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RAPID DIAGNOSIS OF PNEUMOCOCCAL PNEUMONIA AMONG HIV-INFECTED ADULTS WITH URINE ANTIGEN DETECTION

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Abstract

Objectives—*Streptococcus pneumoniae* is the leading cause of bacterial pneumonia and associated bacteremia during HIV infection. Rapid diagnostic assays may limit inappropriate therapy.

Methods— Clinical signs and symptoms and sera and urine were collected prospectively from 70 adults with pneumococcal pneumonia, including 47 with HIV co-infection. Pneumococcal C-polysaccharide antigen was detected in urine using the Binax[®] immunochromatographic test (ICT). A systematic review of 24 published studies was conducted.

Results— Clinical symptoms, signs, and laboratory parameters except leukocytosis, were similar in HIV-infected and HIV-seronegative pneumonia. The performance of the urine antigen ICT was independent of HIV-status (sensitivity 81%, specificity 98%, positive (PPV) and negative predictive values (NPV) 98%, and 82%, respectively). The sensitivity of sputum Gram's stain was 58% [34/59] with sputum unable to be provided by 16%. The CRP response was identical in HIV-infected (mean \pm SD) 133 \pm 88 vs. seronegative 135 \pm 104 mg/L (p=0.9). In the systematic review, the ICT performance revealed 74% sensitivity (95% CI: 72% to 77%) and 94% specificity (95% CI: 93% to 95%). Urine antigen testing increases etiologic diagnosis by 23% (Range: 10% –59%) when testing adults with community acquired pneumonia of unknown etiology.

Conclusions— Urinary antigen detection provides a credible rapid diagnostic test for pneumococcal pneumonia regardless of HIV-status. CRP response to acute infection is similar in HIV co-infection and increases diagnostic certainty.

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Binax was not involved in the design and conduct of this study. We thank Binax® NOW for providing the test kits for the study."

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Keywords

HIV; pneumonia; community acquired pneumonia; streptococcus pnuemoniae; pneumococcal; diagnosis; c-reactive protein; sensitivity

INTRODUCTION

Streptococcus pneumoniae is the leading cause of community acquired pneumonia (CAP), the 6th leading cause of death overall and the leading cause of infectious disease deaths. ^{1,2,3,4,5} In England and Wales, the rates of invasive pneumococcal disease are at near 10 per 100,000 general population.⁶ Among HIV-infected adults, rates of CAP are 5–10 fold higher than in age-matched populations, and bacteremic pneumonia occurs 35–50 times more commonly.⁷, ^{8,9,10} Rates in an HIV population approach 0.4–8 per 100 patient years in the U.S. and 1–4 per 100 person years in Sub-Saharan Africa. ^{8,10,11,12} Despite the beneficial effects of HAART and the paediatric conjugate pneumococcal vaccination on the reducing the rates of CAP in the past decade, HIV-infected adults still comprise 24% of U.S. invasive pneumococcal disease and 28% of pneumonia deaths in British young-adults ^{13,14}

The persistently high incidence and morbidity of HIV-associated pneumococcal infections highlight the need for tools for rapid and accurate diagnosis. In part, this limitation is related to the suboptimal availability and poor quality of sputum specimens provided to laboratories. ¹⁵ Rates of pathogen detection in CAP are only 30–50%. ^{16,17,18} Musher et al. emphasized the importance of collecting adequate sputum specimens promptly, prior to antibiotic administration for diagnosis of pneumococcal pneumonia, ¹⁸ but such specimens are often neither sought nor provided. ¹⁵ Moreover, the decline in accessible microbiologic facilities in developed countries and the dearth of microbiological laboratories in much of the developing world, where the burden of HIV disease predominates, highlights the need for rapid diagnostic tests for *S. pneumoniae* infection at the point of care. The inability to establish specific aetiologic diagnoses leads expert panels to recommend broad-spectrum empiric antimicrobials for CAP. These well-intentioned efforts likely increase cost and facilitate the emergence of resistant organisms.

The Binax NOW[®] immunochromatographic test (ICT) is a rapid urine antigen test with reasonable performance characteristics among adults with clinical pneumonia.^{19,20} However, immunocompromised persons, including those with HIV-infection, have been largely excluded from prior urine antigen studies. In this report we characterize the performance characteristics of this ICT among HIV-infected patients with documented *S. pneumoniae* pneumonia and bacteremia.

PATIENTS AND METHODS

Study population and setting

We prospectively enrolled 230 adult patients from a study of the natural history of pulmonary complications of HIV infection at San Francisco General Hospital during 1990–92, as previously described. 10,21,22 As a sub-study, urine was collected from 70 o these patients for urine antigen testing. After written informed consent, patients were selected for inclusion if they presented with pneumococcal pneumonia, defined by a compatible clinical scenario, a chest radiograph demonstrating parenchymal opacities, and microbiologic criteria. The microbiologic criteria included either: 1) *S. pneumoniae* cultured from a sterile site, i.e. blood, CSF, or tracheal aspirate; 2) acceptable sputum with Gram's stain revealing a predominance of Gram-positive cocci in pairs and chains at 1,000x magnification with or without a positive *S. pneumoniae* sputum culture. Sputum samples were considered acceptable with >25 WBC

and <10 epithelial cells per 400x field. Individuals were identified prospectively via microbiologic reporting.

Cases were defined as either 1) pneumococcal pneumonia or 2) pneumococcal bacteremic pneumonia. Exclusion criteria for this study included: lack of urine collection (n= 160) at hospital admission, nosocomial pneumonia or history of prior pneumococcal pneumonia, bacteremia, or hospitalization for *S. pneumoniae*. Control samples were obtained from 17 HIV + and 46 HIV– healthy ambulatory patients without clinical evidence of pneumococcal infection. An exclusion criterion for controls was the presence of an active, undiagnosed pulmonary infection at the study visit. Clinical characteristics as well as serum, urine, sputum and blood for culture were collected at time of diagnosis. C-reactive protein (CRP) was measured from collected serum by antigen-antibody agglutination (Abbott). Urine samples were immediately frozen and stored at -70° C for a median of 13.5 years. Convalescent urine was collected from 64% [45/70] one month after illness. The urine was collected with the express *a priori* intent for urine antigen testing; however, this was not possible until after August 31, 1999 when the current ICT became available.

Urine antigen detection

The Binax[®] NOW ICT (Binax Inc. Portland, Maine) was performed per the manufacturer's instructions.²³ Unconcentrated aliquots (1 mL) of urine were thawed to room temperature. Briefly, the manufacturer-provided swab was dipped into a gently swirled urine sample and inserted into the test device. The Binax ICT utilizes a colloidal gold-labeled antibody immobilized on a nitrocellulose membrane. When the device is closed, the sample is brought in contact with the test strip. Any pneumococcal antigen present binds with conjugated rabbit anti-C-polysaccharide *S. pneumoniae* antibody creating an antigen-antibody complex. A colorimetric reaction occurs when those complexes are captured by anti-*S. pneumoniae* antibodies immobilized on the sample line (i.e. positive) on the test device. Photography documented results at 15 minutes, and three masked readers read the results independently. This step was undertaken to determine inter-observer variability, previously suggested as a cause of variation in ICT performance.²⁴ Readers quantified test results based on their visual intensity as: negative (no detectable test band), weak positive (any visible test band <control band intensity), or strong positive (test band ≥control band intensity) [Appendix 1].

To exclude false negative reactions, all presumed false negative results (ICT negative but with bacteremic pneumococcal infection) were confirmed first by repeat urine ICT. In addition, the urine was acidified with buffered NH_4Cl to dissociate potential antigen-antibody complexes as previously described, ²⁵ then pH neutralized, and the samples were re-tested.

Systematic Review

We identified and reviewed study design, performance characteristics and the microbiologic "gold-standard" used in 20 studies in adults from the PubMed database (1950–June, 2007) identified using the keywords Binax, urine antigen, and *Streptococcus pneumoniae* and 5 additional studies from cross-references. Selected studies showed heterogeneity in the populations and gold-standards used to assess the test, but most used standard sputum Gram's stain, sputum culture, and blood culture for diagnosis. In the published studies, when the ICT was tested in patients with pneumonia of unknown aetiology, this is reported separately. Following the STARD initiative recommendations, we present positive and negative likelihood ratios (LR+, LR–). Data were analyzed using Meta-DiSc software (v1.4).²⁶ Summary values of the test performance were calculated from pooled Mantel-Haenszel weighted means.

Data Analysis

The sensitivity and specificity of the Binax[®] urinary ICT for diagnosing pneumococcal infection was determined, stratified by HIV-status. A double blind was maintained with test performance, readers, and data analyses. Statistical analysis was performed with SPSS 14.0 (Chicago, IL). Continuous, normally distributed variables are expressed as mean \pm standard deviation (SD) with comparisons via Student's *t*-test. For categorical variables, Fisher's exact test was used. For comparisons between inter-observer or multiple measurements, the Kappa statistic is presented. Statistical significance was defined as *P*<.05.

RESULTS

Diagnosis

Seventy pneumococcal infections occurred during the study (47 HIV+, 23 HIV–). Sixty infections were bacteremic pneumonia. Diagnostic methods of included sputum Gram's stain (n=34), sputum culture (n=32), and/or both bacteremia and sputum positive by either Gram's stain and/or culture (n=32) [Figure 1].

Clinical features of pneumococcal infection

Presenting symptoms included: cough, tactile fever, pleuritic chest pain and chills were present in \geq 75% of each group (Table 1). The majority of patients with and without HIV infection showed symptoms at comparable frequencies. The median duration of symptoms prior to admission was 4 days (IQR: 2–5 days). Cough was present longer than other symptoms by 1.1 days (95% CI: 0.4–1.8 days; P=.0004). No patient reported receiving antibiotics in the 30 days prior to admission, including trimethoprim-sulfamethoxazole prophylaxis.

HIV-infected patients with pneumonia were equally likely to generate an inflammatory response to the infection (fever, elevated CRP), although the proportion with elevated neutrophil counts was lower (Table 2). Chest radiographs revealed parenchymal opacities in all patients. These opacities were multi-lobar in 44% [31/70] with bilateral involvement in 30% [21/70] of persons with pneumococcal pneumonia. The most commonly infected lobes included the left lower (n=37) and right lower (n=36) lung lobes. The frequency and distribution of infiltrates was similar regardless of HIV-status. The interval between collection of blood for culture and final identification of positive cultures averaged 2.1 ± 1.2 days with 33% [24/60] formally identified on hospital day two or three. A majority of HIV-infected patients with pneumonia had advanced HIV disease (56% < 200 CD4⁺ T cells/µL).

Sputum Specimens

Among pneumonia patients, sputum Gram's stains were obtained in 59 (84%) subjects, and acceptable sputum cultured in 57 (81%) subjects (Table 2). The sensitivity of the sputum Gram's stain was 58% (34/59) among all pneumonia patients and 51% (25/49) in bacteremic pneumonia. Only 37% (18/49) had sputum positive both by Gram's stain and positive by sputum culture among bacteremic pneumonia cases and 41% (22/54) among all pneumonia patients (Kappa=0.42). Sputum collected >24 hours after antibiotic therapy rarely yielded either a positive Gram's stain [4/12] or culture [4/11] compared with those collected within 24 hours of admission, 68% (30/44) for Gram's stain (P=.03). Timing of sputum collection was unavailable for three subjects. Over one-third of patients with bacteremic pneumonia submitting sputum had non-diagnostic sputum specimens (18/50; 36%) either by Gram's stain or culture and an additional 16% (10/60) of bacteremic pneumonias submitted no sputum sample, suggesting the need for an adjunctive diagnostic test at the point of care when making therapeutic decisions.

Urine antigen detection (ICT)

Among 70 cases of pneumococcal pneumonia, the urine ICT was positive in 81% (57/70), including 85% (51/60) of bacteremic pneumonias. The urine antigen performance was similar regardless of HIV status: 83% HIV+ vs. 78% HIV– (Table 2). Among control subjects without clinical evidence of current pneumococcal disease, the ICT showed positive results in none of 46 HIV-negative control subjects and 6% (1/17) HIV-infected control subjects. Overall, the urine antigen ICT sensitivity was 81% (95% CI: 71% to 89%) for pneumococcal infection, and specificity was 98% [62/63] (95% CI: 92% to 100%), with similar sensitivity among HIV-infected (83%) as seronegative (78%) persons. The urine antigen ICT LR+ was 51.3 (95% CI: 7.3 to 360), and the LR– was 0.19 (95% CI: 0.12 to 0.31). *S. pneumoniae* serotypes 1, 3, 4 (n=3), 9, 12 (n=3), 14 (n=4), 18, 19 (n=2), 20 (n=2), and 23 were detected by the Binax ICT. One bacteremic episode with serotype 16 yielded a false negative. There were no differences in urine antigen sensitivity based on the duration of symptoms nor on the time to positive blood culture (83% on hospital day \leq 1 versus 87% on days \geq 2). Proteinuria did not affect the ICT performance. The inter-observer validity was excellent with 99% agreement on the Binax ICT positivity among three readers.

False Negatives

Among patients with pneumococcal pneumonia, false negative results did not differ by clinical characteristics (symptom duration, fever), bacteremia, WBC count, or CRP level. Urine immune complexes were not responsible for false negative results as no new positive results were detected after urine acid dissociation of potential antibody-antigen complexes.

Antigen Clearance

Persistently positive results in convalescent samples collected at one month were detected in 40% (18/45) of pneumonia patients tested. Persistent antigen was more likely detected among those with initial strong positive results (58% [14/24]; RR 3.1, 95% CI: 1.4 to 6.9, P = .01) compared with those with initial weak positives (19% [4/21]) The frequency of persistent antigenuria at one month was comparable in pneumonia patients with and without HIV infection (40% [12/30] vs. 42% [6/14], respectively).

C-reactive protein

Results of CRP and ICT paralleled one another. Among patients with pneumonia, CRP was elevated (mean 132 \pm 93 mg/L, range 35–479 mg/L; 90% >40 mg/L). The frequency and magnitude of CRP responses were independent of HIV status (*P*=.6) (Figure 2). Using both tests together (Binax ICT and CRP >40 mg/L) increased the test performance modestly (Table 3) by raising the NPV (83% to 96%) and only slightly lowering the sensitivity (81% to 75%) compared with ICT alone. A negative Binax ICT and CRP <40 mg/L effectively excluded pneumococcal pneumonia (NPV 96%).

Systematic Review

Among the 25 published studies in HIV-negative adults, the pooled sensitivity for the Binax urine ICT was 74% (95% CI: 72% to 77%) and specificity 94% (95% CI: 93% to 95%) compared with traditional microbiology in adults with pneumococcal infection (Figure 3). The summary PPV is 79% (95% CI: 70% to 88%) and NPV is 92% (95% CI: 89% to 96%) using a gold standard of a positive sputum and/or blood culture with a consistent clinical syndrome. The LR+ for the ICT is 17 (95% CI: 11 to 26), and the LR– is 0.29 (95% CI: 0.24 to 0.39). Among the 17 studies testing the urine ICT test among pneumonias of unknown aetiology, an additional 23 \pm 14% were putatively diagnosed as *S. pneumoniae* pneumonia by urinary antigen testing (Table 4).

DISCUSSION

Among HIV-infected and seronegative patients with acute pneumococcal pneumonia, we demonstrate that *S. pneumoniae* infections can be diagnosed with reasonable sensitivity and excellent specificity with a simple test available at the point of care. Just as symptoms and signs of pneumococcal pneumonia are comparable regardless of HIV-status,⁴⁹ the Binax *S. pneumoniae* urine antigen test has comparable performance characteristics among both groups. This test may provide reliable, rapid diagnosis for pneumococcal infection in the absence of an acceptable sputum specimen or readily-accessible microbiologic facilities. The absence of microbiologic laboratories may be encountered not only in resource-limited regions, where HIV infection is prevalent, but also in primary care clinics, urgent visit centers, or after-hours in developed countries when trained microbiology technicians are unavailable.

An advantage of urine antigen testing is that prior antibiotics do not rapidly decrease the sensitivity of ICT.¹⁸ Thus, patients having received antibiotics may still receive a specific diagnosis despite the rapidly diminishing yield of sputum examination. In the U.S., more rapid delivery of antibiotics is being encouraged. One U.S. Medicare quality improvement measure is the "number of pneumonia patients who received their first antibiotic dose within four hours from hospital arrival."⁵⁰ Antigenuria can persist for days to weeks after therapy, thereby enhancing the potential role of urine antigen detection for research purposes, such as vaccination trials.²⁸ Additionally, C-polysaccharide, the antigen detected in urine, appears to be cold stable at -70° C for extended periods (\geq 13 years in this study), a characteristic that will enhance epidemiologic studies.

Of greater consequence, false negative results occur in $\approx 20\%$ of patients with pneumococcal pneumonia. Causes for false negative results include aetiologic misclassification, low levels of C-polysaccharide antigen present, or sequestration of the antigen by antibodies in serum or urine as immune-complexes. Misclassification, by inclusion of non-pneumococcal pneumonia cases, does not account for all the false negatives as 15% of bacteremic pneumonias yielded false negative urine antigen results. In our study, false negatives did not differ by duration of symptoms, temperature, WBC count, or CRP, thus "early" infection is an unlikely explanation. We hypothesize that sequestration of antigen-antibody immune complexes in serum may decrease antigen shedding into the urine and yield false negative results. In prior studies, urine concentration improves ICT sensitivity by 5–20% as low levels antigenuria may occur in some individuals.^{20,28,29} Immune complexes were not present in the urine as acid dissociation did not improve detection. A broad array of different serotypes were detected in this study, thus the antigenic variability of the C-polysaccharide appears low. This is important as the previous latex agglutination urine antigen tests, which were based on the capsular polysaccharide, were limited in being serotype specific.⁵¹

In contrast to the high frequency of false positives in children,⁵² false positive tests occurred in one HIV-infected adult control with a prior pulmonary infection of unknown aetiology. Sources of false positives are two-fold. First, *S. viridans* species of *S. mitis* and *S. oralis* contain the same C-polysaccharide as *S. pneumoniae*.⁵³ Second, antigen shedding continues for weeks as 40% in this study and 70% in a Spanish cohort had detectable urine antigen one month following illness.²⁰ This urine antigen test should not be used in persons with a recent pneumococcal infection. Marcos et al demonstrated that nasopharyngeal pneumococcal carriage in eight clinically stable HIV-infected adults, CD4 count unspecified, did not yield any false positive results.²⁰

One difficulty with definitive CAP diagnosis is that there is no perfect gold standard for the aetiologic diagnosis. Even employing a range of diagnostic tests in an academic centre, a

specific aetiology cannot be determined in 50–70% of cases.^{16,17,18} Because expectorated sputum samples were diagnostic in $\leq 60\%$ of patients in this study, a new approach is necessary.

With traditional microbiological techniques, a 24–48 hours delay typically precedes confirmation of the aetiologic diagnosis. In this study, the average for microbiologic results was 2.1 days. This diagnostic delay may prompt physicians to initiate and continue broader spectrum antimicrobial agents or order more diagnostic tests in the interim. With shorter hospitalizations now commonplace, physicians may be reluctant to narrow the antibiotic spectrum 2–3 days into therapy near time of discharge. As a result, broader, more expensive antimicrobial therapy may well be continued to complete a therapeutic course.

β-lactam antibiotics remain the mainstay therapy for pneumococcal infection and non-severe CAP.⁵⁴ In a meta-analysis of 18 trials with 6749 adults with non-severe CAP, treatment failure with β-lactams was identical to broader spectrum agents having atypical coverage (RR 0.97; 95% CI: 0.87 to 1.07).⁵⁴ Prompt aetiologic diagnosis within 15 minutes should promote narrow spectrum antibiotics, e.g. amoxicillin 1000mg three times daily, sparing broader spectrum, more expensive antibiotics, e.g. "respiratory" fluoroquinolones or azithromycin/ clarithromycin, in non-severe CAP.⁵⁵ The utility of rapid diagnosis-driven therapy at the point of care has been prospectively verified among military recruits.⁵⁶ The cost of Binax Now[®] ICT (~£15, US\$30) outweighs the cost differential for 8 days of fluoroquinolones vs. amoxicillin (~£35, US\$70).⁵⁷ The European and U.S. cost of the Binax ICT is still exponentially more expensive than what is practical for use in resource-limited areas.

For calculating the assay's specificity, controls without acute pulmonary infection were chosen. Among studies utilizing other non-pneumococcal CAP patients as controls, "false positives" are reported in up to 29%.^{20,32,33,35} Because the frequency of the pneumococcal pneumonia is high in HIV selecting non-pneumococcal hospitalized CAP controls is difficult.^{7,8} With such a high pre-test probability, a culture-negative CAP still has a high PPV for pneumococcal pneumonia. In this study, 47% [28/60] of bacteremic pneumonia did not have a positive sputum Gram's stain or culture. Even with viral pneumonias, mixed infections with pneumococcus may occur in up to one-third of children.⁵⁸

Burel et al. have suggested that more work is required to establish the true specificity of the Binax[®] NOW test by studying its performance using urine from healthy subjects.⁴² Other studies have used various controls with CAP of unknown aetiology or with positive serologic tests for Chlamydia, Mycoplasma, or Legionella to define the control group. Among HIV-infected patients without consistent history, clinical and laboratory features of pneumococcal pneumonia, "false positive" urine antigen detection may represent prior, recent CAP infection. In our study, we eliminated this confounder by utilizing controls without pulmonary infection. Although this choice of controls may be viewed as a limitation, confounding of results by a high rate of cases without a specific aetiologic diagnosis was avoided.

Conversely, whether these "false positives" represent actual false positives or mixed infections with unconfirmed *S. pneumoniae* is unclear. Emerging data suggest that even in viral childhood pneumonias, *S. pneumoniae* may contribute to symptoms as a co-pathogen.^{58,59} These putative "false positives" lower the specificity and PPV in some studies reviewed.^{33,45}

In summary, urine ICT is a useful rapid diagnostic test for *S. pneumoniae* pneumonia and bacteremia in adults with HIV-infection. The urine antigen ICT may be quite valuable for establishing an aetiologic diagnosis in the absence of traditional microbiological testing, such as outpatient or rural clinical settings, for research in resource-limited nations to determine appropriate point-of-care algorithms, or for epidemiologic studies. Use of the urine antigen in non-severe pneumonia is also promising to promote the use of narrower spectrum, less expensive antibiotics.^{54,56}

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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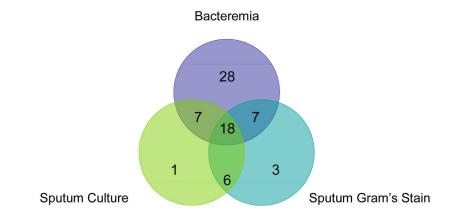
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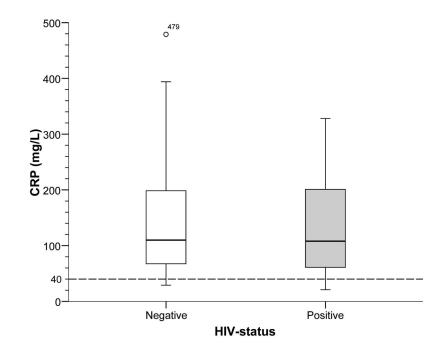
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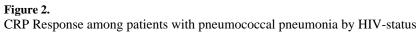
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Distribution of the microbiologic determination of pneumococcal pneumonia





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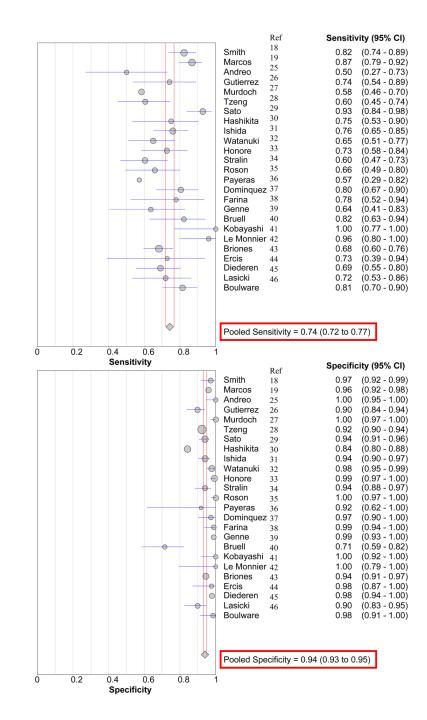


Figure 3. Forest plot of the sensitivity and specificity of Binax urine antigen ICT performance.

Table 1

Clinical characteristics of patients with pneumococcal pneumonia and control subjects.

Demographic	HIV-infected S. pneumoniae Pneumonia	HIV-negative S. pneumoniae Pneumonia	HIV- infected Control Subjects	HIV- negative Controls
No. of patients	47	23	17	46
Age (mean \pm SD)	35 ± 6	38 ± 7	34 ± 6	42 ± 15
Sex (M:F)	39:8	18:5	12:5	29:17
Intravenous drug use	21 (45%)	11 (48%)	11 (64%)	0%
Smoking, current ^{$\frac{1}{2}$}	72%	86%	79%	
MSM	26 (55%)*	1 (4%)	6 (35%)	0%
CD4 ⁺ T cells/µL (mean ±SD)	218 ± 207	593 ± 316	360 ± 230	
CD4 ⁺ T cells <200/ μ L (%)	27 (56%)	5 (22%)	6 (33%)	
Clinical Symptoms & Signs				
Cough	93%	95%	28%	N/A^{\dagger}
Fever (subjective)	90%	89%	11%	N/A
Pleural pain	91%	80%	0%	N/A
Chills	74%	83%	0%	N/A
Haemoptysis	51%	36%	0%	N/A
Temperature >38°C	63%	71%	0%	N/A

* p<05

Numbers (%) unless otherwise specified. MSM = men who have sex with men

 † Clinical symptoms and signs were not collected from HIV-negative controls.

[‡]Tobacco status known for 43 HIV-positive with pneumonia, 21 HIV-negative with pneumonia, 14 HIV-controls, and 1 HIV-negative control.

\$7 HIV pneumonia patients and 3 HIV controls patients were being treated with zidovudine (AZT) monotherapy.

Table 2

Presenting Laboratory Findings:

Laboratory Parameters	Pneumococcal Pneumonia: HIV-infected (n = 47)	Pneumococcal Pneumonia: HIV- negative (n = 23)	Healthy: HIV- infected Controls (n = 17) $\stackrel{\neq}{\neq}$
Haematologic parameters			
$\label{eq:WBC} \begin{split} WBC \ cells \times 10^6 / \mu L \ (mean \pm SD) \\ WBC > 15 \ cells \times 10^6 / \mu L \\ Absolute \ neutrophil \ count \ \times 10^6 / \mu L (mean \pm SD) \\ CRP \ mg/L \ (mean \pm SD) \ \S \\ CRP > 40 \ mg/L \ \$ \end{split}$	$11.0 \pm 6.9^{*}$ $31\%^{*}$ $8.9 \pm 5.9^{*}$ 133 ± 88 90%	$\begin{array}{c} 15.6 \pm 6.6 \\ 54\% \\ 14.4 \pm 7.2 \\ 135 \pm 104 \\ 96\% \end{array}$	$\begin{array}{c} 4.7 \pm 1.7 \\ 0\% \\ 2.9 \pm 1.3 \\ 5 \pm 6 \\ 1\% \end{array}$
Microbiology			
Bacteremia Diagnostic Sputum Gram's Stain [†] Unable to provide acceptable sputum [†] Sputum culture % [positive / acceptable [†] collected] Binax Urine Antigen positive	41 (87%) 57% [24/41] 7 (15%) 55% [22 / 40] 39 (83%)	19 (83%) 56% [10/18] 6 (26%) 59% [10/17] 18 (78%)	1 (6%) **

*p<.05 compared with HIV-seronegative patients with pneumococcal pneumonia

** The frequency of Binax positive urine antigen was 0% [0/46] in HIV-seronegative control subjects for whom other laboratory values were not available.

 † Diagnostic sputum Gram's stain had a predominance of Gram positive cocci in pairs/chains in acceptable sputum. An acceptable sputum specimen for culture had >25 WBC and <10 epithelial cells per 400x field.

 ‡ Data obtained for CRP from 90 HIV+ controls, WBC values from 63 HIV+ controls.

 $^{\$}$ CRP normal reference value <8 mg/L

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Performance of Diagnostic Methodologies

Test	Number of Samples	Number Positive	Sensitivity %	Specificity %	PPV %	% AdN
Sputum Gram's Stain	59	34	58	100*	100^*	I
Sputum Culture	57	32	56	100^{*}	100^*	I
Binax Urine Antigen	70	57	81	98	98	83

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* By inclusion definition; in a clinical setting, the specificity is not 100% due to potential *S. pneumoniae* colonization. **NIH-PA** Author Manuscript
 Table 4

 Review of Binax Immunochromatographic urinary antigen test performance in adults

Study	Z	Methodology	Clinical Scenario	Gold Standard Microbiology technique	Sensitivity %	Specificity %	% Add	% AdN	Additional Yield %
Smith 19	213	Ь	Bacteremia	Blood culture	82	67	76	84	N/A
Marcos ²⁰	295	Ь	CAP, 68 HIV	Blood or sputum	87	96	94	91	25
ndrao 27	34	ط	CAP	cuture Blood smittim culture	50	100	100	87	6
utierrez 28	183	, c.	CAP	Blood or sputum*	20	06	85	86	26
Murdoch ²⁹	192	. Д	CAP	Blood or sputum	58	100	100	80	23
Tzeng ³⁰	859	Ч	CAP	culture Blood or sputum culture	64	92	27	98	10
Sato ³¹	384	Ч	CAP	Blood culture	93	94	LT	98	42
asikita ³²	372	Ч	CAP	Blood or sputum*	75	84	25	98	16
Ishida ³³	349	Ч	CAP	Blood or sputum*	76	94	91	83	39
7atanuki ³³	313	Ч	CAP	Sputum Gram's stain	65	98	86	93	N/A
onore ³⁴	304	R/P	Pneumonia	Blood or sputum*	LL	98	95	95	59
Strålin ³⁵	191	Ч	CAP	Blood or sputum	60	94	86	74	N/A
osón ³⁶	173	Ь	CAP	Sputum Gram's stain	99	100	100	66	26
averas 37	163	Р	CAP	Sputum culture	57	92	42	95	15
Dominquez 38	122	Я	Pneumonia	Blood or sputum*	80	97	95	87	25
Farina ³⁹	104	Ч	CAP	Blood or sputum*	78	66	93	96	N/A
enne ⁴⁰	103	Ч	CAP	Blood or sputum*	64	66	93	91	24
ruell ⁴¹	91	Ь	CAP	Blood, sputum, pleural fluid	82	71	62	06	22
Kobayashi 42	58	Ч	CAP	Blood or sputum*	100	100	100	100	57
Le Monnier 43	41	Ч	Empyema	Pleural fluid culture, PCR	96	100	100	94	N/A
Briones ⁴⁴	566	Ч	CAP	Blood or sputum*	68	88	64	90	26
rics ⁴⁵	52	R	CAP	Blood or sputum*	73	98	89	93	N/A
Diederen ⁴⁶	194	R	CAP	Blood or sputum*	69	98	93	88	N/A
Lasocki ⁴⁷	140	R	ICU	Blood, Sputum, BAL	72	90	68	92	12
Boulware	133	Ч	HIV CAP	Blood or sputum*	81	98	98	83	N/A
Total	5679				74	94	62	60	73

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CAP=community acquired pneumonia; Pneumonia= CAP + nosocomial pneumonia; sputum* = Gram's stain and culture

Methodology: P = Prospective, R = Retrospective

Totals reflected pooled data;