



SPECIAL REPORT

Cloned and transfected P2Y₄ receptors: characterization of a suramin and PPADS-insensitive response to UTPSteven J. Charlton, Colin A. Brown, *Gary A. Weisman, †John T. Turner, *Laurie Erb & ¹Michael R. Boarder

Department of Cell Physiology and Pharmacology, University of Leicester, University Road, Leicester LE1 9HN and Departments of *Biochemistry and †Pharmacology, University of Missouri-Columbia, Columbia, MO 65212 U.S.A.

The P2Y family of receptors are G protein-coupled receptors for ATP, ADP, UTP and UDP. Recently several members of this family have been cloned, including the P2Y₄, which is activated by UTP but not by ATP. In the present report, using receptors stably transfected into 1321N1 cells, we show that suramin acts as an antagonist at cloned P2Y₁ and (less potently) P2Y₂ receptors, but not at the cloned P2Y₄ receptor. Furthermore, PPADS (pyridoxal-phosphate-6-azophenyl-2',4'-disulphonic acid), a potent antagonist at the P2Y₁ receptor, is a relatively ineffective antagonist at the cloned P2Y₄ receptor. This work moves us closer to the goal of classifying the native P2Y receptors on the basis of agonist and antagonist profiles.

Keywords: P2Y receptors; P_{2Y}-purinoceptors; purinoceptors; pyrimidinoceptors

Introduction Recently a number of G protein-coupled P2 receptors (receptors for adenine and uridine nucleotides) have been cloned, including P2Y₁ (also called P_{2Y}) (e.g. Filtz *et al.*, 1994); P2Y₂ (P_{2U}) (e.g. Parr *et al.*, 1994); P2Y₃ (preferentially activated by UDP) (Webb *et al.*, 1996a); P2Y₄ (activated by UTP but not ATP) (e.g. Nguyen *et al.*, 1995); P2Y₅ (Webb *et al.*, 1996b); and P2Y₆ (UDP but not ATP) (Communi *et al.*, 1996). One of the current challenges in the P2 receptor field is to relate the cloned receptors to the diverse native responses using the relative potencies of various agonists and antagonists. Here we show that when compared with transfected P2Y₁ and P2Y₂ receptors, the transfected P2Y₄ receptor has a unique profile with respect to antagonist activity. This provides the potential to relate further native and cloned receptors, and shows that previously described suramin-insensitive responses to UTP may be due to action at P2Y₄ receptors.

Methods Procedures were essentially as described in Charlton *et al.* (1996). Lines of 1321N1 cells stably transfected with turkey P2Y₁ receptors (t-P2Y₁-1321N1 cells), human P2Y₂ receptors (h-P2Y₂-1321N1 cells) or human P2Y₄ receptors (h-P2Y₄-1321N1 cells) have been described previously (Parr *et al.*, 1994; Filtz *et al.*, 1994; Nguyen *et al.*, 1995; Charlton *et al.*, 1996). Cells were labelled for 24 h with [³H]-inositol (1 µCi ml⁻¹, 0.5 ml per well) in medium M199 and stimulated by addition of agonists to this medium. When included, antagonists were present as described previously, except that in some cases the preincubation period was for 1 h instead of 10 min. No difference in results was seen between these two procedures. Total [³H]-inositol (poly)phosphates ([³H]InsP_x) were extracted into 0.5 M trichloroacetic acid, and after ether washes to remove the acid, were purified on small Dowex-1 (Cl⁻) columns. The details of the experimental procedures and data analysis were as in Charlton *et al.* (1996); curve fitting and parameter derivation were by Graph-Pad Prism.

Results Figure 1 shows the concentration-response curves for stimulation of [³H]-InsP_x by UTP, UDP, ATP, ADP and 2MeSATP acting at the h-P2Y₄-1321N1 cells. The potency of UTP (log EC₅₀ = -5.94 ± 0.07; EC₅₀ = 1.17 µM) was considerably greater than that for the other agonists. UDP was

significantly less potent than UTP, and ATP did not reach a maximal response with the highest concentration used.

Figure 2 (a–c) compares suramin as an antagonist at P2Y₁, P2Y₂ and P2Y₄ receptors. Concentration-response curves for stimulation of the t-P2Y₁-1321N1 cells by 2MeSATP (Figure 2a) and h-P2Y₂-1321N1 cells by UTP (Figure 2b) generated Schild plots with slopes of 0.86 ± 0.04 and 0.66 ± 0.05, and pA₂ values of 5.75 ± 0.46 and 4.26 ± 1.09 respectively. UTP concentration-response curves for stimulation of h-P2Y₄-1321N1 cells in the presence of increasing concentrations of suramin are shown in Figure 2c. There was no effect of suramin on maximum responses, and pooled across 3 separate experiments the log EC₅₀ values for the UTP response were -6.21 ± 0.13, -6.70 ± 0.38, -6.63 ± 0.16, and -6.58 ± 0.06 (0, 30, 100 and 300 µM suramin respectively). There were no significant effects of suramin, as determined by analysis of variance and Dunnett's multiple range tests. The tendency of suramin to shift curves slightly to the left may be due to inhibition of UTP breakdown in the absence of any antagonism at the receptor. We have shown with these cells a very small effect of agonist breakdown by ectonucleotidases on the concentration effect curves (Charlton *et al.*, 1996). The main conclusion from this series of experiments is that suramin is not an antagonist at the transfected h-P2Y₄ receptor.

Figure 2 (d–e) examines the effect of PPADS on the h-P2Y₄ receptor. Pooled across 3 separate experiments the log EC₅₀ values for UTP with h-P2Y₄-1321N1 cells were 6.15 ± 0.005 in the absence of PPADS and 6.82 ± 0.17 in the presence of 30 µM PPADS (*P* < 0.05; Student's paired *t* test), showing that PPADS produced a slight shift of the curve to the left (Figure 2d). There was also a small effect on the maximal response (to 100 µM UTP), which was reduced to 82.5% by 30 µM PPADS (*P* < 0.025; Student's *t* test). This is consistent with the effect of increasing concentrations of PPADS when the h-P2Y₄-1321N1 cells were stimulated with 10 µM UTP (Figure 2e).

Discussion The nature of the receptor transfected into the h-P2Y₄-1321N1 cells was confirmed by data in Figure 1 showing a rank order of agonist potency of UTP > UDP > ATP, consistent with the results of Nguyen *et al.* (1995). This unique rank order of agonist potencies confirms this receptor as a pyrimidinoceptor. Thus, responses to UTP in native systems may be at receptors for both ATP and UTP, such as the P2Y₂ receptors, or alternatively at receptors for UTP but not ATP, such as P2Y₄. We have shown in this report that these two P2Y receptor subtypes may be distinguished from

¹ Author for correspondence at: Department of Cell Physiology and Pharmacology, P.O. Box 138, Medical Sciences Building, University Road, Leicester LE1 9HN.

each other, and from P2Y₁, by the antagonist profile of suramin and PPADS.

The PPADS data presented here extends earlier reports that the P2Y₁ receptor was sensitive to antagonism by PPADS with a pA₂ of 6, while the P2Y₂ receptor was unaffected by the

presence of 30 μM PPADS (Brown *et al.*, 1995; Charlton *et al.*, 1996; Ralevic & Burnstock, 1996). Here we show that PPADS has a modest influence on the cloned and transfected P2Y₄ receptor, producing both a small shift of the concentration-response curve to the left and a small reduction in the maximum response. The reduction in EC₅₀ indicates that the compound lacks any competitive antagonist action, and may be due to a residual effect on attenuating agonist breakdown, as explored in Charlton *et al.* (1996). These results show that PPADS can distinguish responses at the P2Y₄ receptor from those at P2Y₁ but not those at P2Y₂ receptors.

We show here that suramin acts as an antagonist at P2Y₁ and P2Y₂ receptors with pA₂ values of 5.7 and 4.3, respectively, confirming the differential potency of suramin at these two receptors reported by Charlton *et al.* (1996). In this report we also show that suramin is not an antagonist at the P2Y₄ receptor. This distinguishes the P2Y₄ receptor from the recently cloned receptor for UTP (P2Y₃), where suramin had an estimated pA₂ value of 5 (Webb *et al.*, 1996a), and from the possibly related P2Y₆ receptor, which has also been reported to show some suramin sensitivity (Chang *et al.*, 1995). Thus, the characterization of the cloned receptors so far indicates that where ATP and UTP are both effective agonists and suramin is a low potency antagonist, this is likely to be the P2Y₂ receptor. When UTP/UDP but not ATP are effective agonists a suramin-insensitive response is likely to be P2Y₄, but a suramin-sensitive response may be P2Y₃, or P2Y₆. The P2Y₄ and P2Y₃/P2Y₆ receptors may also be distinguished by the relative potency of UTP and UDP.

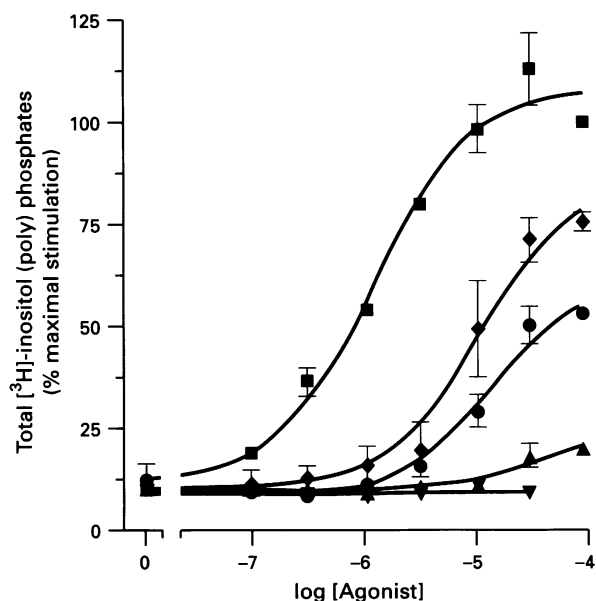


Figure 1 Concentration-response curves for various agonists at the transfected P2Y₄ receptor: UTP (■), UDP (◆), ATP (●), ADP (▲), and 2MeSATP (▼). Data are mean ± s.e. mean from 3 separate experiments each in triplicate.

We thank the Wellcome Trust and the Medical Research Council for financial support.

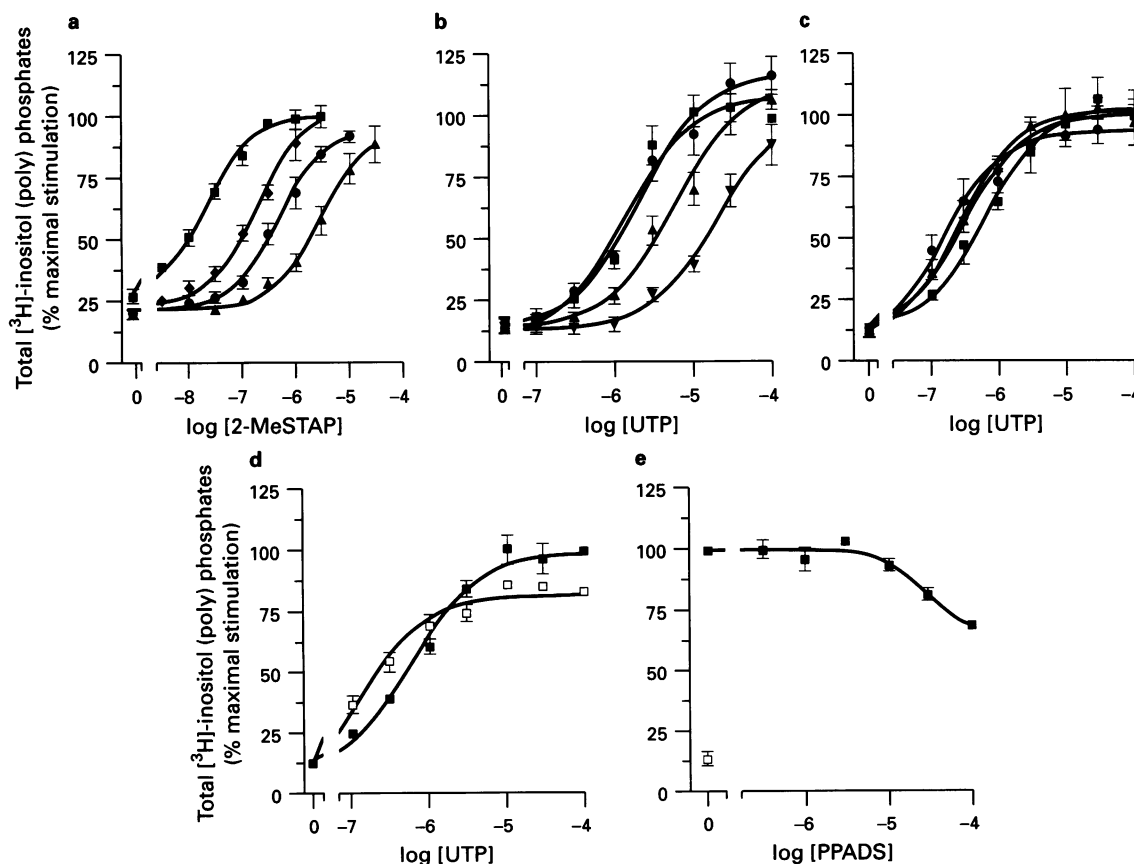


Figure 2 Antagonism by suramin and PPADS at transfected P2Y purinoceptors. (a–c) Show concentration-response curves for 2-MeSATP at P2Y₁ (a), and UTP at P2Y₂ (b) and P2Y₄ (c) receptors with no antagonist (■) or in the presence of suramin at 10 (◆), 30 (●), 100 (▲) or 300 (▼) μM. (d) Concentration-response curves to UTP at the P2Y₄ receptor in the presence (□) or absence (■) of 30 μM PPADS; (e) response to 10 μM UTP in the presence of increasing concentrations of PPADS (■); data from no UTP and no PPADS control are also shown (□). In each case data are pooled from 3 or 4 separate experiments (mean ± s.e. mean) each in triplicate.

References

- BROWN, C., TANNA, B. & BOARDER, M.R. (1995). PPADS: an antagonist at endothelial P_{2Y}-purinoceptors but not P_{2U}-purinoceptors. *Br. J. Pharmacol.*, **116**, 2413–2416.
- CHANG, K., HANAOKA, K., KUMADA, M. & TAKUWA, Y. (1995). Molecular cloning and functional analysis of a novel P₂ nucleotide receptor. *J. Biol. Chem.*, **270**, 26152–26158.
- CHARLTON, S.J., BROWN, C.A., WEISMAN, G.A., TURNER, J.T., ERB, L. & BOARDER, M.R. (1996). PPADS and suramin as antagonists at cloned P_{2Y}- and P_{2U}-purinoceptors. *Br. J. Pharmacol.*, **118**, 704–710.
- COMMUNI, D., PARMENTIER, M. & BOEYNAEMS, J.M. (1996). Cloning, functional expression and tissue distribution of the human P_{2Y6} receptor. *Biochem. Biophys. Res. Commun.*, **222**, 303–310.
- FILTZ, T.A., LI, Q., BOYER, J.L., NICHOLAS, R.A. & HARDEN, T.K. (1994). Expression of a cloned P_{2Y}-purinergic receptor that couples to phospholipase C. *Mol. Pharmacol.*, **48**, 8–14.
- NGUYEN, T., ERB, L., WEISMAN, G.A., MARCHESE, A., HENG, H.H.Q., GARRAD, R.C., GEORGE, S.R., TURNER, J.T. & O'DOWD, B.F. (1995). Cloning, expression, and chromosomal localisation of the human uridine nucleotide receptor gene. *J. Biol. Chem.*, **270**, 30845–30848.
- PARR, C.E., SULLIVAN, D.M., PARADISO, A.M., LAZAROWSKI, E.R., BURCH, L.H., OLSEN, J.C., ERB, L., WEISMAN, G.A., BOUCHER, R.C. & TURNER, J.T. (1994). Cloning and expression of a human P_{2U} nucleotide receptor, a target for cystic fibrosis pharmacotherapy. *Proc. Natl. Acad. Sci. U.S.A.*, **91**, 3275–3279.
- RALEVIC, V. & BURNSTOCK, G. (1996). Discrimination by PPADS between endothelial P_{2Y}- and P_{2U}-purinoceptors in the rat isolated mesenteric arterial bed. *Br. J. Pharmacol.*, **118**, 428–434.
- WEBB, T.E., HENDERSON, D., KING, B.F., WANG, S., SIMON, J., BATESON, A.N., BURNSTOCK, G. & BARNARD, E.A. (1996a). A novel G protein-coupled P₂ purinoceptor (P_{2Y3}) activated preferentially by nucleoside diphosphates. *Mol. Pharmacol.*, (in press).
- WEBB, T.E., SUNDICK, R. & BARNARD, E.A. (1996b). Identification of 6HI as a P_{2Y} purinoceptor P_{2Y5}. *Biochem. Biophys. Res. Commun.*, **219**, 105–110.

(Received July 29, 1996)

Accepted August 27, 1996)