

# NIH Public Access

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*Mitochondrion*. Author manuscript; available in PMC 2008 June 1.

Published in final edited form as: *Mitochondrion.* 2007 June ; 7(Suppl): S122–S126.

## CoQ<sub>10</sub> deficiency diseases in adults

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## Abstract

Deficiency of Coenzyme  $Q_{10}$  (Co $Q_{10}$ ) in muscle has been associated with a spectrum of diseases including infantile-onset multisystemic diseases, encephalomyopathies with recurrent myobinuria, cerebellar ataxia, and pure myopathy. Co $Q_{10}$  deficiency predominantly affects children, but patients have presented with adult-onset cerebellar ataxia or myopathy. Mutations in the CoQ10 biosynthetic genes, *COQ2* and *PDSS2*, have been identified in children with the infantile form of Co $Q_{10}$ deficiency; however, the molecular genetic bases of adult-onset Co $Q_{10}$  deficiency remains undefined.

> A lipid-soluble component of virtually all cell membranes, coenzyme  $Q_{10}$  (Co $Q_{10}$ ) or ubiquinone is an isoprenylated benzoquinone. Co $Q_{10}$  transports electrons from complexes I and II to complex III in the mitochondrial respiratory chain and is essential for the stability of complex III (Santos-Ocana et al, 2002). It is also an antioxidant (Villalba et al, 2000) and is involved in multiple aspects of cellular metabolism (Turunen et al, 2004).

> Primary  $CoQ_{10}$  deficiency causes clinically heterogeneous diseases: 1) encephalomyopathy characterized by the triad of recurrent myoglobinuria, brain involvement and ragged-red fibers (Ogasahara et al, 1989;Sobreira et al, 1997;Boitier et al, 1998;DiGiovanni et al, 2001;Aure et al, 2004); 2) severe infantile multisystemic disease (Rötig et al, 2000;Rahman et al, 2001;Salviati et al, 2005); 3) cerebellar ataxia (Musumeci et al, 2001;Lamperti et al, 2003;Gironi et al, 2004; Artuch et al, 2006); 4) Leigh syndrome with growth retardation, ataxia and deafness (Van Mardergem et al, 2002); and 5) isolated myopathy (Lalani et al, 2005;Horvath et al, 2006). These disorders are transmitted as autosomal recessive traits and in most cases respond to  $CoQ_{10}$  supplementation. In most of the reported patients, the exact site and nature of the defects in the biosynthesis of  $CoQ_{10}$  have not yet been identified. Because ubiquinone biosynthesis is complex and not fully defined, identification of the molecular genetic defect is not straightforward (Figure 1).

The pathogenic molecular defect has been identified in only 3 patients with the infantile form of primary  $CoQ_{10}$  deficiency: a homozygous mutation in the COQ2 gene, which encodes 4-para-hydroxybenzoate:polyprenyl transferase, in two siblings with nephropathy and encephalopathy (Salviati et al, 2006; Quinzii et al, 2006), and compound heterozygous mutations in the *PDSS2* gene, which encodes subunit 2 of polyprenyl diphosphate synthase, the first enzyme of the  $CoQ_{10}$  biosynthetic pathway (Figure 1), in an infant with lactic acidosis, Leigh syndrome, and nephropathy (Lopez Garcia et al, 2006). The renal disease in all three cases manifested as steroid unresponsive nephrotic syndrome.

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Among reported patients with presumed primary  $CoQ_{10}$  deficiency, 12 are adults (Table 1) (Musumeci et al, 2001, Van Maldergem et al, 2002; Lamperti et al, 2003; Gironi et al, 2004; Horvath et al, 2006). In some adult patients, onset of the disease was during childhood, but Lamperti and colleagues described four with adult-onset ataxia among a cohort of eighteen patients with cerebellar ataxia and low  $CoQ_{10}$  levels in muscle (Lamperti et al, 2003) while Gironi et al. and Horvath et al. reported four patients who presented at ages ranging from 29 to 39 years old (Gironi et al, 2004; Horvath et al, 2006).

The adult patients with  $CoQ_{10}$  deficiency reported by Musumeci et al. and Lamperti et al. had phenotypes similar to children with the ataxic form of  $CoQ_{10}$  deficiency, namely, cerebellar ataxia and atrophy, associated with seizures in 37% of the patients, pyramidal signs, mental retardation, weakness and motor development delay. Notably, the 3 adult patients reported by Lamperti and colleagues did not respond to  $CoQ_{10}$  supplementation, whereas the 3 young adults described by Musumeci and colleagues showed dramatic improvements. All three affected siblings were wheelchair-bound, with alternating esotropia, severe limb ataxia with the slightest purposeful movement, peripheral neuropathies, and scoliosis. One had generalized seizures and another had dystonia. After the proband began  $CoQ_{10}$  supplementations at age 20 years, his strength and ataxia improved and he became able to walk a few steps. His siblings showed similar improvements. In addition, seizures in the affected sister disappeared on CoQ<sub>10</sub> therapy and her anti-convulsant medication was discontinued (Musumeci et al, 2001). In these 3 patients, we demonstrated that  $CoQ_{10}$  deficiency was secondary to a stop codon mutation in the APTX gene, which is known to cause ataxia-oculomotor-aprataxia 1 (AOA1) (Quinzii et al, 2005, Date et al, 2001; Moreira et al, 2001). Results from measuring  $CoQ_{10}$ concentration in skeletal muscle from 12 additional patients from 6 different families with AOA1 confirmed this data (data not published). Intriguingly, both  $CoQ_{10}$  and cholesterol share a common biosynthetic pathway, therefore, in AOA1, altered levels of these molecules could be due to aberrant biosynthesis. There is no obvious link between aprataxin and regulation of  $CoQ_{10}$  synthesis or catabolism. Nevertheless, we did not detect mutations in APTX genes in other 13 patients with cerebellar ataxia and CoQ10 deficiency (Quinzii, DiMauro, Hirano, unpublished observation).

Van Maldergam and colleagues reported a 31-year-old woman and her older sister with the typical neuroradiological features of Leigh syndrome encephalopathy, growth retardation, infantilism, ataxia, deafness, and lactic acidosis, but unusually prolonged survival into adulthood. Both clinical and biochemical abnormalities improved remarkably with  $CoQ_{10}$  supplementation (Van Maldergam et al, 2002).

Gironi and colleagues described two brothers with hypergonadotropic hypogonadism and progressive cerebellar ataxia, which started in the fourth decade of life. The late onset of ataxia and associated low levels of testosterone distinguish these patients from those previously reported by Musumeci et al and Lamperti et al (Gironi et al, 2004). As the synthesis of steroid hormones starts with cholesterol, which has the same biosynthetic pathway as  $CoQ_{10}$ , it is conceivable that  $CoQ_{10}$  and testosterone deficiencies may coexist; moreover, it is possible that hypergonadotrophic hypogonadism has been under-recognized as endocrine studies were not reported in described patients with the ataxic form of CoQ<sub>10</sub> deficiency. Both patients responded to  $CoQ_{10}$  supplementation; they showed improved postural stability, gait and speech articulation, and testosterone returned to the normal range. The clinical improvements may have been related to increased muscle strength rather than amelioration of ataxia (Gironi et al, 2004). The lack of improvement of cerebellar functions with  $CoQ_{10}$  supplementation may be due to initiation of therapy after irreversible structural changes have occurred in the brain. Alternatively, insufficient tissue distribution of CoQ<sub>10</sub>, in particular its limited ability to cross the blood-brain barrier may explain the absence of therapeutic benefit in the CNS in these as well as in other patients subsequently reported (Aure et al, 2004, Lopez Garcia et al, 2006).

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Initial studies in rodents suggested that oral  $CoQ_{10}$  supplementation increased levels in plasma, spleen, and liver, but not CNS (Reahal et al, 1992,Zhang et al, 1996); however, subsequent publications demonstrated that long-term  $CoQ_{10}$  administration increased brain mitochondrial concentrations, particularly in aged rodents (Matthews et al, 1998,Kwong et al, 2002).

The most dramatic improvement after  $CoQ_{10}$  supplementation in adults has been associated with the pure myopathic form of  $CoQ_{10}$  deficiency (Lalani et al, 2005;Horvath et al, 2006). The clinical presentation of this variant appears to be homogeneous, with subacute (3 to 6 months) onset of exercise intolerance and proximal weakness affecting predominantly the hip and shoulder girdle muscles. Serum CK and lactate levels were markedly increased. Histologic examination of skeletal muscle revealed a lipid storage myopathy with subtle signs of mitochondrial dysfunction. Biochemical measurement of the respiratory chain enzymes showed reduced activities of complexes II and III (<50% of control mean) secondary to  $CoQ_{10}$  deficiency, as observed in all the variants of  $CoQ_{10}$  deficiency, and increased activity of citrate synthase, in keeping with mitochondrial proliferation. In the myopathic form,  $CoQ_{10}$  levels are low only in muscle, whereas in the infantile multi-systemic and in the cerebellar ataxic forms as well as in the patients described by Van Maldergam,  $CoQ_{10}$  is also reduced in fibroblasts.

CoQ<sub>10</sub> deficiency can be also a secondary consequence of drugs, such as statins (3-hydroxy-3methylglutaryl coenzyme A reductase inhibitors). Statins have been used for the treatment of hypercholesterolemia and coronary artery disease and for the prevention of stroke. Their mechanism of action is the inhibition of cholesterol synthesis at the level of mevalonic acid. The biosynthetic inhibition is not selective, because statins impair the synthesis of other compounds that share mevalonate as precursor, such as dolichols and  $CoQ_{10}$ . For this reason, statin-related myopathy, manifesting as myalgia, muscle necrosis, and myoglobinura, has been hypothesized to be due to a partial deficiency of  $CoQ_{10}$  (Folkers et al., 1985;Rundek et al. 2004). Indirect support for this hypothesis comes from the first reported cases of  $CoQ_{10}$ deficiency, which presented as exercise intolerance, recurrent myoglobinuria, and encephalopathy (mental retardation and seizures) (Ogashara et al, 1989;Sobreira et al, 1997;Boitier et al, 1998;Di Giovanni et al, 2001;Aure et al, 2004). Several groups have studied the effects of statins on the blood concentration of CoQ10 in patients with hypercholesterolemia and healthy subjects and there are several reports showing that various statins partially decrease CoQ<sub>10</sub> levels in blood of patients with hypercholesterolemia and controls, although the number of subjects studied and the severity of CoQ10 deficiency varied markedly (Folkers et al, 1985;Rundek et al, 2004). Recently, Lamperti et al. address the question of whether levels of CoQ10 were also decreased in muscles of patients with statin-related myopathy (Lamperti et al, 2006). The authors measured CoQ10 concentration and respiratory chain enzyme activities in biopsied muscle from 18 patients with statin-related myopathy. Moreover, they looked for evidence of mitochondrial myopathy or morphologic evidence of apoptosis using the TUNEL assay. Their studies revealed a mild decrease in muscle CoQ10 concentration without histochemical or biochemical evidence of mitochondrial myopathy or morphologic evidence of apoptosis in most patients (Lamperti et al, 2006).

Finally, it noteworthy that reduced levels of  $CoQ_{10}$  in blood and mitochondria have been reported in Parkinson disease (PD) by a number of investigators (Shults CW, 2005). These data, together with the implication of oxidative damage and mitochondrial dysfunction in Parkinson disease and other neurodegenerative diseases, such as amyotrophic lateral sclerosis (ALS), Huntington disease (HD), and Friedreich's ataxia (FRDA) have stimulated interest in potential therapeutic effects of  $CoQ_{10}$  as an antioxidant (Beal F, 2004). Initial small clinical trials have suggested beneficial effects in Parkinson disease and FRDA (Shults et al, 2002; Schapira, 2006). However, larger studies are necessary to better define the role of  $CoQ_{10}$  as primary or adjunctive therapy in neurodegenerative diseases.

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The molecular bases and pathogenic mechanisms of the various primary and secondary forms of  $CoQ_{10}$  deficiency remain largely unknown. To date, primary  $CoQ_{10}$  deficiency has been genetically and biochemically proven just in few patients with infantile multi-systemic severe diseases, where nephropathy and encephalopathy seems to be the most consistent feature. However,  $CoQ_{10}$  deficiency should be considered in the differential diagnosis of subacute exercise intolerance and weakness and of all genetically undefined adult-onset cases of cerebellar ataxia, as well as in patients with AOA1, because  $CoQ_{10}$  supplementation seems to improve muscle weakness and other associated symptoms in some individuals. Further studies are likely to shed new insights into causes and to improve therapies for the multiple variants of  $CoQ_{10}$  deficiencies in adults and children.

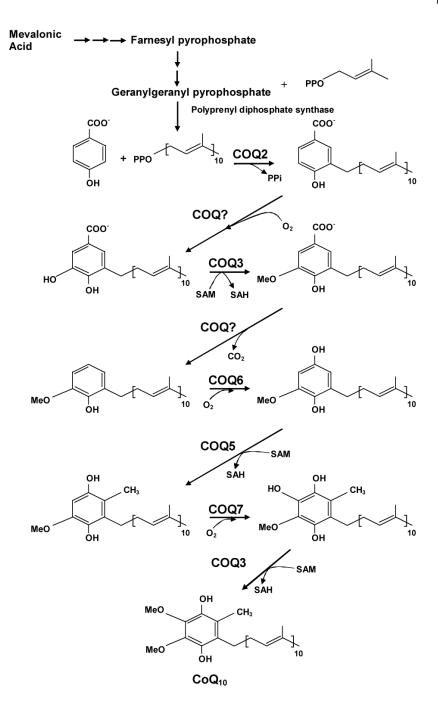
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#### Figure.

 $CoQ_{10}$  biosynthetic pathway with eight known biosynthetic enzymes denoted as polyprenyl diphosphosphate synthase (COQ1) and COQ2-8.  $CoQ_{10}$  is composed of a benzoquinone and a decaprenyl side chain. While the quinone ring is derived from amino acids tyrosine or phenylalanine, the isoprenoid side chain is produced by addition of isopentenyl pyrophosphate molecules to geranylgeranyl pyrophosphate (derived from mevalonate pathway) by decaprenyl diphosphate synthase. After para-hydroxybenzoate and decaprenyl pyrophosphate are produced, at least seven enzymes (encoded by *COQ2-8*) catalyze condensation, methylation, decarboxylation, and hydroxylation reactions to synthesize  $CoQ_{10}$ 

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Patient muscle CoQ <sub>10</sub> levels         Control muscle CoQ <sub>10</sub> levels $7.4^I$ $2.5\pm3.5^I$ $6.6^I$ $7.1^I$ $7.1^I$ $7.1^I$ $7.1^I$ $7.1$ $7.1^I$ $7.1$ $7.1^I$ $7.5\pm3.5^I$ $7.1^I$ $7.1$ $7.1^I$ $7.1$ $7.1^I$ $7.5$ $7.1^I$ $7.5$ $9.2^I$ $9.2^I$ $8.7^I$ $27.6\pm4.4^I$ $8.7^I$ $14.8^I$ $1.4.8^I$ $27.6\pm4.4^I$ $9.2^I$ $9.2^I$ $9.2^I$ $27.6\pm4.4^I$ $9.8^I$ $2.6\pm3^I$ $0.6^3$ $2.7.5\pm4.4^I$	Age (years)Patient muscle CoQ <sub>10</sub> levelsControl muscle CoQ <sub>10</sub> levels20 $74^1_1$ $25\pm3.5^1_1$ 2125 $6.6^1_1$ $25\pm3.5^1_1$ 24 $7.1^1_1$ $7.1^1_1$ $25\pm3.5^1_1$ 24 $7.1^1_1$ $7.3^2_1$ $793^2_1$ 25 $9.2^1_1$ $9.2^1_1$ $27.6\pm4.4^1_1$ 27 $9.2^1_1$ $9.2^1_1$ $27.6\pm4.4^1_1$ 35 $14.8^1_1$ $27.6\pm4.4^1_1$ 33 $0.6^3_3$ $2.7.7^3_1$ n fresh tissue (control values=mean ±standard deviation). $0.8^3_3$ $2.7.7^3_1$					
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$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Patient 1	20	$7.4^{I}$	$25\pm 3.5^{I}$	Musumeci et al., 2001
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Patient 2	25	$6.6^{I}$		
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$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Patient 5	31	43 <sup>2</sup>	793 <sup>2</sup>	Van Maldergem et al., 2002
35 $9.2^{I}$ 27 $8.7^{I}$ 30 $14.8^{I}$ 31 $14.8^{I}$ 32 $15.8^{I}$ 33 $15.8^{I}$ 33 $0.6^{3}$ 27.6 ± $4.4^{I}$ 33 $0.6^{3}$ 27.7 $^{3}$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Patient 6	24	$12.8^{I}$	$27.6 \pm 4.4^{I}$	Lamperti et al., 2003
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccc} 27 & 8.7^{I} \\ 30 & 14.8^{I} \\ 48 & 15.8^{I} \\ 35 & 13.5^{I} \\ 27.6 \pm 4.4^{I} \\ 35 & 13.5^{I} \\ 29 & 0.8^{3} \\ 29 & 0.8^{3} \\ 27.7^{3} \\ 27.7^{3} \\ 29 \\ 0.8 \end{array}$	Patient 7	35	$9.2^{I}$		
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccc} 30 & 14.8^{I} \\ \hline 48 & 15.8^{I} \\ 35 & 13.5^{I} \\ \hline 35 & 13.5^{I} \\ 29 & 0.8^{3} \\ n \ fresh \ tissue \ (control \ values=mean \ \pm standard \ deviation), \end{array}$	Patient 8	27	$8.7^I$		
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Patient 9	30	$14.8^{I}$		
35 $13.5^{I}$ 33 $0.6^{3}$ $2.7-7^{3}$ $29$ $0.8^{3}$ $2.7-7^{3}$	35 $13.5^{I}$ 33 $0.6^{3}$ $2.7-7^{3}$ n fresh tissue (control values=mean ±standard deviation), n rototin (control value=mean) and	Patient 10	48	$15.8^{I}$	$27.6 \pm 4.4^{I}$	Gironi et al., 2004
$\begin{array}{cccc} 33 & 0.6^3 & 2.7_7^3 \\ 29 & 0.8^3 & 0.8^3 \end{array}$	33 0.6 <sup>3</sup> 2.7-7 <sup>3</sup> 29 0.8 <sup>3</sup> 2.7-7 <sup>3</sup> n fresh tissue (control values=mean ±standard deviation), n motein (control value=mean) and	Patient 11	35	$13.5^{I}$		
29 0.8 <sup>3</sup> n fresh rissue (control values=mean +standard deviation)	29 0.8 <sup>3</sup> n fresh tissue (control values=mean ±standard deviation), n nrotein (control value=mean) and	Patient 11	33	0.6 <sup>3</sup>	2.7-7 <sup>3</sup>	Horvath et al., 2006
] uo/om fresh hissue (control values=mean +standard deviation)	l μg/gm fresh tissue (control values=mean ±standard deviation), 2 u«/om mrotein (control value=mean) and	Patient 12	29	0.83		
	2 10/m motein (control value=mean), and	<i>I</i> μg/gm fresh tissue (control va	ues=mean ±standard deviation),			
		$\beta$ nmol/unit citrate synthase (control=range)	trol=range)			