

Pharmacokinetic and pharmacodynamic studies with two α -adrenoceptor antagonists, doxazosin and prazosin in the rabbit

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- 1 The cardiovascular effects of doxazosin, a quinazoline derivative related to prazosin were investigated and compared to prazosin in the rabbit.
- 2 Radioligand binding studies using rabbit cerebral membranes showed that both doxazosin and prazosin were roughly equipotent at displacing [³H]-prazosin from specific binding sites. However, the lower pA₂ value for doxazosin at α_1 -adrenoceptors in isolated thoracic aorta preparations suggests a lower potency compared to prazosin.
- 3 The dose-related pressor effects of intravenous phenylephrine were used to assess vascular α_1 -adrenoceptor antagonism *in vivo*. There was a close agreement between α_1 -adrenoceptor antagonist potency and maximum hypotensive effects with both doxazosin and prazosin. The α_1 -adrenoceptor antagonist effects of doxazosin were more prolonged than those of prazosin.
- 4 Studies using either radioligand binding or pressor responses to B-HT 920 showed that doxazosin did not show any significant affinity for the α_2 -adrenoceptor. Similarly, no direct vasodilator effects were observed either in animals administered angiotensin II or in isolated thoracic aorta spiral strip preparations contracted with potassium.
- 5 Doxazosin has a longer terminal elimination half-life than prazosin. The pharmacokinetics of doxazosin were linear over the dose range examined.
- 6 Following pharmacological 'autonomic blockade' and treatment with prazosin, doxazosin did not cause any further fall in blood pressure.
- 7 These observations suggest that doxazosin, like prazosin, appears to exert its hypotensive action through α_1 -adrenoceptor antagonism. The prolonged fall in blood pressure and well sustained α_1 -adrenoceptor antagonism after doxazosin raise the possibility of an active metabolite which also has α_1 -adrenoceptor blocking properties.

Introduction

Doxazosin is a quinazoline derivative related to prazosin. Like prazosin, it is a relatively selective peripheral postsynaptic α_1 -adrenoceptor antagonist in animals (Karamat Ali *et al.*, 1980; Timmermans *et al.*, 1980; Cambridge & Davey, 1980; Wilson *et al.*, 1981; Vincent *et al.*, 1983a; Downing *et al.*, 1983); in human vascular preparations *in vitro* (Stevens & Moulds, 1981); and in man *in vivo* (Singleton *et al.*, 1980; 1982; Elliott *et al.*, 1982; Vincent *et al.*, 1983b).

This paper describes further *in vitro* and *in vivo* studies in the rabbit using doxazosin and prazosin. The affinity and specificity of these agents for the α_1 -adrenoceptor is compared as is their potency and

duration as α_1 -adrenoceptor antagonists. In addition, the relationship between pharmacokinetic and pharmacodynamic profiles and α_1 -adrenoceptor antagonism is investigated together with other possible receptor and non-receptor mediated mechanisms of blood pressure reduction.

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Methods

Male New Zealand white rabbits weighing 2.2–3.3 kg (Hyline Commercial Rabbits, Northwich, Cheshire) were housed individually in cages and fed a diet of

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rabbit pellet and water *ad libitum*. An arterial cannula was inserted into the central artery of the ear under local anaesthesia (Korner, 1965; Reid, 1979) for the measurement of mean arterial pressure using a Statham P23 ID pressure transducer and a Grass polygraph model 7B. Heart rate was measured directly from the arterial pressure trace. A venous cannula placed in a marginal ear vein under local anaesthesia was used for drug administration. The animals were allowed to recover from this minor surgical procedure unrestrained in individual cages for at least one hour before any measurements were made.

In preliminary experiments, 0.3 mg kg⁻¹ doxazosin and 0.1 mg kg⁻¹ prazosin gave comparable α_1 -adrenoceptor antagonism as assessed by the shift of the dose-response curve for the pressor effects of phenylephrine. For subsequent experiments, except where indicated, doxazosin 0.3 mg kg⁻¹ and prazosin 0.1 mg kg⁻¹ were used. Six to eight rabbits were studied in each experimental group. In all experiments, active treatment was compared with parallel control groups treated with 0.9% w/v NaCl solution (saline).

Pharmacokinetics and pharmacodynamics

Doxazosin 0.3 mg kg⁻¹ or prazosin 0.1 mg kg⁻¹ or an equivalent volume of saline vehicle (control) was given by intravenous bolus injection through a cannula in a marginal vein of the ear. Arterial blood samples were collected and mean arterial pressure and heart rate measured at corresponding intervals all through the day of the experiment. The total volume of blood removed did not exceed 16 ml over 8 h. The volume of blood removed was replaced by an equivalent volume of saline.

Doxazosin and prazosin were assayed in whole blood using h.p.l.c. with fluorescence detection (Yee *et al.*, 1979; Rubin *et al.*, 1980). The drug concentrations were appropriately fitted to a two compartment model, using computer assisted least squares non-linear regression with inverse weighting for drug concentrations, and the pharmacokinetic parameters subsequently derived.

Pressor responses and duration of α_1 -adrenoceptor antagonism

Pressor response curves were constructed using bolus intravenous doses of phenylephrine (0.001–0.3 mg kg⁻¹), a selective α_1 -adrenoceptor agonist, (Wikberg, 1978) before and after the administration of 0.3 mg kg⁻¹ doxazosin, 0.1 mg kg⁻¹ prazosin or saline vehicle. To determine the duration of α_1 -adrenoceptor antagonism, pressor responses to intravenous phenylephrine were repeated 15, 60, 120, 180, 240, 300 and 360 min following drug or vehicle administration. At least 4 doses of phenylephrine were studied on each occasion.

In a separate series of experiments, pressor responses to B-HT 920 (0.01–0.2 mg kg⁻¹) a selective α_2 -adrenoceptor agonist (Kobinger & Pichler, 1980; Hammer *et al.*, 1980) or angiotensin II (1–10 μ g kg⁻¹), a direct vasoconstrictor, were evaluated before and at 15 and 240 min after doxazosin 0.3 mg kg⁻¹ or prazosin 0.1 mg kg⁻¹. Again, at least 4 doses of agonist were studied at each time. The pressor dose-responses were analysed by the method described by Sumner *et al.* (1982) and all data points fitted to a quadratic function. The dose of phenylephrine required to cause a rise of 40 mmHg mean arterial pressure (PD₄₀) was derived. Similarly, the PD₂₀ and PD₆₀ were derived for B-HT 920 and angiotensin II respectively.

Non-receptor-mediated responses

Non-receptor-mediated responses were evaluated *in vivo* in animals following pharmacological 'total' autonomic blockade (TAB). Each animal received propranolol 2 mg kg⁻¹ and this dose was repeated hourly (Reid, 1979). In addition, each animal received atropine 2 mg kg⁻¹ followed by an infusion of 0.1 mg kg⁻¹ min⁻¹ (Korner *et al.*, 1968) and phenoxybenzamine 5 mg kg⁻¹ (Hamilton *et al.*, 1983). This drug regime was adequate to prevent any chronotropic and depressor responses to isoprenaline 2.5 μ g kg⁻¹ occurring, to markedly attenuate the pressor response to B-HT 920 0.2 mg kg⁻¹ and to shift the dose-response curve for the pressors effects of phenylephrine by more than 1000 fold. Thirty minutes after phenoxybenzamine, each animal received intravenous bolus injections of doxazosin (3 mg kg⁻¹) or prazosin (1 mg kg⁻¹) or saline vehicle.

In a separate experiment, with a similar protocol, after phenoxybenzamine treatment each animal received 0.5 mg kg⁻¹ prazosin followed 20 min later by 3 mg kg⁻¹ doxazosin or saline.

In vitro experiments: rabbit isolated aortic strips

Rabbits were killed with intravenous pentobarbitone sodium (60 mg kg⁻¹) and the descending thoracic aorta removed. Helical strips were prepared in the manner described by Furchgott & Bhadrakom (1953). Strips measuring 1.5 × 0.3 cm were suspended in 5 ml organ baths containing Krebs bicarbonate solution gassed with a mixture of 95% O₂ and 5% CO₂. Cocaine (10⁻⁵M) was added to block neuronal uptake, 17- β -oestradiol (10⁻⁵M) to block extra-neuronal uptake and propranolol (10⁻⁶M) to block β -adrenoceptor-mediated vasodilatation.

The pH of the bathing medium was 7.4. The temperature was maintained at 37°C by means of a Churchill thermostatic flow pump. An initial tension of 2 g was placed on the aortic strip and the tissue allowed to equilibrate over 90 min while the bathing medium was changed frequently. Isometric tension

was recorded via a Lectromed 4151 force transducer displayed on a Lectromed MX 412 type recorder. Cumulative dose-response curves were constructed (Van Rossum, 1963) using phenylephrine (10^{-9} to 10^{-2} M) before and after incubating the tissue with doxazosin 10^{-9} to 10^{-7} M or prazosin 10^{-10} to 10^{-8} M for 15 min. pA_2 values were calculated by the method of Arunlakshana & Schild (1959).

In a separate experiment tissues were contracted with 40 mM KCl. The effects of doxazosin 10^{-4} to 10^{-2} M and prazosin 3×10^{-4} to 10^{-2} M were evaluated on the potassium-induced contractile response. In the Krebs medium for this experiment, propranolol, cocaine and 17- β -oestradiol were omitted.

Radioligand binding studies

Rabbit brain membranes were prepared as described by Karliner *et al.* (1979). Aliquots of the membrane suspension (0.8 ml) were incubated for 25 min at 25°C with doxazosin 10^{-9} to 10^{-4} M and prazosin 10^{-9} to 10^{-4} M in the presence of either 3 nM [3 H]-prazosin (Greengrass & Bremner, 1979) or 7.5 nM [3 H]-clonidine (U'Pritchard & Snyder, 1979). The reaction was terminated by rapid vacuum filtration through GF/B glass fibre filters. The filters were washed twice with ice cold 0.05 M Tris buffer, pH 7.5, and left to dry at room temperature overnight. Eight ml of fisofluor containing 0.1% Triton X100 was then added to the dry filters and the bound tritium label counted in a liquid scintillation counter.

Total binding of the radiolabelled compound was determined in the absence of any displacing agent. Non-specific binding was defined as the amount of [3 H]-prazosin or [3 H]-clonidine bound in the presence of 10^{-5} M phentolamine.

Using these assay conditions, the stereo-specificity, pharmacological specificity and kinetics of the ligands [3 H]-prazosin and [3 H]-clonidine were consistent with binding to α_1 - and α_2 -adrenoceptors respectively.

Statistics

Results are presented as the mean \pm standard deviation (s.d.). Statistical analysis was by repeated measures analysis of variance for mean arterial pressure and heart rate responses and Student's *t* test for paired data, with Bonferroni correction elsewhere.

Drugs

Radiolabelled and unlabelled prazosin and doxazosin were obtained from Pfizer Ltd, Sandwich; B-HT 920 (6-allyl-2-amino-5,6,7,8-tetrahydro-4H-Thiazolo-[4,5-d]-azepine dihydrochloride) from Boehringer Ingelheim Ltd; angiotensin II from Ciba; phenoxybenzamine from Smith, Kline and French Ltd; phenylephrine and 17- β -oestradiol from Sigma; propranolol

from the Imperial Chemical Industries Ltd; and atropine sulphate from Antigen Limited, Roscrea, Ireland. Radiolabelled clonidine was obtained from the Radiochemical Centre, Amersham. All drugs were made up fresh in saline or in distilled water before being added to the Krebs solution.

Results

Blood pressure and heart rate

Blood pressure fell significantly ($P < 0.05$) after both doxazosin and prazosin at the doses used. The maximum fall in mean arterial pressure occurred between 5–7 h with doxazosin. Within the first 30 min, mean arterial pressure fell more with prazosin than doxazosin but between 4–8 h the mean fall with doxazosin was greater than in either the prazosin-treated or control animals. There was no significant increase in heart rate with either drug at the doses used compared to that in control animals (Figure 1).

Pharmacokinetics

In the rabbit, the mean terminal elimination half-life for doxazosin was 171 ± 56 min compared to 112 ± 38 min for prazosin. Doxazosin also had a

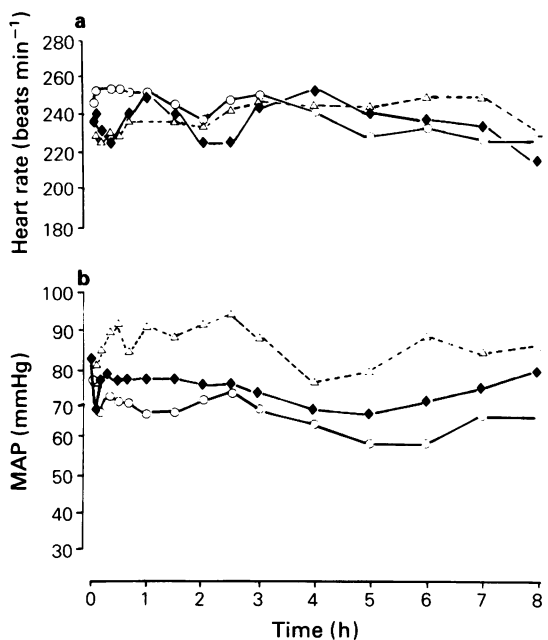


Figure 1 Mean arterial pressure (MAP; b) and heart rate (a) following the intravenous administration of doxazosin (O), prazosin (◆) and saline control (Δ) to 8 conscious rabbits.

Table 1 Derived pharmacokinetic parameters after intravenous administration of doxazosin and prazosin to the rabbit

	Doxazosin (0.3 mg kg ⁻¹)	Doxazosin (1 mg kg ⁻¹)	Prazosin (0.1 mg kg ⁻¹)
AUC (ng ml ⁻¹ min)	4641.6 ± 1120	23336 ± 2325.7*	4124.9 ± 834.6
βt _{1/2} (min)	171.9 ± 55.9	178.3 ± 83.8*	112.8 ± 38.1
Cl (ml min ⁻¹)	191.5 ± 33.9	185.8 ± 24.3*	92.7 ± 22.6
Vd _{ss} (l)	45.1 ± 13.5	40.1 ± 9.6*	14.0 ± 4.8

Data shown are mean ± s.d.; n = 8, *n = 4.

AUC = area under the concentration-time curve; βt_{1/2} = terminal elimination half-life; Cl = clearance; Vd_{ss} = apparent volume of distribution at steady state.

larger volume of distribution and more rapid clearance than prazosin in the rabbit (Table 1). At the higher dose of doxazosin (1 mg kg⁻¹) the half-life, clearance and volume of distribution were unchanged suggesting linear kinetics at these doses (Table 1). No metabolites were identified on the chromatogram of

the whole blood samples assayed for either prazosin or doxazosin.

Radioligand binding to rabbit cerebral membranes

In vitro ligand binding to membranes from rabbit cerebral cortex with [³H]-prazosin (α₁ specific ligand) and [³H]-clonidine (α₂ specific ligand) showed that both doxazosin and prazosin were roughly equipotent at displacing the radioligand [³H]-prazosin. The IC₅₀ value (the molar concentration of drug required to displace 50% of the bound ligand) was 5.2 × 10⁻⁸M for doxazosin and 8.9 × 10⁻⁸M for prazosin (Figure 2). For both drugs, the IC₅₀ for the displacement of [³H]-clonidine was greater than 10⁻⁵M suggesting that both drugs have relatively little affinity for the α₂-adrenoceptor in this preparation.

Isolated thoracic aorta preparation

Doxazosin caused a concentration-dependent shift of the contractile responses to cumulative doses of phenylephrine. The pA₂ value obtained from Schild regression analysis was 7.6 ± 1.5 with a slope of 0.94 ± 1.0 for doxazosin (Figure 3). This value contrasts with the pA₂ of 8.99 ± 0.8 and a slope of 0.98 ± 0.3 for prazosin. Incubation for up to 45 min did not alter the pA₂ for either prazosin or doxazosin.

Both doxazosin and prazosin in concentrations up to 10⁻²M failed to relax contractile responses induced by potassium chloride.

Pressor responses to intravenous phenylephrine and duration of α₁-adrenoceptor antagonism.

There was a parallel rightward shift of the dose-response curve to intravenous phenylephrine after

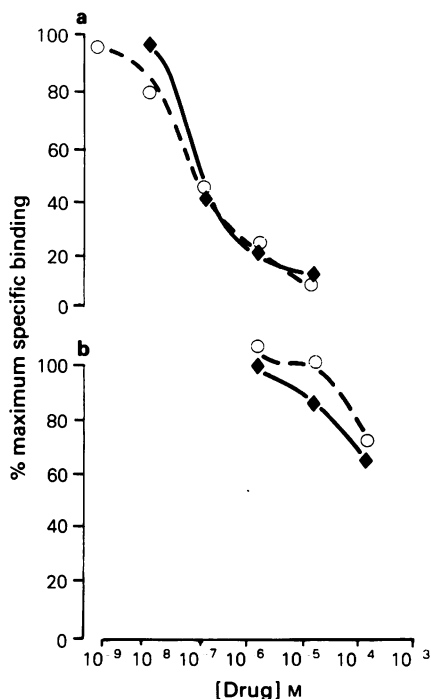


Figure 2 Radioligand displacement of [³H]-prazosin (a) and [³H]-clonidine (b) by doxazosin (O) and prazosin (◆) in isolated cerebral membranes of the rabbit.

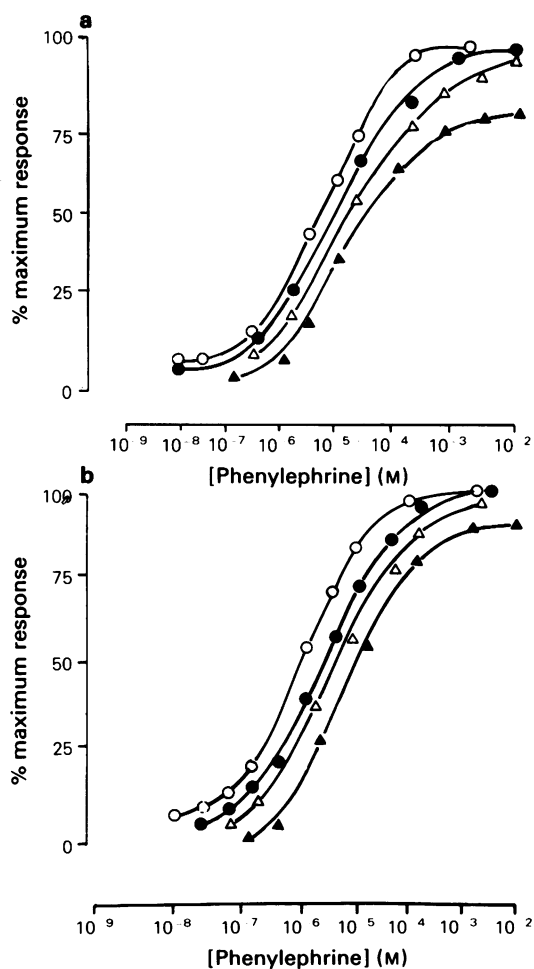


Figure 3 Contractile responses of rabbit isolated thoracic aorta to phenylephrine. (a) Shows responses to control (O); prazosin 10^{-10} M (●), 10^{-9} M (Δ) and 10^{-8} M (▲); while (b) shows responses in the presence of saline control (O), doxazosin 10^{-9} M (●), 10^{-8} M (Δ) and 10^{-7} M (▲). Each point is the mean of 6 observations.

either doxazosin or prazosin administration. The maximum shift of the dose-response curve occurred after 15 min with prazosin, with a dose-ratio of 3.8 ± 1.4 compared to 3.0 ± 0.6 at this time after doxazosin (Figure 4). α_1 -Adrenoceptor antagonism was more prolonged after doxazosin. A progressive increase to a maximum shift at 4 h (dose-ratio 4.3 ± 2.7) was observed in contrast to prazosin which showed a gradual recovery with a dose-ratio of 2.3 ± 0.7 at 4 h (Table 2).

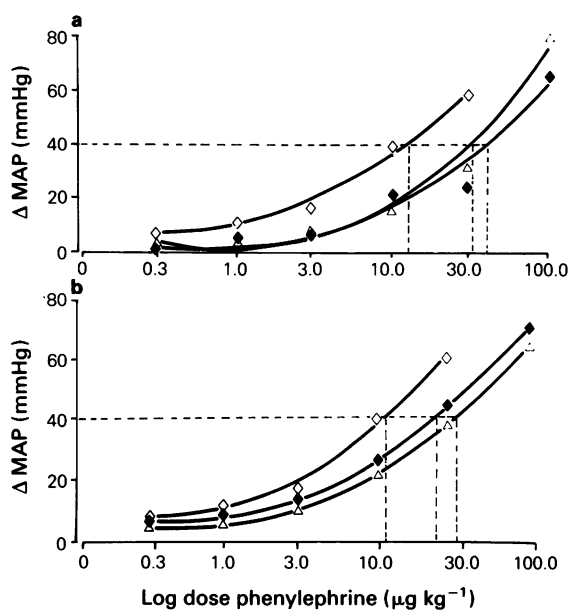


Figure 4 The pressor responses (change in mean arterial pressure (MAP)) to phenylephrine in animals given either saline (◇), doxazosin 0.3 mg kg^{-1} (Δ), or prazosin 0.1 mg kg^{-1} (◆), either 15 min (a) or 4 h (b) previously.

Pressor responses to B-HT 920 and angiotensin II

There was considerable inter animal variability in responses to B-HT 920 and neither doxazosin nor prazosin caused a significant shift of the pressor responses to intravenous B-HT 920 in the rabbit (Table 3).

Doxazosin and prazosin also failed to cause a significant shift of the pressor responses to the direct vasoconstrictor agent, angiotensin II, in the rabbit (Table 3).

Total autonomic blockade

In conscious rabbits following pharmacological 'total' autonomic blockade, doxazosin 3 mg kg^{-1} caused a rise in mean pressure of $16 \pm 5.2 \text{ mmHg}$ lasting 2–3 min. Using a similar protocol, prazosin 1 mg kg^{-1} caused a fall in pressure of $44.2 \pm 8.2 \text{ mmHg}$ lasting for 1.5–2 min followed by a pressor response of $19.0 \pm 7.4 \text{ mmHg}$ lasting 2–3 min. With both treatments, there was no change in heart rate confirming the adequacy of autonomic blockade (Figure 5). The animals treated with prazosin were subsequently given a further 1 mg prazosin after 30 min which resulted in a mean depressor response of $26.5 \pm 13.4 \text{ mmHg}$ lasting 1.5–2 min followed by a mean pressor effect of

Table 2 Dose of phenylephrine (mg kg^{-1} , i.v.) needed to raise mean arterial pressure 40 mmHg (PD_{40}) before and after $300 \mu\text{g kg}^{-1}$ doxazosin or $100 \mu\text{g kg}^{-1}$ prazosin or saline vehicle

Time	PD_{40}	Saline vehicle (dose-ratio)	PD_{40}	Doxazosin (dose-ratio)	PD_{40}	Prazosin (dose-ratio)
Control	12.0 ± 2.2	1.0	11.3 ± 4.3	1.0	12.0 ± 2.0	1.0
15 mins	12.5 ± 3.3	1.0 ± 0.2	$32.8 \pm 9.6^*$	3.0 ± 0.6	$44.1 \pm 10.9^*$	3.8 ± 1.4
60 mins	10.7 ± 3.0	0.9 ± 0.2	$30.3 \pm 5.2^*$	2.9 ± 0.9	$33.2 \pm 5.1^*$	2.8 ± 0.6
120 mins	13.7 ± 2.9	1.2 ± 0.2	$32.0 \pm 9.5^*$	2.9 ± 0.4	$28.0 \pm 4.7^*$	2.3 ± 0.5
180 mins	13.7 ± 4.1	1.2 ± 0.3	$33.1 \pm 8.6^*$	3.1 ± 0.9	$26.9 \pm 6.4^*$	2.3 ± 0.5
240 mins	13.7 ± 3.7	1.1 ± 0.3	$41.8 \pm 19.8^*$	4.3 ± 2.7	$26.3 \pm 6.5^*$	2.3 ± 0.7

Data shown are mean \pm s.d.; $n = 8$ rabbits.

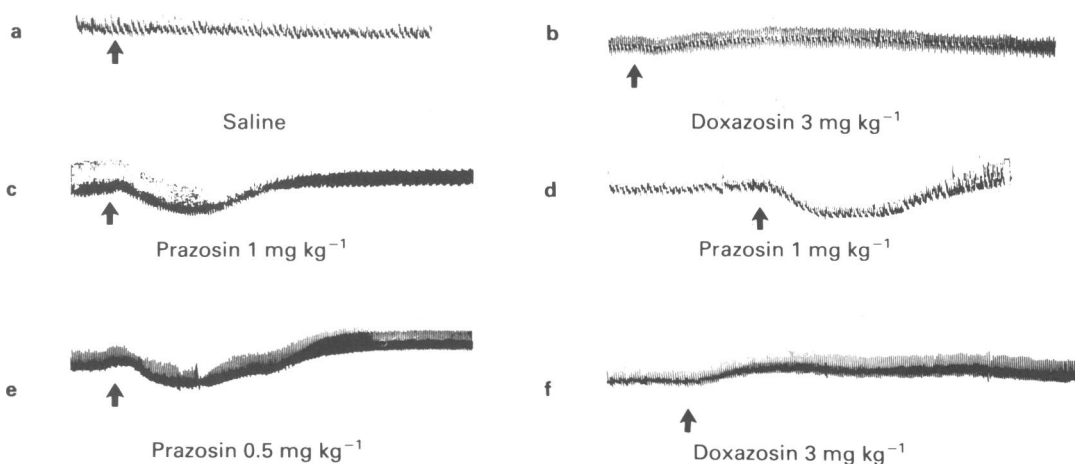
* $P < 0.05$ compared with control values.

Table 3 Doses of (a) angiotensin II and (b) B-HT 920 ($\mu\text{g kg}^{-1}$, i.v.) needed to raise mean arterial pressure 60 mmHg (PD_{60}) (a) and 20 mmHg (PD_{20}) (b) 15 min and 4 h after $300 \mu\text{g kg}^{-1}$ doxazosin or $100 \mu\text{g kg}^{-1}$ prazosin

a	PD_{60}	
	Doxazosin	Prazosin
Control	2.9 ± 0.9	3.3 ± 1.8
15 min	3.3 ± 1.2	4.7 ± 5.2
4 h	2.8 ± 1.6	3.5 ± 1.6

b	PD_{20}	
	Doxazosin	Prazosin
Control	42.0 ± 9.1	52.3 ± 9.9
15 min	52.3 ± 18.9	132.0 ± 69.4
4 h	49.1 ± 25.4	95.1 ± 55.9

Data shown are mean \pm s.d.; $n = 6$ rabbits.

**Figure 5** The effect of treatment with doxazosin and prazosin after pharmacological autonomic blockade; (a) response to saline; (b) response to doxazosin 3 mg kg^{-1} ; (c) response to prazosin 1 mg kg^{-1} ; (d) response to prazosin 1 mg kg^{-1} given 30 min after pretreatment with prazosin 1 mg kg^{-1} ; (e) response to prazosin 0.5 mg kg^{-1} ; (f) response to doxazosin 3 mg kg^{-1} after pretreatment with prazosin 0.5 mg kg^{-1} .

$8.1 \pm 3.8 \text{ mmHg}$ lasting 2–3 min; and no associated change in heart rate was observed.

In a separate group of animals, following 'total' autonomic blockade, prazosin 0.5 mg kg^{-1} caused a mean depressor response of $29.2 \pm 13.9 \text{ mmHg}$ followed by a pressor effect of $30.0 \pm 12 \text{ mmHg}$. Subsequent administration of doxazosin 3 mg kg^{-1} caused only a pressor effect of $12.0 \pm 5.7 \text{ mmHg}$ with no change in heart rate (Figure 5).

Discussion

Following intravenous administration of doxazosin and prazosin, there was a fall in mean arterial pressure compared to that in control animals. After prazosin there was a rapid fall which was maximal within 30 min with recovery over 2 h. With doxazosin, the response was more gradual in onset and was maximal

about 4–7 h later. With both treatments, there was no significant increase in heart rate at the doses used.

Doxazosin has a somewhat longer terminal elimination half-life than prazosin. It is more extensively distributed and also cleared faster than prazosin. However, these pharmacokinetic observations are not consistent with its apparent sustained α_1 -adrenoceptor antagonist effect. The observation that its maximum hypotensive effect occurs about 6 h later, by which time drug levels are low, would suggest either the involvement of an active metabolite or that doxazosin exerts its haemodynamic action in a different manner from prazosin or at a different site from that at which drug concentration was measured.

For both doxazosin and prazosin, maximum α_1 -adrenoceptor antagonism corresponded with maximum hypotensive response. At the doses used, the extent of α_1 -adrenoceptor antagonism was similar 15 min after treatment. Whereas there was a gradual recovery with prazosin, there was a paradoxical late increase and more prolonged α_1 -adrenoceptor antagonism with doxazosin. These observations can only in part be accounted for by the longer terminal elimination half-life of doxazosin and it is difficult to explain the delayed hypotensive response and paradoxical increase in α_1 -adrenoceptor antagonism considering the pharmacokinetic parameters. However, the results do suggest the involvement of an active metabolite with substantial α_1 -adrenoceptor antagonist properties, although no metabolite was detected by our assay technique under the conditions used. Alternatively, doxazosin could bind to the receptor in a partially covalent manner, but there is no evidence to support this hypothesis at present.

Doxazosin, like prazosin, failed to antagonize the pressor responses to B-HT 920 or angiotensin II. The pressor response to B-HT 920 results from activation of postsynaptic vascular α_2 -adrenoceptors (Kobinger & Pichler, 1980). The existence of postsynaptic α_2 -adrenoceptors has been demonstrated *in vivo* in animals (Drew & Whiting, 1979; Timmermans *et al.*, 1979; Reid & Hamilton, 1980; Docherty & McGrath, 1980; Starke & Docherty, 1980; McGrath, 1982) and in man (Elliott & Reid, 1983). The lack of effect of prazosin and doxazosin on pressor responses to B-HT 920 suggests that these drugs at the doses used have no effect on α_2 -adrenoceptors. This is consistent with previous observations of the lack of effect of prazosin and doxazosin on clonidine-induced responses and the failure of both drugs to increase transmitter release in rabbit pulmonary artery preparations (Cambridge & Davey, 1980). Similarly, the absence of any significant effect on the pressor responses to angiotensin II or potassium chloride suggests the lack of a direct smooth muscle relaxant action with both drugs at the doses used and under the conditions of the study.

The radioligand displacement studies further con-

firmed the α_1 -adrenoceptor selectivity of the drugs. Both doxazosin and prazosin were equipotent at displacing [3 H]-prazosin suggesting that they have a similar affinity for the α_1 -adrenoceptor. Neither doxazosin nor prazosin have any appreciable affinity or specificity for the α_2 -adrenoceptor, as suggested by the high molar concentration required to displace 50% of bound [3 H]-clonidine. However, the pA_2 values for doxazosin and prazosin were 7.6 ± 1.5 and 8.89 ± 0.8 , respectively. A high pA_2 value suggests a greater specificity and potency of an antagonist for a particular receptor. The explanation for the disparity between the affinity obtained from ligand binding and the pA_2 values derived from the Schild regression analysis is not clear. Alternatively, there may be differences between the α_1 -receptor population in cerebral membranes compared to the peripheral vasculature. The fact that the slopes of the Schild plots were not significantly different from unity suggests that both drugs are competitive α_1 -adrenoceptor antagonists. However, the observation that at high concentration, both doxazosin and prazosin tended to depress the maximum response (E_{max}) suggests that specificity for the α_1 -adrenoceptor may be dose-related. A similar observation has been made by Stevens & Moulds (1981) who described significant depression of E_{max} when using human vascular preparations. This may also be related to the ability of doxazosin and prazosin to block non-competitively the 'fast' but not the 'slow' component of the contractile responses (Downing *et al.*, 1983).

Pharmacological 'total' autonomic blockade allows the study of mechanisms not involving sympathetic and parasympathetic activity (Korner *et al.*, 1973; West *et al.*, 1975) and in particular α_1 -adrenoceptor-independent responses. In this preparation, doxazosin caused a pressor effect. Administration of doxazosin to animals pretreated with prazosin also did not lead to any further fall in mean arterial pressure. This suggests that doxazosin, prazosin and phenoxybenzamine act on a common receptor to mediate a depressor response. Pressor effects have been described with other α_1 -adrenoceptor antagonists when used in high doses in animals where the autonomic nervous system has been interrupted (Benfey & Varma, 1962; Hilliard *et al.*, 1972; Erker & Chan, 1977); and with prazosin (Drew & Whiting 1979). Recently, Angus & Lew (1984) observed an agonist effect of phentolamine in the rabbit, and attributed it to activation of vascular receptors that mediate vasodepressor responses which are sensitive to benextramine and yohimbine. It is possible that doxazosin in high doses also activates these receptors. Further investigations of the mechanism of this pressor effect are indicated.

Prazosin caused a depressor response followed by a pressor effect in this preparation. Constantine & Lebel

(1980) described pressor responses to noradrenaline mediated by vascular α_2 -adrenoceptors in phenoxybenzamine- and propranolol-treated dogs. Whether the depressor response to prazosin involves these vascular α_2 -adrenoceptors or is related to another mechanism of action is not clear. The depressor responses were followed by a pressor effect, similar to those observed with other α -adrenoceptor antagonists. Neither doxazosin nor prazosin had a sustained depressor effect in this preparation, suggesting that they do not exert a direct vasodilator effect of any importance. However, prazosin but not doxazosin caused a depressor response in this preparation by an α_1 -adrenoceptor independent mechanism.

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