

Different responsiveness of prostaglandin D₂-sensitive systems to prostaglandin D₂ and its analogues

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1 Prostaglandin D₂ (PGD₂) and six PGD₂ analogues were used to classify responsiveness of several PGD₂-sensitive systems. The analogues used were 9β-PGD₂, 5-(6-carboxyhexyl)-1-(3-cyclohexyl-3-hydroxypropyl)hydantoin (BW245C), 17-phenyl-18,19,20-trinor-PGD₂ (17-phenyl-PGD₂), PGD₂ amide, PGD₂ *N*-monomethylamide and 11-keto-15α-hydroxy-Δ^{5,9,12}-prostenoic acid (9-deoxy-Δ^{9,12}-PGD₂). The PGD₂-sensitive systems examined were human platelets, rat peritoneal mast cells, rabbit transverse stomach strip, guinea-pig tracheal ring chain and helical strip of the dog cerebral artery.

2 PGD₂, 9β-PGD₂ and BW245C inhibited the aggregation of human platelets, increased adenosine 3':5'-cyclic monophosphate (cyclic AMP) in rat mast cells and relaxed the rabbit stomach strip. The rank order of potency was BW245C > PGD₂ > 9β-PGD₂. PGD₂ amide and PGD₂ *N*-monomethylamide were inactive in the former two systems but elicited relaxant activity on the rabbit stomach strip. 17-Phenyl-PGD₂ was virtually inactive in the above three systems.

3 PGD₂ and 17-phenyl-PGD₂ contracted the guinea-pig tracheal ring chain and the helical strip of dog cerebral arteries with almost equal potency. 9β-PGD₂ and BW245C antagonized competitively the contractile action of PGD₂. PGD₂ amide and PGD₂ *N*-monomethylamide showed weak agonistic actions in the tracheal preparation.

4 9-Deoxy-Δ^{9,12}-PGD₂, showing stronger growth inhibition than PGD₂ on cultured tumour cells, was inactive in human platelets, rat mast cells and guinea-pig trachea, and elicited contractile response in the rabbit stomach strip.

5 These results indicate the presence of three groups of PGD₂-sensitive systems that respond differently to PGD₂ and its analogues.

Introduction

Prostaglandin D₂ (PGD₂) is one of the prostaglandin which is ubiquitously formed and present in mammalian tissues (Nugteren & Hazelhof, 1973; Ikai *et al.*, 1984). Various actions of this prostaglandin have been reported, which include the anti-aggregatory activity on platelets of several species (Whittle *et al.*, 1978), the contraction of airway smooth muscle (Hamberg *et al.*, 1975; Schneider & Drazen, 1980), inflammatory effects (Flower *et al.*, 1976; Soter *et al.*, 1983), and the action on mast cells (Holgate *et al.*, 1980). PGD₂ also induces either contractions or relaxations in several vascular and gastrointestinal smooth muscle preparations (Horton & Jones, 1974; Whittle *et al.*, 1979; Toda, 1982a). Recently CNS effects of this prostaglan-

din such as hypothermia and induction of sleep have been reported in rodents (Ueno *et al.*, 1982a, b). In addition, inhibition of tumour cell proliferation by PGD₂ and its dehydration derivatives, 9-deoxy-Δ⁹-PGD₂ and 9-deoxy-Δ^{9,12}-PGD₂ has also been described (Fukushima *et al.*, 1982a,b; Kikawa *et al.*, 1984). However, whether these actions are mediated via a single and identical PGD₂ receptor or several PGD₂ receptors exist for each action has remained unclarified. It is also likely that PGD₂ acts on other prostaglandin or thromboxane receptors to elicit responses in some of the systems. A specific PGD₂ receptor was identified on human platelets by two groups using the radioligand receptor assay (Schafer *et al.*, 1979; Siegl *et al.*, 1979), and PGD₂ blocks aggregation of platelets possibly by stimulating adenylate

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cyclase via this receptor (Mills & Macfarlane, 1974; Feinstein *et al.*, 1983). In other tissues, the presence of PGD₂-specific receptor(s) has been suggested by the studies comparing its potency with those of other naturally occurring prostaglandins; these include arterial vascular bed of sheep (Jones, 1978), mast cells (Holgate *et al.*, 1980) and rabbit stomach (Whittle *et al.*, 1979). Recently, 5-(6-carboxyhexyl)-1-(3-cyclohexyl-3-hydroxypropyl)hydantoin (BW245C) has been identified as an agonist for the PGD₂ receptor on blood platelets (Town *et al.*, 1983). In addition, a number of PGD₂ analogues such as 9 β -PGD₂, 17-phenyl-PGD₂ and 9-deoxy- $\Delta^{9,12}$ -PGD₂ have been synthesized chemically (Bundy *et al.*, 1983; Kikawa *et al.*, 1984).

The present study was undertaken to examine responses of several PGD₂-sensitive tissues and cells to these synthetic analogues and to classify PGD₂-sensitive systems into groups on the basis of their responsiveness. The results are discussed in relation to receptors involved and underlying mechanisms in these systems.

Methods

Experiments on isolated tissue preparations

The rabbit transverse stomach strip was prepared as described by Whittle *et al.* (1979). Japanese albino rabbits were killed by exsanguination, and a transverse segment (approximately 2 cm \times 1 cm) of the middle region of the greater curvature was excised from the stomach. After washing, the gastric mucosa was peeled off and the remaining muscle layer was cut to form a strip (approximately 0.3 cm \times 8 cm). The strip was suspended in a 20 ml organ bath containing Krebs-Henseleit solution (NaCl 118, KCl 4.7, MgSO₄ 1.2, KH₂PO₄ 1.2, CaCl₂ 2.5, NaHCO₃ 25, glucose 11 mmol l⁻¹). A combination of antagonists (atropine sulphate, 1.4 \times 10⁻⁷ M; pyrilamine maleate, 2.5 \times 10⁻⁷ M; methysergide maleate, 1 \times 10⁻⁶ M; phentolamine mesylate, 2.6 \times 10⁻⁷ M; propranolol hydrochloride, 6.8 \times 10⁻⁶ M) were added to the bathing solution, which was gassed with 95% O₂:5% CO₂ at 37°C. Tension changes were recorded with an isometric transducer T7-8 (Toyo Baldwin, Tokyo, Japan) coupled to an amplifier type 7236 (San-ei Instrument, Tokyo). The strip was allowed to equilibrate for 60 to 90 min, during which period the bathing solution was replaced every 20 min. Relaxation responses to papaverine hydrochloride, 10⁻⁵ M, were first obtained. After washing the preparations several times and a further 40 min equilibration period, a cumulative concentration-effect curve was obtained for each PGD₂ analogue by increasing the concentration of a compound. At the end of each

experiment, papaverine 10⁻⁵ M was added, to attain the standard relaxation. Then, the strip was washed several times and allowed to equilibrate for 30 to 40 min until the initial tension was restored. Relaxations induced by PGD₂ and PGD₂ analogues were expressed as relative values to those induced by papaverine.

The guinea-pig tracheal ring chain was prepared as follows. A guinea-pig weighing about 350 g was killed by cervical dislocation, the trachea dissected out and cut transversely between the segments of cartilage. A chain was suspended in a 20 ml organ bath containing the Krebs solution and a mixture of antagonists described above. Indomethacin, when included, was added to the bathing solution at a final concentration of 3 \times 10⁻⁶ M. After washing the preparation in the bathing solution several times and equilibration for 60 to 90 min, standard contractions to 30 mM KCl were first obtained and a cumulative concentration-effect curve was obtained for each compound. When antagonistic actions of BW245C and 9 β -PGD₂ were tested, the preparations were incubated for 15 min with the test drugs and then a cumulative concentration-effect curve for PGD₂ was obtained. Contractions induced by PGD₂ and PGD₂ analogues were expressed as relative values to those induced by 30 mM KCl.

Experiments on helical strips of the dog cerebral (basilar and middle cerebral) artery were carried out by the procedures previously described (Toda, 1982a). The strips were fixed vertically in an organ bath containing the modified Ringer-Locke solution, which was maintained at 37 \pm 0.3°C and gassed with a mixture of 95% O₂:5% CO₂. The resting tension was adjusted to 1.5 g and isometric contractions were recorded via a force-displacement transducer (Nihon Koden Kogyo Co., Tokyo, Japan).

Measurement of adenosine 3':5'-cyclic monophosphate (cyclic AMP) content in rat peritoneal mast cells and rabbit stomach strip

A Wistar rat weighing 250 to 350 g was killed by decapitation. After exsanguination, the skin over the abdomen was peeled off and 20 to 25 ml of calcium-free Krebs solution containing 0.1% gelatin and heparin, 0.3 mg ml⁻¹, was injected into the peritoneal cavity. The abdomen was massaged for 1 min and the peritoneal lavage was drained out; cell pellets were resuspended in calcium-free Dulbecco's phosphate-buffered saline (PBS) (Dulbecco & Vogt, 1954) in a concentration of 5 \times 10⁷ cells ml⁻¹. The cell suspension was mixed with the same volume of 40% Ficoll 400 in PBS, and 2 ml of the mixture was layered on 4 ml of 30% Ficoll-PBS solution. After centrifugation at 700 g for 25 min at 4°C, mast cells were recovered at the bottom of the 30% Ficoll layer. The cells were

washed once with PBS and finally suspended in Krebs solution oxygenated with 95% O₂:5% CO₂ at a concentration of 3×10^6 cells ml⁻¹. The yield of mast cells was about 5×10^5 cells per rat. Cyclic AMP accumulation in mast cells induced by PGD₂ and PGD₂ analogues was examined at 37°C in the oxygenated Krebs solution. The incubation mixture containing 1.5×10^5 mast cells and 1 mM 3-isobutyl-1-methylxanthine (IBMX) in 225 μl of Krebs was preincubated at 37°C for 15 min. Reaction was started by the addition of a compound in 25 μl of Krebs and carried out for 1 min. After the reaction was terminated by the addition of 50 μl of 50% trichloroacetic acid (TCA), the mixture was frozen and thawed, and centrifuged at 700 g for 5 min. The supernatant fluid was washed three times with 2 ml of water-saturated diethyl ether, and cyclic AMP in the aqueous solution was measured by radioimmunoassay as described by Honma *et al.* (1977).

Content of cyclic AMP in the rabbit stomach strip was determined as follows. Strips were suspended and incubated in an organ bath as described above. After the endogenous tension had stabilized, PGD₂ was added to the test tissues at a final concentration of 1.4 μM, and vehicle was added to the controls. After incubation for 3 min, the tissues were immediately frozen in liquid nitrogen. The frozen tissues were weighed, cut into small pieces and homogenized in 6% TCA by Polytron. Cyclic AMP content in the TCA extracts was measured as described. When the effect of IBMX was examined, strips were first incubated with 100 μM IBMX for 3 min and then subjected to PGD₂ treatment.

Aggregometry of human platelet-rich plasma

Blood was collected by venipuncture from healthy volunteers who had taken no drugs for at least two weeks, and immediately mixed with one tenth volume of 3.8% trisodium citrate. Platelet-rich plasma (PRP) was recovered by centrifugation of citrated blood at 180 g for 15 min. PRP, 200 μl, was placed and stirred at 37°C in an aggregometer Hema Tracer 1 Model PAT-2M (Niko Bioscience, Tokyo). Platelet aggregation was initiated by the addition of 5 μl of ADP (200 μM). Compounds were dissolved in 50 mM Tris-HCl, pH 7.4, and added 1 min before the addition of ADP. Aggregation was followed as change in light transmission according to the method of Born (1962).

Drugs

PGD₂, 9β-PGD₂, PGD₂ amide, PGD₂ N-monomethylamide, 9-deoxy-Δ^{9,12}-PGD₂, PGE₂ and PGF_{2α} were kindly provided by Ono Pharmaceutical Co., Osaka, Japan. 17-Phenyl-PGD₂ and BW245C were gifts from the Upjohn Company and the Wellcome Research

Laboratories, respectively. IBMX was obtained from Aldrich. Ficoll 400 was obtained from Pharmacia. Cyclic AMP radioimmunoassay kit was purchased from Yamasa Shoyu Co., Choshi, Japan. Detection limit of the assay was 10 fmol per tube. The intra- and interassay coefficients of variation were 5.5% and 10.5%, respectively. PGD₂ and PGD₂ analogues were stored in ethanol at a concentration of 1 mg ml⁻¹ at -20°C. For preparation of aqueous solutions, a small volume of stock ethanolic solution was evaporated under N₂ gas, and the dried residues were dissolved in Krebs solution. Purity of PGD₂ and PGD₂ analogues was examined by silica gel thin layer chromatography before use. The R_F values of PGD₂, 9β-PGD₂, PGD₂ amide, PGD₂ N-monomethylamide, 17-phenyl-PGD₂, 9-deoxy-Δ^{9,12}-PGD₂ and BW245C were 0.19, 0.13, 0.08, 0.08, 0.19, 0.47 and 0.26, respectively, with a solvent system of benzene/ethyl acetate/acetic acid (50/50/2). After development, the compounds were visualized by exposure to iodine vapour. All the compounds gave a single spot upon this chromatography.

Statistical analyses

Results shown in the text and figures are expressed as mean ± s.e.mean. Statistical analyses were made by Tukey's method after one-way analysis of variance (Wallenstein *et al.*, 1980) (Figure 5) and by Student's *t* test.

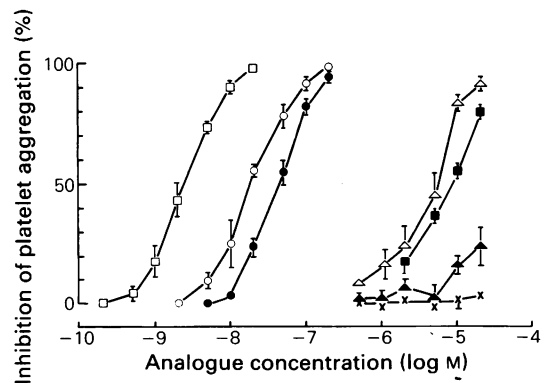


Figure 1 Concentration-inhibition curves for PGD₂ and PGD₂ analogues on ADP-induced platelet aggregation. (○) PGD₂ (*n* = 5); (●) 9β-PGD₂ (*n* = 5); (□) BW245C (*n* = 5); (■) 17-phenyl-PGD₂ (*n* = 4); (△) 9-deoxy-Δ^{9,12}-PGD₂ (*n* = 5); (▲) PGD₂ amide (*n* = 4); (x) PGD₂ N-monomethylamide (*n* = 4). Mean values are shown. S.e.mean is shown by a vertical line when it exceeds the size of a symbol.

Results

Human platelets and rat peritoneal mast cells

The ability of PGD₂ and PGD₂ analogues to inhibit ADP-induced platelet aggregation was determined *in vitro* in human platelet-rich plasma. The concentration-effect curves are shown in Figure 1. PGD₂ inhibited platelet aggregation with an IC₅₀ value of 18.6 ± 1.1 nM (*n* = 7). Among the analogues, BW245C and 9β-PGD₂ were potent agonists. BW245C was seven times more potent than PGD₂ (mean IC₅₀, 2.5 vs 18 nM). 9β-PGD₂ was about half as potent as PGD₂ (mean IC₅₀, 42 nM). 9-Deoxy-Δ^{9,12}-PGD₂ and 17-phenyl-PGD₂ showed full agonist activities but they were more than two orders of magnitude weaker than PGD₂ and had mean IC₅₀ values of 5.8 and 8.4 μM. Conversion of the carboxyl group to an amide almost completely abolished the anti-aggregatory activity. PGD₂ amide showed anti-aggregatory activity only above 10 μM, and PGD₂ *N*-monomethylamide was inactive up to 20 μM. Thus, the rank order of potency of PGD₂ analogues in inhibiting ADP-induced platelet aggregation was BW245C > PGD₂ > 9β-PGD₂ >> 9-deoxy-Δ^{9,12}-PGD₂ > 17-phenyl-PGD₂ > PGD₂ amide >> PGD₂ *N*-monomethylamide.

The same rank order of potency among PGD₂ analogues was observed on their abilities to accumulate cyclic AMP in rat peritoneal mast cells. PGD₂ induced cyclic AMP accumulation in these cells as described (Holgate *et al.*, 1980), and in the presence

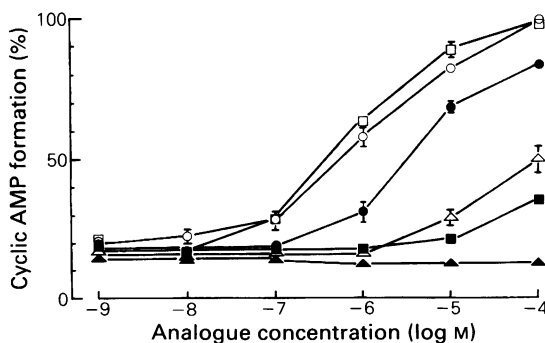


Figure 2 Concentration-dependent cyclic AMP accumulation in rat mast cells induced by PGD₂ and PGD₂ analogues. Cyclic AMP formation is expressed as a percentage of PGD₂- (100 μM) induced cyclic AMP formation in the same batches of mast cells (1988 ± 262 fmol per 10⁵ cells, mean ± s.e.mean, *n* = 7). Mean values are shown. S.e.mean is shown by a vertical line when it exceeds the size of a symbol. Symbols are the same as in Figure 1. Results with PGD₂ *N*-monomethylamide are not shown, since these overlapped almost completely those of PGD₂ amide.

of IBMX (1 mM) such accumulation reached plateau at 1 min after the addition of a prostaglandin. We, therefore, compared cyclic AMP levels in these cells 1 min after the addition of various concentrations of the analogues. Figure 2 represents the concentration-effect curves. PGD₂ stimulated cyclic AMP production with an EC₅₀ value of 0.60 ± 0.12 μM, and the maximal level produced was 1988 ± 262 fmol per 10⁵ cells (*n* = 7). BW245C and 9β-PGD₂ were again good agonists. BW245C was always more potent than PGD₂ in five experiments in which paired comparison of the activities was made; the equipotent molar ratio was 0.68. 9β-PGD₂ was a little less potent than PGD₂ with the molar ratio of 4.1. 9-Deoxy-Δ^{9,12}-PGD₂ and 17-phenyl-PGD₂ were weak agonists. They induced cyclic AMP accumulation only above 10 μM; therefore, the full concentration-effect curves were not obtained. The equipotent molar ratios for 9-deoxy-Δ^{9,12}-PGD₂ and 17-phenyl-PGD₂ were 190 and 590, respectively. PGD₂ amide and PGD₂ *N*-monomethylamide were completely inactive up to 100 μM in this system.

Rabbit transverse stomach strip

The concentration-effect curves of the rabbit stomach strip for PGD₂ and PGD₂ analogues are shown in Figure 3. PGD₂ relaxed this tissue in a concentration-dependent manner with the EC₅₀ value of 0.10 ± 0.02 μM (*n* = 5). Contrary to the findings of Whittle *et al.* (1979), PGE₂ elicited contractile responses in this tissue dose-dependently at concentrations

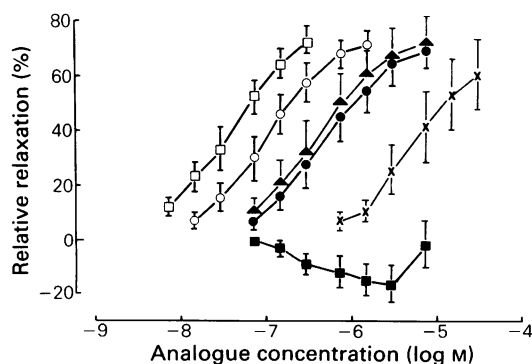


Figure 3 Concentration-relaxation curves for PGD₂ and PGD₂ analogues in rabbit stomach. Results are expressed as a percentage of the standard relaxation induced by 10⁻⁵ M papaverine. Mean values are shown. S.e.mean is shown by a vertical line when it exceeds the size of a symbol. Minus percentage denotes a contraction. Symbols are the same as in Figure 1. *n* = 5 for PGD₂, BW245C and 17-phenyl-PGD₂, *n* = 4 for 9β-PGD₂ and *n* = 3 for PGD₂ amide and PGD₂ *N*-monomethylamide.

above 10 nM ($n = 4$). PGF_{2 α} also evoked dose-dependent contractions over 20 nM ($n = 2$) (data not shown). Responsiveness of this tissue to PGD₂ analogues was similar to that of human platelets and rat mast cells in that BW245C and 9 β -PGD₂ were good agonists. BW245C was more potent than PGD₂; its equipotent molar ratio to PGD₂ was 0.35, and the mean EC₅₀ value was 35 nM. 9 β -PGD₂ also elicited full agonist activity but was about five times less potent than PGD₂. Thus, the rank order of potency is the same in three PGD₂-sensitive systems described so far; BW245C > PGD₂ > 9 β -PGD₂.

Since the actions of PGD₂ on the former two systems i.e. human blood platelets and rat mast cells are both linked to elevation of the intracellular cyclic AMP (Whittle *et al.*, 1978; Holgate *et al.*, 1980), we investigated a possible link of PGD₂-induced relaxation of the rabbit stomach strip to the adenylate cyclase system. To examine whether PGD₂ actually increased the cyclic AMP level, we rapidly froze in liquid nitrogen the strips which had been maximally relaxed by PGD₂ 1 μ M and determined the cyclic AMP content of these strips and of control strips similarly treated but without PGD₂. In the absence of IBMX, the concentrations of cyclic AMP in the control and relaxed tissues were 0.26 ± 0.02 and 0.43 ± 0.05 pmol mg⁻¹ wet tissue, respectively ($n = 4$ for each, $P < 0.05$), and in the presence of IBMX (10^{-4} M), the values were 0.79 ± 0.07 and 1.21 ± 0.03 pmol mg⁻¹ wet tissue, respectively ($n = 4$ for each, $P < 0.01$). The involvement of cyclic AMP in the relaxation of this tissue was further supported by the observation that forskolin, an adenylate cyclase activator (Seamon *et al.*, 1981), added to the organ bath elicited relaxation of the tissue dose-dependently (data not shown).

The rabbit stomach strip was different from the other two systems in the following respects. First, PGD₂ amide and PGD₂ *N*-monomethylamide elicited a concentration-dependent relaxation. PGD₂ amide was as potent as 9 β -PGD₂; its mean EC₅₀ value was 0.70 μ M. PGD₂ *N*-monomethylamide was much weaker than PGD₂ amide, and the mean EC₅₀ was about 10 μ M. Second, 17-phenyl-PGD₂ showed dual actions on this tissue, depending on the concentration used, and 9-deoxy- $\Delta^{9,12}$ -PGD₂ elicited contractile rather than relaxant effects. 17-Phenyl-PGD₂ contracted the tissue dose-dependently at concentrations from 0.1 to 5 μ M, and elicited dose-dependent relaxation over 5 μ M (Figure 3). 9-Deoxy- $\Delta^{9,12}$ -PGD₂ produced strong contractions of the tissue over 5 μ M. Since the contractions at each concentration continued to increase for more than 30 min, a concentration-effect curve for this compound could not be obtained.

Guinea-pig trachea

Cumulative concentration-effect curves of the guinea-

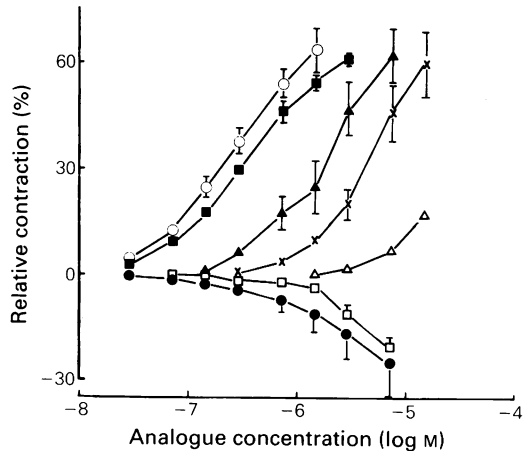


Figure 4 Concentration-contracture curves for PGD₂ and PGD₂ analogues in guinea-pig trachea. Contractions are expressed as a percentage of the KCl (30 mM)-induced maximal response in the same preparations. Mean values are shown. S.e.mean is shown by a vertical line when it exceeds the size of a symbol. Minus response denotes a relaxation. Symbols are the same as in Figure 1. $n = 5$ for PGD₂, $n = 4$ for 9 β -PGD₂, BW245C, 17-phenyl-PGD₂, PGD₂ amide and PGD₂ *N*-monomethylamide, and $n = 3$ for 9-deoxy- $\Delta^{9,12}$ -PGD₂.

pig trachea for PGD₂ and PGD₂ analogues are shown in Figure 4. PGD₂ contracted this tissue; maximum contraction, $69.4 \pm 5.1\%$ of 30 mM KCl-induced contraction, was attained at 2.84 μ M, and a 50% maximum response was elicited at a mean bath concentration of 0.24 ± 0.03 μ M ($n = 5$). 17-Phenyl-PGD₂ was almost as potent as PGD₂. The mean EC₅₀ value averaged 0.38 μ M. PGD₂ amide and PGD₂ *N*-monomethylamide were also full agonists with respective mean EC₅₀ values of 2.0 and 4.8 μ M. 9-Deoxy- $\Delta^{9,12}$ -PGD₂ elicited a weak agonist activity at concentrations higher than 2 μ M. BW245C and 9 β -PGD₂ did not produce any contractile response but relaxed the tissue at concentrations higher than 0.5 μ M.

In order to investigate antagonism of BW245C and 9 β -PGD₂ to PGD₂-induced contraction, we treated the tracheal preparations with 3 μ M indomethacin. The indomethacin treatment diminished the intrinsic tone of the preparations significantly. PGD₂ (2.8 μ M) contracted the tissues maximally to $156 \pm 8.3\%$ of the KCl- (30 mM) induced contraction after treatment and the EC₅₀ value was 0.44 ± 0.05 μ M ($n = 7$). Neither BW245C nor 9 β -PGD₂ evoked a response in the indomethacin-treated preparations up to concentrations of 28.4 μ M. Under these conditions, we analysed the antagonism of the two compounds by obtaining cumulative concentration-response curves for PGD₂ in the presence of three (3.55, 7.1 and 14.2 μ M) and four (3.55, 7.1, 14.2 and 28.4 μ M) concentrations of 9 β -

PGD₂ and BW245C, respectively. Both BW245C (Figure 5a) and 9 β -PGD₂ (Figure 5b) caused a parallel shift to the right of the log concentration-effect curve for PGD₂. Plots of log (dose ratio - 1) versus log molar concentration were constructed. The slopes of the lines fitted by least square regression for BW245C

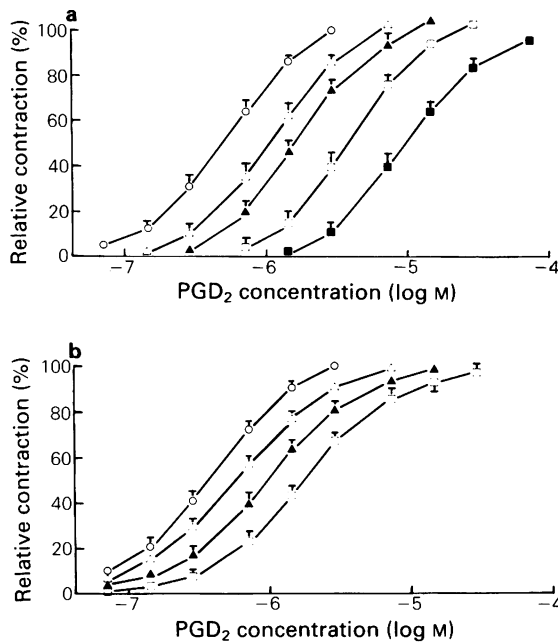


Figure 5 Concentration-contraction curves for PGD₂ in guinea-pig trachea in the absence and presence of BW245C (a) and 9 β -PGD₂ (b). BW245C and 9 β -PGD₂ were added at concentrations of 0 (○), 3.55 (△), 7.10 (▲), 14.2 (□) and 28.4 (■, in (a) only) μ M. Contractions are expressed as a percentage of the contraction induced by PGD₂ (10^{-5} M) in the control. Experiments were carried out in the presence of indomethacin 3 μ M. Pyrilamine maleate was omitted from the incubation solution. The mean ED₅₀ values for PGD₂ in the control and presence of 3.55, 7.10, 14.2 and 28.4 μ M BW245C were 0.49 ± 0.08 , 1.07 ± 0.17 , 1.63 ± 0.23 , 3.75 ± 0.43 , 9.30 ± 1.17 μ M, respectively ($n = 4$). The F ratio obtained from the analysis of variance, 39.77, is greater than the $P = 0.005$ critical value. The values in preparations treated with 14.2 and 28.4 μ M BW245C were significantly different from the control values by Tukey's method ($P < 0.01$, and $P < 0.01$, respectively). The mean ED₅₀ values for PGD₂ in the control and in the presence of 3.55, 7.10 and 14.2 μ M 9 β -PGD₂ were 0.37 ± 0.05 , 0.56 ± 0.06 , 0.95 ± 0.12 , 1.69 ± 0.26 μ M ($n = 3$). The F ratio obtained from the analysis of variance, 18.77, is greater than $P = 0.005$ critical value. The values in 14.2 μ M 9 β -PGD₂-treated preparations were significantly different from the control by Tukey's method ($P < 0.01$).

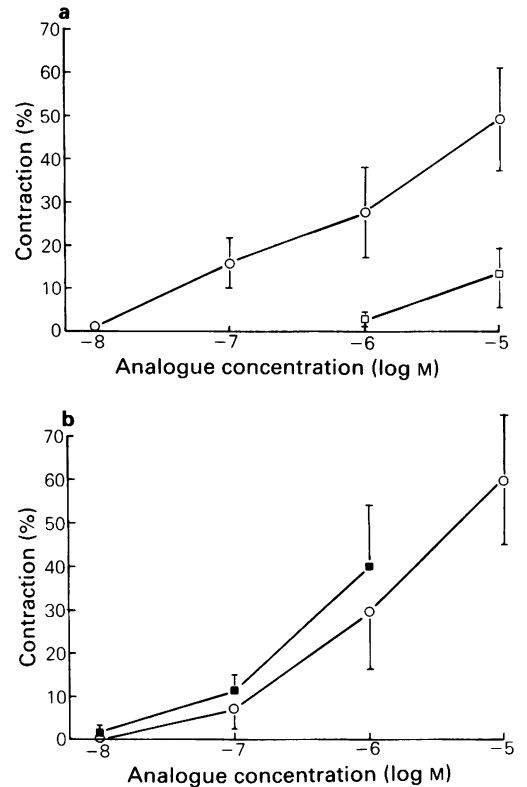


Figure 6 Concentration-contraction curves for PGD₂ and BW245C (a) or 17-phenyl-PGD₂ (b) in the dog cerebral artery strips. Results are expressed as a percentage of KCl- (30 mM) induced contraction in the same preparations. Symbols are the same as in Figure 1. $n = 7$ in (a) and $n = 5$ in (b).

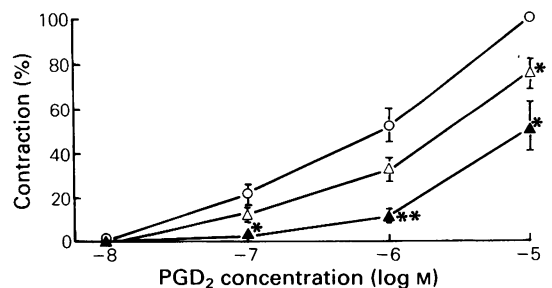


Figure 7 Modification by BW245C of the contractile response of the dog cerebral artery strips to PGD₂. Contractions induced by PGD₂ (10^{-5} M) in the control media were taken as 100%. Concentrations of BW245C were 0 (○), 1 (△) and 10 μ M (▲). $n = 7$. * Significantly different from the control: $P < 0.01$; ** $P < 0.001$.

and 9 β -PGD₂ were 1.36 and 1.36, respectively. The pA₂ values were found to be 5.48 and 5.26, respectively. Effects of BW245C and 9 β -PGD₂ were also examined on histamine-induced contraction of the tissue. These compounds reduced the histamine-induced contraction only slightly at the highest concentrations (28.4 μ M BW245C and 14.2 μ M 9 β -PGD₂); average reductions were 14.8 \pm 1.7% and 17.9 \pm 0.9%, respectively.

Dog cerebral artery strips

The addition of PGD₂ in concentrations ranging from 10⁻⁸ to 10⁻⁵ M produced a dose-related contraction in helical strips of dog cerebral (basilar and middle cerebral) arteries. BW245C did not significantly alter the cerebroarterial tone in concentrations up to 10⁻⁶ M but contracted the arteries at 10⁻⁵ M (Figure 6a). 17-Phenyl PGD₂ (10⁻⁸ to 10⁻⁶ M) elicited a concentration-dependent contraction, which tended to be greater than that induced by PGD₂ (Figure 6b).

Pretreatment with BW245C (10⁻⁶ and 10⁻⁵ M) attenuated the contractile response of cerebral artery strips to PGD₂ (Figure 7), but did not affect contractions of the strips induced by 5-hydroxytryptamine (5-HT) (data not shown). BW245C in these concentrations relaxed the artery strips ($n = 3$) precontracted with PGD₂ (10⁻⁶ M) but in contrast, contracted the arteries ($n = 3$) precontracted with K⁺ (12 to 15 mM).

Discussion

The results of the present study are summarized in Table 1. The five PGD₂-sensitive tissues and cells studied here can be classified into two groups. To one group belong human blood platelets, rat mast cells and rabbit stomach strip. They are characterized by (i) agonistic activities of two PGD₂ analogues i.e. BW245C and 9 β -PGD₂; (ii) linking of PGD₂ action to activation of adenylate cyclase; and (iii) PGD₂ evoking responses in these systems as potently as or more potently than other natural prostaglandins (Whittle *et al.*, 1978; 1979; Holgate *et al.*, 1980; this study). A specific PGD₂ receptor has been identified by radioligand receptor assay on one of them, i.e. human platelets (Schafer *et al.*, 1979; Siegl *et al.*, 1979) and BW245C has been shown to act on that receptor (Town *et al.*, 1983). It, therefore, seems likely that responses of the other two tissues are mediated by the same type of PGD₂ receptor. However, there were some differences in responsiveness to other PGD₂ analogues between the former two systems and the rabbit stomach strip. One difference was that platelets and mast cells did not respond to PGD₂ amide and PGD₂ *N*-monomethylamide, whereas the rabbit stomach strip responded to them with relaxation. Therefore, PGD₂ receptors in human platelets, rat mast cells and in the rabbit stomach strip do not appear to share all the same properties. The other

Table 1 Activities of prostaglandin D₂ analogues on human platelets, rat peritoneal mast cells, isolated preparations of rabbit stomach, guinea-pig trachea and dog cerebral artery, and murine L-1210 leukaemia cells

	Equipotent molar ratios (standard agonist, PGD ₂ = 1.0)					
	PGD ₂	BW245C	9 β -PGD ₂	17-phenyl PGD ₂	PGD ₂ -amide	9-deoxy- Δ ^{9,12} -PGD ₂
Inhibition of human platelet aggregation	1.0 (18 nM)	0.14	1.9	> 400	> 2,000	> 300
Cyclic AMP elevation in rat mast cells	1.0 (0.60 μ M)	0.68	4.1	> 500	> 500	190
Relaxation of rabbit stomach strip	1.0 (0.10 μ M)	0.35	4.8	Contraction	3.9	Contraction
Contraction of guinea-pig trachea	1.0 (0.24 μ M)	No effect relaxation	No effect relaxation	1.4	8.5	150
Contraction of dog cerebral artery	1.0 (1.1 μ M)	> 100	NT	0.46	NT	NT
Growth inhibition of L-1210 cells	1.0 (6.9 μ M)	No effect	NT	NT	NT	0.3

NT not tested.

Results with PGD₂ monomethylamide are omitted from the table.

Values on growth inhibition of L-1210 cells were obtained from Fukushima *et al.* (1982b) and Kikawa *et al.* (1984).

group of PGD₂-sensitive tissues consists of the guinea-pig trachea and dog cerebral artery. These tissues are characterized as follows; (i) PGD₂ and 17-phenyl-PGD₂ produce contractions with almost equal potency and (ii) BW245C and 9β-PGD₂ produce no response or only a slight contractions but antagonize the contractile action of PGD₂. It is evident that these contractions by PGD₂ are mediated via prostaglandin receptor(s) for the following two reasons. First, Toda (1982a, b) previously found that polyphlorethin and diphlorethin phosphates (prostaglandin antagonists) inhibit the contractile responses to PGD₂, PGF_{2α}, PGE₂ and carbocyclic thromboxane A₂ (cTXA₂) of dog cerebral artery but not the response to other vasoconstrictors such as K⁺ and 5-HT. Second, in the present study we have found that BW245C as well as 9β-PGD₂ effectively antagonizes the contractile responses of the tissues to PGD₂ but attenuates very little such responses to histamine or 5-HT. However, the above results does not necessarily imply that the receptor(s) involved is specific for PGD₂. Since PGF_{2α} and cTXA₂ constrict a helical strip of the dog cerebral artery more potently than PGD₂ (Toda, 1982a, b), it is likely that PGD₂ acts on the receptors of PGF_{2α} and/or thromboxane to elicit contraction. Jones *et al.* (1982) also postulated from their studies using EP-045 (a presumed thromboxane antagonist) that PGD₂ acts as a partial agonist on both contractile PGE₂ and thromboxane receptors to elicit contraction in the guinea-pig trachea. On the other hand, Feigen & Chapnick (1979) suggested separate contractile receptors for PGD₂ and PGF_{2α} in the canine mesenteric vascular bed and Jones (1978) suggested a potent contractile PGD₂ receptor in the sheep arterial vascular bed. Whether the responses we observed were mediated via interaction of PGD₂ on other prostaglandin and thromboxane receptors or via action on a PGD₂-specific contractile receptor(s) cannot be determined, unless highly selective antagonists against

actions of each of the prostaglandin and thromboxane are introduced.

Another system which is sensitive to PGD₂ analogues but has not been analyzed in the present study is cultured tumour cells. Inhibition of cultured tumour cell growth by PGD₂ and its analogues has been reported by several groups on more than 20 tumour cell lines (Narumiya *et al.*, 1985). 9-Deoxy-Δ^{9,12}-PGD₂ is the most potent PGD₂ analogue and has a growth inhibitory activity several times stronger than PGD₂ itself (Kikawa *et al.*, 1984). Interestingly, this compound is a poor agonist of PGD₂ in all of the four PGD₂-sensitive systems examined in this study. On the other hand, BW245C which showed significant actions on the rabbit stomach, human platelets and rat mast cells (Table 1) has been reported to lack such growth-inhibitory activity (Fukushima *et al.*, 1982b). Thus, the growth inhibition by PGD₂ analogues does not appear to be mediated via mechanisms mediating PGD₂ responses in the tissues and cells used in the present study, and cultured tumour cells may belong to another group of PGD₂-sensitive systems.

In conclusion, the present study suggests the presence of three group of PGD₂-sensitive systems which respond differently to synthetic PGD₂ analogues. These PGD₂ analogues can be used to modulate selectively these PGD₂-sensitive systems.

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