Frequency of cytochrome P450 2C9 mutant alleles in a Korean population

Young-Ran Yoon, Ji-Hong Shon, Moon-Kyung Kim, Young-Chai Lim,¹ Hye-Rang Lee, Ji-Young Park, In-June Cha & Jae-Gook Shin

Department of Pharmacology, Inje University College of Medicine and Clinical Pharmacology Center, Pusan Paik Hospital, ¹Department of Pharmacology, Chonnam National University College of Medicine, Pusan, Kwangju, Korea

> Aims To determine the frequencies of CYP2C9 variants in the Korean population and compare them with the frequencies in other ethnic populations.

> Methods Genotyping of CYP2C9*2 and CYP2C9*3 allelic variants was carried out in 574 Korean subjects by PCR and restriction fragment length pattern analysis.

> Results Thirteen of 574 subjects (2.3%) were heterozygous for CYP2C9*3 (Ile359Leu), but no subjects with a CYP2C9*2 allele or homozygous for $CYP2C9*3$ were identified. The allele frequency of $CYP2C9*3$ in Korean subjects $(0.0113, 95\% \text{ CI } 0.0066-0.0193)$ was similar to that of other East Asian populations, but was considerably lower than that of Caucasian populations.

> **Conclusions** CYP2C9 \star 3 seems to be an allelic variant related to the functional polymorphism of CYP2C9, but this variant is rarely seen among Koreans compared with Caucasians. Routine genotyping of the CYP2C9*2 allele is considered to be unnecessary in Korean and East Asians, because this allele appears to be extremely rare or absent in these populations.

Keywords: CYP2C9, genotype, Korean

Introduction

Cytochrome P450 2C9 (CYP2C9) is a genetically polymorphic enzyme that is involved in the metabolism of phenytoin, S-warfarin, tolbutamide, losartan, torasemide, and many nonsteroidal anti-inflammatory drugs, including diclofenac, ibuprofen, and flurbiprofen [1]. Several different CYP2C9 cDNA sequences have been reported. Two amino acid variants, Arg₁₄₄Cys, which result from a $C_{430} \rightarrow T$ nucleotide substitution in exon 3 $(CYP2C9 \star 2)$, and Ile₃₅₉Leu, produced by an A₁₀₇₅ \rightarrow C substitution in exon 7 (CYP2C9*3) have reduced catalytic activity compared with the wild type $CYP2C9*1$ [2, 3]. The catalytic activity of the CYP2C9*3 encoded enzyme is much lower than those of CYP2C9*1 and CYP2C9*2 [4]. The unbound oral clearance of S-warfarin in vivo was reduced by 66% and 90% among subjects heterozygous

Correspondence: Jae-Gook Shin, MD, PhD, Associate Professor of Pharmacology, Inje University College of Medicine, Clinical Pharmacology Center/IJUPH, #633-165, Kaekum-Dong, Jin-Ku, Pusan 614-735, Korea. Tel.: + 82-51-890-6709; Fax: $+82518937761$; E-mail: phshinjg@ijnc.inje.ac.kr

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and homozygous for the CYP2C9*3 allele, respectively, compared to homozygous CYP2C9*1 subjects [5].

The frequency of CYP2C9 allelic variants has been reported to differ among Caucasian, African, and Asian populations. The allele frequencies of CY2C9*2 and CYP2C9*3 tend to be greater in Caucasian populations than in African-American and Asian populations $[3, 6-8, 1]$ 10]. In contrast to Caucasians and African-Americans, the CYP2C9*2 allelic variant was not found in East Asians including Chinese and Japanese [8, 10]. However, no information on the genotype of CYP2C9 allelic variants is available for Korean populations. Since subjects with CYP2C9 allelic variants are rare, especially in East Asian populations, study population size can affect the accuracy of estimations of the frequency of allelic variants. This is the first report on the frequency of CYP2C9 allelic variants in a Korean population, and is the largest CYP2C9 genotyping study published to date.

Methods

Subjects

A group of 574 unrelated Korean subjects (388 males and 186 females) were recruited for genotyping after obtaining informed written consent. Whole blood (7 ml) was drawn from 480 healthy subjects and 94 outpatients who took phenytoin for epilepsy. The aliquot of whole blood was stored at -70° C until used for CYP2C9 genotyping. This study was approved by the Institutional Review Board of Inje University Pusan Paik Hospital, Pusan, Korea.

Identification of subjects with a mutant $CYP2C9$ allele

The CYP2C9 genotype was determined by PCR-RFLP as described by Nasu et al. [8] with slight modification. DNA was prepared from $600 \mu l$ of whole blood using a standard phenol/chloroform extraction method [9] and PCR was performed in 20 μ l volumes with 1.5 mm MgCl₂, 1×PCR buffer (TaKaRa Shuzo Co., Ltd, Shiga, Japan), 200 µm dNTP (TaKaRa Shuzo Co., Ltd), 10 pmol of the forward and reverse primers for CYP2C9*2 or CYP2C9*3, 1 U of Taq DNA polymerase (Amersham Pharmacia Biotech, Uppsala, Sweden), and 60 ng of genomic DNA. The sequences of the forward and reverse primers used were 5'-GTA TTT TGG CCT GAA ACC CAT A-3' and 5'-GGC CTT GGT TTT TCT CAA CTC-3' and for the CYP2C9*2 genotype, and 5'-TGC ACG AGG TCC AGA GGT AC-3' and 5'-ACA AAC TTA CCT TGG GAA TGA GA-3' for the CYP2C9*3 genotype. PCR amplification to detect CYP2C9*2 was performed using a Gene Amp PCR System 2400 (Perkin Elmer, Foster City, CA, USA) with an initial denaturation at 94° C for 5 min, followed by 30 cycles of denaturation at 94° C for 45 s, annealing at 64° C for 45 s, and extension at 72° C for 1 min. A final 5-min extension at 72° C followed. To genotype CYP2C9*3, 35 PCR cycles were used and the annealing temperature was 60° C. Aliquots of each PCR product (15 μ) were digested with restriction enzymes (AvaII for CYP2C9*2, KpnI for CYP2C9*3, Boehringer Mannheim GmbH, Mannheim, Germany) at 37° C for 1.5 h. The DNA fragments were separated by 2.5% agarose gel electrophoresis and were detected by ethidium bromide staining. Alleles for $Tyr_{358}Cys$ and $Gly_{417}Asp$ were also determined using polymerase chain reaction with restriction fragment length polymorphism as described by Nasu et al. [8]. Because of absence of these alleles in 310 Korean subjects screened, no further screening of these alleles were continued to other subjects.

Statistical analysis

Data were compiled according to the genotype and allele frequencies were estimated from the observed numbers of each specific allele. The population frequency of each allele is given together with the 95% confidence intervals.

Table 1 Frequency distribution of the Ile359Leu variants in CYP2C9 observed in the 574 Korean subjects.

	(He359He)	$CYP2C9*1/*1$ $CYP2C9*1/*3$ $CYP2C9*3/*3$ (He359Leu)	(Leu359Leu)	Total
Observed	561	13	$\left(\right)$	574
number	(97.7%)	(2.3%)		
Expected	561.1	12.83	0.07	574
$number^{\star}$	(97.75%)	(2.24%)	(0.01%)	

*Calculated by the Hardy-Weinberg law. *One subject homozygous for CYP2C9*3 is expected in every 7,700 Korean subjects.

Results

No mutant CYP2C9*2 allele was found in any of the 574 Korean subjects genotyped in this study. In addition, no subject homozygous for CYP2C9*3 was found. Only 13 subjects (2.3%) heterozygous for $CYP2C9 \star 3$ (Ile₃₅₉leu) were identified (Table 1). Using the Hardy-Weinberg law, the frequency of the homozygous CYP2C9*3 genotype (q^2) was estimated to be 0.013% (about 1 of 7700 subjects) in the Korean population. The allele frequency of the CYP2C9*3 in the 574 Korean subjects was 0.0113 (95% CI 0.0066-0.0193, Table 2).

Discussion

In this study, no subjects with a mutant CYP2C9*2 allele were identified in 574 Korean subjects. This is consistent with the previous studies of the CYP2C9 genotype in other East Asian subjects, including Japanese [8], Taiwan Chinese [3], and Han Chinese [10] populations. These results suggest that the CYP2C9*2 genotype is absent or at least very rare in Koreans and other East Asian populations, and routine genotyping for this mutant allele is not necessary to determine the effects of this variant on catalytic activity of CYP2C9 in these Asian populations. In addition, many authors have failed to find $Tyr_{358}Cys$ and Gly417Asp, other reported allelic variants of CYP2C9, in Asian or Caucasian subjects [3, 8, 10, 11]. In our study, we tested 310 of the 574 Korean subjects for the Tyr₃₅₈Cys and Gly₄₁₇Asp alleles and did not find subjects with these mutant alleles (data not shown), suggesting that these allelic variants are not involved in the genetic polymorphism of CYP2C9.

For the CYP2C9*3 genotype, 13 subjects (2.3%) were heterozygous for the Ile₃₅₉Leu allele, but none of the 574 Korean subjects was homozygous for this allele. The frequencies of the Ile₃₅₉ and Leu₃₅₉ alleles in this Korean population were 0.989 and 0.011, respectively. This result was in good accordance with an expected frequency of 0.011 for the Leu₃₅₉ allele, calculated by the

Table 2 Comparison of allele frequencies of CYP2C9 reported from different ethnic populations.

Population	$n*$	Ile359	Leu359	$n*$	Arg144	Cys144	Reference
Asian							
Korean	1148	0.987 $(0.981 - 0.993)$	0.011 $(0.007 - 0.019)$	620		$\overline{0}$	Present study
Japanese	436	0.979 $(0.966 - 0.992)$	0.021 $(0.008 - 0.034)$	436	$\mathbf{1}$	$\overline{0}$	[8]
Han Chinese	230	0.983 $(0.956 - 0.993)$	0.017 $(0.007 - 0.044)$	106	1	$\overline{0}$	$[10]$
Taiwan Chinese				164		θ	$[10]$
Taiwan Chinese	196	0.974 $(0.942 - 0.989)$	0.026 $(0.011 - 0.058)$	196		θ	$[3]$
Caucasian							
American	200	0.940 $(0.907 - 0.973)$	0.060 $(0.027 - 0.093)$	200	0.920 $(0.882 - 0.958)$	0.080 $(0.042 - 0.118)$	$[3]$
British	200	0.915 $(0.876 - 0.954)$	0.085 $(0.046 - 0.124)$	200	0.875 $(0.829 - 0.921)$	0.125 $(0.079 - 0.171)$	$[13]$
Swedish	860	0.926 $(0.906 - 0.941)$	0.074 $(0.056 - 0.091)$	860	0.893 $(0.872 - 0.913)$	0.107 $(0.086 - 0.127)$	$[7]$
Turkish	998	0.900 $(0.880 - 0.917)$	0.100 $(0.079 - 0.123)$	998	0.894 $(0.873 - 0.911)$	0.106 $(0.089 - 0.127)$	$[12]$
African							
African-American	200	0.995 $(0.985 - 1.005)$	0.005 $(-0.005 - 0.015)$	200 $(0.976 - 1.004)$	0.990 $(-0.004 - 0.024)$	0.010	$\lceil 3 \rceil$

*Number of alleles studied.

Hardy-Weinberg equation (Table 1). On average one subject homozygous for CYP2C9*3 is expected in every 7700 Korean subjects. This finding is similar to that for a Japanese population, in which the frequency of subjects homozygous for CYP2C9*3 allele is also expected to be extremely low (about 1 in 2200) [8]. No subject homozygous for $CYP2C9*3$ was identified in our 574 Korean subjects or in any other East Asian subjects, including 218 Japanese, 98 Taiwan Chinese and 115 Han Chinese [3, 8, 10]. Unlike East Asian populations, several subjects homozygous for $CYP2C9*3$ have been identified in Caucasian and Turkish populations [3, 6, 7, 12].

The allele frequency of CYP2C9*3 among 574 Korean subjects was 0.011, a value slightly lower than in Japanese (0.021) [8] and in Chinese $(0.017-0.026)$ [3, 10], but higher than in African-Americans (0.005) [3] (Table 2). The allele frequencies of CYP2C9*3 in Koreans and East Asians (0.015, a value pooled from Korean, Japanese, and Chinese populations) [present study, 3, 8, 10] is considerably lower than in Caucasians (0.085, a value pooled from American, British, Swedish, and Turkish populations) [3, 7, 12, 13].

The allele frequency of CYP2C9*2 in Caucasian populations is 0.106 (a value pooled from references 3, 7, 12, 13). In contrast, no subject with a CYP2C $9*2$ allele has been found in 761 East Asian subjects from four different studies [present study 3, 8, 10].

In summary, CYP2C9*3, an allelic variant related to the functional polymorphism of CYP2C9, is relatively rare in Koreans and seems to occur at a lower frequency than in Caucasians. The CYP2C9*2 variant appears absent in East Asians including Koreans or present at an even lower frequency than the $CYP2C9*3$ allele, suggesting that routine genotyping for the CYP2C9*2 allele is unnecessary in these populations.

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References

- 1 Miners JO, Birkett DJ. Cytochrome P450 2C9: an enzyme of major importance in human drug metabolism. Br J Clin Pharmacol 1998; 45: 525-538.
- 2 Rettie AE, Wienkers LC, Gonzalez FJ, Trager WF, Korzekwa KR. Impaired S-warfarin metabolism catalyzed by R144C allelic variant of CYP2C9. Pharmacogenetics 1994; 4: 39-42.
- 3 Sullivan-Klose TH, Ghanayem BI, Bell DA, et al. The role of the CYP2C9-Leu³⁵⁹ allelic variant in the tolbutamide polymorphism. Pharmacogenetics 1996; 6: 341-349.
- Yamazaki H, Inoue K, Chiba K, et al. Comparative studies on the catalytic roles of Cytochrome P450 2C9 and its Cys- and Leu-variants in the oxidation of warfarin, flurbiprofen, and diclofenac by human liver microsomes. Biochem Pharmacol 1998; 56: 243-251.
- 5 Takahashi H, Kashima T, Nomoto S, et al. Comparisons between in-vitro and in-vivo metabolism of (S)-warfarin: catalytic

activities of cDNA-expressed CYP2C9, its Leu₃₅₉ variant and their mixture versus unbound clearance in patients with the corresponding CYP2C9 genotypes. Pharmacogenetics 1998; 8: 365±373.

- 6 London SJ, Daly AK, Leathart JB, Navidi WC, Idle JR. Lung cancer risk in relation to the CYP2C9*1/CYP2C9*2 genetic polymorphism among African-Americans and Caucasians in Los Angeles County, California. Pharmacogenetics 1996; 6: $527 - 533$.
- 7 Yasar U, Eliasson E, Dahl M-L, Johansson I, Ingelman-Sundberg M, Sjöqvist F. Validation of methods for CYP2C9 genotyping: frequencies of mutant alleles in a Swedish population. Biochem Biophys Res Comm 1999; 254: 628±631.
- 8 Nasu K, Kubota T, Ishizaki T. Genetic analysis of CYP2C9 polymorphism in a Japanese population. Pharmacogenetics 1997; 7: 405±409.
- 9 Blin N, Stafford DW. A general method for isolation of high molecular weight DNA from eukaryotes. Nucl Acid Res 1976; 3: 2303±2308.
- 10 Wang SL, Huang J, Lai MD, Tsai JJ. Detection of CYP2C9 polymorphism based on the polymerase chain reaction in Chinese. Pharmacogenetics 1995; 5: 37-42.
- 11 Kimura M, Ieiri I, Mamiya K, Urae A, Higuchi S. Genetic polymorphism of cytochrome P450s, CYP2C19, and CYP2C9 in a Japanese population. Ther Drug Monit 1998; 20: 243±247.
- 12 Aynacioglu AS, Brockmöller J, Bauer S, et al. Frequency of cytochrome P450 CYP2C9 variants in a Turkish population and functional relevance for phenytoin. Br J Clin Pharmacol 1999; 48: 409-415.
- 13 Stubbins MJ, Harries LW, Smith G, Trabit MH, Wolf CR. Genetic analysis of the cytochrome P450 CYP2C9 locus. Pharmacogenetics 1996; 6: 429-439.