# $\beta$ -Adrenoceptor heterogeneity in guinea-pig airways: comparison of functional and receptor labelling studies

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1 The distribution of  $\beta$ -adrenoceptor subtypes in guinea-pig airways has been studied by radioligand binding assays and analysis of mechanical responses.

2 Binding studies with the ligands  $[3H]$ -dihydroalprenolol and  $[125]$ -cyanopindolol, revealed that  $\beta$ -adrenoceptors were unevenly distributed throughout the airways with the highest density located in the parenchyma and the lowest density in the trachea.

3 The relative proportion of  $\beta_1$ :  $\beta_2$ -adrenoceptor binding sites was assessed by computer-assisted analysis of the inhibition curves generated by selective agents. It was virtually identical in each region and in the order of 15: 85%.

 $4$   $\beta$ -Adrenoceptor agonists caused concentration-dependent relaxations of both tracheal spirals and parenchymal lung strips. This response appeared to be mediated by both  $\beta_1$ - and  $\beta_2$ adrenoceptors in tracheal spirals as the  $pA_2$  value for the  $\beta_1$ -selective antagonist, atenolol, varied depending upon which agonist was used, and, in the presence of the  $\beta_2$ -adrenoceptor antagonist ICI 118,551, noradrenaline and isoprenaline produced biphasic concentration-effect curves. In parenchymal lung strips only the one subtype was involved as antagonist  $pA_2$  values were not dependent on the agonist used and the properties were consistent with those expected for a  $\beta_2$ -adrenoceptor.

# Introduction

The sub-classification of  $\beta$ -adrenoceptors into  $\beta_1$ and  $\beta_2$ -subtypes was first suggested by Lands, Arnold, McAuliff, Luduena & Brown (1967) on the basis of relative responses to sympathomimetic amines in different tissues, and led to the receptors of adipose tissue being classified as  $\beta_1$  and those in bronchial and vascular smooth muscle as  $\beta_2$ . Further support for this hypothesis has come from the use of antagonists which are selective for each subtype. Using these selective drugs Carlsson, Ablad, Brandstrom & Carlsson (1972) proposed that both  $\beta_1$ - and  $\beta_2$ -adrenoceptors may co-exist within the same organ or tissue and, under certain circumstances, may serve the same function. This has been clearly demonstrated for catecholamine-induced relaxation in guinea-pig tracheal smooth muscle by several studies, including those of Zaagsma, Oudhof, Van der Heijden & Plantje (1979) and O'Donnell & Wanstall (1979), which have shown that both  $\beta_1$ - and  $\beta_2$ -adrenoceptors mediate the relaxation. However, it should be noted that Zaagsma et al. (1979) showed that this heterogeneity is not present in guinea-pig parenchymal lung strips where the relaxant response

appears to be mediated solely by the  $\beta_2$ adrenoceptor.

Considerable support for  $\beta$ -adrenoceptor heterogeneity and for the co-existence of  $\beta_1$ - and  $\beta_2$ -adrenoceptors in the same tissue has come from radioligand receptor binding studies (Nahorski, 1981). Using this approach, experiments in our laboratory (Rugg, Barnett & Nahorski, 1978) first revealed that in the rat both  $\beta_1$ - and  $\beta_2$ -adrenoceptors are present in peripheral lung tissue. This would at first sight appear to conflict with the evidence from functional studies in the guinea-pig. However, it remained possible that the differences relate to species variation. Also, since binding studies are performed on homogenates of tissue, the heterogeneity found in these studies might relate to  $\beta_1$ - and  $\beta_2$ -adrenoceptors being located on functionally distinct cell types, with  $\beta_1$ -receptors not being involved in smooth muscle relaxation. In the present study, the distribution of  $\beta$ -adrenoceptor subtypes has been determined by comparing results from radioligand binding assays with functional studies in different regions of guinea-pig airways. Some of

these results have been presented to the British Pharmacological Society (Carswell & Nahorski, 1982a).

## Methods

Male guinea-pigs  $(250-300 \text{ g})$  were killed by a blow to the head and exsanguinated.

## Membrane preparation

The trachea and lungs were dissected on ice to give three separate preparations of parenchyma, bronchi and trachea, as described previously (Carswell & Nahorski, 1982b). Each preparation was homogenized in 30vols. of ice-cold 50mM Tris-HCl buffer (pH 7.8) with a Polytron homogenizer at setting 5 for  $2 \times 10$ s bursts. Homogenates were then filtered through a double layer of cheesecloth and centrifuged at 48,000 g for 15 min at 4°C in a Sorvall RC5 refrigerated centrifuge. Each pellet was washed three times by rehomogenization and centrifugation, followed by resuspension in buffer and stored at  $-40^{\circ}$ C. Protein was determined by the method of Lowry, Rosebrough, Farr & Randall (1951) using bovine serum albumin as standard.

## Radioligand binding;assays

Binding studies were performed with either tritiated dihydroalprenolol ([<sup>3</sup>H]-DHA) or iodinated cyanopindolol ([1251]-Cyp) in a final incubation volume of  $250 \mu$ l with all constituents made up in 50 mM Tris-HCl, pH 7.8. For agonists the buffer also contained  $0.1\%$  ascorbate as an antioxidant.

Membranes were incubated with [3H]-DHA to equilibrium (22°C, 30 min) and then terminated by addition of <sup>1</sup> ml ice-cold buffer and collection on Whatman GF/B filters under a vacuum pressure of about 500 mmHg. The filters were rapidly washed with  $3 \times 5$  ml of buffer and the radioactivity on filters was extracted into 4 ml of Fisofluor '1' scintillation fluid and counted in a Packard liquid scintillation spectrometer at an efficiency of about 40%.

Incubations with  $[125]$ -Cyp were carried out at

37°C for 50 min then terminated by the addition of <sup>1</sup> ml ice-cold buffer and collection on Whatman GF/B filters under a low vacuum pressure of 250 mmHg. The filters were then washed with  $1 \times 30$  ml buffer and radioactivity on the filters counted in an Ames Gammacord II gamma-counter.

## Functional studies

Parenchymal lung strips measuring approximately  $30 \times 3 \times 3$  mm were cut from the peripheral edge of the lower lobes of the lung and placed in ice-cold Krebs-Ringer. The trachea from the same animal was kept overnight in Krebs-Ringer at 4°C, gassed with 95%  $O_2$  and 5%  $CO_2$ , and then cut into a spiral the next day and divided into two strips of equal length (trachea kept overnight in this way responded to  $\beta$ -agonists in the same way as fresh tissue).

Paired preparations were suspended in two 50 ml organ baths which contained Krebs-Ringer solution at 37°C and continuously gassed with 95%  $O_2$  and  $5\%$  CO<sub>2</sub>. The solution also contained desmethylimipramine  $1 \times 10^{-6}$ M to block neuronal catecholamine uptake and phenoxybenzamine  $5 \times 10^{-6}$ M was added directly to the baths to block a-adrenoceptors and extra-neuronal agonist uptake, then washed out after 30 min. An initial tension of  $250$  mg was applied to parenchymal lung strips and 5 g to tracheal spirals, throughout an initial equilibrium period of <sup>1</sup> h. During this time the preparations were rinsed with fresh solution every 20 min.

Concentration-effect (C/E) curves to agonists were constructed by use of a cumulative dosing technique (Van Rossum, 1963). The effect of antagonists was studied by adding the antagonist to one strip with the other serving as <sup>a</sup> control. A <sup>60</sup> min equilibrium period was allowed for each antagonist before the C/E curve was repeated on both the control and treated strips.

Responses were measured isotonically by Washington Type TII isotonic lever transducers, and recorded on a Washington oscillograph.  $pD_2$  values  $(-\log EC_{50})$  for agonists were estimated by analysing the C/E curves with a computer-assisted curve fitting procedure (De Lean, Munson & Rodbard, 1978).

**Table 1** Distribution of  $\beta$ -adrenoceptors in guinea-pig airways identified by [<sup>3</sup>H]-dihydroalprenolol ([<sup>3</sup>H]-DHA) and  $[$ <sup>125</sup>I]-cyanopindolol ( $[$ <sup>125</sup>I]-Cyp)

	$B_{max}[^3H]$ -DHA $(fmol$ mg <sup>-1</sup> protein)	$B_{\text{max}}$ [ <sup>125</sup> <i>I</i> ]-Cyp $(fmolmg^{-1}$ protein)	$K_D$ $\beta$ H <sub>l</sub> -DHA (nM)	$K_D l^{125}$ Il-Cyp (m)
Trachea	$46 \pm 4$	$59 + 13$	$0.14 \pm 0.04$	$16.0 \pm 3.2$
Bronchi	$196 \pm 14$	$125 \pm 12$	$0.21 \pm 0.03$	$23.1 \pm 1.4$
Parenchyma	$366 \pm 31$	$400 \pm 25$	$0.20 \pm 0.03$	$21.2 \pm 2.6$

 $B_{max}$  values were estimated from Scatchard plots and  $K_D$  values from Hill plots. Values are the mean + s.e.mean of three separate determinations

**Table 2**  $K_i$  values of selective antagonists for  $\beta_1$ - and  $\beta_2$ -adrenoceptors and the proportions of each receptor in the trachea, bronchi and parenchyma, from radioligand binding displacement studies

	Trachea		<b>Bronchi</b>		Parenchyma	
Drug	β,	$\beta_2$	ß1	$\beta_2$		ß2
Atenolol	$4.6 \times 10^{-8}$	$2.2 \times 10^{-5}$	$4.6 \times 10^{-8}$	$1.1 \times 10^{-5}$	$1.8 \times 10^{-8}$	$5.5 \times 10^{-6}$
	15.6%	84.4%	14.7%	85.3%	16.8%	83.2%
ICI 118,551	$2.0 \times 10^{-7}$	$6.5 \times 10^{-9}$	$4.0 \times 10^{-7}$	$2.9 \times 10^{-9}$	$2.0 \times 10^{-7}$	$6.3 \times 10^{-9}$
	15.3%	84.7%	$9.6\%$	90.4%	12.8%	87.2%

The means of three separate inhibition curves were analysed using a computer-assisted iterative curve-fitting procedure. K<sub>i</sub> values (molar) were calculated using the equation  $K_i = IC_{50}/1 + L/K_D$  and the percentages indicate the proportions of  $\beta_1$ - and  $\beta_2$ -receptors. The above data were obtained using [1251]-cyanopindolol, but virtually identical results were also obtained with  $[{}^{3}H]$ -dihydroalprenolol

For strips treated with antagonists, the response is expressed as % control maximum response and apparent  $pA_2$  values were calculated from a Schild plot (Arunlakshana & Schild, 1959) of log (dose ratio  $-1$ ) vs log [antagonist] by linear regression analysis.

Drugs used were  $(-)$ -noradrenaline bitartrate,  $(-)$ -adrenaline bitartrate,  $(-)$ -isoprenaline bitartrate (Sigma Chemical Co.); terbutaline sul-<br>phate (Astra): (-)-timolol hydrochloride phate (Astra); (-)-timolol hydrochloride<br>(Leo Laboratores): atenolol hydrochloride (Leo Laboratores); atenolol and ICI 118,551 erythro-DL-1 (7-methylindan-4 yloxyl-3-isopropylaminbutan-2-ol, ICI Pharmaceuticals); desmethylimipramine (Geigy Pharmaceuticals) and phenoxybenzamine (Smith Kline & French).  $(-)$ -[<sup>3</sup>H]-dihydroalprenolol (103 Ci/mmol) and  $(\pm)$ - $\left[1^{25}\right]$ -cyanopindolol (2200 Ci/mmol) were both obtained from Amersham International. The iodinated ligand was only used within its half-life (60 days).

#### **Results**

### Radioligand binding studies

Specific binding, defined as the binding displaceable by  $200 \mu M$  (-)-isoprenaline (Nahorski & Richardson, 1979) of  $[3H]$ -DHA and  $[125]$ -Cyp, to homogenates of guinea-pig airways was saturable, produced linear Scatchard plots and had Hill coefficients of close to unity. The dissociation constant  $(K<sub>D</sub>)$  for both radioligands was virtually identical between each area (Table 1), but although both ligands labelled a virtually identical number of sites within each area, there were significant regional differences in  $B_{max}$  values. The highest density of  $\beta$ adrenoceptors was found in the parenchyma, with the lowest density in the trachea (Table 1).

Binding of both ligands was displaced stereoselectively by the  $(+)$ - and  $(-)$ -isomers of propranolol in

Table 3  $pA_2$  values and slopes of Schild plots of timolol, atenolol and ICI 118,551 for the antagonism of f-adrenoceptor mediated relaxation of guinea-pig tracheal spirals and lung strips



Cumulative agonist concentration-effect curves were performed in the absence and presence of increasing concentrations of antagonist as described in Methods. Values represent the mean  $\pm$  s.e.mean,  $n = 3$  (\* $n = 12$ )

all regions, with the  $(-)$ -isomer being approximately 100 times more potent than the (+ )-isomer (data not shown), and was also displaced by timolol, another non-selective  $\beta$ -antagonist. These drugs all produced monophasic displacement curves and had slope factors close to unity. The selective antagonists, atenolol ( $\beta_1$ -selective; Barrett, 1977) and ICI 118,551 ( $\beta_2$ selective; Bilski, Dorries, Fitzgerald, Jessup, Tucker & Wale, 1980) however, generated displacement curves deviating from law of mass action behaviour. Analysis of these curves by a computer-assisted curve-fitting procedure (Dickinson, Richardson & Nahorski, 1981) to a two-site model, indicated that they represented displacement of radioligand from a population of  $\beta$ -adrenoceptors which consisted of approximately 85% of the  $\beta_2$  subtype and 15% of the  $\beta_1$  subtype. The relative proportion of these two subtypes was very similar between the different regions (Table 2).

## Functional studies

Both tracheal spirals and parenchymal lung strips relaxed in response to the agonists isoprenaline (Iso, non-selective), noradrenaline (NA,  $\beta_1$ -selective) and terbutaline (Ter,  $\beta_2$ -selective). In the trachea, the rank order of potency for these agonists was  $Iso > NA > Ter$ , but in parenchymal lung strips Ter was more potent than NA (Table 3).

The non-selective antagonist, timolol, produced parallel shifts to the right of the C/E curves for all three agonists, in both tracheal spirals and parenchymal lung strips. Schild plots obtained from these curves had slopes close to unity and the  $pA_2$  value was the same irrespective of the preparation or which agonist was used (Table 3).

In the presence of the  $\beta_1$ -selective antagonist atenolol, agonist C/E curves were again shifted to the right, but when NA was used as the agonist, the size of the shift in the trachea was much greater than the shift produced by the same concentration of atenolol in the lung strip (Figure 1). This resulted in the apparent inhibitory potency of atenolol being  $100 \times$ greater when the agonist was NA than when either Iso or Ter were used in tracheal spirals, but only a 3 fold difference in parenchymal lung strips (Table 3). The slopes of the Schild plots for the antagonism of NA by atenolol were also significantly different from unity.

Pretreatment of tracheal spirals with the highly selective  $\beta_2$ -antagonist, ICI 118,551, transformed the sigmoidal C/E curves for both NA (Figure 2) and Iso (not shown) into biphasic curves. Low concentrations  $(2 \times 10^{-8} \text{M})$  of this antagonist produced shifts to the right of the upper portion of the C/E curve and had no effect on the lower portion of the curve. At higher concentrations of ICI 118,551 ( $2 \times 10^{-7}$  M),



Figure 1 Antagonism of noradrenaline (NA) by the  $\beta_1$ -selective antagonist, atenolol. NA concentrationeffect curves were produced as described in the Methods, in the presence of increasing concentrations of atenolol, with 1h equilibration time between each curve. The data represent a single experiment only on (a) tracheal spiral and (b) parenchymal strip of guineapig, but similar results were obtained on at least two separate occasions. (O) Control; ( $\bullet$ ) 2 × 10<sup>-6</sup>M; ( $\square$ )  $2 \times 10^{-5}$  M; (1)  $2 \times 10^{-4}$  M atenolol.

the whole C/E curve was displaced to the right. In parenchymal lung strips, however, the same concentrations of ICI 118,551 produced parallel shifts to the right of the complete C/E curve for all three agonists, giving Schild plots that had slopes of close to unity and similar  $pA_2$  values (Table 3).

# Discussion

Substantial recent evidence from both functional and receptor binding studies suggests that both  $\beta$ adrenoceptor subtypes can co-exist in tissues such as heart (Carlsson et al., 1972; Hedberg, Minneman & Molinoff, 1980), lung (Furchgott, 1976; Rugg et al., 1978; Zaagsma et al., 1979; O'Donnell & Wanstall,



Figure 2 Antagonism of noradrenaline (NA) by the  $\beta_2$ -selective antagonist, ICI 118,551. Concentrationeffect curves for NA were produced as described in the Methods, in the presence of increasing concentrations of ICI 118,551, with <sup>1</sup> h equilibration time between each curve. The data represent a single experiment on (a) tracheal spiral and (b) parenchymal strip of guinea-pig, but similar results were obtained on at least two separate occasions. (O) Control; ( $\bullet$ ) 2 × 10<sup>-8</sup>M; ( $\Box$ ) 5 × 10<sup>-8</sup>M; and ( $\Box$ )  $2 \times 10^{-7}$  M ICI 118,551.

1979), rat jugular vein (Cohen & Wiley, 1978) or cat nictitating membrane (Varma & Nickerson, 1981). However, there is no reason to expect that the direct identification of both  $\beta$ -adrenoceptor subtypes in a tissue should necessarily be accompanied by functional heterogeneity and there are good examples of this kind of discrepancy in guinea-pig atria (O'Donnell & Wanstall, 1979; Hedberg et al., 1980) and in the present study, at least in the case of peripheral lung tissue. It is very likely that  $\beta$ -receptor heterogeneity identified in binding studies may merely reflect the heterogeneous cellular nature of the tissue in question. Indeed, it is only with functional receptor heterogeneity that one can suggest that since both receptors mediate the same function, they may co-exist on the same cell.

The present study confirms and extends earlier

studies (Zaagsma et al., 1979; O'Donnell & Wanstall, 1979) that suggest that relaxation of tracheal smooth muscle of the guinea-pig can be mediated by  $\beta_1$ - and  $\beta_2$ -adrenoceptors whereas in the finer peripheral airways, a homogeneous  $\beta_2$  population mediates this function. The use of both selective agonists and antagonists has been critical here. Thus, the non-selective antagonist, timolol, had very similar affinity irrespective of the agonists in both areas, but with the selective antagonists, atenolol and ICI 118,551, this was only true in peripheral lung strips. In tracheal spirals the apparent affinity for atenolol was dependent on the agonist used; atenolol was  $100 \times$  more potent with NA than with Iso and Ter as agonists. NA therefore probably produces relaxant responses by stimulating different receptors from those stimulated by the other agonists and, as the slope of the Schild plot was much less than unity, more than one type of receptor could be involved. In the presence of atenolol, higher concentrations of NA are required to produce <sup>a</sup> response and as its selectivity for  $\beta_1$ -receptors (from ligand binding studies) is only around 30 fold, this agonist may also stimulate  $\beta_2$ -receptors. This could result in an apparent decrease in the potency of atenolol at higher NA concentrations (where the latter acts at  $\beta_2$ -receptors) and thus produce a Schild plot with a regression line of low slope. In parenchymal strips the low value of the slope for atenolol with NA may, in part, be due to difficulties in accurately measuring the C/E curve for NA in this tissue owing to its low potency; and it should be noted that the low slope will increase the error in estimating the  $pA_2$  value.

The involvement of both  $\beta$ -adrenoceptor subtypes in the response of the trachea received support from the studies with the highly  $\beta_2$ -selective antagonist, ICI 118,551, which produced biphasic C/E curves with both Iso and NA. These data confirm and extend those of Zaagsma, Van der Heijden, Van der Schaar & Bank (1983) with this antagonist. The findings may be explained by ICI 118,551 producing a greater inhibitory effect at  $\beta_2$ -receptors than  $\beta_1$ -receptors, thereby increasing the apparent selectivity of NA for  $\beta_1$ -receptors and making Iso a  $\beta_1$ -selective agonist. The opposite is true for atenolol, which will have a greater inhibitory effect at  $\beta_1$ -receptors than  $\beta_2$ receptors, thus reducing the selectivity of NA for  $\beta_1$ -receptors and explaining why biphasic curves to NA were not seen in the presence of atenolol. The apparent parallel shifts in the C/E curves for Iso in the presence of atenolol are, however, more difficult to explain, but they may reflect some functional interaction between the two subtypes whereby  $\beta_2$ receptors have the capacity to produce the full maximum response when  $\beta_1$ -receptors are inhibited, but  $\beta_1$ -receptors alone cannot produce the full maximum response. From the curves for NA in the presence of

ICI 118,551 it appears that  $\beta_1$ -receptors may only be able to produce 50-60% of the maximum response, but this appears to vary from animal to animal (unpublished observations, see also Furchgott & Wakade, 1976).

Whatever the relationship between the two subtypes, it is clear that whilst both  $\beta_1$  and  $\beta_2$ -receptors mediate the relaxant response to catecholamines in the trachea, only the  $\beta_2$ -subtype appears to be involved in the response of parenchymal lung strips. This is particularly intriguing since these strips possess several potential contractile elements including bronchiolar and vascular smooth muscle (Lulich, Mitchell & Sparrow, 1976) and alveolar interstitial myofibroblasts (Kapanci, Assimacopoulos, Irle, Zwahlen & Gabbiani, 1974). The distribution of functional  $\beta_1$ -adrenoceptors appears to correspond to the density of noradrenergic innervation in the airways (O'Donnell, Saar & Wood, 1978) and thus tends to support the concept of Ariens & Simonis (1976) that  $\beta_1$ -receptors respond primarily to noradrenaline released from sympathetic nerve endings, whereas  $\beta_2$  sites are humoral receptors responding mainly to circulating adrenaline. However, the direct

### References

- AINSWORTH, G.A., GARLAND, L.G. & PAYNE, A.N. (1981). Modulation of bronchoconstrictor responses to histamine in pithed guinea-pigs by sympathetic nerve stimulation. Br. J. Pharmac., 77,249-254.
- ARIENS, E.J. & SIMONIS, A.M. (1976). Receptors and receptor mechanisms. In Beta-Adrenoceptor Blocking Agents. The Pharmacological Basis of Clinical Use ed. Saxena, P.R. & Forsyth, R.P. Amsterdam: North-Holland Publ. Co.
- ARUNLAKSHANA, 0. & SCHILD, H.O. (1959). Some quantitative uses of drug antagonists. Br. J. Pharmac. Chemother., 14, 48-58.
- BARNES, P.J., BASBAUM, C.B., NADEL, J.A. & ROBERTS, J.M. (1982). Localisation of  $\beta$ -adrenoceptors in mammalian lung by light microscope autoradiography. Nature, 299, 444-447.
- BARRETT, A.M. (1977). The pharmacology of atenolol. Postgrad. Med. J., 53, (Suppl. 3), 58-64.
- BILSKI, A., DORRIES, S., FITZGERALD, J., JESSUP, R., TUCKER, H. & WALE, J. (1980). ICI 118,551, a potent  $\beta_2$ -adrenoceptor antagonist. Br. J. Pharmac., 69, 292-293P.
- CARLSSON, E., ABLAD, B., BRANDSTROM, A. & CARLSSON, B. (1972). Differential blockade of the chronotropic effects of various adrenergic stimuli in the cat heart. Life Sci., 11, 953-958.
- CARSWELL, H. & NAHORSKI, S.R. (1982a). The distribution of  $\beta$ -adrenoceptor subtypes in guinea-pig airways. Br. J. Pharmac., 77, Proc. Suppl., 480P.
- CARSWELL, H. & NAHORSKI, S.R. (1982b). Distribution and characteristics of histamine  $H_1$ -receptors in guineapig airways identified by  ${}^{3}$ H-mepyramine. Eur. J. Pharmac., 81, 301-307.

identification of  $\beta$ -adrenoceptors using ligand binding techniques suggests that the relative proportion of  $\beta_1$ -adrenoceptors is similar throughout the airways. If we are to accept that these  $\beta_1$  sites mediate functions other than smooth muscle relaxation, we may have to consider that they could respond to NA which may overflow from sympathetic nerves that innervate pulmonary blood vessels (see Ainsworth, Garland & Payne, 1982). Clearly the most urgent objective is to obtain much more precise information on the cellular localisation of  $\beta$ -adrenoceptor subtypes in the airways. The recent use of light microscope autoradiography in ferret lung with  $[3H]$ dihydroalprenolol (Barnes, Basbaum, Nadel & Roberts, 1982) has revealed that high densities of P-adrenoceptors are associated with alveoli and airway smooth muscle, particularly the smooth muscle of the smaller airways (bronchioles). It will be important now to extend this approach to examine receptor subtypes in the airways of several species.

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- COHEN, M.L. & WILEY, K.S. (1978). Beta<sub>1</sub> and beta<sub>2</sub> receptor mechanisms in rat jugular veins: differences between norepinephrine and isoproterenol-induced relaxation. Life Sci., 23, 1997-2006.
- DE LEAN, A., MUNSON, P.J. & RODBARD, D. (1978). Simultaneous analysis of families of sigmoid curves: applications to bioassay, radioligand assay and physiological dose-response curves. Am. J. Physiol., 4, E97-E102.
- DICKINSON, K.E.J., RICHARDSON, A. & NAHORSKI, S.R. (1981). Homogeneity of beta<sub>2</sub>-adrenoceptors on rat erythrocytes and reticulocytes: a comparison with heterogeneous rat lung beta-adrenoceptors. Mol. Pharmac., 19, 194-204.
- FURCHGOTT, R.F. (1976). Postsynaptic adrenergic receptor mechanisms in vascular smooth muscle. In Vascular Neuroeffector Systems, ed. Bevan, J.A. pp. 131-142. Basel: Karger.
- FURCHGOTT, R.F. & WAKADE, T.D. (1976). Evidence for both  $\beta_1$  and  $\beta_2$  receptors in guinea-pig tracheal smooth muscle and variation of the  $\beta_1 : \beta_2$  ratio in different animals. 6th Int. Congress Pharmac., Abst. Helsinki, p. 622.
- HEDBERG, A., MINNEMAN, K.P. & MOLINOFF, P.B. (1980). Differential distribution of beta<sub>1</sub> and beta<sub>2</sub> adrenergic receptors in cat and guinea-pig heart. J. Pharmac. exp. Ther., 212, 503-508.
- KAPANCI, Y., ASSIMACOPOULOS, A., IRLE, C., ZWAHLEN, A. & GABBIANI, G. (1974). 'Contractile interstitial cells' in pulmonary alveolar septa: A possible regulator of ventilation/perfusion ratio? J. cell. Biol., 60, 375-392.
- LANDS, A.M., ARNOLD, A., McAULIFF, J.P., LUDUENA,

F.P. & BROWN, T.G. (1967). Differentiation of receptor systems activated by sympathomimetic amines. Nature, 214, 597-598.

- LOWRY, O.H., ROSEBROUGH, N.J., FARR, A.L. & RAN-DALL, R.J. (1951). Protein measurement with the folin phenol reagent. J. biol. Chem., 193, 265-275.
- LULICH, K.M., MITCHELL, H.W. & SPARROW, M.P. (1976). The cat lung strip as an in vitro preparation of peripheral airways. Br. J. Pharmac., 58, 71-79.
- NAHORSKI, S.R. (1981). Identification and significance of beta-adrenoceptor subtypes. Trends Pharmac. Sci., 2, 95-98.
- NAHORSKI, S.R. & RICHARDSON, A. (1979). Pitfalls in the assessment of the specific binding of  $(-)-[^3H]$ dihydroalprenolol to beta-adrenoceptors. Br. J. Pharmac., 66, 469P.
- O'DONNELL, S.R., SAAR, N. & WOOD, L.J. (1978). The density of adrenergic nerves at various levels in the guinea-pig lung. Clin. exp. Pharmac. Physiol., 5, 325 -332.
- O'DONNELL, S.R. & WANSTALL, J.C. (1979). The importance of choice of agonist in studies designed to predict  $\beta_2$ :  $\beta_1$  adrenoceptor selectivity of antagonists from PA<sub>2</sub> values on guinea-pig trachea and atria. Naunyn-Schmiedebergs Arch. Pharmac., 308, 183-190.
- RUGG, E., BARNETr, D.B. & NAHORSKI, S.R. (1978). Coexistence of beta<sub>1</sub> and beta<sub>2</sub> adrenoceptors in mammalian lung: evidence from direct binding studies. Mol. Pharmac., 14,996-1005.
- VAN ROSSUM, J.N. (1963). Cumulative dose-response curves. II. Techniques for the making of dose-response curves in isolated organs and the evaluation of drug parameters. Archs. Int. Pharmacodyn. Thér., 143, 299-330.
- VARMA, D.R. & NICKERSON, M. (1981). Betaadrenoceptors of the cat nictitating membrane. J. auton. Pharmac., 1, 291-297.
- ZAAGSMA, J., OUDHOF, R., VAN DER HEIJDEN, P.J.C.M. & PLANTJE, J.F. (1979). Subheterogeneity of  $\beta$ adrenoceptors in the pulmonary and the cardiac system of the guinea-pig. In Catecholamines: Basic and Clinical Frontiers ed. Usdin, E., Kopin, I.J. & Barchas, J. pp. 435 -437. Oxford: Pergamon Press.
- ZAAGSMA, J., VAN DER HEIJDEN, P.J.C.M., VAN DER SCHAAR, M.W.G. & BANK, C.M.C. (1983). Comparison of functional *β*-adrenoceptor heterogeneity in central and peripheral airway smooth muscle of guinea-pig and man. J. Receptor Res., (in press).

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