemokines known to bind to this receptor. Further advances in this direction are eagerly awaited.

REFERENCES

- 1 Griffiths-Johnson DA, Collins PD, Rossi AG, Jose PJ, Williams TJ. The chemokine, eotaxin, activates guinea-pig eosinophils *in vitro* and causes their accumulation into the lung *in vivo*. Biochem Biophys Res Commun 1993; **197**:1167–72.
- 2 Jose PJ, Griffiths-Johnson DA, Collins PD *et al.* Eotaxin: a potent eosinophil chemoattractant cytokine detected in a guinea pig model of allergic airways inflammation. J Exp Med 1994; **179**:881–7.
- 3 Jose PJ, Adcock IM, Griffiths-Johnson DA, Berkman N, Wells TNC, Williams TJ, Power CA. Eotaxin: cloning of an eosinophil chemoattractant cytokine and increased mRNA expression in allergenchallenged guinea-pig lungs. Biochem Biophys Res Commun 1994; 205:788–94.
- 4 Kitaura M, Nakajima T, Imai T, Harada S, Combadiere C, Tiffany HL, Murphy PM, Yoshie O. Molecular cloning of human eotaxin, an eosinophil-selective CC chemokine and identification of a specific eosinophil eotaxin receptor, CC chemokine receptor 3. J Biol Chem 1996; 271:7725–30.
- 5 Garcia-Zepeda EA, Rothenberg ME, Weremowicz S, Sarati MN, Morton CC, Luster AD. Genomic organization, complete sequence, and chromosomal location of the gene for human eotaxin (SCYA11) an eosinophil-specific CC chemokine. Genomics 1997; 4:471–6.
- 6 Hein H, Schluter C, Kulke R, Christophers E, Schroder JM, Bartels J. Genomic organization, sequence, and transcriptional regulation of the human eotaxin gene. Biochem Biophys Res Commun 1997; 237:537– 42.
- 7 Forssmann U, Uguccioni M, Loetscher P, Dahinden CA, Langen H, Thelen M, Baggiolini M. Eotaxin-2, a novel CC chemokine that is selective for the chemokine receptor CCR3, and acts like eotaxin on human eosinophil and basophil leukocytes. J Exp Med 1997; 185:2171–6.
- 8 White JR, Imburgia C, Dul E *et al.* Cloning and functional characterization of a novel human CC chemokine that binds to the CCR3 receptor and activates human eosinophils. J Leuk Biol 1997; **62**:667– 75.
- 9 Mochizuki M, Bartels J, Mallet AI, Christophers E, Schroder JM. IL-4 induces eotaxin: a possible mechanism of selective eosinophil recruitment in helminth infection and atopy. J Immunol 1998; 160:60–68.
- 10 Alam R. Chemokines in allergic inflammation. J Allergy Clin Immunol 1997; 99:273–7.
- 11 Ugoccioni M, Mackay CR, Ochensberger B *et al.* High expression of the chemokine receptor CCR3 in human blood basophils. Role in activation by eotaxin, MCP-4, and other chemokines. J Clin Invest 1997; **100**:1137–43.
- 12 Heath H, Qin S, Rao P, Wu L, LaRosa G, Kassam N, Ponath PD, Mackay CR. Chemokine receptor usage by human eosinophils: the

© 1999 Blackwell Science Ltd, Clinical and Experimental Immunology, 116:1-3

importance of CCR3 demonstrated using an antagonistic monoclonal antibody. J Clin Invest 1997; **99**:178–84.

- 13 Sallusto F, Lenig D, Mackay CR, Lanzavecchia A. Flexible programs of chemokine receptor on human polarized T helper 1 and 2 lymphocytes. J Exp Med 1981; 187:875–83.
- 14 Sallusto F, Mackay CR, Lanzavecchia A. Selective expression of the eotaxin receptor CCR3 by human T helper 2 cells. Science 1997; 277:2005–7.
- 15 He J, Chen Y, Farzan M et al. CCR3 and CCR5 are co- receptors for HIV-1 infection of microglia. Nature 1997; 385:645–7.
- 16 Corrigan CL, Kay AB. T cells and eosinophils in the pathogenesis of asthma. Immunol Today 1992; 13:501–7.
- 17 Coffins PD, Marleau S, Griffiths-Johnson DA, Jose PJ, Williams TJ. Cooperation between interleukin-5 and the chemokine eotaxin to induce eosinophil accumulation *in vivo*. J Exp Med 1995; **182**:1169–74.
- 18 Palframan RT, Collins PD, Williams TJ, Rankin SM. Eotaxin induces a rapid release of eosinophils and their progenitors from the bone marrow. Blood 1998; 91:2240–8.
- 19 Peled A, Gonzalo JA, Lloyd C, Gutierret-Ramos J-C. The chemotactic cytokine Eotaxin acts as a granulocyte-macrophage colony-stimulating factor during lung inflammation. Blood 1998; **91**:1909–16.
- 20 Mould AW, Matthaei KI, Young IG, Foster PS. Relationship between interleukin-5 and eotaxin in regulating blood and tissue eosinophilia in mice. J Clin Invest 1997; 99:1064–71.
- 21 Burke-Gaffney A, Hellewell PG. Eotaxin stimulates eosinophil adhesion to human lung microvascular endothelial cells. Biochem Biophys Res Commun 1996; 227:35–40.
- 22 Hohki G, Terada N, Hamano N *et al.* The effects of eotaxin on the surface adhesion molecules of endothelial cells and on eosinophil adhesion to microvascular endothelial cells. Biochern Biophys Res Commun 1997; **241**:136–41.

- 23 Sanz MJ, Ponath PD, Mackay CR *et al*. Human eotaxin-induced alpha4 and beta2 integrin-dependent eosinophil accumulation in rat skin *in vivo*: delayed generation of eotaxin in response to IL-4. J Immunol 1998; **160**:3569–76.
- 24 Lundahl J, Moshfegh A, Gronneberg R, Hallden G. Eotaxin increases the expression of CD11b/CD18 and adhesion properties in IL-5, but not fMLP-prestimulated human peripheral blood eosinophils. Inflammation 1998; 22:123–35.
- 25 Ying S, Robinson DS, Meng Q *et al.* Enhanced expression of eotaxin and CCR3 mRNA and protein in atopic asthma. Association with airway hyperresponsiveness and predominant co-localization of eotaxin mRNA to bronchial epithelial and endothelial cells. Eur J Immunol 1997; 27:3507–16.
- 26 Mattoli S, Stacey MA, Sun G, Bellini A, Marini M. Eotaxin expression and eosinophilic inflammation in asthma. Biochem Biophys Res Commun 1997; 236:299–301.
- 27 Lilly CM, Nakamura H, Kesselman H *et al.* Expression of eotaxin by human lung epithelial cells. Induction by cytokines and inhibition by glucocorticoids. J Clin Invest 1997; **99**:1767–73.
- 28 MacLean JA, Ownbey R, Luster AD. T cell-dependent regulation of eotaxin in antigen-induced pulmonary eosinophilia. J Exp Med 1996; 184:1461–9.
- 29 Garcia-Zepeda EA, Rothenberg ME, Ownbey RT, Celestin J, Leder P, Luster AD. Human eotaxin is a specific chemoattractant for eosinophil cells and provides a new mechanism to explain tissue eosinophilia. Nature Med 1996; 2:449–56.
- 30 Rothenberg ME, MacLean JA, Pearlman E, Luster AD, Leder P. Targeted disruption of the chemokine eotaxin partially reduces antigeninduced tissue eosinophilia. I Exp Med 1997; 185:785–90.