THE LEUCOCYTE ADHERENCE INHIBITION TEST IN CANCER OF THE LARGE BOWEL

P. R. ARMITSTEAD AND G. GOWLAND

From the University of Leeds, Department of Immunology, The General Infirmary, Leeds, LS1 3EX

Received 27 June 1975. Accepted 28 July 1975

Summary.—A recently introduced assay of cell mediated immunity and humoral inhibitory factors has been evaluated in colorectal cancer patients. Using a perchloric acid extract of adenocarcinoma of the large bowel as antigen, 16/27 patients with colorectal cancer had significant cellular reactivity when their separated peripheral leucocytes were tested in homologous AB serum. In autologous serum only 7/27 had significant reactivity; 6/20 patients with a variety of other malignancies showed sensitization to the colorectal antigen preparation.

It is concluded that the leucocyte adherence inhibition test may offer a simple method of assaying for serum blocking factors in sequential studies but will be of little value in the diagnosis of large bowel cancer.

In vitro techniques for assessing immunological reactivity have facilitated the study of the immune response to human cancer and of factors which modify its expression. Such information is of much more than theoretical importance as it could provide guidelines for planning suitable immunotherapy or monitoring the post-operative clinical course of patients.

Lymphocyte cytotoxicity (Hellström et al., 1971a), lymphocyte transformation (Vanky et al., 1971) and leucocyte migration inhibition techniques (Anderson et al., 1970; Guillou and Giles, 1973) have been used to show specific cell mediated immunity to tumour associated antigens in a variety of human malignancies. These tests are, however, time consuming, may require complex equipment and are often difficult to interpret.

Recently, the leucocyte adherence inhibition (LAI) test has been described by Halliday, Maluish and Isbister (1974a) as both a simple and rapid in vitro test of cellular reactivity in human cancer. The test was first used for experimental murine tumours (Halliday, Maluish and Isbister, 1974b) and then adapted for

human malignancies (Maluish and Halliday, 1974).

The principle of the test is that when specifically sensitized lymphocytes are incubated with the antigens, a lymphokine is released which decreases the normal tendency of leuocytes to adhere to a glass surface. This is analogous to the macrophage migration system in which a product of antigen lymphocyte interaction causes inhibition of macrophage migration.

To date the LAI test has been used only to investigate small groups of patients with a specific cancer. The present study was an attempt to evaluate this potentially useful test in a group of 27 patients with colorectal cancer.

PATIENTS AND METHODS

Twenty-seven patients (mean age 65.6 years) with primary adenocarcinoma of the large bowel were studied pre-operatively. These were compared with 19 control subjects (12 healthy volunteers and 7 patients with benign disease, mean age 57.1 years) and 20 patients (mean age 68.2 years) with various other malignancies.

A perchloric acid extract of surgically resected adenocarcinomata of large bowel was

prepared after the method of Freed and Taylor (1972). None of the tumours used for this extract were autochthonous with respect to the patients studied. The tumour extract was sterile on culture and furthermore an antiserum raised against the extract failed to agglutinate the common enteric bacteria. As used in the LAI test this extract contained 10 µg protein/ml (Kjeldahl).

The technique of the LAI test was essentially as originally described (Halliday et al., 1974a). Leucocytes were obtained by sedimentation for 1 h at 37 °C from heparinized blood, any remaining red cells being lysed by brief treatment with 0·15 mol/l ammonium chloride. After washing, the leucocytes were finally resuspended to a concentration of $2 \times 10^7/\text{ml}$ in tissue culture medium 199 (Wellcome) with 10% foetal calf serum added.

Equal volumes (0.05 ml) of the leucocyte suspension, tumour extract (when required) and undiluted pooled AB or autologous serum were mixed with TC 199 + 10% foetal calf serum, to give a final volume of 0.2 ml. Each of the mixtures was set up in duplicate. The tubes were randomized, coded and then incubated at 37°C for 30 min with constant mixing. In view of the short incubation time in this assay, antibiotics were not added to the culture medium.

Leucocyte adherence was then determined by introducing each mixture into the 2 chambers of a haemacytometer (Optical

American Corporation). After a further 60 min incubation, the nucleated cells in a predetermined pattern of squares (16 squares/haemacytometer) were counted. The coverslips were then floated off and slides subjected to a gentle washing procedure; the remaining, adherent cells were then counted in the same squares.

The mean percentage adherence for each mixture was determined and a leucocyte adherence index calculated.

Leucocyte adherence index

 $= \frac{\text{Mean \% adherence with antigen}}{\text{Mean \% without antigen}}$

RESULTS

The mean percentage adherence for each mixture and the leucocyte adherence indices for the control subjects, colorectal and other malignancies are shown in Tables I, II and III respectively.

The control subjects exhibited little or no reduced adherence in the presence of the tumour extract either in homologous AB serum (mean index $1\cdot024\pm0\cdot0547$ s.d.) or autologous serum (mean index $1\cdot0421\pm0\cdot0866$ s.d.). As there was no significant difference between the healthy subjects and the patients with benign disease, they have been treated as one group. No nonspecific or toxic effects of

Table I.—Results of LAI Test in Control Patients

Patients			Me	an % adher	Leucocyte adherence index		
Age	Sex	Diagnosis	AB serum	${ m AB \ serum} \ + { m antigen}$	Auto serum +antigen		Auto
30	\mathbf{M}	$\mathbf{Healthy}$	53	49	47	$0 \cdot 92$	$0 \cdot 9$
61	\mathbf{M}	$\mathbf{Healthy}$	73	74	71	$1 \cdot 01$	0.98
49	\mathbf{M}	Healthy	57	64	63	$1 \cdot 12$	$1 \cdot 1$
69	\mathbf{M}	Healthy	62	66	57	$1 \cdot 06$	$0 \cdot 92$
74	\mathbf{M}	Healthy	$46 \cdot 5$	47	54	$1 \cdot 01$	$1 \cdot 16$
45	\mathbf{F}	Healthy	59	60	72	$1 \cdot 02$	$1 \cdot 22$
52	\mathbf{M}	Healthy	75	71	79	0.95	$1 \cdot 05$
53	\mathbf{F}	Healthy	$67 \cdot 5$	$70 \cdot 5$	$71 \cdot 5$	$1 \cdot 04$	$1 \cdot 06$
58	\mathbf{F}	Healthy	70	71	67	$1 \cdot 01$	$0 \cdot 96$
61	\mathbf{F}	Healthy	$65 \cdot 5$	$63 \cdot 5$	69	0.97	$1 \cdot 05$
55	\mathbf{F}	Healthy	$71 \cdot 5$	$71 \cdot 3$	76	0.99	$1 \cdot 06$
59	\mathbf{F}	Healthy	63	61	69	0.97	$1 \cdot 09$
75	M	Urethral stricture	44	48	50	$1 \cdot 09$	$1 \cdot 13$
40	M	Spontaneous pneumothorax	69	71	67	$1 \cdot 03$	$0 \cdot 97$
71	M	Angina	56	64	57	$1 \cdot 14$	$1 \cdot 07$
45	M	Gastric ulcer	57	60	52	$1 \cdot 05$	$0 \cdot 91$
66	M	Diverticulitis	51	55	52	$1 \cdot 07$	$1 \cdot 02$
72	M	Osteoarthritis	59	58	68	$1 \cdot 02$	$1 \cdot 15$
5 0	M	Fissure-in-Ano	73	72.5	73	$0 \cdot 99$	$1 \cdot 0$

Table II.—Results of LAI Test in Patients with Colorectal Cancer

Patients				Mean % adherence			Leucocyte adherence index	
Age	Sex	Diagnosis	Stage*	AB serum	${ m AB\ serum}\ +{ m antigen}$	Auto serum		Auto
45	\mathbf{F}	Ca colon	D	$73 \cdot 5$	36	31	$0 \cdot 49$	$0 \cdot 43$
52	M	Ca colon	$\bar{\mathbf{D}}$	62	69	65	1.1	$1 \cdot 03$
68	\mathbf{F}	Ca colon	\mathbf{D}	58	74	43	$1 \cdot 27$	0.74
76	M	Ca rectum	$\overline{\mathbf{D}}$	79	70	59	0.85	0.75
69	M	Ca rectum	D	46	48	67	$1 \cdot 04$	$1 \cdot 45$
67	M	Ca rectum	$\overline{\mathbf{c}}$	72	50	30	0.69	0.41
80	F	Ca colon	$\dot{\mathbf{c}}$	62	50	78	0.8	$1 \cdot 26$
65	M	Ca colon	$\dot{\mathbf{C}}$	58	49	49	0.84	0.84
58	\mathbf{F}	Ca colon	Ď	66	57	84	0.86	$1 \cdot 26$
49	M	Ca rectum	\mathbf{c}	69	54	72	0.78	$1 \cdot 05$
69	M	Ca rectum	Ď	87	64	84	$0 \cdot 74$	0.97
78	M	Ca rectum	\mathbf{C}	75	74	84	0.99	$1 \cdot 12$
53	\mathbf{F}	Ca colon	Ď	80	77	81	0.96	$1 \cdot 01$
70	M	Ca colon	A	55	58	57	1.05	$1 \cdot 03$
68	\mathbf{F}	Ca colon	\mathbf{c}	57	42	60	$0 \cdot 74$	$1 \cdot 05$
77	\mathbf{F}	Ca rectum	В	66	55	77	0.83	$1 \cdot 16$
65	M	Ca rectum	D	48	42	38	0.87	0.81
73	\mathbf{M}	Ca rectum	D	67	62	79	$0 \cdot 93$	$1 \cdot 17$
53	M	Ca colon	\mathbf{B}	68	59	64	0.87	0.94
79	F	Ca rectum	\mathbf{A}	41	44	44	$1 \cdot 07$	$1 \cdot 07$
67	M	Ca colon	${f B}$	70	77	67	1.1	0.96
54	\mathbf{M}	Ca colon	D	64	54	65	$0 \cdot 78$	$1 \cdot 01$
67	\mathbf{M}	Ca colon	\mathbf{D}	53	41	43	$0 \cdot 76$	0.81
71	\mathbf{F}	Ca colon	\mathbf{c}	52	55	62	$1 \cdot 05$	$1 \cdot 19$
68	\mathbf{F}	Ca colon	\mathbf{D}	83	73	84	0.88	$1 \cdot 02$
62	${f F}$	Ca rectum	${f A}$	50	48	67	$0 \cdot 97$	$1 \cdot 3$
69	\mathbf{M}	Ca rectum	В	69	56	65	0.81	0.94

Table III.—Results of LAI Test in Patients with Other Malignancies

Patients				M	ean % adhe	rence	T		
A	Sex		St*	AB		Auto serum	·	adherence index	
\mathbf{Age}		Diagnosis	Stage*	\mathbf{serum}	+antigen	+antigen	\mathbf{AB}	Auto	
66	M	Seminoma	${f L}$	72	68	65	$1 \cdot 07$	$1 \cdot 05$	
75	\mathbf{F}	Ca breast	M	71.5	59	83	0.8	$1 \cdot 1$	
62	\mathbf{F}	Ca thyroid	\mathbf{M}	49	47	64	0.96	$1 \cdot 3$	
57	\mathbf{M}	Ca lung	\mathbf{M}	86	64	51	0.74	0.59	
58	\mathbf{F}	Rectal melanoma	\mathbf{M}	83	75	88	0.90	$1 \cdot 06$	
55	\mathbf{F}	Ca breast	M	89	85	88	0.96	$0 \cdot 99$	
63	\mathbf{M}	Ca lung	${f L}$	86	85	91	0.99	$1 \cdot 06$	
67	\mathbf{M}	Ca lung	${f L}$	52	51	55	0.99	$1 \cdot 06$	
75	M	Ca oesophagus	M	52	52	59	1.0	$1 \cdot 13$	
83	\mathbf{M}	Ca stomach	M	88	81	80	$0 \cdot 92$	$0 \cdot 91$	
69	\mathbf{F}	Ca stomach	\mathbf{M}	77	76	79	0.99	$1 \cdot 03$	
74	\mathbf{F}	Ca cervix	${f L}$	$62 \cdot 5$	66	76	$1 \cdot 05$	$1 \cdot 2$	
73	\mathbf{M}	Ca prostate	M	77	77.5	81	$1 \cdot 0$	$1 \cdot 05$	
73	\mathbf{F}	Ca stomach	\mathbf{M}	81	68	74	0.83	0.91	
74	\mathbf{F}	Anal melanoma	\mathbf{M}	69	58	43	0.83	$0 \cdot 63$	
67	\mathbf{F}	Ca biliary tract	M	73	57	45	0.8	0.61	
75	\mathbf{F}	Ca stomach	M	65	70	60	$1 \cdot 07$	$0 \cdot 92$	
81	\mathbf{F}	Ovarian ca	M	$78 \cdot 5$	79	$77 \cdot 5$	$1 \cdot 00$	0.99	
73	\mathbf{F}	Ca breast	${f L}$	82	79	83	0.96	$1 \cdot 01$	
73	\mathbf{M}	Ca bladder	${f L}$	68	65	78	0.96	$1 \cdot 15$	

^{*} L = Localized; M = Metastatic.

 $[\]begin{array}{l} {}^*A = \text{Confined to bowel.} \\ {} B = \text{Invading para-colonic tissues, nodes} - \text{ve.} \\ {} C = \text{Involvement of lymph nodes.} \\ {} D = \text{Distant metastases.} \end{array}$

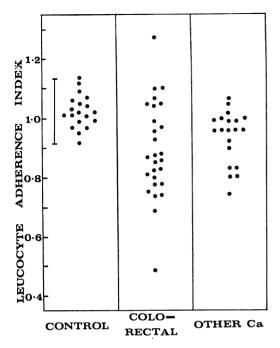


Fig. 1.—Leucocyte adherence inhibition indices of separated peripheral leucocytes from patients with colorectal cancer, various other carcinomata and control subjects when exposed to a perchloric acid extract of colorectal tumour tissue in pooled homologous AB serum. Vertical line indicates the mean $\pm 2 \times \text{s.d.}$ of results obtained from the control group.

the tumour extract were apparent in any of these tests.

Therefore, in this study, significantly reduced leucocyte adherence in the presence of the tumour extract is said to have occurred when the index falls below the 95% confidence limits (mean -2 s.d.) calculated from all the control subject observations. The results obtained in AB serum are shown in Fig. 1 and those in autologous serum in Fig. 2.

In the patients with colorectal cancer of all stages 16/27 ($59\cdot25\%$) showed a significantly reduced index in homologous AB serum, whereas in autologous serum only 7(26%) showed a significantly reduced index. Of the 20 patients with other malignancies, 6(30%) showed a significantly reduced index in homologous AB serum and 3(15%) in their autologous

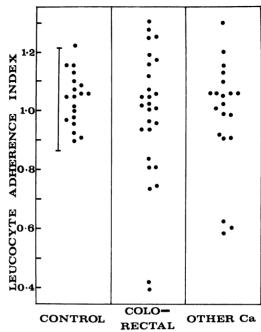


Fig. 2.—Leucocyte adherence inhibition indices of separated peripheral leucocytes from patients with colorectal cancer, various other carcinomata and control subjects when exposed to a perchloric acid extract of colorectal tumour tissue in autologous serum. Vertical line indicates the mean $\pm 2 \times \text{s.d.}$ of results obtained from the control group.

serum. The patients showing the sensitization in this group had metastatic cancer of the breast, lung, stomach, rectal melanoma (2) or biliary tracts. Patients without metastases were uniformly negative.

DISCUSSION

This study has confirmed that the interaction between human leucocytes from patients with colorectal cancer and an extract of pooled colorectal tumours causes a reduced adherence of the cells to a glass surface. This reduced adherence does not appear to be due to toxicity of the tumour extract since adherence of the control subjects' leucocytes was not affected. Similarly, as 4 multiparous female control subjects showed no reduced adherence, presensitization to alloantigens

seems unlikely. As the assay mixtures were incubated only for a short time, *in vitro* sensitization by histocompatibility antigens possibly present in the extracts can be excluded.

Unfortunately the LAI test as performed here appears to be of little use in the positive diagnosis of large bowel cancer since only 16 of the 27 patients studied exhibited significant reactivity to the colorectal tumour extract. A different type of extract may increase the sensitivity but it is of interest that similar proportions of patients are reported to show sensitization to tumour associated antigens in large bowel malignancy and other cancers by the leucocyte migration technique (Andersen et al., 1970; Guillou and Giles, 1973).

No difference in reactivity in homologous AB serum was seen between patients whose tumours were locally confined (Dukes Stage A, B and C) and those with distant spread (Stage D). This clinical-pathological staging, although useful, probably does not truly represent the natural history of the disease. However, 4/4 with histologically de-differentiated tumours showed reactivity, 6/11 with moderately differentiated tumours and 6/12 with well differentiated growths.

A humoral inhibitory effect of autologous sera was present in 10/16 patients who showed reactivity when tested in homologous AB serum. This is analogous to the serum blocking factors reported using the lymphocyte cytotoxicity techniques (Hellström et al., 1971b) and might represent one means by which the cellular immunological effects of sensitized leucocytes might be inhibited in vivo in the cancer patient. In our opinion, it is as an assay for serum blocking that the LAI test has its greatest potential use, particularly in sequential studies of postexcision patients, as the appearance of blocking factors is reported to herald recurrent growth.

The LAI test is claimed to be a rapid and simple *in vitro* method of detecting sensitization to tumour associated antigens (Maluish and Halliday, 1974). In our experience the tedious visual cell counting and technical dexterity necessary for consistent results might limit its usefulness. The modification of the test by a radioisotope technique, recently described by Peirce and Devald (1974) in animals, could overcome these difficulties if successfully applied to man.

In this study the tumour antigen preparation was extracted with perchloric acid, an established method for preparing carcinoembryonic antigen (CEA). Indeed, our extract has been shown to contain CEA, among other antigens, by the use of heterologous antiserum and a purified preparation (Medical Research Council, CEA C73/601). The adherence inhibition was specific for patients with tumours but not organ specific. other carcinomata which were reactive were of types which can produce elevated serum CEA levels (Pusztaszeri and Mach, 1972). It is of interest to speculate on its immunogenic role, although Lejlenvi, Freedman and Gold (1971) found no evidence for a cell mediated immune reaction to purified CEA using a lymphocyte transformation technique. ström, Hellström and Shepard (1970) demonstrated that lymphocytes from patients with carcinoma of the colon, which reacted specifically with colon cancer cells in vitro, were also cytotoxic to foetal gut and liver epithelial cells; also, a delayed skin reaction to extracts of foetal human gut tissue containing CEA was found in patients with carcinoma of the colon and rectum (Hollinshead et al., 1970), however purified CEA elicited no skin reaction in the one patient tested. $\overline{ ext{between}}$ The relationship embryonic macromolecules found in tumour tissue and antigens involved in anti-tumour immunity still remain to be resolved.

We thank the surgeons of Leeds (St James's) University Hospital for allowing us to study patients under their care and also the Medical Research Council for their gift of CEA.

REFERENCES

- Andersen, V., Bjerrum, O., Bendixen, G., Schiødt, T. & Dissing, I. (1970) Effect of Autologous Mammary Tumour Extracts on Human Leukocyte Migration in vitro. Int. J. Cancer, 5, 357.
- Cancer, 5, 357.

 FREED, D. L. G. & TAYLOR, G. (1972) Carcinoembryonic Antigen in Facces. Br. med. J., i, 85.
- Guillou, P. J. & Gilles, G. R. (1973) Inhibition of Leucocyte Migration by Tumour Associated Antigens of Colon and Rectum. Gut, 14, 733.
- HALLIDAY, W. J., MALUISH, A. & ISBISTER, W. H. (1974a) Detection of Anti-tumour Cell-mediated Immunity and Serum Blocking Factors in Cancer Patients by the Leucocyte Adherence Inhibition Test. Br. J. Cancer, 29, 31.
- HALLIDAY, W. J., MALUISH, A. & ISBISTER, W. H. (1974b) Blocking and Unblocking of Cell-mediated Immunity in Mice as detected by the L.A.I. Test. Cell. Immun., 10, 467.
- Hellström, I., Hellström, K. E. & Shepard, T. H. (1970) Cell-mediated Immunity against Antigens Common to Human Colonic Carcinomas and Fetal Gut Epithelium. Cancer, N.Y., 6, 346.
- HELLSTRÖM, I. E., HELLSTRÖM, K. E., SJÖGREN, H. O. & WARNER, G. A. (1971a) Demonstration of Cell Mediated Immunity to Human Neoplasms of Various Histological Types. *Int. J. Cancer*, 7, 1. HELLSTRÖM, I., SJÖGREN, H. O., WARNER, G. &

- HELLSTRÖM, K. E. (1971b) Blocking of Cellmediated Tumour Immunity by Sera from Patients with growing Neoplasms. *Int. J. Cancer*, **7**, 226.
- HOLLINSHEAD, A., GLEW, D., BUNNAG, B., GOLD, P. & HERBERMAN, R. (1970) Skin Reactive Soluble Antigen from Intestinal Cancer-cell-membranes and Relationship to Carcinoembryonic Antigens. *Lancet*, i, 1191.
- LEJLENYI, M. C., FREEDMAN, S. O. & GOLD, P. (1971) Response of Lymphocytes from Patients with Gastro-intestinal Cancer to CEA of the Human Digestive System. *Cancer*, N.Y., 28, 115.
- MALUISH, A. & HALLIDAY, W. G. (1974) Cell Mediated Immunity and Specific Serum Factors in Human Cancer: The LAI Test. J. natn. Cancer Inst., 52, 1415.
- PIERCE, G. E. & DEVALD, B. L. (1974) Leucocyte Adherence Inhibition as Measured by a Radioisotope Technique for Detection of Cell-mediated Tumour Immunity. *Int. J. Cancer*, **14**, 833.
- Pusztaszeri, G. & Mach, J. P. (1972) Tumour Immunity. Carcinoembryonic Antigen in Nondigestive Cancerous and Normal Tissues. *Im*munochemistry, 10, 197.
- VANKY, F., STJERNSWARD, G., KLEIN, G. & NILSONNE, V. (1971) Serum Mediated Inhibition of Lymphocyte Stimulation by Autochthonous Human Tumors. J. natn. Cancer Inst., 47, 95.