

# Local and Distant Sequences Are Required for Efficient Readthrough of the Barley Yellow Dwarf Virus PAV Coat Protein Gene Stop Codon†

CHRIS M. BROWN, S. P. DINESH-KUMAR‡ AND W. ALLEN MILLER\*

*Departments of Plant Pathology and Biochemistry & Biophysics, and Molecular, Cellular and Developmental Biology Program, Iowa State University, Ames, Iowa 50011*

Received 12 February 1996/Accepted 17 May 1996

**Many viruses use stop codon readthrough as a strategy to produce extended coat or replicase proteins. The stop codon of the barley yellow dwarf virus (PAV serotype) coat protein gene is read through at a low rate. This produces an extended polypeptide which becomes part of the virion. We have analyzed the *cis*-acting sequences in the barley yellow dwarf virus PAV genome required for this programmed readthrough in vitro in wheat germ extracts and reticulocyte lysates and in vivo in oat protoplasts. Two regions 3' to the stop codon were required. Deletion of sections containing the first 5 of the 16 CCN NNN repeats located 3' of the stop codon greatly reduced readthrough in vitro and in vivo. Surprisingly, readthrough also required a second, more distal element that is located 697 to 758 bases 3' of the stop codon within the readthrough open reading frame. This element also functioned in vivo in oat protoplasts when placed more than 2 kb from the coat protein gene stop in the untranslated region following a GUS reporter gene. This is the first report of a long-range readthrough signal in viruses.**

Viruses in the luteovirus group utilize a variety of unusual translational control mechanisms to express their essential genes (reviewed in references 36 and 38). These include ribosomal frameshifting (7), stop codon readthrough (16), leaky scanning (17), and the use of a 3' translational enhancer (64). Luteoviruses can be divided into two subgroups (34, 38), with barley yellow dwarf virus (BYDV) PAV serotype being the best characterized member of subgroup I. In all luteoviruses, there is a block of three open reading frames (ORFs) which are expressed from a single subgenomic RNA (sgRNA1) and make up the 3' half of the genome. These encode the coat protein (CP), a protein of about 17 kDa (the 17K protein) that is nested within the coat protein sequence but in a different reading frame, and an ORF of about 50 kDa following the coat protein (RT) (39). RT is expressed by readthrough of the CP stop codon as a CP-RT fusion protein (16). The CP and a carboxy-terminally truncated form of CP-RT are the structural proteins of the virion (11, 18, 20, 63), with the RT domain located on the surface of the virion (46). The ratio of fusion protein to CP in purified virus preparations differs markedly between luteoviruses, serotypes of BYDV (20, 63), and individual virus preparations but has been estimated to be between 1:100 and 1:4 (2, 8, 11, 18, 20, 63). The CP-RT fusion protein is required for the aphid transmission of members of both subgroups of luteoviruses (8, 10) and the luteo-like enamovirus pea enation mosaic virus (PEMV) (13).

Stop codon readthrough is used as a regulatory strategy by a large number of plant, animal, and bacterial viruses (24, 67). The eukaryotic release factor complex normally decodes stop

signals efficiently (37, 52, 53, 69). However, at certain stop codons, the competition between the eukaryotic release factor complex and suppressor tRNA for the stop signal shifts in favor of the tRNA. The tRNAs involved are likely to be naturally occurring isoacceptors with near-cognate anticodons (25, 57, 68). This phenomenon was first reported for a eukaryotic virus in tobacco mosaic virus (41) and has been subsequently observed or proposed for members of the alpha-, carmo-, enamo-, furo-, hordei-, luteo-, machlo-, necro-, tobamo-, tobra-, tombus-, tymo-, and retroviruses (6, 24, 25, 38, 65, 67). Interference with the readthrough process in plant or human pathogenic viruses may also interfere with the viral life cycle.

In a few cases, the signals that permit or promote readthrough have been identified, but the mechanisms by which they act are not understood. The simplest signal is that of the mammalian alphaviruses (54). In vitro, a UGA C stop signal is sufficient to permit 10% readthrough, with the C providing an important determinant (33). In vivo, each of the three stop codons can be suppressed at this location in the Sindbis virus genome (32). Interestingly, it has been shown that termination is profoundly affected by the base following the stop codon (5, 37, 42). Stop codons followed by pyrimidine residues were poorly recognized as stop codons, with UGA C being one of the weakest signals (37). Thus, the combination of this weak stop signal and the presence of a UGA-suppressing tRNA might permit readthrough of the alphavirus stop (56). For tobacco mosaic virus RNA, a larger signal, UAG CAR YYA, is necessary and sufficient for 5% readthrough in vivo (49, 50, 58). The wild-type signal, UAG CAA UUA, also promotes 21% readthrough in yeast cells and 2% readthrough in mammalian cells (51) but does not function in *Escherichia coli* (50). A more elaborate signal is required for UAG readthrough by murine leukemia virus, including a pseudoknot spaced 8 nucleotides from the UAG stop, as well as specific sequences within the spacer and loops (19, 65).

A large number of luteoviral structural gene sequences are available because of the economic importance of this group of

\* Corresponding author. Mailing address: Department of Plant Pathology, 351 Bessey Hall, Iowa State University, Ames, IA 50011. Phone: (515) 294-2436. Fax: (515) 294-9420. Electronic mail address: wamiller@iastate.edu.

† Paper J-16735 of the Iowa Agriculture and Home Economics Experiment Station, project 3330.

‡ Present address: USDA Plant Gene Expression Center, Albany, CA 94710.

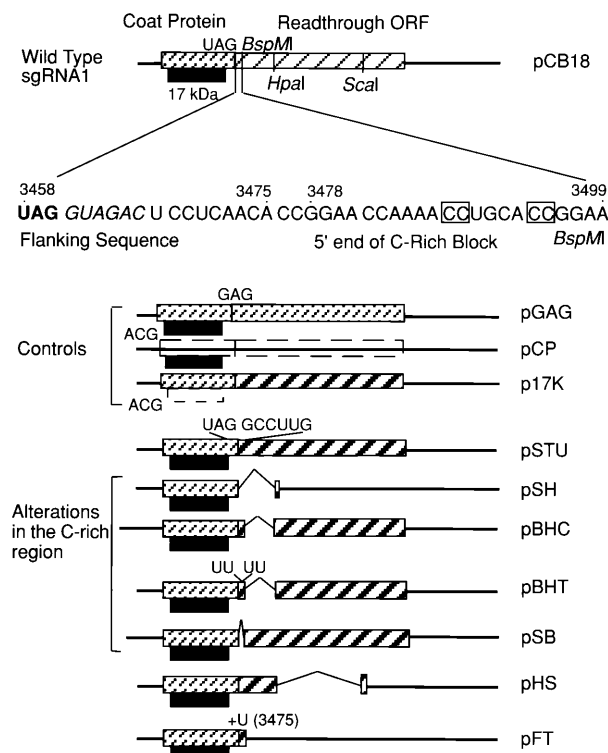


FIG. 1. Maps of BYDV-PAV sgRNA1 (pCB18) and derived transcripts. The sequence between the CP stop codon (UAG) and the *BspMI* site is numbered as in reference 39. The two pairs of C's altered to U's in the pBHT derivative are boxed. ORFs for which the AUG has been changed to ACG are shown as dashed boxes.

viruses. In all known luteovirus sequences, the CP stop codon is flanked by the sequence CN AAA UAG GUA GAC (Fig. 1). This conserved block is entirely different from all known readthrough contexts. After a short spacer of 6 to 15 bases, this sequence is followed by a C-rich block containing 7 to 16 tandem repeats of CCN NNN (Fig. 1; also see Fig. 6). The length of the spacer puts the CCN of these repeats in frame, encoding prolines. This region, in which every second amino acid is proline, has been proposed to form a hinge between the CP and the remainder of the RT domain (21). In all luteoviruses, at least one of the CCN NNN repeats contains the sequence CCCC (38). No conserved secondary structures have been identified within this region. Here, we determine which of these elements following the stop are required for readthrough *in vitro* and *in vivo*. Two regions, part of the C-rich block and an additional element located some distance from the CP stop codon, were found to be required for readthrough.

#### MATERIALS AND METHODS

**Full-length infectious clones.** All plasmids were constructed by standard techniques (47) and site-specific mutagenesis (30). Plasmid pPAVGUSRT1 is derived from a full-length BYDV PAV infectious clone pPAV6 (14) and contains the *uidA* reporter gene (without its initiation codon), which encodes  $\beta$ -glucuronidase (GUS). The *ApaI* (in the N-terminal GUS coding region)-*AflI* (following the GUS stop codon, blunted) fragment of pAGUS1 (49) was used to replace bases 3593 to 3787 (*HpaI*) of pPAV6 to generate pPAVGUSRT1 (see Fig. 4). In pPAVGUSRT2, the fragment was placed after nucleotide 3477, and in pPAVGUSRT15, it was placed after nucleotide 3534. In pPAVGUSRT3, pPAVGUSRT4, and pPAVGUSRT21, GAG sense codons replace the TAG stop codon of pPAVGUSRT1, pPAVGUSRT2, pPAVGUSRT20, respectively.

Plasmids pPAVGUSRT9 and pPAVGUSRT10 contain deletions, in the coat protein ORF, of bases 2985 to 3345 and 2985 to 3423, respectively. pPAVGUS

RT20 is a derivative of pPAVGUSRT15 with a T inserted after 3475 and the final T (base 3534) before the *ApaI* site deleted to maintain the frame. pPAVGUSRT6, pPAVGUSRTTHN, pPAVGUSRTNA, and pPAVGUSRTAS are derivatives of pPAVGUSRT1 with deletions of 3788 (*HpaI* blunt) to 4515 (*ScaI* blunt), 3788 (*HpaI* blunt) to 4120 (*NdeI* filled), 4123 (*NdeI* filled) to 4154 (*Acc65I* filled), and 4155 (*Acc65I* filled) to 4515 (*ScaI* blunt), respectively.

**Plasmids derived from full subgenomic RNA1.** The parent vector pCB18 contains the complete sgRNA1 sequence adjacent to a T7 promoter. Maps are shown in Fig. 1. A T7 promoter was added to the subgenomic RNA start site (italics) by PCR with primer SG1 (GGTCTAGATAATACGACTCACTATAG TGAAGGTGACGACTCCACATC) and a 3' primer. An *XbaI-SalI* fragment of this product was ligated into *XbaI-SalI*-cut pSP18 (16). In pGAG, the CP stop codon, UAG, was altered to GAG. Templates for *in vitro* transcription of sgRNAs in which the start codons of the CP or the 17K protein had been altered to ACG, pCP, and p17K, were generated by PCR with SG1 and a primer complementary to the 3' end of the genome (SMA: GGGTTGCCGAAGTGC TCITTC) from full-length genomic clones harboring these mutations, PAV31 and PAV33 (40). pSTU contains UAG GCCTTG in place of the UAG GUA GAC at the coat protein stop, adding a *StuI* site (italics). The *StuI-HpaI* fragment of pSTU was deleted to give pSH. The *BspMI* (3-base overhang, filled)-*HpaI* fragment of pCB18 was deleted to give pBHC. Two pairs of C's (3488 to 3489, 3494 to 3495) in pBHC were altered to T's in pBHT. The *StuI-BspMI* (3-base overhang, filled) fragment of pSTU was deleted in pSB. The *HpaI-ScaI* fragment was deleted in pHS. The *SalI-BspMI* fragments of pPAVGUSRT20 and pPAVGUSRT21 were subcloned into pCB18 to generate pFT and pFG, respectively. pFNA and pFAS were derived by subcloning *BglII-PstI* fragments from pPAVGUSRTNA and pPAVGUSRTAS into pFT. pFT was cut with *HpaI-NdeI* and filled with Klenow DNA polymerase, which fortuitously produced a mutant containing a slightly larger than expected deletion (3788 to 4129), pFHN. pFG contains a GAG sense codon in place of the TAG stop. A template for transcription truncated after position 4219 was generated by PCR with primers SG1 and 4219 (ITCAGCGTGCCTTGTTATTC). All plasmids were sequenced in relevant regions with an Applied Biosystems model 377 automated sequencer at the Iowa State University Nucleic Acids Facility.

***In vitro* transcription.** Transcripts were synthesized from linearized plasmid templates or PCR products with T7 RNA polymerase by the method described by Promega (43). Capped transcripts were synthesized with MessageMachine kits (Ambion). The RNA concentration was determined with a GeneQuant spectrophotometer and by ethidium bromide staining following electrophoresis.

***In vitro* translation.** Translation in wheat germ extracts was done essentially as recommended by the manufacturer (Ambion). RNAs (usually 0.4  $\mu$ g) were translated in 20- $\mu$ l reaction mixtures containing 150 mM potassium acetate, 2.5 mM magnesium acetate, 15 U of RNasin (Promega), 1  $\mu$ l of master mix minus methionine, 10  $\mu$ l of wheat germ extract, and [ $^{35}$ S]methionine (New England Nuclear). Translation products (1  $\mu$ l) were separated on 6% stacking-15% resolving polyacrylamide gels as described previously (29), except that twice the concentration of resolving gel buffer was used (0.75 M). Three sets of  $M_r$  markers were used (Bio-Rad, Bethesda Research Laboratories, and Amersham); they were separated on a polyacrylamide gel, and the  $M_r$  was determined for each marker from each set. The mean of these three determinations was used as the  $M_r$  indicated (Fig. 2). Gels were fixed for 6 h before being processed for fluorography (Amplify; Amersham). Radioactivity in the gels was quantified with a Phosphorimager 400E and ImageQuant 3.3 software (Molecular Dynamics). Readthrough percentages given in the text represent the mean of three or more independent experiments. Adjustments were made for the relative number of methionines in each product (CP, 4; 17K protein, 3; CP-RT, 12). Translation in reticulocyte lysates was done as described previously (16).

**Protoplast transfection and analysis.** Oat protoplasts were isolated, transfected, and analyzed as described previously (17, 40). They were electroporated with 20  $\mu$ g of full-length transcript and then incubated in the dark for 24 h before being harvested for analysis. Total GUS activity (picomoles of 4-methylumbelliferone [MU] per minute per milligram of protein) was determined as described previously (26). GUS activity was detected following concentration of total protein and separation on semidenaturing 7.5% polyacrylamide gels as described previously (7). Protein concentrations were determined with Bradford protein assay kits (Bio-Rad). RNA analysis was performed as described previously (40). Total RNA was extracted by a procedure in which aurintricarboxylic acid was used as an RNase inhibitor (62), separated on 1% denaturing gels, transferred to a nylon membrane, and probed with an RNA probe complementary to the 3' end of the BYDV PAV genome (from pSP10 [17]). Double-antibody sandwich enzyme-linked immunosorbent assays (ELISA) were done as described previously (22).

#### RESULTS

**Translation *in vitro*.** For these studies, uncapped BYDV PAV subgenomic RNA1 derivatives were transcribed and translated *in vitro*. Different 5' ends of sgRNA1 had been reported for the Illinois (nucleotide 2769) and Australian (nucleotide 2670) isolates of BYDV PAV (16, 28). The 5' end

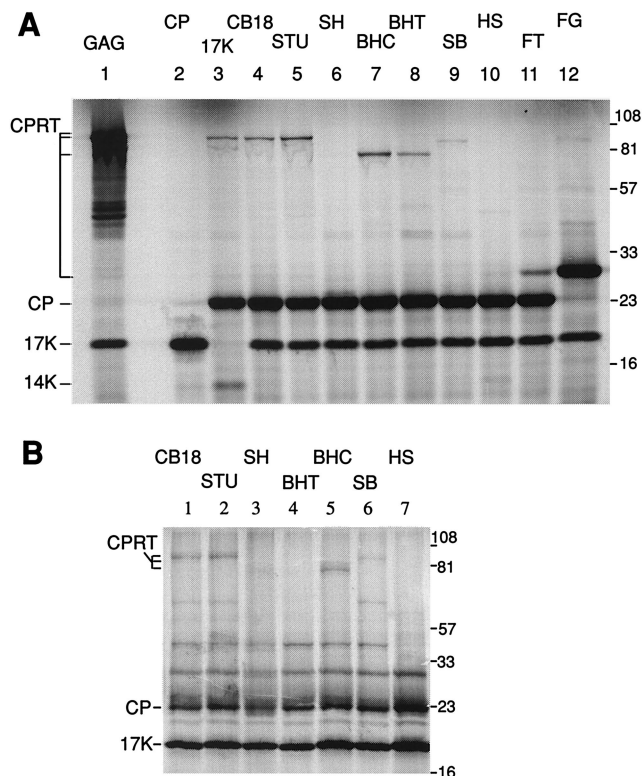


FIG. 2. (A) In vitro translation of sgRNA1 derivatives in wheat germ extracts. The positions of sgRNA1 translation products are indicated on the left. The expected sizes of the CP-RT products are 72 kDa (lanes 1 to 5), 23 kDa (lane 6), 61 kDa (lanes 7 and 8), 71 kDa (lane 9), 35 kDa (lane 10), and 25 kDa (lanes 11 and 12). The sizes of the  $M_r$  markers are indicated on the right. (B) In vitro translation of sgRNA1 derivatives in rabbit reticulocyte lysates. Aliquots (0.4  $\mu$ g) of selected RNAs were translated in rabbit reticulocyte lysates.

described by Kelly et al. was used for this study because the sgRNA1 region of the infectious clone used here is derived from the Australian isolate (14). RNAs were translated in vitro in systems from two organisms. Wheat germ extracts that had been supplemented with wheat germ tRNAs were used for in vitro translation (1), because wheat is a natural host of the virus. For comparison, we also used reticulocyte lysates supplemented with calf liver tRNAs, as used previously for BYDV PAV sgRNA1 translation (16).

Three products were synthesized from sgRNA1 transcripts in wheat germ. Translation of wild-type sgRNA1 is shown in Fig. 2A, lane 4. The most abundant product was 22-kDa CP. A second protein migrating at  $\sim$ 18 kDa, which corresponded to the product of the nested 17K protein ORF within the CP, was also seen. About 1/10 as much of the 17K product which initiates from the second AUG in the mRNA was made compared with CP. It had been shown previously that this 17K protein is synthesized by a leaky scanning mechanism in vitro in rabbit reticulocyte lysates and in vivo in oat protoplasts (17). A third high- $M_r$  protein corresponding to the CP-RT fusion (RT) was also seen. The average readthrough percentage determined from multiple experiments was 0.8% for the wild-type sequence.

Despite slow migration of the 17K and CP-RT proteins, the identities of the products were confirmed by translation of transcripts in which either the CP or 17K AUGs had been changed to ACG (pCP, p17K) or the CP stop codon had been changed to GAG (pGAG; Fig. 2A). When the CP AUG was

altered (lane 2), no CP or CP-RT fusion was detected, as expected. When the 17K protein AUG was altered (lane 3), the CP and CP-RT proteins were unaffected and the 17K protein was not visible. A new, lower- $M_r$  protein (the 14K protein) appeared as predicted by initiation from the next AUG in the mRNA which would give an N-terminally truncated product of the 17K protein ORF with a  $M_r$  of 14,079. When the CP stop was changed to GAG, a prominent fusion protein appeared (lane 1). The additional species in this lane of  $\sim$ 50 kDa probably correspond to internal initiations near the 3' end of the CP gene.

**Sequences required for readthrough.** The 6-base sequence following the stop is the same in all luteoviruses (GUA GAC). To test if it is required for readthrough, we altered five of the bases to create GCC UUG (this introduces a unique *Stu*I site). Readthrough was not reduced; indeed, a modest increase in readthrough was observed (Fig. 2A, lane 5).

We made deletions and alterations in the C-rich region following the stop to test if these sequences were needed for readthrough. Bases 3478 to 3787 (numbered as in reference 39), which include 14 of the 16 CCN NNN repeats, were deleted (pSH). This alters the reading frame, giving an expected RT product with an  $M_r$  of only 22,902. No RT product was detected from this construct. However, a smaller deletion (3500 to 3787) did not affect the amount of readthrough (pBHC). This construct encoded a slightly smaller than wild-type CP-RT product that maintains the wild-type frame ( $M_r$  61,466 [lane 7]). It retained five CC pairs, four of which are evenly spaced as CCN NNN repeats. Alteration of the last two of these CC doublets in this construct to UU caused a significant reduction in readthrough (to 40% of the wild-type control; pBHT [lane 8]). The effect of deletion of a region including these five CC pairs (3462 to 3499) was also tested. This placed the remaining 12 pairs closer to the stop and maintained the reading frame (pSB,  $M_r$  70,641 [lane 9]). This alteration also reduced the amount of readthrough product (to 10%). These data indicate that a portion of the C-rich element is required for readthrough in vitro.

To investigate whether the CCN NNN repeats needed to be in frame to function, a single U was inserted after base 3475, 15 nucleotides after the stop (pFT). This places the repeats in a different reading frame (NCC NNN). A product of the expected size ( $M_r$  24,821) was visible (Fig. 2A, lane 11). Lane 12 contained a derivative of pFT in which the UAG was changed to GAG (pFG). These data indicate that the C-rich region does not need to encode prolines or be in a particular reading frame to function.

BYDV PAV readthrough had been noted previously (16), and leaky scanning has been characterized (17) in rabbit reticulocyte lysates. For comparison, the mutant sgRNA1 transcripts were also translated in these lysates (Fig. 2B). Translation was only one-quarter as efficient on rabbit ribosomes. Translation of sgRNA1 gave the same three products, but their relative amounts differed (Fig. 2B, lane 1). There were also additional products, which are likely to result from initiation at AUGs within the RT ORF (also noted in reference 16). In reticulocyte lysate, the ratio of CP to 17K protein was reversed and the percent readthrough (at about 10%) was higher. Deletions and alterations in the sgRNA had similar effects on readthrough in reticulocyte lysates. Deletion of the C-rich element (lanes 3 and 6) or an alteration to it (lane 5 compared with lane 4) also reduced readthrough in reticulocyte lysates, and changing the conserved bases after the stop had no effect (lane 2).

**A second distal element.** Surprisingly, deletion of a more distal region (3788 to 4515), nearly 700 bases from the stop

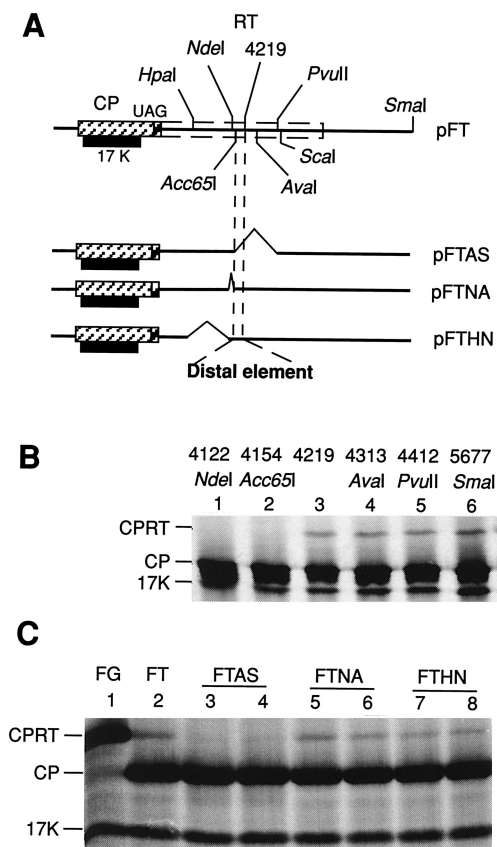


FIG. 3. Effect of the 3' distal element on translation. (A) Sites of linearization with restriction enzymes or deletions within the RT reading frame (dashed box) of pFT. The CP-RT product terminates out of frame before the 3' element because the constructs contain a single-base insertion (Fig. 1). (B) In vitro translation of 3'-truncated RNAs. Transcripts from pFT that had been linearized with the enzymes indicated terminate at the nucleotide shown above each lane (numbering as in reference 39). The positions of the CP-RT (expected size, 25 kDa), CP (22 kDa), and 17K protein (17 kDa) products are indicated at the left. (C) In vitro translation of RNAs with deletions in the distal element. Two independent constructs were tested for each mutation. A derivative which had a GAG sense codon in place of the UAG CP stop codon was translated as a CP-RT marker (lane 1).

codon, reduced readthrough to an undetectable level in both translation systems (expected  $M_r$  35,269 [Fig. 2A, lane 10; Fig. 2B, lane 7]). This suggested that an additional element required for readthrough may be located within this region. Truncations from the 3' end were made in the sgRNA1 to identify the 3' border of this putative element. Capped transcripts were used because the 3' translation enhancer (64) was also removed. This sequence in the 3' end of the PAV genome is required for translation of genomic (64) and subgenomic (9) uncapped RNAs. Truncations were derived from the construct with a U inserted at +15, causing CP-RT translation to terminate only 50 bases after the CP stop (pFT). In these derivatives, the distal element is in the untranslated region (Fig. 3A,  $M_r$  24,821).

Truncations could be made 3' of base 4219 without significantly affecting the amount of RT translation product (5667, 4412, 4313, and 4219 [Fig. 3B, lanes 3 to 6]), but the CP-RT product was undetectable following 3' truncations 5' of base 4154 (4122 and 4154 [lanes 1 and 2]). For the control "in-frame" derivative in which the UAG was replaced by a sense codon (GAG), these truncations had little effect on CP-RT translation (data not shown).

Three internal deletions were made spanning the distal region (3788 to 4515). A deletion (Fig. 3A, 4155 to 4515; pFTAS) that encompassed the 3' border of the distal element (4219) reduced readthrough to less than 10% of wild-type levels (Fig. 3C, lanes 3 and 4). A 32-base central deletion (4123 to 4154; pFTNA) had no effect (Fig. 3C, lanes 5 and 6), whereas a larger 5' deletion (3788 to 4129; pFTHN) reduced readthrough modestly (to 65% of wild-type levels [lanes 7 and 8]). This indicates that a distal RNA element located between nucleotides 4155 and 4219 (697 to 758 nucleotides from the CP stop) is required for readthrough. Furthermore, this element can function when located within or 3' of the readthrough ORF.

**Replicating viral constructs in vivo.** To determine whether the same readthrough signals mapped above are required in vivo, we inserted a modified GUS reporter gene downstream of the CP stop codon in a BYDV PAV full-length infectious clone (14). The GUS ORF lacking the start codon (1894 bases) was inserted in place of bases 3593 to 3787 of the RT ORF (194 bases) (Fig. 4A). This region is dispensable for efficient readthrough in vitro (Fig. 2, pBHC). This places the distal readthrough element in the noncoding region following the GUS reporter gene, 2.4 rather than 0.7 kb from the stop codon. This also places the 3' translation enhancer 1.7 kb further away (64). The resulting hybrid genome of 7.4 kb is 30% larger than the 5.7-kb BYDV PAV genome. Expression of GUS in these constructs requires viral replication for synthesis of its mRNA, sgRNA1. The GUS reporter gene can be expressed only by readthrough of the CP stop codon.

Full-length genomic RNAs were transcribed in vitro and electroporated into oat protoplasts. Accumulation of viral RNAs was measured by Northern (RNA) blot analysis (Fig. 4B), accumulation of virus particles was measured by ELISA (Fig. 4B), and the relative amount of readthrough was measured by monitoring GUS activity (Fig. 4C). Hybrid viral RNA containing the GUS reporter gene without other alterations replicated (pPAVGUSRT1 [Fig. 4B, lane 3]), but to lower levels than that of the full-length infectious clone (pPAV6 [Fig. 4B, lane 6]). The probe was complementary to the 3' end of the genome and detected the full-length genomic RNA and sgRNA1 at the expected greater lengths (Fig. 4B, 7.4 and 4.7 kb [lane 3]; cf. 5.7 and 3.0 kb [lane 6]). Viral antigen (coat protein) accumulated to half the control levels (Fig. 4B, lane 3 compared with 6). These data indicate that the larger hybrid genome can replicate in oat protoplasts, although not as efficiently as the wild-type genome. This also verifies previous observations that the RT ORF is not required for BYDV PAV RNA replication in protoplasts (20, 40).

Readthrough of the CP stop was detected by assaying for GUS activity. An extract from protoplasts infected with the reporter construct gave 89 GUS units versus 1 unit in mock-infected cells (Fig. 4C; lane 5 compared with lane 3). To ensure that the GUS activity was not due to internal initiation or translation of fragmented RNA, the size of the protein giving GUS activity was determined following separation of infected protoplast proteins on a semidenaturing polyacrylamide gel (Fig. 4C). GUS activity was found at a mobility corresponding to the expected size of the full length CP-GUS fusion protein (95 kDa [Fig. 4C, lane 5]). These data indicate that readthrough occurred in vivo in infected protoplasts.

To determine if the C-rich element was required in vivo, a construct lacking all but two of the CC doublets was tested (pPAVGUSRT2 [Fig. 4A]). It accumulated viral antigen and RNAs to levels similar to that of the construct containing the C-rich element (Fig. 4B, lane 2 compared with lane 3). However, GUS activity was only 9% of the control (Fig. 4C, lane 2

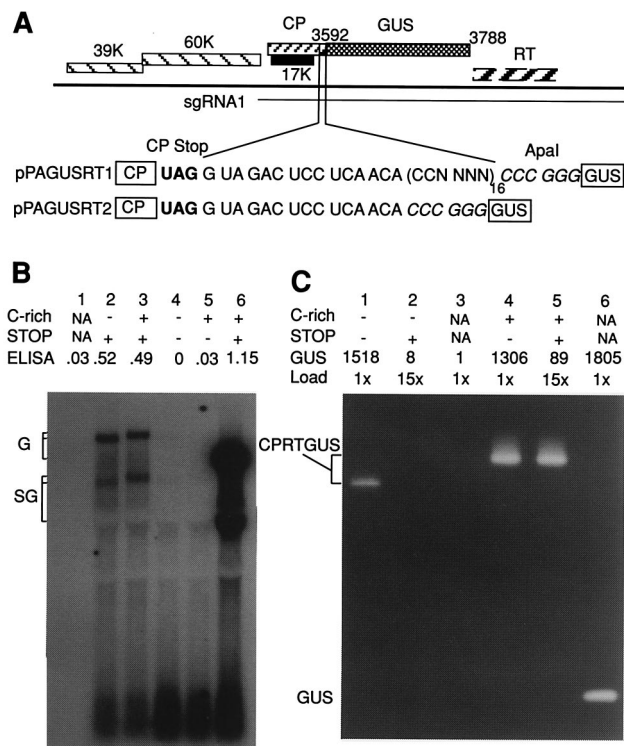


FIG. 4. Replication and GUS expression of full-length infectious reporter constructs in vivo. (A) Schematic diagram of the pPAVGUSRT series of constructs. The genome organization of pPAVGUSRT1 and pPAVGUSRT2 is shown with the region following the CP stop expanded to show sites of GUS ORF insertion. 39K, 60K, ORFs encoding the 39- and 60-kDa (putative RNA polymerase) products. The 3' end of the RT ORF following the GUS ORF is indicated as a dashed box labeled RT. (B) Northern blot hybridization of total RNA from cells transfected with reporter constructs. Cells were electroporated with salmon sperm DNA (lane 1) or with T7 polymerase transcripts from the following *Sma*I linearized plasmids: pPAVGUSRT2 (lane 2), pPAVGUSRT1 (lane 3), pPAVGUSRT4 (lane 4), pPAVGUSRT3 (lane 5), and pPAV6 (lane 6). The presence (+) or absence (-) of the C-rich region in these constructs is indicated as a dashed box labeled RT. (C) Total protein from protoplasts transfected with transcripts from the following plasmids was separated by polyacrylamide gel electrophoresis and stained for GUS activity (7): pPAVGUSRT4 (lane 1), pPAVGUSRT3 (lane 2), salmon sperm DNA (lane 3), pPAVGUSRT2 (lane 4), pPAVGUSRT1 (lane 5), and pAGUS1 (lane 6). The GUS activity (picomoles of MU per minute per milligram of protein) in a lysate of duplicate infected samples is also shown. Fifteen times as much lysate was loaded in lanes 2 and 5 as indicated (15 $\times$ ). The positions of the GUS protein (GUS) and different CP-RT-GUS fusion proteins are indicated on the left. The expected mobilities are 90 kDa in lanes 1 and 2, 95 kDa in lanes 4 and 5, and 73 kDa in lane 6.

compared with lane 5), indicating that the C-rich element was required for readthrough in vivo. The expected size of the product was 90 kDa.

We constructed control in-frame derivatives of these two constructs in which the UAG stop was changed to a GAG sense codon (pPAVGUSRT3 and pPAVGUSRT4). These control viral RNAs should produce no free CP, only a CP-RT-GUS fusion. Surprisingly, these alterations reduced accumulation of viral full-length and subgenomic RNAs dramatically (Fig. 4B, lanes 4 and 5). No intact virions were detected by ELISA (Fig. 4B, lanes 4 and 5), but there was a large amount of GUS activity (Fig. 4C, lanes 1 and 4) which migrated at the expected  $M_r$ , indicating the presence of CP-RT-GUS protein (Fig. 4C, lanes 1 and 4). Because the total GUS activity was lower in constructs with the stop, 15 times as much sample was

loaded to give approximately the same activity in lanes 4 and 5 (Fig. 4C, lanes 2 and 5). Because the GAG constructs did not replicate as well as UAG constructs, we could not calculate the absolute rate of readthrough. However, we can compare relative readthrough activities of constructs with the CP stop codon.

Other deletions and alterations were tested in replicating viral RNAs (Fig. 5). The GUS activities measured in different experiments are shown, with the average value presented as a percentage of the wild-type value in the final column. Deletions of most of the coat protein did not affect readthrough significantly (Fig. 5A). Deletion of the entire C-rich element reduced readthrough, as indicated above (Fig. 4), but the first 10 CCN NNN repeats were sufficient (Fig. 5B). Placing the C-rich region in the +1 frame so that it no longer encoded prolines did not affect the readthrough rate (Fig. 5C), as observed in vitro (Fig. 2).

Deletion of a large section containing the distal element identified in vitro reduced readthrough 10-fold (Fig. 5D, pPAVGUSRT6). Viral antigen accumulated to 76% of wild-type levels (data not shown), indicating that replication and virion formation were occurring. Effects of smaller deletions (Fig. 5D) were similar to those seen in vitro. The greatest reduction was caused by a deletion of nucleotides 4155 to 4515 (to 7%), whereas lesser effects were seen with deletions from 4123 to 4154 (to 61%) and 3788 to 4129 (to 42%). Deletion of both the C-rich and distal elements reduced readthrough 10-fold (8%, Fig. 5E). These results indicate that this distal element detected in vitro is also required in vivo and that key determinants lie between bases 4154 and 4515.

## DISCUSSION

**The C-rich element.** The conserved block around the luteoviral stop codons (CN AAA UAG GUA GAC, Fig. 6) is not well conserved in PEMV (13) and was not sufficient for readthrough in vivo (Fig. 4). A C-rich region following the CP stop codon was required for readthrough. All the luteovirus and PEMV sequences contain multiple CCN NNN repeats located 6 to 15 bases after the CP stop codon, but there is little homology other than the CC doublets (Fig. 6). For BYDV PAV, the sequence containing the first 5 CCN NNN repeats was sufficient, but the sequence containing the next 11 repeats would not functionally replace it. Also, although mutation of the fourth and fifth CC doublets to UU weakened the element, they did not completely abolish its activity. These data may indicate a requirement for optimal spacing from the stop or for sequences between the CC doublets. Because the element functioned when placed in a different reading frame, the signal is in the RNA itself rather than in the proline-rich protein it encodes.

It had been noted that for luteoviruses, at least one of the repeats contained CCCC (38). This sequence is also located at variable distances after the suppressible stops of other plant viruses (38). This was not sufficient for readthrough. Deletion of regions containing the CCCC had no effect on readthrough. Deletion of the natural C-rich region, which fortuitously placed the same sequence (CCCC) within the reporter gene (GUS) at the same distance from the stop codon, resulted in a mutant that had no read-through activity (pPAVGUSRT2 [Fig. 4 and 5]).

**The distal readthrough element.** A region normally located nearly 700 bases 3' of the stop was also required for readthrough. The unexpected distal element is more distant than any readthrough signal hitherto reported and functioned over a wide range of distances. It functioned in vitro when located

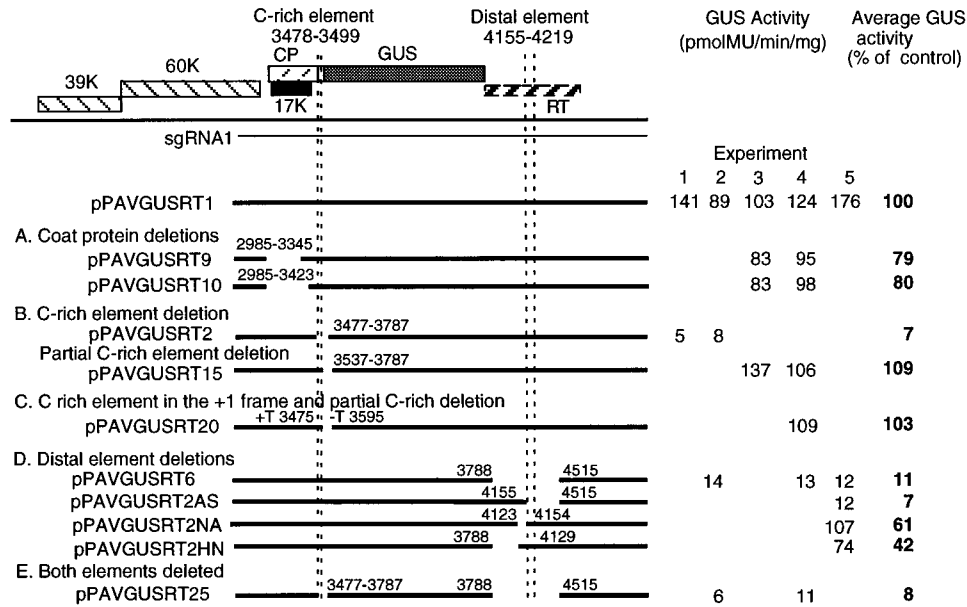


FIG. 5. GUS activities from cells transfected with PAV reporter derivatives. Duplicate aliquots of protoplasts were transfected with full-length, infectious RNAs from the indicated plasmids. The average GUS activity is shown for each experiment. The final column contains means from all the experiments expressed as a percentage of the activity of the control wild-type construct pPAVGUSRT1. Because pPAVGUSRT20 is a derivative of pPAVGUSRT15, the latter plasmid was used as a control for pPAVGUSRT20, to single out the effects of the frame change in pPAVGUSRT20.

0.4 to 0.7 kb after the stop in the 3' untranslated region (3' LTR) of a truncated RT protein (Fig. 3) and in vivo when located in the 3' UTR 2.4 kb from the CP stop codon (Fig. 5). The 5' portion of the distal element (4154 to 4219) is well conserved in all luteoviruses and PEMV (Fig. 7), as is the amino acid sequence (13, 36). It is possible that the distal element interacts with the C-rich element by long-distance base pairing. Downstream secondary structures are needed for other types of readthrough (65). There are two conserved GG doublets in the 5' region of the motif which could potentially base pair with CC doublets in the C-rich region. There is also potential base-pairing between bases 4184 to 4203 in the distal element and 3466 to 3485 in the C-rich element (bases indicated in Fig. 6 and 7). This pairing is conserved in the PAV and

MAV strains of BYDV but not in SGV strains or other luteoviruses.

The CP-RT protein is required for aphid transmissibility (8, 10), so that alterations to the elements identified in this study could reduce the amount of RT product and aphid transmission. For example, deletion of the *HpaI-ScaI* segment (which contains the distal element) from pPAV6 gave no detectable truncated CP-RT protein from Western blot (immunoblot) analysis of infected protoplasts and no aphid transmission (10). We looked for a correlation between the sequence of the homologous elements from seven potato leafroll virus (PLRV) isolates and aphid transmissibility (27) but found none. They differed in one base in the C-rich element (underlined in Fig. 6) and three in the distal element (underlined

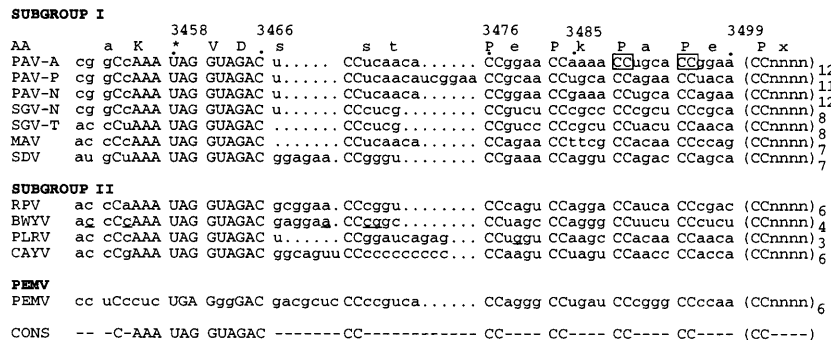


FIG. 6. Nucleotide alignment of sequences flanking the CP stop codon of luteoviruses and PEMV. The positions of key nucleotides mentioned in the text are indicated. The amino acid sequence and base numbering for PAV-A are shown above the nucleotide sequence. Capital letters indicate conserved amino acids in 10 of 12 sequences. The two pairs of C's mutated in the pBHT variant are boxed. PAV-A, BYDV-PAV-Australia (39); PAV-P, BYDV-PAV-Purdue (66); PAV-N, BYDV-PAV-New York (63); SGV-N, BYDV-SGV-New York (31); SGV-T, BYDV-SGV-Texas (31); MAV, BYDV-MAV (61); SDV, soybean dwarf virus (45); RPV, BYDV-RPV-New York (60); BWYV (59); bases that differ in any of 12 other isolates (12) are underlined. For these variants, only the first 23 bases of the RT ORF were available. For PLRV (35), sites of variations in seven other isolates are underlined (27). CAYV, cucurbit aphid-borne yellows luteovirus (21); PEMV (GenBank accession number Z48507) (13); CONS, consensus sequence (bases conserved in 10 of 12 sequences are shown). The beginning (3466) and end (3485) of the PAV sequence that could base-pair with bases 4184-4203 in the distal element are indicated.

```

SUBGROUP I
                4154                4184                4203                4219
AA   Y T n k r M E f G t i s V t c D . E s d v e a e r i r h a e t p i
PAV-A UacACaaAcAag cgaAUGGAaUUC GGuaccAUauCu GUGaCauguGAU GAAuccgauguu gagGcagaacga auaacaaggcac gcugaaacgccc
PAV-P UacACaaAcAaa cggAUGGAaUUu GGuaccAUauCu GUGaCcuGUaU GAAuccgauguu gagGcagaacgc auaacaaggcau gcugaaacaccc
PAV-N UacACaaAcAaa cgaAUGGAaUUu GGuaccAUauCu GUGaCauguGAU GAAuccgauguu gagGcagaacgc auaacaaggcac gcugaaacaccc
SGV-N UacACuAgAag cgaAUGGAaUUu GGuaccAUcuCc GUGaCcauGAU GAAagaaacgc gagGca... aga ucacaguggcac acgcuacaacc
SGV-T UacACcAgAaa cgaAUGGAaUUu GGaaccAUcuCu GUGaCcauGAU GAAagaaacgc gagGca... aga ucacaauggcac acacagcaaac
MAV   UacACagAuAaa agaAUGGAaUUu GGaaccAUauCu GUGaCcuGUaU GAAuccgauguu gagGcagaacgc uauagcaggcac acgagcaaggc
SDV   UggACagAuAga gauAUGGAaUUu GGuauGGUauCg GUGgugcuGAU AGcac... uaa gaaGgugcaagg aguucucaguau guuagaaaaucg

SUBGROUP II
RPV   UggACggAcAga gacAUGGAaUUC GGauG AUuaCa GUcaCguuGAU GAGaaacguggc ucgGguuccccc acucgcaaaagc uua...cgcgcu
BPV   UggACagAucgg augAUGGAaUa GGaugAUcgcC aUagCacuuGAU GAAcaaggcuca uccGguuccgua aagacagaaaga ccaagagaguu
PLRV  UggACggAcCga gacAUGGAaUcG GGcugAUcaCc GUGgugcuGAU GAAca... uuua gaaGgcacuggu ucggcuagcaga gucgcgccggcu
CAYV  UacACggAgAaa uauugcGAgUgg GGggcaAUuuCg GUauCaucGAc GAAgaaauuuu gggaacgagcca agaagaaauacca cgcagggggggu

PEMV
PEMV  UacACcgAgAaa ucgAUuGaaUgg GGgucugUcuCu aUauCaaauGAU GAAgugaauca acgGcgucacgc agucccuggagg ggcagggcucga

CONS  U--AC--A-A-- ---AUGGA--UU- GG----AU--C- GU--C-----GAU GAA-----G-----

```

FIG. 7. Nucleotide alignment of regions homologous to the distal readthrough element. Sequences are labeled as in Fig. 6. Bases that differ within seven isolates of PLRV are underlined (27). The two bases that are different (C's) in the non-aphid-transmissible PEMV variant (13) are also underlined. The beginning (4184) and end (4203) of the PAV sequence that could base pair with bases 3466 to 3485 in the C-rich element are indicated.

in Fig. 7). They also differed in protein sequences in several places in the RT protein, and this may account for the alterations in transmissibility as proposed previously (27). For PEMV, the sequence of a strain that does not read through differs at two positions (13) in the distal-element homolog (underlined in Fig. 7). These changes do not alter amino acid sequence, but might abolish readthrough.

One example is known in which decoding of a stop codon is affected by the sequence located kilobases downstream in the 3' untranslated region (3' UTR) (4). This is the incorporation of selenocysteine (SeCys) at UGA "sense" codons in specific mammalian cellular genes. This might be considered a special case of readthrough (23). A large bulged stem-loop located in the 3' UTR directs SeCys incorporation at UGA codons within mammalian mRNAs (3, 48). This structure is thought to specifically bind a complex of selenocysteinyl-tRNA and a special elongation factor, channeling this complex into the decoding site on the ribosome. This mechanism is specific for SeCys and UGA codons, but an analogous mechanism might be possible for readthrough. However, computer analysis did not detect any large conserved stem-loops in the distal readthrough element of the BYDV-PAV genome.

**Translation in vitro in wheat germ lysates.** We used the wheat germ translation system because wheat is a natural host of the virus and some plant translational control signals function only in plants. The predominance of CP over the 17K protein was unexpected. In reticulocyte lysates, the 17K protein predominates (Fig. 2A) (16). It had been shown previously with DNA reporter constructs in vivo in protoplasts that the 17K protein AUG context gives a higher rate of initiation for both BYDV-PAV (17) and PLRV (55). However, in those in vivo studies, less than 200 bases of the CP and 17K protein initiation codon contexts was fused to reporter genes (17, 55). Our finding of better initiation at the first CP AUG in wheat germ could reflect greater fidelity in this system or could mean that additional wheat germ-specific signals are found further away in the sgRNA.

The low (~1%) readthrough rate in vitro in wheat germ contrasts with the heterologous reticulocyte lysate system in which readthrough occurred at 7 to 15% (Fig. 2) (16). This difference could result from the ribosomes and termination factors used or the tRNAs available. The wheat germ extracts used were supplemented with wheat germ tRNAs, whereas the reticulocyte lysate was supplemented with calf liver tRNAs. However, wheat germ extracts supplemented with calf liver tRNAs also exhibited low readthrough (9, 44). Thus, the difference in readthrough is not likely to result from differences in tRNA populations.

The percent readthrough in phloem cells, in which virus replication occurs, has not been determined accurately. For the subgroup II luteovirus PLRV, the readthrough efficiency was estimated to be 0.9 to 1.3% in vivo in tobacco and potato protoplasts. The DNA reporter constructs contained only 18 bases before and 21 bases after the CP stop (55), so that these constructs lacked both elements we identified in this report. By using DNA reporter constructs similar to those of Tacke et al., <0.2% readthrough of BYDV PAV CP stop codon was detected in oat protoplasts (15).

**Replication of chimeric viruses in protoplasts.** We found that hybrid BYDV PAV viruses containing the GUS gene were able to replicate and accumulate virions in oat protoplasts. Deletion of most of the RT protein does not affect replication (10, 40). Unexpectedly, replication of full-length infectious transcripts containing the GAG sense codons in place of the UAG stop was barely detectable by Northern blot analysis or ELISA. These clones were constructed to make none of the normal CP of the virion, only the CP-RT-GUS fusion. The construct with the C-rich deletion would have only four RT-encoded amino acids and so would be essentially a CP-GUS fusion. Similarly, it was shown recently that a mutant BYDV PAV construct which contained a UAC sense codon in place of the CP stop also did not accumulate viral RNAs or antigen (20). This could be interpreted to indicate that free CP is required for virus replication. However, mutants that do not express CP accumulate significant levels of RNA (40), albeit at a reduced level, unlike the stop to sense codon mutants. Thus, 100% fusion of CP to RT reduced RNA accumulation more than the lack of CP alone. However, some replication must be occurring, because GUS activity was detected.

In vivo and in both in vitro translation systems, both readthrough elements were required, indicating that they are biologically relevant and do not function exclusively in plants. We have identified them as two equally essential elements; however, they may function in concert, possibly linked directly by base pairing or through an additional factor. We are currently identifying more precisely the key features of these elements.

#### ACKNOWLEDGMENTS

C.M.B. is a long-term fellow of the Human Frontier Science Program Organisation. This study was supported by Hatch Act and State of Iowa funds. S.P.D.K. was supported by USDA National Research Initiative grant 91-37303-6424 to W.A.M.

#### REFERENCES

1. Ambion Inc. 1995. *In vitro* translation kits. Ambion, Austin, Tex.
2. Bahner, I., J. Lamb, M. A. Mayo, and R. T. Hay. 1990. Expression of the



- genome of potato leafroll virus: readthrough of the coat protein termination codon *in vivo*. *J. Gen. Virol.* **71**:2251–2256.
3. **Berry, M. J., L. Banu, J. W. Harney, and P. R. Larsen.** 1993. Functional characterization of the eukaryotic SECIS elements which direct selenocysteine insertion at UGA codons. *EMBO J.* **12**:3315–3322.
  4. **Berry, M. J., J. W. Harney, T. Ohama, and D. L. Hatfield.** 1994. Selenocysteine insertion or termination: factors affecting UGA codon fate and complementary anticodon:codon mutations. *Nucleic Acids Res.* **22**:3753–3759.
  5. **Bonetti, B., L. W. Fu, J. Moon, and D. M. Bedwell.** 1995. The efficiency of translation termination is determined by a synergistic interplay between upstream and downstream sequences in *Saccharomyces cerevisiae*. *J. Mol. Biol.* **251**:334–345.
  6. **Boonham, N., C. M. Henry, and K. R. Wood.** 1995. The nucleotide sequence and proposed genome organization of oat chlorotic stunt virus, a new soil-borne virus of cereals. *J. Gen. Virol.* **76**:2025–2034.
  7. **Brault, V., and W. A. Miller.** 1992. Translational frameshifting mediated by a viral sequence in plant cells. *Proc. Natl. Acad. Sci. USA* **89**:2262–2266.
  8. **Brault, V., J. F. J. M. Van den Heuvel, M. Verbeek, G. V. Ziegler, A. Reutenauer, E. Herrbach, J. C. Garaud, H. Guillely, K. Richards, and G. Jonard.** 1995. Aphid transmission of beet western yellows luteovirus requires the minor capsid read-through protein P74. *EMBO J.* **14**:650–659.
  9. **Brown, C. M.** 1995. Unpublished data.
  10. **Chay, C., U. B. Umasinge, S. P. Dinesh-Kumar, W. A. Miller, and S. L. Gray.** 1996. Aphid transmission and systemic spread of barley yellow dwarf virus-PAV are mediated by the coat protein read-through domain and 17K protein, respectively. *Virology* **219**:57–65.
  11. **Cheng, S. L., L. L. Domier, and C. J. D'Arcy.** 1994. Detection of the read-through protein of barley yellow dwarf virus. *Virology* **202**:1003–1006.
  12. **de Miranda, J., M. Stevens, E. de Bruyne, H. Smith, C. Bird, and R. Hull.** 1995. Sequence comparison and classification of beet western yellows isolates. *Ann. Appl. Biol.* **127**:113–124.
  13. **Demler, S. A., and G. A. de Zoeten.** 1991. The nucleotide sequence and luteovirus-like nature of RNA 1 of an aphid non-transmissible strain of pea enation mosaic virus. *J. Gen. Virol.* **72**:1819–1834.
  14. **Di, R., S. P. Dinesh-Kumar, and W. A. Miller.** 1993. Translational frameshifting by barley yellow dwarf virus RNA (PAV serotype) in *Escherichia coli* and in eukaryotic cell-free extracts. *Mol. Plant-Microbe Interact.* **6**:444–452.
  15. **Dinesh-Kumar, S. P.** 1993. Gene expression strategies of barley yellow dwarf virus. Ph.D. thesis. Iowa State University, Ames.
  16. **Dinesh-Kumar, S. P., V. Brault, and W. A. Miller.** 1992. Precise mapping and *in vitro* translation of a trifunctional subgenomic RNA of barley yellow dwarf virus. *Virology* **187**:711–722.
  17. **Dinesh-Kumar, S. P., and W. A. Miller.** 1993. Control of start codon choice on a plant viral RNA encoding overlapping genes. *Plant Cell* **5**:679–692.
  18. **Domier, L. L.** 1995. Genome structure and function of barley yellow dwarf viruses, p. 181–202. *In C. J. D'Arcy and P. A. Burnett (ed.), Barley yellow dwarf—40 years of progress.* APS Press, St. Paul, Minn.
  19. **Feng, Y. X., H. Yuan, A. Rein, and J. G. Levin.** 1992. Bipartite signal for readthrough suppression in murine leukemia virus mRNA—an 8-nucleotide purine-rich sequence immediately downstream of the GAG termination codon followed by an RNA pseudoknot. *J. Virol.* **66**:5127–5132.
  20. **Filichkin, S. A., R. M. Lister, P. F. McGrath, and M. J. Young.** 1994. *In vivo* expression and mutational analysis of the barley yellow dwarf virus read-through gene. *Virology* **205**:290–299.
  21. **Guillely, H., S. C. Wipf, K. Richards, H. Lecoq, and G. Jonard.** 1994. Nucleotide sequence of cucurbit aphid-borne yellows luteovirus. *Virology* **202**:1012–1017.
  22. **Hampton, R., E. Ball, and S. De Boer.** 1990. Serological methods for the detection and identification of viral and bacterial plant pathogens. APS Press, St. Paul, Minn.
  23. **Hatfield, D., and A. Diamond.** 1993. UGA—a split personality in the universal genetic code. *Trends Genet.* **9**:69–70.
  24. **Hatfield, D. L., J. G. Levin, A. Rein, and S. Oroszlan.** 1992. Translational suppression in retroviral gene expression. *Adv. Virus Res.* **41**:193–239.
  25. **Hatfield, D. L., D. W. E. Smith, B. J. Lee, P. J. Worland, and S. Oroszlan.** 1990. Structure and function of suppressor transfer-RNAs in higher eukaryotes. *Crit. Rev. Biochem. Mol. Biol.* **25**:71–96.
  26. **Jefferson, R. A.** 1987. Assaying chimeric genes in plants: the GUS gene fusion system. *Plant Mol. Biol.* **5**:387–405.
  27. **Jolly, C. A., and M. A. Mayo.** 1994. Changes in the amino acid sequence of the coat protein readthrough domain of potato leafroll luteovirus affect the formation of an epitope and aphid transmission. *Virology* **201**:182–185.
  28. **Kelly, L., W. L. Gerlach, and P. M. Waterhouse.** 1994. Characterisation of the subgenomic RNAs of an Australian isolate of barley yellow dwarf luteovirus. *Virology* **202**:565–573.
  29. **Laemmli, U. K.** 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature (London)* **227**:680–685.
  30. **Landt, O., H. P. Grunert, and U. Hahn.** 1990. A general method for rapid site-directed mutagenesis using the polymerase chain reaction. *Gene* **96**:125–128.
  31. **Lei, C.-H., R. M. Lister, J. R. Vincent, and M. N. Karanjkar.** 1995. SGV serotype isolates of barley yellow dwarf virus differing in vectors and molecular relationships. *Phytopathology* **85**:820–826.
  32. **Li, G. P., and C. M. Rice.** 1989. Mutagenesis of the in-frame opal termination codon preceding nsP4 of Sindbis virus: studies of translational readthrough and its effect on virus replication. *J. Virol.* **63**:1326–1337.
  33. **Li, G. P., and C. M. Rice.** 1993. The signal for translational readthrough of a UGA codon in Sindbis virus RNA involves a single cytidine residue immediately downstream of the termination codon. *J. Virol.* **67**:5062–5067.
  34. **Martin, R. R., and C. J. D'Arcy.** 1995. Taxonomy of barley yellow dwarf viruses, p. 203–214. *In C. J. D'Arcy and P. A. Burnett (ed.), Barley yellow dwarf—40 years of progress.* APS Press, St. Paul, Minn.
  35. **Mayo, M. A., D. J. Robinson, C. A. Jolly, and L. Hyman.** 1989. Nucleotide sequence of potato leafroll luteovirus RNA. *J. Gen. Virol.* **70**:1037–1051.
  36. **Mayo, M. A., and V. Ziegler-Graf.** 1996. Molecular biology of luteoviruses. *Adv. Virus Res.* **46**:413–460.
  37. **McCaughan, K. K., C. M. Brown, M. E. Dalphin, M. J. Berry, and W. P. Tate.** 1995. Translational termination efficiency in mammals is influenced by the base following the stop codon. *Proc. Natl. Acad. Sci. USA* **92**:5431–5435.
  38. **Miller, W. A., S. P. Dinesh-Kumar, and C. P. Paul.** 1995. Luteovirus gene expression. *Crit. Rev. Plant Sci.* **14**:179–211.
  39. **Miller, W. A., P. M. Waterhouse, and W. L. Gerlach.** 1988. Sequence and organisation of barley yellow dwarf virus genomic RNA. *Nucleic Acids Res.* **16**:6097–6111.
  40. **Mohan, B. R., S. P. Dinesh-Kumar, and W. A. Miller.** 1995. Genes and *cis*-acting sequences involved in replication of barley yellow dwarf virus-PAV RNA. *Virology* **212**:186–195.
  41. **Pelham, H. R. B.** 1978. Leaky UAG termination codon in TMV RNA. *Nature (London)* **272**:469–471.
  42. **Poole, E. S., C. M. Brown, and W. P. Tate.** 1995. The identity of the base following the stop codon determines the efficiency of *in vivo* translational termination in *Escherichia coli*. *EMBO J.* **14**:151–158.
  43. **Promega Corp.** 1992. Large scale RNA synthesis. Promega Notes **39**:12–18.
  44. **Promega Corp.** 1992. Wheat germ extract. Promega, Madison, Wis.
  45. **Rathjen, J. P., L. E. Karageorgos, N. Habili, P. M. Waterhouse, and R. H. Symons.** 1994. Soybean dwarf luteovirus contains the third variant genome type in the luteovirus group. *Virology* **198**:571–579.
  46. **Rizzo, T. M., and S. M. Gray.** 1992. Localization of a surface domain of the capsid protein of barley yellow dwarf virus. *Virology* **186**:300–302.
  47. **Sambrook, J., E. F. Fritsch, and T. Maniatis.** 1989. Molecular cloning: a laboratory manual, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
  48. **Shen, Q., J. L. Leonard, and P. E. Newburger.** 1995. Structure and function of the selenium translation element in the 3'-untranslated region of human cellular glutathione peroxidase mRNA. *RNA* **1**:519–525.
  49. **Skuzeski, J. M., L. M. Nichols, and R. F. Gesteland.** 1990. Analysis of leaky viral translation termination codons *in vivo* by transient expression of improved beta-glucuronidase vectors. *Plant Mol. Biol.* **15**:65–79.
  50. **Skuzeski, J. M., L. M. Nichols, R. F. Gesteland, and J. F. Atkins.** 1991. The signal for a leaky UAG stop codon in several plant viruses includes the 2 downstream codons. *J. Mol. Biol.* **218**:365–373.
  51. **Stahl, G., L. Bidou, J. Rousset, and M. Cassan.** 1995. Versatile vectors to study recoding: conservation of rules between yeast and mammalian cells. *Nucleic Acids Res.* **23**:1557–1560.
  52. **Stansfield, I., K. M. Jones, V. V. Kushnirov, A. R. Dagkesamanskaya, A. I. Poznyakovski, S. V. Pauskin, C. R. Nierras, B. S. Cox, M. D. Ter-Avanesyan, and M. F. Tuite.** 1995. The products of the SUP45 (eRF1) and SUP35 genes interact to mediate termination in *Saccharomyces cerevisiae*. *EMBO J.* **14**:4365–4373.
  53. **Stansfield, I., and M. F. Tuite.** 1995. The end in sight: terminating translation in eukaryotes. *Trends Biochem. Sci.* **20**:489–491.
  54. **Strauss, J. H., and E. G. Strauss.** 1994. The alphaviruses: gene expression, replication, and evolution. *Microbiol. Rev.* **58**:491–562.
  55. **Tacke, E., D. Prufer, F. Salamini, and W. Rohde.** 1990. Characterization of a potato leafroll luteovirus subgenomic RNA: differential expression by internal translation initiation and UAG suppression. *J. Gen. Virol.* **71**:2265–2272.
  56. **Tate, W. P., and C. M. Brown.** 1992. Translational termination: “stop” for protein synthesis or “pause” for regulation of gene expression. *Biochemistry* **31**:2443–2450.
  57. **Urban, C., and H. Beier.** 1995. Cysteine tRNAs of plant origin as novel UGA suppressors. *Nucleic Acids Res.* **23**:4591–4597.
  58. **Valle, R. P. C., G. Drugeon, M. D. Devignesmorch, A. B. Legocki, and A. L. Haenni.** 1992. Codon context effect in virus translational readthrough—a study *in vitro* of the determinants of TMV and Mo-MuLV amber suppression. *FEBS Lett.* **306**:133–139.
  59. **Veidt, I., H. Lot, M. Leiser, D. Scheidecker, H. Guillely, K. Richards, and G. Jonard.** 1988. Nucleotide sequence of beet western yellows virus RNA. *Nucleic Acids Res.* **16**:9917–9932.
  60. **Vincent, J. R., R. M. Lister, and B. A. Larkins.** 1991. Nucleotide sequence analysis and genomic organization of the NY-RPV isolate of barley yellow dwarf virus. *J. Gen. Virol.* **72**:2347–2355.
  61. **Vincent, J. R., P. P. Ueng, R. M. Lister, and B. A. Larkins.** 1990. Nucleotide sequences of coat protein genes for three isolates of barley yellow dwarf virus



- and their relationships to other luteovirus coat protein sequences. *J. Gen. Virol.* **71**:2791–2799.
62. **Wadsworth, G. J., M. G. Redinbaugh, and J. G. Scandalios.** 1988. A procedure for the small-scale isolation of plant RNA suitable for RNA blot analysis. *Anal. Biochem.* **172**:279–283.
63. **Wang, J. Y., C. Chay, F. E. Gildow, and S. M. Gray.** 1995. Readthrough protein associated with virions of barley yellow dwarf luteovirus and its potential role in regulating the efficiency of aphid transmission. *Virology* **206**:954–962.
64. **Wang, S., and W. A. Miller.** 1995. A sequence located 4.5 to 5 kilobases from the 5' end of the barley yellow dwarf virus (PAV) genome strongly stimulates translation of uncapped mRNA. *J. Biol. Chem.* **270**:13446–13452.
65. **Wills, N. M., R. F. Gesteland, and J. F. Atkins.** 1994. Pseudoknot-dependent read-through of retroviral GAG termination codons: importance of sequences in the spacer and loop 2. *EMBO J.* **13**:4137–4144.
66. **Young, M. J., L. Kelly, P. J. Larkin, P. M. Waterhouse, and W. L. Gerlach.** 1991. Infectious in vitro transcripts from a cloned cDNA of barley yellow dwarf virus. *Virology* **180**:372–379.
67. **Zaccomer, B., A. L. Haenni, and G. Macaya.** 1995. The remarkable variety of plant RNA virus genomes. *J. Gen. Virol.* **76**:231–247.
68. **Zerfass, K., and H. Beier.** 1992. Pseudouridine in the anticodon GPsiA of plant cytoplasmic transfer RNA (Tyr) is required for UAG and UAA suppression in the TMV-specific context. *Nucleic Acids Res.* **20**:5911–5918.
69. **Zhouravleva, G., L. Frolova, X. Le Goff, R. Le Guellec, S. Inge-Vechtomov, L. Kisselev, and M. Philippe.** 1995. Termination of translation in eukaryotes is governed by two interacting polypeptide chain release factors, eRF1 and eRF3. *EMBO J.* **14**:4065–4072.