COMMENTARY

Renal mesangial cells: moving on sphingosine kinase-1

D Meyer zu Heringdorf and KH Jakobs

Institut für Pharmakologie, Universität Duisburg-Essen, Essen, Germany

Sphingosine kinase-1 (SphK1) catalyses the phosphorylation of sphingosine to sphingosine-1-phosphate (S1P), which acts on at least five specific G-protein-coupled receptors and also intracellularly. SphK1 has been implicated in cell proliferation, cancer growth, chemoresistance, immune cell functions and cell migration. In this issue of the British Journal of Pharmacology, Klawitter et al. demonstrate that extracellular nucleotides stimulate the migration of renal mesangial cells. The nucleotides furthermore upregulated SphK1 expression and activity, and this enzyme was required for nucleotide-induced migration. Together with previous findings, these data raise exciting questions: by which mechanism does SphK1 regulate migration in mesangial cells, how is the interplay of purinoceptors and S1P receptors organized in these cells, and how would SphK1-deficient mice respond to kidney damage?

British Journal of Pharmacology (2007) 150, 255–257. doi:10.1038/sj.bjp.0706986; published online 18 December 2006

Keywords: sphingosine-1-phosphate; sphingosine kinase; purinoceptor; cell migration; mesangial cell

Abbreviations: MAPK, mitogen-activated protein kinase; S1P, sphingosine-1-phosphate; SphK, sphingosine kinase

In renal mesangial cells, extracellular nucleotides acting on purine P2Y receptors stimulate several signalling pathways, thereby promoting proliferation (via p42/44 mitogen-activated protein kinase (MAPK)), stimulating prostaglandin release (via phospholipase A_2) and protecting from apoptosis (via Akt) (see Huwiler et al., 2002; Klawitter et al., 2007 and citations therein). In this issue of the British Journal of Pharmacology, Huwiler's group (Klawitter et al., 2007) demonstrates that extracellular adenosine triphosphate and uridine triphosphate stimulate the migration of renal mesangial cells, and they suggest that this might be an important event in the repair mechanism during inflammatory kidney diseases. Furthermore, the nucleotides upregulated expression and activity of sphingosine kinase-1 (SphK1), and this enzyme was essential for nucleotideinduced mesangial cell migration (Klawitter et al., 2007).

Sphingosine kinases catalyse the phosphorylation of sphingosine to sphingosine-1-phosphate (S1P), which is an ubiquitous lipid mediator, regulating cell growth and survival, migration and adhesion (for reviews, see Spiegel and Milstien, 2003; Sanchez and Hla, 2004; Chalfant and Spiegel, 2005). S1P acts on at least five specific G-proteincoupled receptors $(S1P_{1-5})$ and also intracellularly. Recently, the role of S1P in lymphocyte trafficking, angiogenesis and vascular permeability has gained particular attention as the novel immunosuppressant, FTY720, modulates these conditions by interfering with S1P receptors (Brinkmann et al., 2004; LaMontagne et al., 2006). Two sphingosine kinases, SphK1 and SphK2, and transcriptional variants thereof, have been identified in mammals so far. SphK1 and SphK2 are differentially expressed and appear to have opposing cellular functions, with SphK2 acting preferentially as a pro-apoptotic agent, whereas SphK1 has been implicated in cell proliferation, cancer growth, chemoresistance, immune cell functions and cell migration (for reviews, see Spiegel and Milstien, 2003; Taha et al., 2006).

The formation of S1P by sphingosine kinases is a highly regulated process. Both transcriptional and post-transcriptional regulation of SphK1 activity and/or protein have been reported in several cellular systems (for reviews, see Spiegel and Milstien, 2003; Taha et al., 2006). Acute stimulation of sphingosine kinase activity by $P2Y_2$ receptors, leading to S1P formation within a timescale of minutes, has been shown before (Alemany et al., 2000). Klawitter et al. now demonstrate a specific activation of SphK1, but not SphK2, by purine receptors in mesangial cells. Activation of the enzyme was biphasic, with a rapid phase, most likely during to posttranscriptional activation, and a delayed phase of transcriptional upregulation of SphK1 within hours, which was mediated by protein kinase C and MAPK. This was similar, for example, to epidermal growth factor-stimulated SphK1 expression in human breast cancer cells (Döll et al., 2005) and histamine- and hepatocyte growth factor-stimulated SphK1 expression in endothelial cells (Duan et al., 2004;

Correspondence: D Meyer zu Heringdorf, Institut für Pharmakologie, Universitäts-klinikum Essen, Hufelandstrasse 55, Essen 45122, Germany. E-mail: dagmar.meyer-heringdorf@uni-due.de

Received 30 October 2006; accepted 31 October 2006; published online 18 December 2006

Huwiler et al., 2006). SphK1 was essential for nucleotidestimulated migration. This was demonstrated using (1) short interfering RNA (siRNA) to SphK1 and (2) mesangial cells from SphK1-deficient mice. Interestingly, mice deficient in SphK1 were viable and fertile with no obvious dysfunctions (Allende et al., 2004). Therefore, considering the present data, it would be important to study the response to renal damage in these mice, for example by streptozotocin treatment, which in a recent study was found to cause upregulation of sphingosine kinase activity and increased S1P levels within renal glomeruli (Geoffroy et al., 2005).

Another interesting observation was that deletion of SphK1 did not affect nucleotide-induced proliferation of mesangial cells. As discussed by the authors, mesangial cells differ in this regard from many other cell types in which SphK1 is essential for cell growth. SphK1 has furthermore been implicated in induction of cyclooxygenase-2 (reviewed in Taha et al., 2006); therefore, it would be interesting to determine whether purinoceptor-stimulated prostaglandin production is affected by SphK1 deletion.

The differential cellular response to SphK1 deletion raises the intriguing question, how does SphK1 mediate migration of mesangial cells in response to extracellular nucleotides? In several cellular systems, activation of SphK1 leads to autocrine S1P release and activation of G-protein-coupled S1P receptors. This was shown, for example, in mast cells, in which crosslinking of Fce receptor-I stimulated SphK1 activity, S1P secretion and internalization of $S1P_1$ and $S1P_2$ receptors, with $S1P_1$ mediating migration and $S1P_2$ degranulation of the cells in response to antigen (Jolly et al., 2004). SphK1-siRNA had no effect on S1P-induced chemotaxis in this study. In PC12 cells and dorsal root ganglion cells, SphK1 was required for nerve growth factor-induced neurite extension, and SphK1 plasma membrane translocation and $S1P_1$ cross-activation were involved in this process (Toman et al., 2004). Remarkably, Klawitter et al. also showed that deletion of SphK1 abrogated cell migration induced by extracellular S1P. This highly interesting observation suggests that either the SphK1 protein or its catalytic product, S1P or a shift in the ceramide/sphingosine/S1P-balance acted intracellularly for signalling migration. In this scenario, extracellular S1P would then activate and induce SphK1. Indeed, G-proteincoupled S1P receptors can stimulate sphingosine kinase activity (Meyer zu Heringdorf et al., 2001). However, intracellular targets of S1P remain to be identified.

It has been found that G-protein-coupled S1P receptors differentially regulate migration, with $S1P_1$ and $S1P_3$ stimulating and S1P_2 inhibiting this response (Sanchez and Hla, 2004). Therefore, assuming that S1P inside-out signalling is involved in SphK1-dependent migration of mesangial cells, it is possible for the cells to modulate their response to SphK1 upregulation in a specific manner by up- and downregulating S1P receptors. Mesangial cells express the S1P receptors, S1P_1 , S1P_2 and S1P_3 (Katsuma *et al.*, 2002), which mediate increases in intracellular Ca^{2+} concentrations, activation of MAPK, p38, JNK and Akt kinases, crossactivation of transforming growth factor- β receptor and activation of the Smad signalling cascade (see Xin et al. (2004a) and citations therein). Furthermore, S1P stimulated proliferation (via MAPK activation), inhibited apoptosis (Katsuma et al., 2002) and induced migration (Klawitter et al., 2007) in these cells. Interestingly, purinoceptors are known to interfere with S1P receptor signalling in mesangial cells, causing a heterologous desensitization of S1P-stimulated MAPK activation (Xin et al., 2004b). As SphK1 deletion had no effect on purinoceptor-stimulated proliferation, it might be speculated that the heterologous desensitization serves to shut off certain S1P-receptor signalling pathways that otherwise would be activated by inside-out signalling, including MAPK activation and eventually inhibition of migration by S1P₂, thereby facilitating other pathways such as pro-migratory signalling by $S1P_1$ and/or $S1P_3$. Taken together, the data presented by Klawitter et al. demonstrate differential, cell type-specific signalling by SphK1. Together with previous findings, these data raise challenging questions regarding the mechanism of SphK1-mediated migration and the interplay of purine and G-protein-coupled S1P receptors in mesangial cells.

Acknowledgements

The work of the authors is supported by the Deutsche Forschungsgemeinschaft and the Interne Forschungsförderung Essen.

References

- Alemany R, Sichelschmidt B, Meyer zu Heringdorf D, Lass H, van Koppen CJ, Jakobs KH (2000). Stimulation of sphingosine-1 phosphate formation by the P2Y₂ receptor in HL-60 cells: Ca^{2+} requirement and implication in receptor-mediated Ca^{2+} mobilization, but not MAP kinase activation. Mol Pharmacol 58: 491–497.
- Allende ML, Sasaki T, Kawai H, Olivera A, Mi Y, Echten-Deckert G et al. (2004). Mice deficient in sphingosine kinase-1 are rendered lymphopenic by FTY720. J Biol Chem 279: 52487–52492.
- Brinkmann V, Cyster JG, Hla T (2004). FTY720: sphingosine-1 phosphate receptor-1 in the control of lymphocyte egress and endothelial barrier function. Am J Transplant 4: 1019–1025.
- Chalfant CE, Spiegel S (2005). Sphingosine-1-phosphate and ceramide-1-phosphate: expanding roles in cell signaling. J Cell Sci 118: 4605–4612.
- Döll F, Pfeilschifter J, Huwiler A (2005). The epidermal growth factor stimulates sphingosine kinase-1 expression and activity in the human mammary carcinoma cell line MCF7. Biochim Biophys Acta 1738: 72–81.
- Duan HF, Wu CT, Lu Y, Wang H, Liu HJ, Zhang QW et al. (2004). Sphingosine kinase activation regulates hepatocyte growth factor induced migration of endothelial cells. Exp Cell Res 298: 593–601.
- Geoffroy K, Troncy L, Wiernsperger N, Lagarde M, El Bawab S (2005). Glomerular proliferation during early stages of diabetic nephropathy is associated with local increase of sphingosine-1-phosphate levels. FEBS Lett 579: 1249–1254.
- Huwiler A, Döll F, Ren S, Klawitter S, Greening A, Romer I et al. (2006). Histamine increases sphingosine kinase-1 expression and activity in the human arterial endothelial cell line EA.hy 926 by a PKC-*a*-dependent mechanism. Biochim Biophys Acta 1761: 367–376.
- Huwiler A, Rolz W, Dorsch S, Ren S, Pfeilschifter J (2002). Extracellular ATP and UTP activate the protein kinase B/Akt cascade via the $P2Y_2$ purinoceptor in renal mesangial cells. Br J Pharmacol 136: 520–529.
- Jolly PS, Bektas M, Olivera A, Gonzalez-Espinosa C, Proia RL, Rivera J et al. (2004). Transactivation of sphingosine-1-phosphate receptors by FceRI triggering is required for normal mast cell degranulation and chemotaxis. J Exp Med 199: 959–970.
- Katsuma S, Hada Y, Ueda T, Shiojima S, Hirasawa A, Tanoue A et al. (2002). Signalling mechanisms in sphingosine-1-phosphate-promoted mesangial cell proliferation. Genes Cells 7: 1217–1230.
- Klawitter S, Hofmann LP, Pfeilschifter J, Huwiler A (2007). Extracellular nucleotides induce migration of renal mesangial cells by upregulating sphingosine kinae-1 expression and activity. Br J Pharmacol 150: 271–280 (this issue).
- LaMontagne K, Littlewood-Evans A, Schnell C, O'Reilly T, Wyder L, Sanchez T et al. (2006). Antagonism of sphingosine-1-phosphate receptors by FTY720 inhibits angiogenesis and tumor vascularization. Cancer Res 66: 221–231.
- Meyer zu Heringdorf D, Lass H, Kuchar I, Lipinski M, Alemany R, Rümenapp U et al. (2001). Stimulation of intracellular sphingosine-1-phosphate production by G-protein-coupled sphingosine-1-phosphate receptors. Eur J Pharmacol 414: 145–154.
- Sanchez T, Hla T (2004). Structural and functional characteristics of S1P receptors. J Cell Biochem 92: 913–922.
- Spiegel S, Milstien S (2003). Sphingosine-1-phosphate: An enigmatic signalling lipid. Nat Rev Mol Cell Biol 4: 397–407.
- Taha TA, Hannun YA, Obeid LM (2006). Sphingosine kinase: biochemical and cellular regulation and role in disease. J Biochem Mol Biol 39: 113–131.
- Toman RE, Payne SG, Watterson KR, Maceyka M, Lee NH, Milstien S et al. (2004). Differential transactivation of sphingosine-1-phosphate receptors modulates NGF-induced neurite extension. J Cell Biol 166: 381–392.
- Xin C, Ren S, Kleuser B, Shabahang S, Eberhardt W, Radeke H et al. (2004a). Sphingosine 1-phosphate cross-activates the Smad signaling cascade and mimics transforming growth factor-beta-induced cell responses. J Biol Chem 279: 35255–35262.
- Xin C, Ren S, Pfeilschifter J, Huwiler A (2004b). Heterologous desensitization of the sphingosine-1-phosphate receptors by purinoceptor activation in renal mesangial cells. Br J Pharmacol 143: 581–589.