Phonophoresis and the Absorption of Dexamethasone in the Presence of an Occlusive Dressing

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Context: Phonophoresis is purported to represent a method to apply topical medications through the skin to treat soft tissue injuries and inflammatory conditions. Few data are available to demonstrate the clinical effectiveness of the treatment.

Objective: To determine the effect of ultrasound on the transcutaneous absorption of dexamethasone when occluded with a dressing.

Design: Crossover design.

Setting: University general clinical research center.

Patients or Other Participants: Ten healthy subjects (age = 29.2 ± 8.8 years; height = 170.0 ± 3.9 cm; mass = 67.5 ± 18.4 kg).

Intervention(s): Two grams of 0.33% dexamethasone cream were applied to a 10-cm² area on the anterior forearm. The drug was applied to the skin and occluded with a dressing for 30 minutes before the ultrasound and sham ultrasound treatments. The treatments were applied over the drug and occlusive dressing. Ultrasound treatments were delivered at an intensity of 1.0 W/cm² (50% pulsed) at an output frequency of 3 MHz for 5 minutes and compared with sham ultrasound treatments that were delivered at an intensity of 0.0 W/cm² (50% pulsed) at an

output frequency of 3 MHz for 5 minutes. All subjects received both the ultrasound and sham treatments, and the order in which subjects received the treatments was counterbalanced.

Main Outcome Measure(s): Serum samples were drawn before treatment and immediately posttreatment and at 2, 4, 6, 8, and 10 hours posttreatment. Using high-performance liquid chromatography, we analyzed serum to determine dexamethasone concentrations.

Results: A 2-way repeated-measures analysis of variance (condition \times time) revealed a significant main effect for ultrasound treatment (P = .047). The rate of appearance and the total concentration of dexamethasone in the serum were greater in subjects after phonophoresis than after sham ultrasound. The sham group had only trace amounts of dexamethasone in the serum, indicating that drug absorption was negligible without the ultrasound energy. The effect size of the phonophoresis condition fell within a 95% confidence interval after the baseline measurement.

Conclusions: We found that a phonophoretic effect occurred with dexamethasone when its application saturated the skin.

Key Words: ultrasound, therapeutic ultrasound, skin saturation

Key Points

- Ultrasound enhanced the absorption of dexamethasone when the drug was applied to the skin and covered with an
 occlusive dressing.
- · Drug absorption increased with longer exposure time and use of an occlusive dressing.

Phonophoresis is a technique by which therapeutic ultrasound is used to introduce pharmacologic agents, usually anti-inflammatory or analgesic drugs, through intact skin into the subcutaneous tissues. Theoretically, phonophoresis can provide a safe and painless alternative to injections for treatment of common inflammatory conditions such as bursitis, sprains, strains, and tendinitis. Phonophoresis has been studied in vivo with several anti-inflammatory drugs, including hydrocortisone,¹⁻⁶ benzydamine,⁷ dexamethasone,^{4,8-10} and salicylates,¹¹⁻¹⁴ and with anesthetics, such as lidocaine,¹⁵⁻¹⁷ with variable results. Authors^{18,19} of in vitro studies of the phonophoretic effect of ultrasound reported that ultrasound enabled a greater transport of whole molecules across synthetic or organic semipermeable membranes than was afforded by sham ultrasound. Researchers have noted varying results with regard to the therapeutic benefits of phonophoresis (such as pain relief and improved range of motion) when it was used to treat lateral epicondylitis,^{3,20,21} temporomandibular joint pain,²² and osteoarthritic conditions.^{1,23,24} Most authors^{1,20–22} have shown that, when compared with placebo treatments or ultrasound alone, phonophoresis provides clinical improvement by decreasing pain and increasing function. In contrast, Halle et al²⁰ and Stratford et al³ reported no statistical difference between phonophoresis and ultrasound alone when treating pain and dysfunction in patients with lateral epicondylitis. However, in many of these studies,^{1,4–6,21–23} the inclusion criteria, ultrasound factors, drug dosages, and transmission of ultrasound through the drugs were controlled poorly.

One potential problem with phonophoresis may be related

to the clinical procedure used. Most clinicians apply the drug directly to clean skin, apply ultrasound conductive gel, and deliver the treatment for a period of 4 to 6 minutes, based on the size of the treatment area.²⁵ The drug and gel generally are removed immediately after the ultrasound treatment. Researchers^{6,10} have not shown an increased penetration of a top-ically administered drug in humans when ultrasound is applied with this technique. This technique may not allow adequate saturation of the stratum corneum, which is the rate-limiting barrier to transcutaneous drug absorption.^{26,27} Investigators^{28–31} have reported that a longer contact time and occlusion of the drug with an impermeable film enhance transcutaneous drug penetration.

The absence of a laboratory test to accurately measure the amount and depth of penetration of anti-inflammatory drugs has limited phonophoresis research. An effective technique that accurately can determine drug accumulation in the subcutaneous area of humans has not been established.^{5-7,11,12,14,16} Furthermore, investigators^{1,32} have used various ultrasound frequencies, intensities, and treatment durations that may not be indicated for use on an inflamed area. The commonly recommended factors^{33,34} for the treatment of acute or subacute inflammatory conditions are low intensity, high frequency, and pulsed mode to minimize thermal effects. Therefore, the ultrasound intensity for phonophoresis should be consistent with an intensity used in the treatment of a subacute inflammatory condition. Mitragotri et al³⁵ examined the minimal ultrasound energy for a phonophoretic effect, but the frequency that they suggested (20 kHz) is not consistent with the ultrasound used in physical rehabilitation.

Our purpose was to determine the effect of ultrasound on the transcutaneous absorption of dexamethasone when occluded with a dressing.

METHODS

Pilot Testing

An assay to determine the amount of dexamethasone in the blood is not commercially available. Clinically, endocrinologists use a measure of serum cortisol to determine the cumulative effect of therapy with glucocorticoids. In pilot testing, we attempted to use an enzyme-linked immunosorbent assay (ELISA) that detects steroid dosing in horses before races. When using radiomarked dexamethasone to examine the usefulness and accuracy of the ELISA, we could not control the sensitivity of the assay, especially in the lower ranges that we were anticipating. Although we were unable to power the study a priori, we were able to show significant differences between groups; therefore, retrospectively powering the analysis was not necessary. We did not use the ELISA in our study, but our finding is relevant for developing methods for future studies.

Subjects

Ten healthy subjects (3 men, 7 women; age = 29.2 ± 8.8 years; height = 170.0 ± 3.9 cm; mass = 67.5 ± 18.4 kg) volunteered to participate in our study. A physician examined all subjects before they entered the study. All subjects met the inclusion criteria: they had no skin lesions in the treated area; they had not used oral steroids or topical steroid creams within 1 month before the study; they had not received an intramus-

cular or intra-articular injection within the year before the study; they had no contraindications to the use of therapeutic ultrasound; and they had no history of vascular or heart disease. The institutional review board of the university and the General Clinical Research Center (GCRC) approved the study. All subjects gave informed consent.

Instruments

To administer ultrasound treatments, we used an Ultrasound 2000 (Accelerated Care Plus, Reno, NV) that had a beam uniformity ratio of 4:1 and an effective radiating area of 4.2 cm². The manufacturer calibrated the ultrasonic output of the generator 1 week before testing, and the use of the machine was reserved for this investigation. Blood samples were analyzed via high-performance liquid chromatography (HPLC) using a lambda-max ultraviolet spectrophotometer (model 486; Waters Associates Inc, Milford, MA), an autosampler (model 717 plus; Waters Associates Inc), and a Millenium chromatography manager (Waters Associates Inc). We set the wavelength to 245 N·m and followed the manufacturer's guidelines for use. All solvents and reagents were liquid chromatography grade (Waters Associates Inc). We obtained standards for the dexamethasone and hydrocortisone (product numbers D1756 and H5885, respectively; Sigma Chemical Co, St Louis, MO). The stationary phase was a µBondapak C18 column, and the liquid chromatograph was a pump with an automated gradient controller (model 501; Waters Associates Inc).

Procedures

The subjects were admitted to the GCRC for 1 night on 2 separate occasions. During 1 admission, sham ultrasound was applied to the volar surface of the forearm, and during the other admission, a therapeutic dosage of ultrasound was applied to the same area. All other aspects of the experimental procedure remained identical. The order in which these tests were performed was counterbalanced, and a minimum of 2 weeks elapsed between treatments.

An intravenous line was placed in a vein of the forearm that was not receiving the treatment, and it remained in position for the duration of the subject's stay. The untreated arm was chosen to ensure that a systemic, rather than a local, effect would be measured. If needed, excess hair was clipped carefully from a 10-cm² area on the volar surface of the forearm. A 10-cm² template was traced onto the forearm to ensure proper localization and treatment consistency. An individually prepared and equivalent 2-g dose of 0.33% dexamethasone cream (Vann Healthcare Services Inc, Glasgow, KY) from the same lot was applied to this area at 8:00 PM to control for a possible diurnal effect of the steroid. We used a topical dexamethasone mixture in an inert carbocol gel because Byl et al⁴ and J. C. Castel (unpublished data, 1998) found that ultrasound penetrates well through this drug. An occlusive dressing (Tegaderm; 3M, St Paul, MN) was applied over the drug to enhance the saturation of the stratum corneum with the drug.

The same clinician administered all treatments to ensure consistent ultrasound delivery. The subjects were blinded to their treatment groups. However, the clinician was not blinded because, unlike when the sham treatment was used, she could not disengage the ultrasound treatment timer when the ultrasound energy was used. The laboratory technicians were blind-

Results of Treatment With Ultrasound and Sham Ultrasound

Source	Type III Sum of Squares	Degrees of Freedom	Mean Square	F	P Value
Treatment (sham/ultrasound)	1095.36	1	1095.36	5.28	.047*
Error (treatment)	1868.31	9	207.59		
Time	577.78	6	92.96	3.90	.003*
Error (time)	1288.47	54	23.86		
Treatment $ imes$ time	234.94	6	39.16	1.46	.21
Error (treatment $ imes$ time)	1444.50	54	26.75		

*Significance was set at P < .05.

ed to the condition of each serum sample so that they could not determine group identity.

After a 30-minute exposure to the drug, each subject had the ultrasound or sham ultrasound treatment. Each treatment was performed by applying 4 g of Aquasonic gel (Parker Labs, Fairfield, NJ), which was premeasured, over the dressing that was occluding the drug. The experimental dosage was pulsed ultrasound (50%) applied with an intensity of 1.0 W/cm² at a 3-MHz frequency for 5 minutes. The sham group received the same treatment, but the output intensity was 0.0 W/cm². Both the drug and the ultrasound or sham ultrasound treatment remained within the designated area for the entire treatment. The clinician moved the transducer approximately from 2 cm/s to 4 cm/s; the distance was measured using a metronome during pilot testing. The transducer was moved in a circular pattern throughout the treatment area.

The ultrasound gel, occlusive dressing, and remaining dexamethasone cream were removed after each treatment. Nurses extracted a 10-mL sample of blood before treatment, immediately posttreatment, and at 2, 4, 6, 8, and 10 hours posttreatment. The blood was centrifuged within 30 minutes and frozen so that we could analyze the serum with HPLC. The samples were labeled so that the laboratory would not be able to determine group identity.

Subjects were allowed to eat, sleep, and participate in daily activities during their stay. They were discharged after breakfast.

High-Performance Liquid Chromatography

The HPLC was performed according to the procedure outlined by Caldarella et al.³⁶ We performed the assay with known standards in serum to check the accuracy of the quantification method. Several dilution techniques were tested to ensure that the standard curve was consistent and reliable at very low concentrations (less than 20 ng/dL). The assay was accurate at all concentrations that were more than 5 ng/dL. All standard curves had a correlation of 0.99 to known standards.

For each sample, we prepared the mobile phase of the HPLC in the following manner. We shook 4 mL of serum with methylene chloride to extract steroids. The aqueous phase was removed by aspiration, and the organic phase was washed with sodium hydroxide and water. Five-milliliter aliquots were evaporated to dryness in a water bath with a temperature of 37°C. The dried samples were reconstituted into a mobile phase and injected into the HPLC. Subject samples and controls were compared with a standard curve to determine the actual dexamethasone concentration.

The standard curve was prepared by spiking aliquots of known concentrations of hydrocortisone and dexamethasone. The curve specimens were extracted in the same way as the subject samples. Corrections for sample volumes and total serum volume were incorporated into the data reduction formula.

Statistical Analysis

We used 2-way repeated-measures analysis of variance (ANOVA) (condition \times time) to analyze the data. Post hoc means comparisons were performed if we found significant main effects or interactions. An α level of P < .05 was set for analysis. We used SPSS (version 11.0 for MacIntosh; SPSS Inc, Chicago, IL) to analyze the data.

RESULTS

The 2-way repeated-measures ANOVA indicated that the treatment had a main effect ($F_{1,9} = 5.28$, P = .047), with a significantly greater concentration of dexamethasone in the serum when ultrasound was used than when sham ultrasound was used (Table; Figure 1). We found a significant main effect for time (P = .003) with all posttreatment levels, more so than at the baseline, but at each time, they were not different from each other (Figure 2). No significant treatment \times time interaction was observed.

The main effect for time indicated a difference in the concentrations at various times. Examination by post hoc means comparisons demonstrated that this difference existed at a significant level only from the baseline measurement. The effect size for comparing the phonophoresis treatment condition with the sham condition (-0.5) was within a 95% confidence interval at all times after the baseline measurement (see the Table).

DISCUSSION

Our primary finding was a significantly greater accumulation of dexamethasone concentration in the serum when ultrasound was used than when sham ultrasound was used. The drug concentration increased over time, and it was still elevated 10 hours posttreatment. We found no significant condition \times time interaction. Although our results are inconsistent with the results found by authors^{1,3,6,7,10} of other phonophoresis studies on human subjects, we expected the experimental protocol of the extended exposure time and the use of an occlusive dressing to saturate the stratum corneum and enhance the effect of phonophoresis. We also thought that the choice of drug and the ultrasound factors would optimize the chance of increasing drug penetration. Using the occlusive dressing with sham ultrasound caused only a trace change in concentration of dexamethasone in the serum. Therefore, the massaging effect of repetitively moving the transducer over the oc-

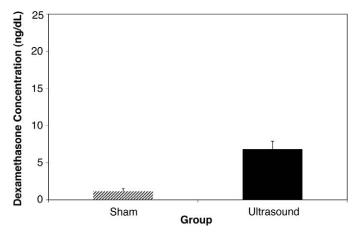


Figure 1. Effect of group on dexamethasone concentration. The mean concentration of dexamethasone in overall measurements of the sham group was well below the 5-ng/dL level that was considered to be an accurately measurable amount. The error bars represent SDs.

cluded drug did not cause an appreciable amount of drug absorption.

We chose treatment factors at the high end of the dosage range to maximize the potential phonophoretic effect. The factors were expected to cause a mild thermal effect, which may not be ideal for an inflammatory condition but which would increase the nonthermal effects of ultrasound.^{37–39} Michlovitz⁴⁰ reported that no investigators have shown evidence for the role of phonophoresis in treating tendinitis. However, we chose the mild thermal dosage based on the premise that the ultrasound energy would cause a phonophoretic effect without treating a potentially inflamed condition with an aggressive thermal dosage.

The serum samples had a high degree of variance. Some subjects had relatively little change in the drug concentrations even when ultrasound was applied, but other subjects had a remarkable increase in absorption. We did not have enough data to determine the variable absorption among subjects. Chien and Liu⁴¹ reported that skin type, age, and hydration can affect topical drug absorption; however, each subject in our study had both treatments, so the variance within subjects was minimized. Sex of the subjects is not a factor in skin permeability.⁴² We did not evaluate factors that may have contributed to variance in topical drug absorption. In addition, although dexamethasone is a steroid, it is not produced endogenously.

We examined the effect of a single exposure to ultrasound. Clinical treatments involving both ultrasound and phonophoresis in athletic training and physical therapy are often delivered several times per week for up to 2 or 3 weeks.²⁴ Franklin et al⁹ examined the effect of phonophoresis with dexamethasone on adrenal function when 8 treatments were delivered during a 2-week period. They found no immunosuppressant effect even with the repetitive treatments; however, they used the typical clinical protocol of a 5-minute exposure. We found an increased absorption of dexamethasone. Investigators must explore the ramifications of repeated treatments because of the risk of an immunosuppressant effect.

With the exception of Franklin et al,⁹ researchers conducting controlled experiments have used only a single application of phonophoresis. The biologic half-life of dexamethasone is from 26 to 54 hours, and we found that greater dexamethasone

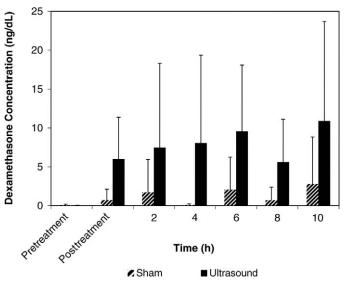


Figure 2. The mean concentrations of dexamethasone as a function of time (ng/dL). Although the drug concentration increased over time, the interaction of the condition and time was not significant. The error bars represent SDs.

levels in the serum were present up to 10 hours after a single exposure of ultrasound than were present after sham ultrasound. Therefore, a cumulative effect could occur if treatments were administered daily. The immunosuppressive effects of steroids have serious ramifications on numerous physiologic systems and must be examined. Clearly, the serum levels of the drug must be monitored to prevent adverse effects, and a physician must prescribe the administration of phonophoresis in this manner.

No clinical test is available to determine the amount of dexamethasone in the serum; the effects of the drug are interpreted from blood cortisol levels. We wanted to determine the amount of absorption of dexamethasone rather than the clinical or therapeutic dosage. The serum values that we measured were low (mean concentration = 6.7 ng/dL), but because a clinical laboratory test for this drug was not available, we could not determine whether this value represented a therapeutic range. More research is needed to evaluate the treatment effect. The effect sizes were calculated to compare the treatment condition with the sham condition and ranged from 1.34 (10 hours) to 61.23 (4 hours). These values exemplify the magnitude of the treatment effect.

Whether taken orally or applied topically, anti-inflammatory drugs have a local target that coincides with the injured part. Topically applied drugs diffuse through the epidermis to the dermis to reach the capillary networks, which causes systemic uptake of the drug.⁴¹ The systemic effects of the drug can be monitored by analyzing the blood serum content or the excretion of the drug in the urine. Researchers^{13,43} have found evidence that some topically administered drugs are absorbed at the treatment site in a greater concentration than in other areas of the body. We measured a systemic effect, but a drug concentration may localize in the area immediately below the administration site. This topical application over the muscle, synovium, ligaments, and tendons also could benefit the patient by providing a therapeutic dosage at the injured area.

CONCLUSIONS

Our results indicate that changes should be implemented with regard to the clinical procedure of phonophoresis. The clinician must choose a drug that enables the transmission of ultrasound. Dexamethasone is a strong anti-inflammatory drug that can be used without attenuating the ultrasound energy as it is delivered to the tissues. Other types of drugs and vehicles should be investigated for transmission of ultrasound before their use in phonophoresis. One of our most significant findings is the use of longer exposure times with an occlusive dressing. The commonly used method of applying the drug to the skin only for the duration of the ultrasound treatment has not produced favorable results in human or animal studies, even when dexamethasone has been used.¹⁰ Therefore, clinical applications that use treatment procedures similar to the procedures we used should produce more effective clinical results.

ACKNOWLEDGMENTS

This research was funded by the University of Virginia General Clinical Research Center and the National Athletic Trainers' Association Research & Education Foundation (Dallas, TX). We thank Frank McCue III, Ethan Saliba, Don Ball, William Webright, Chris Castel, and Chad Starkey for their research assistance and support of this endeavor.

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