

Evidence for pharmacological similarity between α_2 -adrenoceptors in the vas deferens and central nervous system of the rat

J.C. Doxey, B. Gadie, A.C. Lane & I.F. Tulloch

Department of Pharmacology, Reckitt and Colman Limited, Pharmaceutical Division, Dansom Lane, Hull HU8 7DS

- 1 Seven α_2 -adrenoceptor antagonists with diverse chemical structures have been examined for their effects at α_2 -adrenoceptors in the vas deferens and central nervous system of the rat.
- 2 Antagonist potency assessed against the presynaptic α_2 -adrenoceptor agonist action of clonidine in the isolated vas deferens (RX 781094 > Wy 26703 > yohimbine > rauwolscine > piperoxan > mianserin > RS 21361) was highly correlated with the ability of these drugs to displace saturable [3 H]-RX 781094 binding from cerebral cortex membranes.
- 3 Similarly, antagonist potency in the vas deferens was highly correlated with antagonist activity in reversing the centrally-mediated mydriasis induced by the selective α_2 -adrenoceptor agonist, guanoxabenz, in pentobarbitone-anaesthetized rats.
- 4 The results indicate that the presynaptic α_2 -adrenoceptors in the vas deferens are pharmacologically similar to characterized these α_2 -adrenoceptors in the central nervous system of the rat.

Introduction

A substantial body of evidence now exists demonstrating the presence of α_2 -adrenoceptors in a wide variety of mammalian tissues (for reviews see Langer, 1981; Starke, 1981). In the majority of studies these α_2 -adrenoceptors have been classified according to their relative sensitivity to various α_2 -adrenoceptor agonists and antagonists. This approach has allowed the characterization of α_2 -adrenoceptors in a number of peripheral tissues and one in particular, the rat isolated vas deferens, is now widely used for screening drugs with potential α_2 -adrenoceptor activity (Doxey & Roach, 1980). In order to assess the general predictive value of this screen, especially with regard to central activity, it is important to investigate whether the α_2 -adrenoceptors in the vas deferens resemble presumed α_2 -adrenoceptors in the CNS.

Central α_2 -adrenoceptors have been widely investigated by radioligand binding techniques using various selective α_2 -adrenoceptor agents (U'Prichard, Greenberg & Snyder, 1977; Perry & U'Prichard, 1981; Jarrott, Louis & Summers, 1982). Recently RX 781094, a highly selective α_2 -adrenoceptor antagonist (Doxey, Roach & Smith, 1983) has proved to be a particularly useful ligand for studying α_2 -

adrenoceptor sites (Howlett, Taylor and Walter, 1982). In addition, RX 781094 has been used to characterize functional α_2 -adrenoceptors in the CNS (Dettmar, Lynn & Tulloch, 1981), including those mediating the mydriatic effect of α_2 -adrenoceptor agonists (Berridge, Gadie, Roach & Tulloch, 1983).

The availability of RX 781094 and other recently discovered selective α_2 -adrenoceptor antagonists such as RS 21361 (Michel & Whiting, 1981) and Wy 26703 (Lattimer, Rhodes, Ward, Waterfall & White, 1982) makes it possible to perform a detailed characterization of various α_2 -adrenoceptors. This approach, as an alternative to using α_2 -adrenoceptor agonists for the characterization of this receptor subtype, should have the advantage of obviating the interpretative problems caused by differences in intrinsic activity between agonists. The present study has compared the prejunctional α_2 -adrenoceptors in the rat vas deferens with central α_2 -adrenoceptors in the same species; namely (1) the functional receptors responsible for mediating the mydriatic effect of α_2 -adrenoceptor agonists, and (2) the receptor sites in cerebral cortex labelled by [3 H]-RX 781094. Receptor characterization was achieved by determining the relative activities of a range of selective α_2 -

adrenoceptor antagonists at these various receptor sites. A preliminary account of this work has been published (Doxey, Gadie, Lane & Tulloch, 1982).

Methods

α_2 -Adrenoceptor antagonist potency in the CNS

(a) *Antagonism of guanoxabenz-induced mydriasis*
Male Sprague-Dawley rats, 250–350 g, were anaesthetized with sodium pentobarbitone (Sagatal; 60 mg kg⁻¹, i.p.) and subsequent drug experiments were all performed within the following 40 min period. Antagonist potency was determined as described by Berridge *et al.* (1983). Maximal pupil dilation (> 4 mm) was produced by injection of the selective α_2 -adrenoceptor agonist guanoxabenz (0.3 mg kg⁻¹, i.v.). Fifteen minutes later increasing doses (half-log increments) of an antagonist were injected i.v. at 5 min intervals; antagonist potency was assessed by determining the cumulative dose to produce a 50% reversal of the mydriatic response. These AD₅₀ values, expressed as μ mol kg⁻¹, were determined graphically from the log dose-response curves for each antagonist.

(b) *Displacement of [³H]-RX 781094 binding in rat cortex*
The displacement of [³H]-RX 781094 was determined in rat brain cerebral cortex preparations essentially as described by Howlett *et al.*, (1982). Male Sprague-Dawley rats, 250–300 g, were killed by decapitation, the brains quickly removed, and the cerebral cortex dissected free from striatal tissue but including the hippocampus. The tissue was homogenized in ice-cold buffered sucrose (20 ml g⁻¹ tissue, 300 mmol l⁻¹ sucrose in Tris/HCl buffer, 50 mmol l⁻¹, pH 7.4) using a Potter-type glass-teflon homogenizer. A crude synaptosomal P₂ fraction was prepared according to Whittaker (1969). The washed P₂-pellet was resuspended in a physiological salt solution to give a final protein concentration of about 1 mg ml⁻¹. The physiological salt solution had the following composition (mM): NaCl 118, KCl 4.8, CaCl₂ 1.3, KH₂PO₄ 1.2, MgSO₄ 1.2, NaHCO₂ 25 and was equilibrated with 95% O₂; 5% CO₂ (Starke & Montel, 1973).

A typical incubation mixture (in triplicate) contained: membrane preparation (970 μ l), saturating ligand or varying concentrations of an α_2 -adrenoceptor antagonist (10 μ l), and [³H]-RX 781094 (20 μ l, to give a final concentration of 1 nM). Non-saturable binding, representing about 30% of the total binding, was defined by the addition of an excess of phentolamine (1 μ M). Samples were incubated for 15 min at 25°C and then rapidly filtered under vacuum through Whatman GFB filters and

washed with 2 × 4 ml of ice-cold Tris/HCl buffer (0.05 M, pH 7.4). Filters were transferred to scintillation vials and NE 260 micellar scintillation fluid (3.5 ml; Nuclear Enterprises) was added prior to scintillation counting. Competition binding curves using up to eight concentrations of test antagonist were constructed and IC₅₀ values, defined as the concentrations which displaced 50% of saturable [³H]-RX 781094 binding, were determined. Hill plots were constructed for each antagonist. These were linear regressions of log (B - B_L)/B_L against log (antagonist concentration) where B was specific binding of [³H]-RX 781094 in the absence of competing antagonist and B_L was specific binding at a particular concentration of antagonist. Inhibitor constants (K_i values) were derived from the IC₅₀ values according to the method of Cheng & Prusoff (1973). Saturation binding studies were also performed separately under these conditions using a range of [³H]-RX 781094 concentrations (0.1–30 nM). Specific binding of [³H]-RX 781094 was characterized using Scatchard analysis.

Rat vas deferens

The detailed method has been described elsewhere (Doxey, Smith & Walker, 1977). In brief, the prostatic half of the vas deferens was suspended under an initial tension of 0.5 g in an organ bath of 8–10 ml capacity. Tissue was bathed in a Krebs solution (composition, mM: NaCl 118, KCl 4.7, CaCl₂ 2.5, KH₂PO₄ 1.2, MgSO₄ 0.6, NaHCO₃ 25 and glucose 11.1) containing corticosterone (40 μ M), desmethylmipramine (10 nM) and propranolol (100 nM) and gassed with 95% O₂ and 5% CO₂. Bath fluid, at 30°C, was renewed continuously at a rate of 3–5 ml min⁻¹.

The intramural nerves of the vas deferens were stimulated by rectangular 40 V pulses of 3 ms duration at a frequency of 0.1 Hz. Clonidine inhibits this stimulation-evoked twitch response. Cumulative clonidine concentration-response curves were constructed in the absence and presence of a range of concentrations of each antagonist. Antagonists were allowed to equilibrate for 15 min before clonidine administration. Each antagonist (except RX 781094) was tested at three concentrations in the range indicated: RX 781094 (4.2 nM – 4.2 μ M, four concentrations), Wy 26663 (27 nM – 2.7 μ M), yohimbine (26 nM – 260 μ M), rauwolscine (26 nM – 2.6 μ M), piperoxan (4 nM – 400 nM), mianserin (33 nM – 3.3 μ M), RS 21361 (360 nM – 3.6 μ M).

Antagonist concentrations (K_e values in nM) to produce a 2 fold parallel shift of the clonidine concentration-response curve were calculated according to the method of van Rossum (1963). Schild plots of log (clonidine concentration ratio - 1) against log (molar concentration of antagonist) were

also constructed (Arunlakshana & Schild, 1959) and the slopes of the plots determined.

Drugs

The following drugs were used in the study: clonidine hydrochloride (Boehringer Ingelheim), corticosterone (Sigma), desmethylimipramine hydrochloride (Ciba Geigy), guanoxabenz hydrochloride (Roussel), mianserin hydrochloride (Organon), piperoxan hydrochloride (synthesized in the Medicinal Chemistry Department, Reckitt and Colman), (\pm)-propranolol hydrochloride (ICI), rauwolscine hydrochloride (Roth), RS 21361 hydrochloride [2-(1-ethyl-2-imidazolyl methyl)-1,4-benzodioxan] (Syntex), RX 781094 [2-(2(1,4-benzodioxanyl)) Wy 26703 [N-methyl-N-(1,3,4,6,7,11ba-hexahydro-2H-benzo (α)-quinolizin-2 β -yl) isobutanesulphonamide] 2-imidazole hydrochloride] (Reckitt and Colman), Wy 26703 [N-methyl-N-(1,3,4,6,7,11ba-hexahydro-2H-benzo (α)-quinolizin-2 β -yl) isobutanesulphonamide] (Wyeth), yohimbine hydrochloride (Sigma). [^3H]-RX 781094 was prepared by catalytic bromine-tritium exchange (Radiochemical Centre, Amersham) and purified (>99%) by preparative thin layer chromatography on silica gel 60 F₂₅₄ plates (Merck) using chloroform:methanol:880 ammonia (91:8:1 by volume) as the solvent system. The specific activity was 30 Ci mmol⁻¹.

In the binding studies, drug solutions were made up in distilled water or dilute hydrochloric acid. Drugs applied to the vas deferens were dissolved in distilled water or 0.9% w/v sodium chloride solution (saline). For i.v. administration, drugs were dissolved in sterile saline or distilled water (dilutions in sterile saline) and given in a dose-volume of 1 ml kg⁻¹.

Results

[^3H]-RX 781094 binding in rat cerebral cortex

The binding of [^3H]-RX 781094 to rat cerebral cortex membranes was a saturable process over the concentration range studied (0.1 to 30 nM) (Figure 1a). Non-saturable binding, defined by the addition of an excess of phentolamine (10 μM), represented about 30% of the total binding. Scatchard analysis of the data yielded linear plots consistent with binding to a single homogeneous population of binding sites with an equilibrium dissociation constant (K_D) of 4.4 ± 0.4 nM (mean \pm s.e. mean of 5 determinations). The maximal number of binding sites was 144.4 ± 14 fmol mg⁻¹ protein. A typical Scatchard plot is shown in Figure 1 (b).

Comparison between peripheral and central α_2 -adrenoceptor antagonist activity

A structurally diverse group of α_2 -adrenoceptor antagonists covering a fairly wide potency range was assessed against clonidine-induced inhibition of the rat isolated vas deferens, [^3H]-RX 781094 cortical binding and guanoxabenz-induced mydriasis in the anaesthetized rat. The antagonists were: RX 781094, Wy 26703, yohimbine, rauwolscine, piperoxan, mianserin and RS 21361.

In the vas deferens, each antagonist produced a parallel concentration-dependent shift to the right of the clonidine concentration-response curve. Plots of log (clonidine concentration ratio - 1) against log (antagonist concentration) gave linear regressions with slopes close to unity (see Table 1), consistent with competitive antagonism. Schild plots for

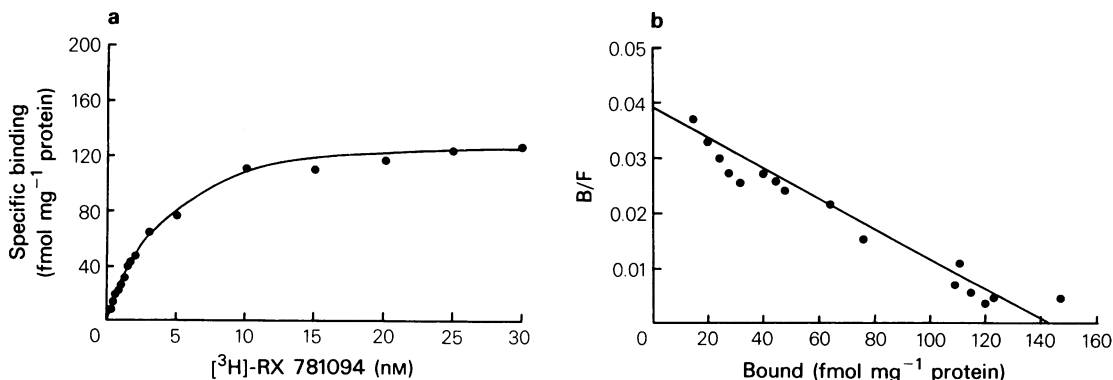


Figure 1 (a) Specific binding of [^3H]-RX 781094 to rat cerebral cortex membranes in physiological salt solution at 25°C. Non-specific binding was determined with 10 μM phentolamine. Each point is the mean of three determinations. (b) Scatchard plot of the data shown in (a). Each point is the mean of three determinations and the line was fitted by linear regression analysis. The maximum number of binding sites (B_{max}) indicated by the intercept on the abscissa scale was 142.7 fmol mg⁻¹ protein and the dissociation constant (minus 1/slope) 3.7 nM. A Hill plot of these data has a slope of 0.94 indicating a lack of cooperativity in binding.

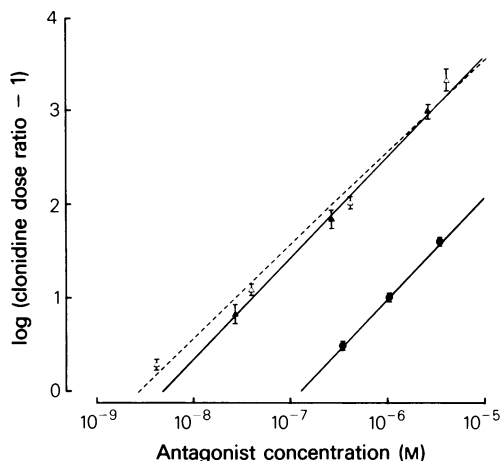


Figure 2 Schild plots for the interaction between selective α_2 -adrenoceptor antagonists and clonidine on the electrically stimulated (0.1 Hz) rat isolated vas deferens. Antagonists: RX 781094 (Δ , dashed line), Wy 26703 (\blacktriangle) and RS 21361 (\bullet). The lines were fitted by linear regression analysis of the data. Each point and vertical bar represents the mean and s.e.mean of a minimum of three determinations. The slope of these Schild plots are presented in Table 1.

RX 781094, Wy 26703 and RS 21361 are shown in Figure 2.

Each antagonist caused a concentration-dependent displacement of [^3H]-RX 781094 binding from cortical membranes. Competition binding curves (Hill plots) were linear with Hill coefficients

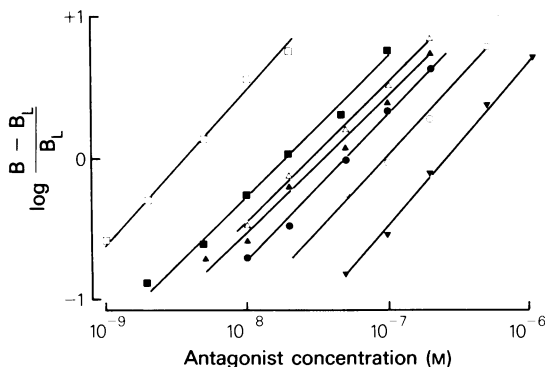


Figure 3 Representative Hill plots showing linear regression of $\log (B - B_L)/B_L$ against \log antagonist concentration. B is specific [^3H]-RX 781094 binding in the absence of competing antagonists and B_L specific binding at a particular concentration of antagonist. Antagonists: RX 781094 (\square), Wy 26703 (\blacksquare), piperoxan (\triangle), yohimbine (\blacktriangle), rauwolscine (\bullet), mianserin (\circ), RS 21361 (\blacktriangledown). Each point is the mean of three determinations. The respective Hill coefficients of these plots are presented in Table 1.

(n_H) close to 1 (Figure 3, Table 1), indicating that all the antagonists were acting in a competitive manner.

The α_2 -adrenoceptor antagonist potencies in the rat vas deferens and central nervous system (assessed against [^3H]-RX 781094 cortical binding and guanoxabenz-induced mydriasis) are presented in Table 1. The relative potencies of the compounds were similar in the three tests. Linear regression analysis of the data revealed that antagonist activity in the vas deferens was positively correlated with the ability of these agents to reverse guanoxabenz-induced mydriasis (Figure 4a) and to displace [^3H]-RX 781094 binding (Figure 4b). There was also a highly significant correlation between the antagonist effects in the two tests of central activity ($r=0.97$; $P<0.001$).

Discussion

This study has compared the pharmacological activities of several structurally heterogeneous α_2 -adrenoceptor antagonists at presumed α_2 -adrenoceptors in the vas deferens and CNS of the rat. As far as is known from published data, all the agents used are thought to act as pure antagonists at α_2 -adrenoceptors, though it should be noted that not all of them are selective for this particular sub-type. RX 781094, RS21361 and Wy 26703 possessed the highest α_2 -adrenoceptor selectivities relative to their effects at α_1 -adrenoceptors (see Introduction for references).

The pharmacological activity of the antagonists in the vas deferens was highly correlated with both their ability to displace [^3H]-RX 781094 binding from cortical sites and their ability to reverse the centrally-mediated mydriasis induced by the selective α_2 -adrenoceptor agonist guanoxabenz (Berridge *et al.*, 1983). These results therefore provide convincing evidence that these peripheral and central α_2 -adrenoceptors are similar in terms of their pharmacological responsiveness. Furthermore the significant correlation between the antagonist activities in the two central tests indicates that [^3H]-RX 781094 binding sites are pharmacologically similar to the functional α_2 -adrenoceptors responsible for mediating mydriasis. Whether this close relationship holds for other functional α_2 -adrenoceptors in the CNS remains to be determined.

Previous studies, mainly employing α_2 -adrenoceptor agonists, have also demonstrated a pharmacological similarity between several identified central and peripheral α_2 -adrenoceptors. For instance, in a study with a series of three agonists, Hammer, Kobinger & Pichler (1980) found central α_2 -adrenoceptor sites labelled by [^3H]-clonidine to be similar to postsynaptic α_2 -adrenoceptors on vascu-

Table 1 α_2 -Adrenoceptor antagonist activity assessed against clonidine-induced inhibition of the rat isolated vas deferens, [3 H]-RX 781094 cortical binding and guanoxabenz-induced mydriasis in the anaesthetized rat

Antagonist	α_2 -Adrenoceptor antagonist potency				
	Vas deferens K_e (nM)	Schild plot slope	[3 H]-RX 781094 binding K_i (nM)	Hill coefficient n_H	Guanoxabenz-induced mydriasis AD_{50} ($\mu\text{mol kg}^{-1}$, i.v.)
RX 781094	3.0 \pm 0.2 (21)	1.00 \pm 0.04	3.1 \pm 0.4 (3)	1.04 \pm 0.10	0.2 \pm 0.05 (5)
Wy 26703	4.1 \pm 0.6 (10)	1.07 \pm 0.08	17.6 \pm 2.5 (3)	0.87 \pm 0.07	0.2 \pm 0.01 (5)
Yohimbine	8.0 \pm 0.7 (8)	0.92 \pm 0.10	40.0 \pm 5.5 (3)	0.86 \pm 0.07	2.2 \pm 0.04 (5)
Rauwolscine	16.0 \pm 3.0 (14)	1.03 \pm 0.08	43.0 \pm 8.2 (3)	0.92 \pm 0.08	3.2 \pm 0.2 (5)
Piperoxan	20.0 \pm 1.6 (14)	0.96 \pm 0.04	34.0 \pm 6.5 (3)	1.03 \pm 0.07	3.1 \pm 0.2 (5)
Mianserin	53.0 \pm 12.0 (6)	0.90 \pm 0.16	82.0 \pm 2.4 (3)	0.95 \pm 0.03	10.0 \pm 0.9 (5)
RS 21361	107.0 \pm 5.0 (10)	1.09 \pm 0.03	190.0 \pm 21.0 (3)	1.02 \pm 0.03	16.2 \pm 2.5 (5)

Values are mean \pm s.e.mean of the number of experiments indicated in parentheses. K_e values for yohimbine, rauwolscine, piperoxan and RS 21361 were calculated from pA_2 values from Doxey *et al.* (1983). AD_{50} values against guanoxabenz-induced mydriasis are from Berridge *et al.* (1983), except Wy 26703.

lar tissue. Similarly, Kobinger & Pichler (1980) concluded on the basis of the rank order of potency of eight clonidine analogues, that central α_2 -adrenoceptors mediating sympatho-inhibition resemble those on vascular tissue.

Attempts to correlate pharmacological activity at central and peripheral α_2 -adrenoceptors may be influenced by factors other than differences in the drug responsiveness of the particular receptors under study. For example one of the major determinants of a drug's potency in the CNS is its ability to penetrate the blood brain barrier. All the antagonists used in this study seemed to have ready access to the CNS as

shown by the excellent correlation between central α_2 -adrenoceptor affinity *in vitro* and central antagonist activity *in vivo*. The ability of these drugs to penetrate into the CNS should allow other putative central α_2 -adrenoceptors to be characterized by use of this pharmacological approach.

There is still considerable uncertainty as to whether central α_2 -adrenoceptors are located predominantly on noradrenergic terminals and cell bodies or on postsynaptic neuronal structures. It has been proposed that activation of presynaptic α_2 -adrenoceptors inhibits K^+ - and electrically-induced [3 H]-noradrenaline release from brain slices (Dis-

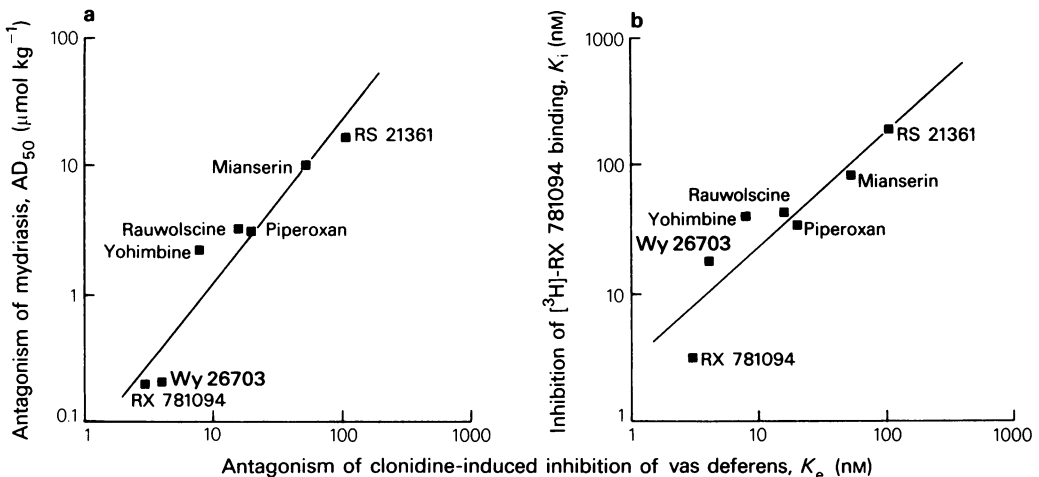


Figure 4 Comparison of the effects of α_2 -adrenoceptor antagonists on clonidine-induced inhibition of the rat vas deferens (K_e values) and either (a) guanoxabenz-induced mydriasis in the pentobarbitone-anaesthetized rat (AD_{50} values) or (b) [3 H]-RX 781094 binding in rat cortical membranes (K_i values). Linear regression analysis of the untransformed data (shown in Table 1) gives positive correlation coefficients of $r = 0.98$ (K_e versus K_i values; $P < 0.001$) and $r = 0.99$ (K_e versus AD_{50} values; $P < 0.001$).

mukes, De Boer & Mulder 1977; Taube, Starke & Borowski, 1977; Wemer, Frankhuyzen & Mulder 1982). The existence of functional presynaptic α_2 -adrenoceptors could also account for the increased turnover of cortical noradrenaline seen after RX 781094 administration (Flockhart, Haynes & Walter, 1982; Scatton, Dedek & Zivkovic, 1983). This would be analogous to the presynaptic, negative feedback mechanism that is thought to modulate noradrenaline release from sympathetic nerve endings in some peripheral tissues (Langer, Adler-Graschinsky & Giorgi 1977; Story, McCulloch, Rand & Stanford-Starr, 1981).

It appears unlikely, however, that [3 H]-RX 781094 labels mainly presynaptic α_2 -adrenoceptor sites in the cerebral cortex. If a proportion of α_2 -adrenoceptors were located presynaptically then removal of cortical noradrenergic terminals would be expected to reduce [3 H]-RX 781094 binding. However, extensive destruction of the noradrenergic innervation to the rat cerebral cortex has no effect on either the number (B_{\max}) or affinity (K_D) of

[3 H]-RX 781094 binding sites in this brain region (Langer, Pimoule & Scatton, 1983; present authors, unpublished observations). This result is partly consistent with the finding that [3 H]-clonidine binding in cerebral cortex is not reduced, but increased, after lesioning the dorsal noradrenergic bundle (U'Prichard, Reisine, Mason, Fibiger & Yamamura 1980). There is thus no evidence to date from radioligand binding studies that α_2 -adrenoceptors are located on noradrenergic terminals in the cerebral cortex. Further studies are clearly necessary in order to determine the anatomical location of these receptor sites and to investigate the mechanism by which they regulate central noradrenergic activity under normal physiological conditions.

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Correspondence to I.F.T.

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