

RESEARCH PAPER

Effects of adenosine on adhesion molecule expression and cytokine production in human PBMC depend on the receptor subtype activated

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Background and purpose: Adenosine suppresses immune responses through adenosine_{2A} (A_{2A}) receptors, by raising intracellular cAMP. Interleukin (IL)-18 up-regulates the expression of intercellular adhesion molecule (ICAM)-1 on monocytes, leading to production of pro-inflammatory cytokines such as IL-12, interferon (IFN)- γ and tumor necrosis factor (TNF)- α by human peripheral blood mononuclear cells (PBMC). We have previously demonstrated that elevation of cAMP inhibits this IL-18-induced expression of adhesion molecules. In the present study, we examined the effect of adenosine on the IL-18-induced up-regulation of ICAM-1 on human monocytes and production of IL-12, IFN- γ and TNF- α by PBMC.

Experimental approach: The expression of ICAM-1 was examined by flow cytometry. IL-12, IFN- γ and TNF- α were determined by ELISA assay.

Key results: Adenosine inhibited the IL-18-induced up-regulation of ICAM-1 on human monocytes and it abolished the IL-18-enhanced production of IL-12, IFN- γ and TNF- α . While an A_{2A} receptor antagonist reversed the action of adenosine, an A₁ or A₃ receptor antagonist enhanced them. An A_{2A} receptor agonist, CGS21680, mimicked the effects of adenosine and its effects were abolished not only by the A_{2A} receptor antagonist but also by A₁ or A₃ receptor agonists. Activation via A_{2A} receptors resulted in elevation of cAMP in monocytes, whereas the stimulation of A₁ or A₃ receptors inhibited it, suggesting that intracellular signal transduction following ligation of A_{2A} receptors might be blocked by activation of A₁ or A₃ receptors.

Conclusions and Implications: Adenosine differentially regulates IL-18-induced adhesion molecule expression and cytokine production through several subtypes of its receptors.

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Keywords: adenosine; interleukin-18; intercellular adhesion molecule-1; human peripheral blood mononuclear cells; cytokine

Abbreviations: COPD, chronic obstructive pulmonary diseases; ICAM, intercellular adhesion molecule; IFN, interferon; IL, interleukin; LFA, lymphocyte function-associated antigen; PBMC, peripheral blood mononuclear cells; PKA, protein kinase A; TNF, tumor necrosis factor; MS, multiple sclerosis; RA, rheumatoid arthritis

Introduction

Adenosine is an endogenous nucleoside that regulates many physiological processes through its four receptor subtypes: A₁, A_{2A}, A_{2B} and A₃. These adenosine receptors belong to the superfamily of seven transmembrane G-protein-coupled receptors. A₁ and A₃ subtypes inhibit adenylate cyclase via Gi protein, whereas A_{2A} and A_{2B} activate adenylate cyclase

via Gs protein (Jacobson *et al.*, 1992; Poulsen and Quinn, 1998). Stimulation via A_{2A} receptors results in elevation of cAMP and the activation of protein kinase A protein kinase A (PKA) in human monocytes (Bshesh *et al.*, 2002). The Gs-coupled high-affinity A_{2A} receptor mediates many of the anti-inflammatory actions of adenosine in monocytes (Link *et al.*, 2000) and is involved in the inhibitory activity of adenosine analogs on tumor necrosis factor (TNF)- α production by lipopolysaccharide-activated monocytes (Khoa *et al.*, 2001; Zhang *et al.*, 2005). Thus, the A_{2A} receptor is likely to be important for the regulation of immune responses.

Interleukin (IL)-18 is a potent inflammatory cytokine secreted by antigen-presenting cells such as macrophages

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and dendritic cells in response to invading pathogens (Okamura *et al.*, 1995; Janeway and Medzhitov, 2002). Serum levels of IL-18 were also found to be elevated during multiple sclerosis multiple sclerosis (MS) (Gomez and Sitkovsky, 2003) and rheumatoid arthritis (RA) (Cronstein *et al.*, 1992; Marabito *et al.*, 1998). Cell-to-cell interactions between monocytes and T cells mediated through intercellular adhesion molecule (ICAM)-1 engagement with their respective ligands play important roles in immune responses (Camacho *et al.*, 2001). Recently, we found that IL-18 upregulated the expression of ICAM-1 on monocytes as well as influencing the production of IL-12, interferon (IFN)- γ and TNF- α in peripheral blood mononuclear cells (PBMC) (Takahashi *et al.*, 2002, 2003). Blockade of the engagement of adhesion molecule by antibodies against ICAM-1 reduced cytokine production by IL-18-treated PBMC (Takahashi *et al.*, 2002; 2003). This suggests that IL-18 induces cytokine production through upregulation of adhesion molecule expression on monocytes. However, little is known about the effect of adenosine on IL-18-initiated immune responses. In the present study, therefore, we investigated the effect of adenosine on the expression of adhesion molecules on monocytes and cytokine production by PBMC treated with IL-18 and the adenosine receptor subtypes what were involved therein.

Methods

Isolation of PBMC and monocytes

Normal human PBMC were obtained from ten healthy volunteers after IRB approval (Okayama University IRB No.106) (Table 1). PBMC were isolated from 20 to 50 ml of venous blood and they were treated with heparin. Monocytes were separated by counterflow centrifugal elutriation, as described previously (Takahashi *et al.*, 2002; 2003). The PBMC and monocytes were then suspended at a final concentration of 1×10^6 cells ml^{-1} in the culture medium, RPMI 1640 medium (Nissui, Co. Ltd, Tokyo, Japan) supplemented with 10% (v/v) heat-inactivated fetal calf serum, $20 \mu\text{g ml}^{-1}$ kanamycin and $100 \mu\text{g ml}^{-1}$ streptomycin and penicillin (Sigma, St Louis, MO, USA), as described previously (Takahashi *et al.*, 2002; 2003). The Figures 1–4 and 6 relate to experiments with PBMC and Figure 5 relates to data from monocytes isolated from PBMC. The PBMC obtained

from volunteers 1, 2, 3, 8 and 9 were used for Figures 1 and 2, and the PBMC obtained from volunteers 1, 2, 3, 4 and 5 were used for Figures 3 and 4. The monocytes obtained from volunteers 1, 3, 7, 8 and 9 were used for Figure 5, and the PBMC obtained from volunteers 1, 2, 3, 6 and 10 were used for Figure 6.

Flow cytometric analysis for adhesion molecule expression

Changes in the expression of human leukocyte antigen, ICAM-1, on monocytes were examined by multi color flow cytometry using a combination of anti-CD14 antibody with anti-ICAM-1 antibody. PBMC at 1×10^6 cells ml^{-1} were treated with adenosine, the selective A_1 (DPCPX), A_{2A} (ZM-241385), A_{2B} (alloxazine) and A_3 (MRS1220) receptor antagonists, the selective A_1 (CPA), A_{2A} (CGS-21680), A_{2B} (NECA) and A_3 (IB-MECA) receptor agonists (Table 2) or a PKA inhibitor, H89, each at concentrations ranging from 0.01 to $100 \mu\text{M}$ in the presence or absence of IL-18 at 100 ng ml^{-1} . Treatment was carried out by incubation for 24 h. The cells were washed once with washing buffer (phosphate-buffered saline supplemented with 2.5% normal horse serum, 0.1% NaN_3 , and 0.01 M HEPES, pH 7.3). The cells were prepared for flow cytometric analysis and analyzed with a FACSCalibur (BD Biosciences, San Jose, CA, USA) as described previously (Takahashi *et al.*, 2002; 2003). The data were processed using the CELL QUEST program. The data were expressed as the relative fluorescent intensities against isotype-matched control.

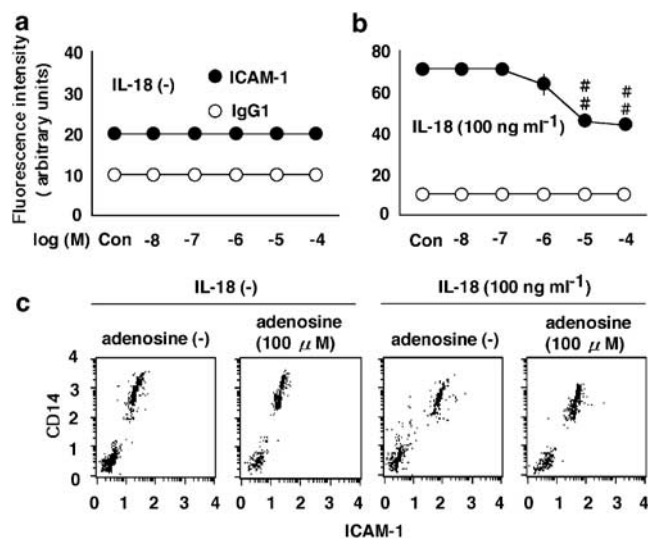


Figure 1 The effects of adenosine on the expression of ICAM-1 on monocytes. (a and b) The effect of adenosine at concentrations ranging from 0.01 to $100 \mu\text{M}$ (Con, no adenosine) on the expression of ICAM-1 in the absence (a) or presence (b) of 100 ng ml^{-1} IL-18. PBMC were treated with adenosine or IL-18, and the expression of ICAM-1 on monocytes was examined by multi color flow cytometry using a combination of anti-CD14 antibody with anti-ICAM-1 antibody. The values shown as IgG₁ represent the IgG₁ isotype-matched control. Results are expressed as the means \pm s.e.m. of triplicate determinations from five donors. $^{##}P < 0.01$ compared with the value for IL-18 alone. When an error bar was within a symbol, the bar was omitted. (c) The dot plots of ICAM-1 are shown. The cells were treated with adenosine at $100 \mu\text{M}$ in the presence or absence of IL-18 at 100 ng ml^{-1} and then incubated for 24 h.

Table 1 Blood donors

Volunteers	Sex	Age (years)
1	Man	47
2	Man	34
3	Man	33
4	Man	28
5	Man	35
6	Man	52
7	Man	30
8	Woman	37
9	Woman	66
10	Woman	28

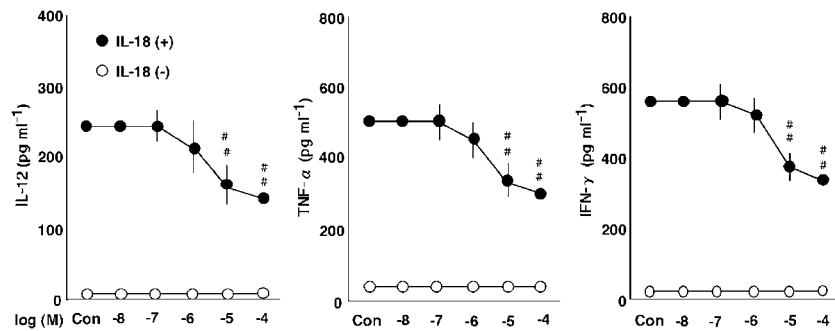


Figure 2 The effects of adenosine on the production of IL-12, TNF- α and IFN- γ by PBMC. The effect of adenosine at concentrations ranging from 0.01 to 100 μM (Con, no adenosine) on the production of IL-12, TNF- α and IFN- γ in the presence (+) or absence (-) of 100 ng ml^{-1} IL-18. Results are expressed as the means \pm s.e.m. of triplicate determinations from five donors. ## $P < 0.01$ compared with the value for IL-18 alone. When an error bar was within a symbol, the bar was omitted.

Enzyme-linked immunosorbent assay (ELISA) assay

PBMC at 1×10^6 cells ml^{-1} were used for analyzing IL-12, IFN- γ and TNF- α production. After culturing for 24 h at 37°C in a 5% CO_2 and air mixture, cell-free supernatant were assayed for IL-12, IFN- γ and TNF- α protein by enzyme-linked immunosorbent assay (ELISA) employing the multiple antibody sandwich principle (IL-12 (p70), TNF- α and IFN- γ reagents were from R&D Systems, Minneapolis, MN, USA). The detection limits of the ELISA for IL-12, IFN- γ and TNF- α were 10 pg ml^{-1} .

Measurement of cAMP production in monocytes

Monocytes at 1×10^6 cells ml^{-1} were incubated with the culture medium at 37°C in a 5% CO_2 and air mixture. After 30 min, trichloroacetic acid at a final concentration of 5% and 3-isobutyl-1-methylxanthine, an inhibitor of phosphodiesterase, at 100 μM was added to cells at 2×10^5 cells. These were then frozen at -80°C until analyzed. Frozen samples were subsequently sonicated and assayed for cAMP using a cAMP enzyme immunoassay kit (Cayman Chemical, Ann Arbor, MI, USA) according to the manufacturer's instructions, for which no acetylation procedures were performed.

Statistical analysis

Statistical significances were evaluated using analysis of variance (ANOVA) followed by Dunnett's test. A probability value of less than 0.05 was considered to indicate statistical significance. The results were expressed as means \pm s.e.m. of triplicate findings from five donors. The IC_{50} is the concentration at which 50% of the maximum inhibition was achieved.

Reagents and drugs

Recombinant human IL-18 was purchased from MBL (Nagoya, Japan). Adenosine, CPA (2-chloro-*N*-cyclopentyladenosine), DPCPX (8-cyclopentyl-1,3-dipropylxanthine), CGS-21680 (2-(*p*-(2-carboxylethyl) phenylethylamino)-5-*N*-ethylcarboxamido adenosine), ZM-241385 (4-(2-(7-amino-2-(2-furyl)(1,2,4-triazolo(2,3-NECA (5'-*N*-ethylcarboxamido adenosine), alloxazine (benzo(g)pteridine-2,4 (1H,3H)-dione),

IB-MECA (1-deoxy-1-(6-((3-iodophenyl) methylamino)-9H-purin-9-yl)-*N*-methyl-D-ribofuranuronamide), MRS1220 (*N*-(9-chloro-2-furan-2-yl-(1,2,4)triazolo (1,5-*c*)quinazolin-5-yl)-2-phenylacetamide) and H89 were purchased from Sigma Chemical. For flow cytometric analysis, fluorescein isothiocyanate (FITC)-conjugated mouse IgG1 monoclonal antibody (mAb) against ICAM-1 (6.5B5), which recognizes the D1 domain of ICAM-1 and phycoerythrin-conjugated anti-CD14 mAb, were purchased from DAKO (Glostrup, Denmark). An FITC-conjugated IgG1 isotype-matched control was obtained from Sigma Chemical.

Results

The effects of adenosine on the expression of ICAM-1 on monocytes and the production of IL-12, TNF- α and IFN- γ by PBMC

We investigated the effect of adenosine on the expression of ICAM-1 on monocytes (Figure 1) and its impact on the production of IL-12, TNF- α and IFN- γ by PBMC (Figure 2). Adenosine had no effect on the expression of ICAM-1 or cytokine production in the absence of IL-18. IL-18 induced the expression of ICAM-1 on monocytes and the production of IL-12, TNF- α and IFN- γ (Takahashi *et al.*, 2002; 2003). Adenosine inhibited the IL-18-enhanced expression of ICAM-1 and production of IL-12, TNF- α and IFN- γ (Figures 1 and 2). The IC_{50} values of adenosine for inhibition of the IL-18-enhanced ICAM-1 expression and IFN- γ production were both 20 μM , with the maximal effect being obtained at 100 μM .

The effect of a selective A_1 , A_{2A} , A_{2B} and A_3 receptor antagonists on the action of adenosine

To investigate the involvement of four different subtype receptors in the action of adenosine, we examined the effects of the selective A_1 , A_{2A} , A_{2B} and A_3 receptor antagonists, DPCPX, ZM-241385, alloxazine and MRS1220, respectively, on the adenosine-induced inhibition of ICAM-1 expression on monocytes and the production of IL-12, TNF- α and IFN- γ by PBMC in the presence of IL-18 (Figure 3a and b). The A_{2A} receptor antagonist, ZM-241385, reversed both the

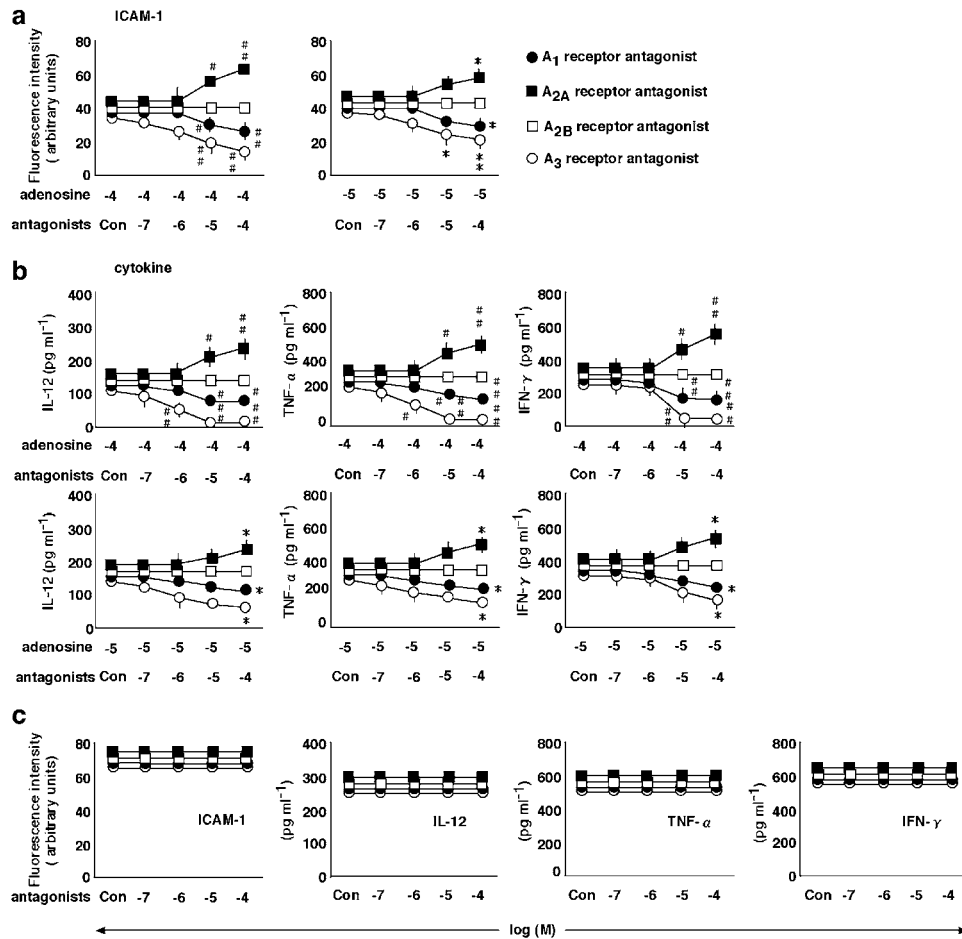


Figure 3 The effects of A₁, A_{2A}, A_{2B} and A₃ receptor antagonists on the action of adenosine. (a) The effects of the selective A₁, A_{2A}, A_{2B} and A₃ receptor antagonists, DPCPX (●), ZM-241385 (■), alloxazine (□) and MRS1220 (○), respectively, at concentrations ranging from 0.1 to 100 μM (Con, no antagonist) on the expression of ICAM-1 in the presence of IL-18 at 100 ng ml⁻¹ and adenosine at 10 (right-hand graph) and 100 μM (left-hand graph). PBMC were treated with various stimulations and the expression of ICAM-1 on monocytes was examined by multi color flow cytometry using a combination of anti-CD14 antibody with anti-ICAM-1 antibody. (b) The effects of A₁, A_{2A}, A_{2B} and A₃ receptor antagonists (Con, no antagonist) on the production of IL-12, TNF-α and IFN-γ in the presence of IL-18 at 100 ng ml⁻¹ and adenosine at 10 (lower graphs) and 100 μM (upper graphs). (c) The effects of antagonists at concentrations ranging from 0.1 to 100 μM (Con, no antagonist) on the expression of ICAM-1 and production of IL-12, TNF-α and IFN-γ by PBMC treated with IL-18 at 100 ng ml⁻¹ in the absence of adenosine were determined. Results are expressed as the means ± s.e.m. of triplicate determinations from five donors. #*P* < 0.05, ##*P* < 0.01 compared with the value for IL-18 and adenosine at 10 μM. **P* < 0.05, ***P* < 0.01 compared with the value for IL-18 and adenosine at 100 μM. When an error bar was within a symbol, the bar was omitted.

adenosine-induced inhibition of expression of ICAM-1 and cytokine production, but the A_{2B} receptor antagonist, alloxazine, had no effect. Conversely, the A₁ and A₃ receptor antagonists, DPCPX and MRS1220, enhanced the actions of adenosine. None of the antagonists had any effect in the absence of adenosine (Figure 3c).

The effects of an A_{2A} receptor agonist on the IL-18-enhanced expression of ICAM-1 on monocytes and production of IL-12, TNF-α and IFN-γ by PBMC

The effects of the selective A₁, A_{2A}, A_{2B} and A₃ receptor agonists, CPA, CGS-21680, NECA and IB-MECA, respectively, on the expression of adhesion molecules and cytokine production in the presence or absence of IL-18 were determined (Figure 4). In the presence of IL-18, the A_{2A} receptor agonist CGS-21680 inhibited the expression of

ICAM-1 and production of IL-12, TNF-α and IFN-γ (Figure 4a). However, it had no effect on the cytokine production in the absence of IL-18 (data not shown). The A₁, A_{2B} and A₃ receptor agonists had no effect in the presence or absence of IL-18 (data not shown). In addition, we investigated the effects of four different receptor antagonists on the A_{2A} agonist-initiated inhibition of ICAM-1 expression and production of IL-12, TNF-α and IFN-γ in the presence of IL-18 (Figure 4b). The A_{2A} receptor antagonist, ZM-241385, but not the other receptor antagonists reversed the action of the A_{2A} receptor agonist, CGS-21680. Moreover, we examined the effect of the A₁, A_{2B} and A₃ receptor agonists on the A_{2A} receptor agonist-initiated inhibition of ICAM-1 expression on monocytes and production of IL-12, TNF-α and IFN-γ by PBMC in the presence of IL-18 (Figure 4c). The A₁ and A₃ receptor agonists, CPA and IB-MECA, but not the A_{2B} receptor

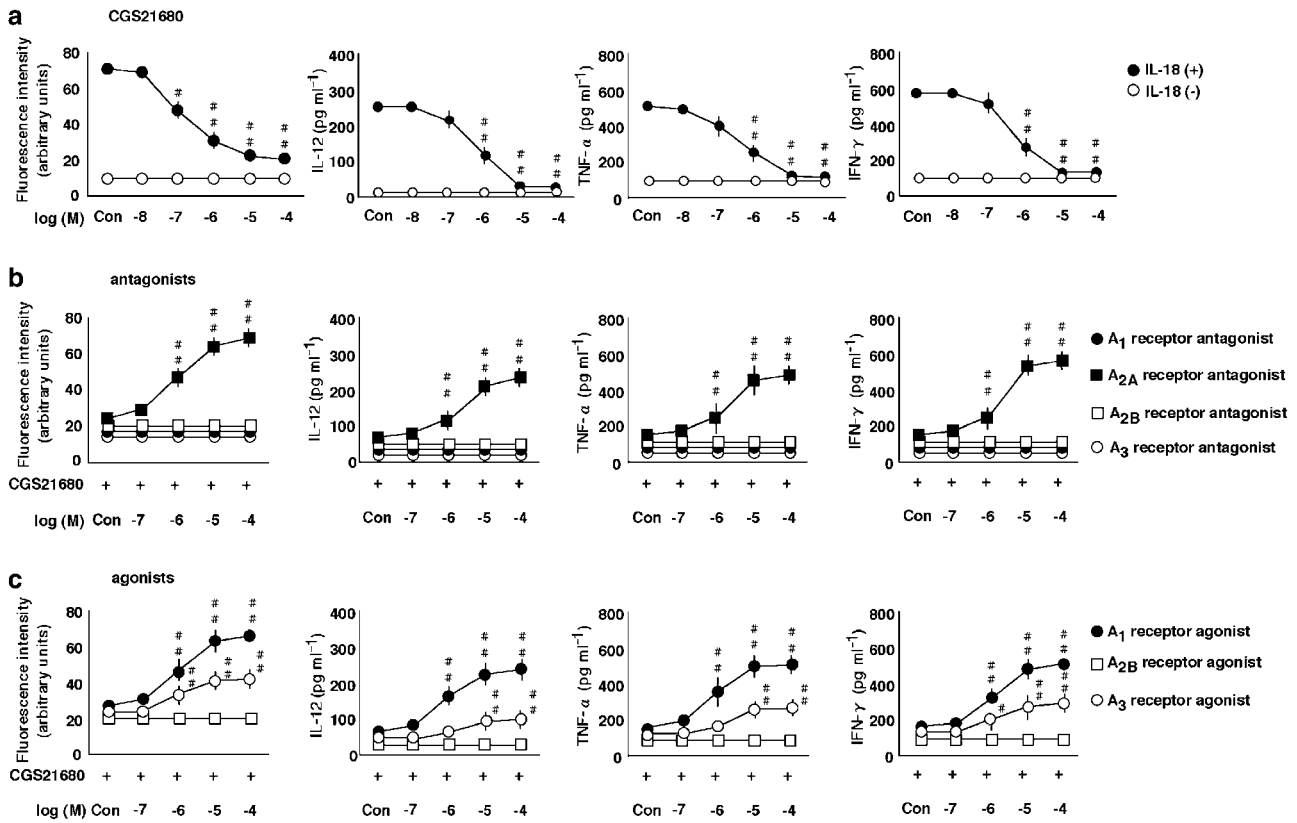


Figure 4 The effects of an A_{2A} receptor agonist on the expression of ICAM-1 on monocytes and production of IL-12, TNF- α and IFN- γ by PBMC and their modulation. (a) The effect of a selective A_{2A} receptor agonist, CGS-21680, at concentrations ranging from 0.1 to 100 μ M (Con, no CGS 21680) on the expression of ICAM-1, and the production of IL-12, TNF- α and IFN- γ in the presence (+) or absence (-) of IL-18 at 100 ng ml⁻¹. The expression of ICAM-1 on monocytes was examined by multi color flow cytometry using a combination of anti-CD14 antibody with anti-ICAM-1. The results are expressed as the means \pm s.e.m. of triplicate determinations from five donors. $^{\#}P < 0.05$, $^{\#\#}P < 0.01$ compared with the value for IL-18 alone. (b) The effect of a selective A_1 , A_{2A} , A_{2B} and A_3 receptor antagonists, DPCPX, ZM-241385, alloxazine and MRS1 220, at concentrations ranging from 0.1 to 100 μ M (Con, no antagonist) on the expression of ICAM-1 and production of IL-12, TNF- α and IFN- γ in the presence of IL-18 at 100 ng ml⁻¹ and CGS-21680 at 100 μ M. $^{\#}P < 0.05$, $^{\#\#}P < 0.01$ compared with the value for IL-18 and CGS-21680. (c) The effect of a selective A_1 , A_{2B} and A_3 receptor agonists, CPA, NECA and IB-MECA, respectively, at concentrations ranging from 0.1 to 100 μ M (Con, no agonist) on the expression of ICAM-1 and production of IL-12, TNF- α and IFN- γ in the presence of IL-18 at 100 ng ml⁻¹ and CGS-21680 at 100 μ M. $^{\#}P < 0.05$, $^{\#\#}P < 0.01$ compared with the value for IL-18 and CGS-21680. When an error bar was within a symbol, the bar was omitted.

agonist, NECA, reversed the action of the A_{2A} receptor agonist, CGS-21680.

The effects of A_{2A} receptor-mediated stimulation on the levels of cAMP in monocytes treated with IL-18

We examined the involvement of the elevation of cAMP in the action of A_{2A} receptor stimulation (Figure 5). IL-18 had no effect on the elevation of intracellular cAMP. Adenosine and the A_{2A} receptor agonist, CGS-21680, but not the A_1 and A_3 receptor agonists, CPA and IB-MECA, induced the elevation of cAMP. The effect of adenosine was reduced by the A_{2A} receptor antagonist, ZM-241385, and it was enhanced by the A_1 and A_3 receptor antagonists, DPCPX and MRS1220. The action of the A_{2A} receptor agonist CGS-21680, was reduced by the A_1 and A_3 receptor agonists, CPA and IB-MECA. Neither the A_{2B} receptor antagonist, alloxazine nor the corresponding agonist, NECA had any effect on A_{2A} receptor stimulation by adenosine. A PKA inhibitor, H89, reversed the inhibitory effect of the A_{2A} receptor agonist, CGS-21680, on the IL-18-elicited expression of ICAM-1 and production of IL-12, TNF- α and IFN- γ (Figure 6).

Discussion and conclusions

We previously reported that blockade of the engagement of ICAM-1 and lymphocyte function-associated antigen (LFA)-1 by anti-ICAM-1 antibody and a statin-derived LFA-1 inhibitor, LFA703, reduced the IL-18-enhanced production of IL-12, TNF- α and IFN- γ by PBMC (Takahashi *et al.*, 2002; 2003; 2005). This suggests that the engagement of ICAM-1 and LFA-1 might play an important role in IL-18-enhanced cytokine production. Here, as shown in Figures 1 and 2, we demonstrate that adenosine inhibited the expression of ICAM-1 and production of IL-12, TNF- α and IFN- γ in the presence of IL-18. This further suggests that adenosine-initiated inhibition of ICAM-1 expression might regulate cytokine production. Adenosine deaminase is known to convert adenosine to inosine by the removal of an amine group. We demonstrated that inosine at concentrations ranging from 0.01 to 100 μ M had no effect on the expression of ICAM-1 and production of IL-12, TNF- α and IFN- γ in the presence of IL-18 (data not shown). Therefore, the actions of adenosine were independent of any conversion to inosine.

Zhang *et al.* (2005) reported the binding affinity profile of agonists and antagonists at human adenosine receptors. Adenosine inhibited monocyte function through activation of A_{2A}, A_{2B} and A₃ receptors (Le Vraux *et al.*, 1993; Bouma *et al.*, 1994; Khoa *et al.*, 2001; Zhang *et al.*, 2005), and, thus, the involvement of adenosine receptor subtypes is controversial. In the present study, we found a complex interaction between adenosine receptor subtypes in mediating the overall effect of adenosine. The inhibitory effect of adenosine was blocked by the A_{2A} receptor antagonist, ZM-241385, and, conversely, the A_{2A} receptor agonist, CGS-21680, mimicked the effect of adenosine (Figures 3 and 4b). Unexpectedly, the inhibitory effect of adenosine was enhanced by the A₁ and A₃ receptor antagonists, DPCPX and MRS1220, respectively (Figure 3). In addition, we demonstrated that the A₁ and A₃ receptor agonists, CPA and IB-

MECA, respectively, reduced the actions of the A_{2A} receptor agonist (Figure 4). Thus, the inhibitory effect of adenosine was owing to the stimulation of A_{2A} receptors and that effect was reduced by stimulation of A₁ and A₃ receptors.

Stimulation through the A_{2A} receptor is known to result in the elevation of cAMP levels and PKA activation in human monocytes (Bshesh *et al.*, 2002). As shown in Figure 5, stimulation via the A_{2A} receptor increased cAMP levels in monocytes, which were again reduced by A₁ and A₃ receptor agonists. We have found that the activation of cAMP inhibited the IL-18-elicited expression of ICAM-1 and the production of IL-12, TNF- α and IFN- γ by PBMC (Takahashi *et al.*, 2002; 2003). A PKA inhibitor, H89 reversed the effect of A_{2A} receptor agonist on the expression of ICAM-1 and production of IL-12, TNF- α and IFN- γ (Figure 6). These results are consistent with the notion that cAMP might be involved in mediating the effects of A_{2A} receptor agonists on the expression of ICAM-1 and production of IL-12, TNF- α and IFN- γ . Moreover, the inhibitory influence of A₁ and A₃ receptor agonists on the effects of A_{2A} receptor agonist might depend on the inhibition of cAMP elevation induced by the latter. However, the expression of B7.2 and CD40 was not changed by adenosine or by the A_{2A} receptor agonist, suggesting that the activation of cAMP was not sufficient for this outcome. Further study on the mechanism of the action of adenosine and A_{2A} receptor agonist will be required to clarify this.

Ohta and Sitkovsky (2001) reported that the stimulation of A_{2A} receptor signaling suppressed the inflammation and inhibited the tissue damage in mouse models of liver injury and endotoxin-induced septic shock. The A_{2A} receptor-initiated inhibition of chemokine and cytokine production also reduced the inflammation and the tissue damage during reperfusion after hepatic ischemia (Harada *et al.*, 2000; Day

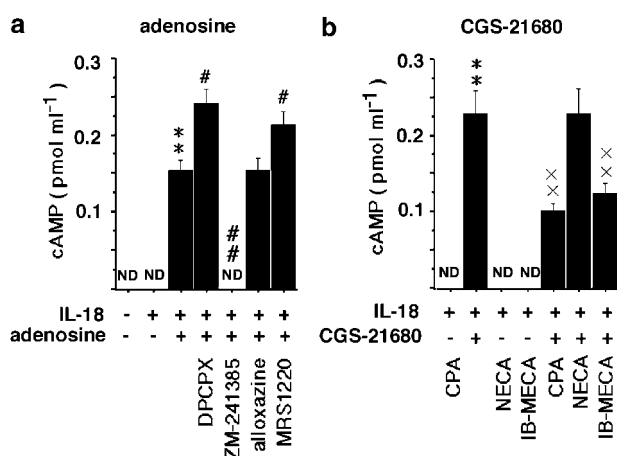


Figure 5 The effects of A_{2A}R stimulation on the levels of cAMP in isolated monocytes treated with IL-18. (a) The effect of adenosine, the selective A₁, A_{2A}, A_{2B} and A₃ receptor antagonists, DPCPX, ZM-241385, alloxazine and MRS1220, respectively, on the production of cAMP in the presence of 100 ng ml⁻¹ IL-18 at 30 min. (b) The selective A₁, A_{2A}, A_{2B} and A₃ receptor agonists, CPA, CGS-21680, NECA and IB-MECA, respectively, at 100 μ M on the production of cAMP in the presence of IL-18. ***P* < 0.01 compared with the value for IL-18 alone. #*P* < 0.05, ##*P* < 0.01 compared with the value for adenosine. ××*P* < 0.01 compared with the value for CGS-21680 and IL-18. The results are the means \pm s.e.m. of triplicate determinations from five donors. ND, not detected.

Table 2 Adenosine receptor antagonists and agonists

	Antagonists	Agonists
A ₁	DPCPX	CPA
A _{2A}	ZM-241385	CGS-21680
A _{2B}	Alloxazine	NECA
A ₃	MRS1220	IB-MECA

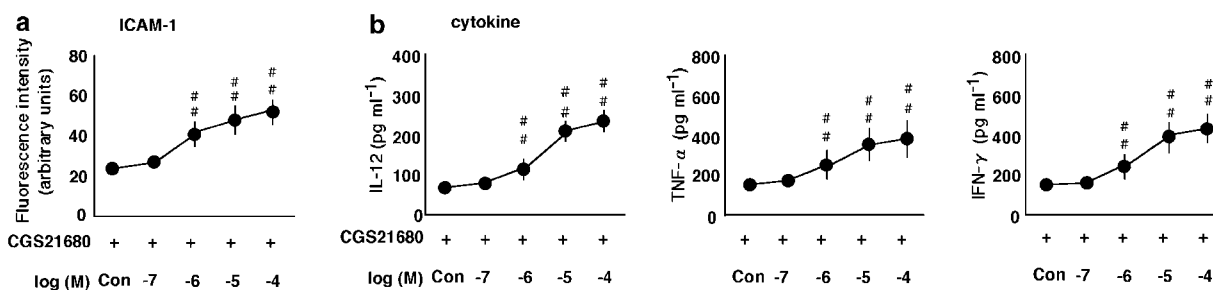


Figure 6 The effect of H89 on CGS-21680-mediated inhibition of IL-18 induced ICAM-1 expression and the cytokine production. (a) The effect of a PKA inhibitor, H89, at concentrations ranging from 0.1 to 100 μ M (Con, no H89) on the expression of ICAM-1 on monocytes in the presence of 100 ng ml⁻¹ IL-18 and 100 μ M of the selective A_{2A} receptor agonist, CGS-21680, at 24 h. PBMC were treated as described and the expression of ICAM-1 on monocytes was examined by multi color flow cytometry using a combination of anti-CD14 antibody with anti-ICAM-1 antibody. (b) The effect of H89 (Con, no H89) on IL-12, TNF- α and IFN- γ production by PBMC in the presence of IL-18 and CGS-21680 after 24 h. Results are expressed as the means \pm s.e.m. of triplicate determinations from five donors. #*P* < 0.05, ##*P* < 0.01 compared with the value for IL-18 and CGS-21680. When an error bar was within a symbol, the bar was omitted.

et al., 2004). Moreover, it has been suggested that adenosine may be useful for treating the chronic inflammatory diseases, including MS and RA (Cronstein *et al.*, 1992; Marabito *et al.*, 1998; Gomez and Sitkovsky, 2003). Adenosine induces bronchoconstriction in animal models and in patients with inflammatory airway diseases such as asthma and chronic obstructive pulmonary diseases (COPD), and adenosine receptors are present in many cell types involved in airway inflammation (Spicuzza *et al.*, 2003). In the lung, A_{2A} receptors activate a protective mechanism playing a critical role in the downregulation of inflammation and tissue damage in different models (Ohta and Sitkovsky, 2001; Thiel *et al.*, 2005). The mechanism of adenosine's actions in these cases may involve IL-18 (Saha *et al.*, 1999; Karni *et al.*, 2002; Yamagata and Ichinose, 2006). The present study may explain, at least in part, the mechanism of action of adenosine on IL-18-initiated diseases, including MS, RA, asthma and COPD. In conclusion, we have demonstrated that adenosine regulates IL-18-enhanced expression of ICAM-1 and the production of IL-12, TNF- α and IFN- γ via the stimulation several subtypes of adenosine receptor.

Conflict of interest

The authors state no conflicts of interest.

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