NOTES

The *lonD* Gene Is Homologous to the *lon* Gene Encoding an ATP-Dependent Protease and Is Essential for the Development of *Myxococcus xanthus*

NOBUKI TOJO,¹ SUMIKO INOUYE,^{2*} AND TERUYA KOMANO¹

Department of Biology, Tokyo Metropolitan University, Minamiohsawa, Hachioji-shi, Tokyo 192-03, Japan,¹ and Department of Biochemistry, Robert Wood Johnson Medical School, University of Medicine and Dentistry of New Jersey, Piscataway, New Jersey 08854²

Received 12 March 1993/Accepted 4 May 1993

Myxococcus xanthus contains two genes (lonV and lonD) homologous to the Escherichia coli lon gene for an ATP-dependent protease. We found that the lonD gene encodes a 90-kDa protein consisting of 827 amino acid residues. The lonD gene product shows 49, 48, and 52% sequence identity to the products of the M. xanthus lonV, E. coli lon, and Bacillus brevis lon genes, respectively. When a lonD-lacZ fusion was used, lonD was expressed during both vegetative growth and development. However, while lonD-disrupted strains were able to grow normally vegetatively, the development of M. xanthus was found to be arrested at an early stage in these strains. The mutant strains were able to form neither fruiting bodies nor myxospores.

Myxococcus xanthus is a unique gram-negative bacterium that undergoes multicellular development involving a cellcell interaction (for a review, see reference 16). Upon nutrient depletion on a solid surface, cells aggregate to form mounds in which rod-shaped cells are converted into round or ovoid myxospores. The mounds of mature myxospores are called fruiting bodies.

We previously isolated six developmental mutants of M. *xanthus* defective in fruiting body formation (Fru⁻) by screening 855 independent kanamycin-resistant (Km^r) clones mutagenized with TnV, a Tn5 derivative transposed carrying the replication origin of Escherichia coli plasmid pSC101 (5). Mutants 221, 328, and 530 were shown to have TnV insertions within a 0.25-kb region of the M. xanthus chromosome. Fifteen-kilobase Sall fragments with TnV insertions were cloned from the chromosomal DNAs of these mutants. As described below, the three insertions were located within a single gene that is essential for fruiting body formation. Here, we show that this gene is homologous to the E. coli lon gene and is thus designated lonD, for the lon gene required for development. We have independently identified another gene homologous to the E. coli lon gene; this gene has been shown to be essential for the vegetative growth of M. xanthus and is thus designated lonV (formerly lon [19]).

The lon gene of E. coli encodes protease La (2, 4), an ATP-dependent protease associated with cellular protein degradation. Mutations in the E. coli lon gene result in pleiotropic phenotypes, such as increased sensitivity to UV irradiation and SOS-inducing agents, filament formation, mucoidy, and reduced degradation of various abnormal proteins and certain normal proteins.

For cloning the wild-type allele of the *lonD* gene, a genomic library of *M. xanthus* DZF1 was screened by use of the nick-translated 15-kb SalI DNA fragment (5) as a probe.

A positive clone, P576, was identified; from it a 13.5-kb EcoRI fragment was subcloned into pUC9 (20) to yield pMXL101. For defining the DNA region of the lonD locus, 13 new insertion mutations were constructed as described previously (11), and their developmental phenotypes were tested (Fig. 1). First, pMXL101 DNA was digested with various restriction enzymes and ligated with a 1.3-kb DNA fragment containing the Km^r gene of Tn5 to generate insertion mutations on pMXL101. P1 incompatibility genes were added to the plasmids, which then were introduced into M. xanthus DZF1 by P1 transduction (18). The wild-type lonD allele of the chromosome was replaced with mutant alleles by double-crossover recombination (18). In each case, replacement-type transductants were selected by screening for Km^r transductants by colony hybridization (15) with vector DNA as a probe. When fruiting body formation was induced on clone fruiting (CF) agar plates (8), of 13 insertion mutants, 4 mutants were normal in fruiting body formation (Fru⁺), 8 mutants were defective (Fru⁻), and 1 mutant was leaky (Fru[±]), as shown in Fig. 1. In the leaky mutant, fruiting bodies of abnormal morphology were formed at a later stage



FIG. 1. Restriction map of the area near the *lonD* gene of *M. xanthus*. The locations of insertion mutations at various restriction sites and TnV insertions are indicated. Symbols: \bigcirc , Fru⁺ mutations; ●, Fru⁻ mutations (the rightmost Sm site yields a Fru⁺ mutation); ▼, TnV insertions with a Fru⁻ phenotype. The solid arrow below the map denotes the *lonD* coding region and its orientation. Ac, AccIII; Ap, ApaI; B, BamHI; E, Eco47III; G, BgIII; M, MluI; P, PstI; Sa, SacII; Sc, ScaI; Sm, SmaI; X, XhoI.

^{*} Corresponding author.

Petl

BamHI Eco47111 BamHI 361 TGCGAGACGCCGCTGCCCTCCTTCAGCGCCTCCAGGAAGGCGGGGCAGCTCCGTGGCGATGGCGTTCTGCTCCTTCCATGGATCCGGAGTTGTCGATGACGAAGAGGATGTCCGTCTTC Accill Taq I 721 TGGAGAGGTGACGGGGGGGGGGGGAAAAACGCACCTGCGT<u>ITGCCA</u>TGTCCGTTTTGGTTGT<u>IGCCGTGTGTGTGTCATCCGCCGCAGAAAACCTGGAAATGTTCACTGGTTGTCGAAGAGAAAAG</u> 721 TGGAGAGGTGACGGGGGGGGGGGAAAAAGGCALACIGUGIIIIGUCAIGIGUGITTIGGTGGTGGCCCCTCCGGGGGCTCATCAACAAGGAAGACATCCCGCAGGTGCTTCCCATCCTCCC Eco47111 830 841 GCTGAATTTCATGTCCGATGAGAAGAAGAAGAAGGGGCTCCGCTGCGAGGCGCTATGCCCACCGCGGGGCCCCTCCGGGGGCTCATCAACAAGGAAGAACATCCCGCAGGTGCTTCCCATCCTCCC M S D E K K K G S A A S A M P T A M A P P G L I N K E D I P Q V L P I L P M S D E K K K G S A A S A M P Smal G V L P L A V G R Q K T I A L I K D A V R D D Q V I G V 1201 CGCCCGCTTCCGCGTGGTGGAGCTGGTCCAGGAAGCGCCCTACCTCAAGGCCCGCGTGGACGCCGTGGAGGACAAGACCTCTTCGGAGAACGTGGAAGTCGAGGCGCTGGGCATCAACCT 118 A R F R V V E L V Q E A P Y L K A R V D A V E D K T S S E N V E V E A L G I N L 1321 CAAGAAGCTGGCGCGCGGGGTCATCGAGCTGATGCCCGAGCTGCCGCCGCCGCCGCCGCCGCGGGGAGGCATCACCCCGCCGCCGCCGCCGACCTGATCGCCGCCACCTGGCGGA 158 K K L A R E V I E L M P E L P A A A T E L V E S I T H P G H L A D L I A A N V D 1441 CGTGCCCATCGAGGAGAAGCAGGCCGTCCTGGAGACGGTGGACCTCAAGGCCCGGATGAAGCTCGTGCTGGAGCTGCTCAACCGGAAGCGGGAGATCCTCAAGGCCCCAACAAGATCGA 198 V P I E E K Q A V L E T V D L K A R M K L V L E L L N R K R E I L K L S N K I D Scal 1561 CICCGCCGTGAAGGGCGAGATGICGAAGACCCAGCGC<u>GAGIACT</u>ACCTGCGCCAGCAGCTCAAGGCCATCAAGGAAGAAGAAGAGGGAGATGGGGGAGAAGAGGAGCTCGACGAG<u>C</u>T Pstl 238 SAVKGEMSKTQRE<u>YY</u>LRQQLKAIKEELGEMGEEEEELDE 1681 <u>GCAG</u>GAGCGCCTGAAGAAGGCCGGCCTGCCGCCCGACGAGGAGAAGGTCCCAACAAGGAGCTCAACCGCCTGAAGAACCATTCCGGCGGCGCCTCCAGCGAGTACACCGTCGCGCGCCACCTA 278 Q E R L K K A G L P P D V E K V A N K E L N R L K T I P A A S S E Y T V A R T Y Bg111 1801 CCTGGATTGGATCGCCGACCTGCGTGGGCGA<u>AGATC</u>TCCGAGGACAACCTCGACATCGAGAACGCGCGCCAGCAGCTGGACAAGGATCACTTCGGCATCAAGAAGGTGAAGAAGCGCAT 318 L D W I A D L P W A K I S E D N L D I E N A R Q Q L D K D H F G I K K V K K R I 1921 CCT6GAGTACCT6GCCGTCCGCAAGCTGAAGAACGACGT6GCCCCATCCTGT6CCTCGTCGGTCCGCCGGCGAAGACGTCGCCGCGAGAGCGT6GCCAAGACGTCGCCGTCGCCAAGACGTCGCCAAGACGTCGCCAAGACGTCGCCAAGACGTCGCCAAGACGTCGCCAAGACGTCGCCAAGACGTCGCCAAGACGTCGC 358 L E Y L A V R K L K N D M R G P I L C L V G P P G V G K T S L G Q S V A K A T G SacII 2281 CCACTACCTCGACGTGCCCTTCGATTTGTCCAAGGTGATGTTCGTCGCCACGGCGAACCAGCTGGACCCCATCGCCGTGCGGTGACGGATGATCGAGGGGATCATCGAGGGGCTA 478 H Y L D V P F D L S K V M F V A T A N Q L D P I P G P L R D R M E I I E L T G Y 518 TFEEKQSIARIHLVPKQLKEHGLSPDHIDITDEALLTLTT 558 A Y T R E A G V R N L E R R I A D I C R A V A V E V A G G K T E K Q T I N A D R Apal EMFYSEVAERTEVPGVATGLAWTAAGGDLLF 638 A T K M A G K G G M T L T G Q L G D V M K E S A T A A L S Y L R S K A E Q L G I Sacii 2881 CAGCCCGAACTTCCTGGAGAAGACGGACCTGCACCTGCACCTGCACCTGCCGCGGGGCTCCATTCCGAAGGACGGGCCTT<u>CCGCGG</u>GCGTCACCAGCGCGCGCCCCACCAGCCTCCCGACGGG 678 SPNFLEKTDLHLHFPAGSIPKDGP Ğ ILTALTSLLT 718 I R V R H D T A M T G E A T L R G L V L P V G G I K E K V L A A H R A G I K R 758 I L P E R C R K D L I D V P D Q A R N E L E F I F V T H M D D V L K A A L E T Smal 3241 TCCCGTCGGCGTGGCGGGGAAC<u>CCCGGG</u>CGGTGAGCCGGGCAAGGAGGCTCCGCTGCCGAAGCCGGCCCGAGGTCCGCCCCTGAGGTCCGCGCCTAGCGCACGGCCTTCCAGCCAATAGGAAG 798 P V G V A G T P G G E P G K E A P L P K P A E S A P E V R A * G G E P G K E A P L P K P A E S A P E V R A 3361 TGACACGGGCAGGTTCCCTTCTTCCGGGAACCTGCCCGTTGTCTTTGCGGGGCAGGTGGCGCTCAGAGCGAGAACAGGAAGTGTCGCGGCTCCGGGACGCGCTTCACCCGCAGGATGTGA

Mlu1 3481 CTGTGCTCCACAGGCAGCCGGGGGATGGCCTCTGGACGGGGAACGTAGAGGATGCCGGGGTCATCCTCTCGGGTGAAGACGTACTGCCCGCGCGAGCCCTTACGCGT

FIG. 2. Nucleotide sequence of the *lonD* gene of *M. xanthus* and its deduced amino acid sequence. Numbers to the left of the top line enumerate the nucleotide bases, and numbers to the left of the bottom line indicate the amino acid residues. Relevant restriction sites are indicated. Duplicated 9-bp sequences at TnV insertion sites are underlined and identified by insertion numbers. Putative promoter sequences are boxed, and a putative Shine-Dalgarno sequence is also underlined.

Mxa	lonD	MSDEKKKGSAASAMPTAMAPPGLINKEDIPQV	VLPI UP LENS VF FRG GVLP FANGEORTHALIKDAV RODOVIGV VTOR RATE FOR 8
Mxa	lonV	MFFGRDDKKEAQKRGLT	TVPLEPLED I IVTPHNVVPLTVGREKSTA AFKDAMAHKGPDDKAVILHA AOKKAKTNDP 7
Eco	lon	MNPERSERIE	IPVINE LED V VVYPHM VIP IPVICE FKS IR COLLAMDHDKK -IMIV JOK TAS TO TO
Bbr	lon	MGERSGKRE	LPLLP INGLIVYPTMVL HIDVAREKSIR ALTONYY DDNVHLIA TOPYYTEP
Mara	lonD		
Mara	lonV		AND KVVEDVQEAPILKAKVDAVODKAS SONVOVSAUGINLKKLAREVNEDNPELOAAA 17
MX4	TOUA	TPDDI FRYGALGRY LOLLPL PDGAY KVLVEQV	/RRAKVKKFHPNDAJENVEVEZVƏZQT-IKTVƏLƏALVRSVHSVFƏAPVKIMRRIPPEM 16
ECO	lon	GANDER INGTANS I TODOREDDEAD KARADED	ORARISALSDNGEHBSAKAEYLEBSPELTDER-BOEVLVRTAISOF BGYLKUNRK IP DEV 15
Bbr	lon	DAEQIYSIGTVARVKOMLKIPNGTI R <u>VIVEGL</u>	<u>qra</u> kieeylqkedygvvsitylkeekaegn-gv padmr sllth pp qy <mark>iki</mark> skkvs <u>pp</u> t
яха	TOUD	TE LVESHTHPGHLADLIAANVDVPIEEKOA VI	etvolkarniklvlellnrkreiliklsnkidsavkgensktorevvilr <u>colkarkee</u> lg 26
Mxa	lonV	on quashd dearladth vahl surl ndkoa lu	JETESPÄKRLEKLYKLNQG <mark>E</mark> IEILQVEKKIRTRVKKQMEKTQKEYYDNEQMOAIQKELG 25
Eco	lon	LTSLNSID DEARLADTHAAHN PURLADKOS VU	ENSDVNER GEYCHANNES <mark>EIDLO</mark> QVER RURNRVKKQME <u>KS</u> QREYYDNEQMKA IQKELC 24
Bbr	lon	gt svo dir figgreadvilasht porm koror io	<u>- Bei</u> vniq erleig ltilnnerev ge lerkignr vkkomer <u>tor</u> eyydreomkaiokeuc 24
Мха	lonD	MGEEEELDE OERKKAGL PDVEKVANNS	ednrekt <u>i</u> paasseyhvartyldwiaderwakisedmudienarqquudkuhyginxvkk 35
Mxa	lonV	Brdef Knbi qbi ee kuknkrm <u>s</u> keat l <mark>k</mark> v k <u>k</u> b	elk kur MMS PMS A BATVVRNY I DWI I SLPWYD I TODRLDV TEALTVLN EDHYGLKKPKI 34
Eco	lon	BNDDAPDENE<u>AU</u>KRK<u>I</u>DAAKN<mark>P</mark>KEAKEKAE<u>A</u>B	LQ KUKMUS PMSAEAD VVRGYID WMVQVPWNARSKVKKDLRQAQ E ILDTDHYGDERVKD 33
Bbr	lon	dkigrqqiy dilira queksdaper i kaki eke	die Rie Rip 8 t <u>sae</u> g svirt <u>y id</u> t ly <u>agew</u> tktt zonddikhae z vudzoh y guzkoka 33
Мха	lonD	RTLEVIAVRELENDERGETLELVGPROVCKTS	
Mxa	lonV	ELEYLANOL WELLGPULC WGPROVCKTS	
Eco	lon	RILEYLAVOS EVNETEGRIL CLUCPROVOETS	
Bbr	lon	RULFYLANO LVNENEGPTLCLVGPPCVCKTS	
	204		29 NOV BRANCHARTEL BACCVADAATIKCHARTELVCAUREK IEDUURUNET INEVE DE
Mxa	lonD	DEIDK <mark>LGA</mark> DFRGDP <mark>SAALLEVLDPEQNNT</mark> FSD	dhyl dvpj dlskvmevatanqldptpcplrdrmeiteltgytytekostarthlvpkol
MXA .	lonv	DEIDKUS TOFRGOPS NALLEWUDPEONH TEND	hy loldy dlskymfi cfantmint pop hodrney ir i xgyteppikistarry lip keq 52
Eco	lon	DEIDKNSSDURGDERSAMMEVIDBEONVRESD	DHYLZYDYDLISDYMEYATSNSH - NTEAPELDRMEYIRLSGYNEDEKUNHAKRHLLPKOI 51
Bbr	lon	Dendikla soerenta sangenting prond resid 	9HVI I I TVOOT NVHHI TOANS LD TOEREULORHEVUS I SCYVERLEH MULEC YFLEHOM 51
Mxa	lonD	KEHGUSPDHIDITUEALLTLTTAYTREAGVR	u der i ad i gra vave - vaggatekot i nadrvke i bopempy <u>se vaertev povato</u> e 62.
Mxa	lonV	EARGESDLKVDISDPALRTIIHRYFRESGVR S	H <u>erei</u> gov jrk iard-vlkngkrdidvdrkmamkflgt pry ry gmaeaed <u>ov g</u> i <u>vtigp</u> 61
Eco	lon	BRNAUKKGELTVDDSAIIGIIRY<u>YWRDAGVR</u>G	LERBESKLORRAYKQLLLDKSLKHIEINGDNLHDYLOVORFDYCRADNENRY GOVARD 60
Bbr	lon	Edh <mark>gl</mark> okdklommed <mark>a</mark> mlklvrlytreagvrn	u <mark>nreaanvorkaa</mark> ki-ivggekkrvvvtaktleal <mark>ig</mark> k p <u>r</u> y r <mark>yg</mark> lækkd o <mark>vg</mark> s <mark>vtgl</mark> 60
Mxa	lonD	AWTA ACCOLOF IBAT KMACKCOMTLTCOLCOV	/MKESATAALSYLRSKAEQLGISENGLEKTDLHLHTEASSIEKDGESAGVHILTALTSL 71
Мха	lonV	Awne loce ilt teat inp <u>gkgkui</u> it ck <u>lo</u> ev	/MQESAQAAMSYVRSRAERIGIDRKVIENYDIHVHLPEGAIPKDGPSAGVTICTALVSA 70
Eco	lon	AWTE VGGDLUT IETACVPGKGKLTYTCSLCEV	/ HQE SIQAAL TVVRARAEK LGINPDFYEKRDIHVHVPEGATPKDGPSACIAMCTALVSC 69
Bbr	lon	<u>ANTO AGGDTLN VEVSILAGKGKUTLT GOLGDV</u>	/EKESAQAAYSYIRSRASENGIOPIPHE KADIHIHVPEGAIPKDGPSACIAAATALVSA 69 •
Мха	lonD	LTCIRVENDTANTGEATLECLVLEVCCIKEKV	BAAHRAGI KRVI LERCKIDELDVED OARNEERTI WTHMDVEKAADE - TEPVGVAG 80
Mxa	lonV	LTRVLIRRDVAMTGEITLRGRVLPIGGLKEKT	LAAHRAGH KEVL IPK ANKKOLKOLPL KIRKOLRIVPVE FVDDVER PALVLEKPEEFGR 79
Eco	lon	LTGN PV RADVANTGEITLRGOVLPLGGLKEKL	A A A HEGG I KAVIL IPP ENK RO DE ELEDNVI A DUDI HEVKRIE EVIT LA LON BESOMHIS 78
Bbr	lon	LTC <mark>IPVKKFVC</mark> MTGEITLRG <mark>R</mark> VLPIGGLKEK <mark>C</mark>	CMSAHRAGL THIILDER DIENER SVERALTEYPYCHLDEVIC HALTKOPYCOKK 77
Mxa	lonD	TPGGEPGKEAPLPKPAESAPEVRA 827	1
Mxa	lonV	KPTTDGGKLGGTTELPASPAVAPA 817	7
Eco	lon	LRRRCSTASTYYWAAS 793	3
Bbr	lon		

FIG. 3. Alignment of four lon amino acid sequences. Mxa lonD, M. xanthus lonD gene; Mxa lonV, M. xanthus lonV gene; Eco lon, E. coli lon gene; Bbr lon, B. brevis lon gene. Identical amino acid residues in three or all four sequences are printed in white on black. Putative ATP-binding sequences are underlined, and a serine residue at the putative active site is marked with a circle. M. xanthus lonV, E. coli lon, and B. brevis lon sequences were taken from Tojo et al. (19), Amerik et al. (2), and Ito et al. (10), respectively.

than normal (data not shown). These results indicate that the *lonD* locus is located within the 2.9-kb *AccIII-MluI* region.

Using some of the insertion mutants described above, we analyzed further other characteristics of the lonD mutations. The lonD mutants appeared to stop their development at a very early stage; they could not aggregate at all, and no spores were found, even after 7 days on CF agar plates. In contrast, no effect on vegetative growth was noted, and the mutants could form spores when 0.5 M glycerol was added to liquid cultures (16). As noted below, since lonD was found to be identical to bsgA (7), extracellular complementation with wild-type cells was expected during development (6, 8, 14). However, when the lonD mutants were mixed with wild-type cells and examined for complementation on CF

agar plates, the *lonD* mutants failed to form spores. This result agrees with the experiment carried out by Kroos and Kaiser to examine extracellular complementation with *bsgA330* (12).

Nucleotide sequence of the lonD gene. The nucleotide sequence of the 3,587-bp PstI-MluI fragment was determined by the dideoxy chain termination method (17) (Fig. 2). The locations of TnV insertions in mutants 221, 328, and 530 were also determined (underlining in Fig. 2). In each TnV insertion, a 9-bp target duplication was found.

Analysis of the nucleotide sequence revealed that there is an open reading frame that shows homology to *E. coli lon*. A putative *lonD* initiation codon was deduced by the coding frame analysis proposed by Hagen and Shimkets (9), on the basis of the observation that the G+C content at the third codon position is very high within the protein coding region of M. xanthus genes. This analysis suggested that the lonD gene starts with the ATG codon at position 851. Construction of a lonD-lacZ fusion gene by use of the Eco47III site at position 884 as described below supported this suggestion. Furthermore, the insertion of 2 nucleotides at the TaqI site at position 828 did not affect the expression of the lonD-lacZ fusion gene. Thus, we conclude that the lonD gene starts with the ATG codon at position 851 and ends with the TAG codon at position 3332. The average G+C contents at positions 1, 2, and 3 of codons in the lonD gene were 68, 39, and 94%, respectively. The putative lonD initiation codon is preceded by a purine-rich Shine-Dalgarno-like sequence (AAGG). Possible promoter sequences, TTGCCA (positions 769 to 774) and TACGTT (791 to 796), that share homology with the *E*. coli σ^{70} consensus promoter sequence were found in the upstream region.

Amino acid sequence of the lonD gene. The proposed lonD coding sequence could encode a protein of 827 amino acids with a calculated molecular weight of 90,433 (Fig. 2). When the deduced amino acid sequence of lonD was compared with known protein sequences, striking similarities to *E. coli* and *Bacillus brevis lon* genes, with 48 and 52% identity, respectively, were found (2, 10). Furthermore, the proposed amino acid sequence of the lonD gene shares 49% identity with that of the *M. xanthus lonV* gene, which was recently identified as a gene essential for vegetative growth by use of the *E. coli lon* gene as a probe (19). Thus, we can conclude that *M. xanthus lonV* gene is essential for vegetative growth (19), and the lonD gene is essential for development.

Figure 3 shows an alignment of four lon amino acid sequences. They share highly conserved sequences encompassing almost the entire region. ATP-binding sequences [segment A, $(G/A)X_4(G/A)(H/K/R)X_{0-1}(T/S/K/R/H)$, and segment B, $(H/K/R)X_{5-8}\Phi X \Phi_2(D/E)$, where X and ϕ stand for any amino acid and a hydrophobic amino acid (4), respectively] were found in the lonD sequence at the same positions as in the others, and the similarity near these sequences was relatively high for the four sequences (Fig. 3, underlining). Recently, an active-site serine residue in the E. coli lon gene was proposed by site-directed mutagenesis (1). This serine residue (indicated with a filled circle) was also conserved in the four sequences (Fig. 3). The similarity in amino acid sequences suggests that the lonD product is an ATP-dependent protease. It is noteworthy that the level of similarity of the amino acid sequence of lonD to those of the other three lon products is rather low in the N-terminal region.

Expression of the *lonD* gene. To investigate the expression of the *lonD* gene, we constructed a fusion of the *lonD* gene with the *lacZ* gene of *E. coli*. The 12th codon of the *lonD* gene was joined in frame to the 8th codon of the *lacZ* gene by insertion of the 666-bp *Eco*47III fragment (positions 221 to 886; Fig. 2) into the *SmaI* site of pMC1403 (3) in the proper orientation. The P1 incompatibility and Km^r genes were added to the resultant plasmid, and the final construct was introduced into *M. xanthus* DZF1 by P1 transduction (18). The *lonD-lacZ* fusion gene was integrated into the homologous region of the *M. xanthus* DZF1 chromosome. The integrated *M. xanthus* strain was merodiploid, containing the *lonD-lacZ* gene under the control of the *lonD* gene (Fig. 4A). The chromosome structure of the integrated strain was



FIG. 4. β -Galactosidase expression during the development of an *M. xanthus* strain carrying a *lonD-lacZ* fusion gene. (A) Chromosome structure of the *M. xanthus* strain carrying the *lonD-lacZ* gene. Unique sequences of the *M. xanthus* chromosome are shown by open bars, and duplicated sequences are shown by stippled bars. A solid line indicates the vector sequence. Arrows indicate *lonDlacZ* and *lonD* coding sequences. (B) Changes in the specific activity of β -galactosidase during development induced on CF agar plates. ONP, *o*-nitrophenol.

confirmed by Southern blot analysis (data not shown). The integrated strain exhibited normal development on CF agar plates. When the strain carrying the *lonD-lacZ* gene was plated on a Casitone-yeast extract (16) agar plate containing 5-bromo-4-chloro-3-indolyl- β -D-galactoside, the colonies turned green, indicating that the *lonD* gene is expressed during vegetative growth. β -Galactosidase specific activity was measured (13) during development induced on CF agar plates (Fig. 4B). β -Galactosidase activity was present at 170 U/mg of protein at the onset of development and increased gradually during development. These results suggest that the *lonD* gene is expressed during vegetative growth and that its expression increases gradually during development.

On the basis of the present results together with our previous findings (19), we conclude that M. xanthus bears at least two essential lon-related genes, lonV for vegetative growth and lonD for development. The lonV and lonD products may degrade many regulatory proteins during vegetative growth and development. However, the specificities of lonV and lonD proteases may be different.

Recently, the nucleotide sequence of the bsgA gene of M. xanthus was determined by Gill et al. (7), and it was found that the lonD gene is identical to the bsgA gene.

Nucleotide sequence accession number. The nucleotide sequence data shown in Fig. 2 will appear in the EMBL, GenBank, and DDBJ nucleotide sequence data bases under accession number D13204.

We are grateful to M. Inouye for discussions and T. Furuichi for plasmids. We thank Susan Eagle for critical reading of the manuscript.

This work was partially supported by a grant from the National Institutes of Health (GM 26843 to S.I.).

REFERENCES

- Amerik, A. Y., V. K. Antonov, A. E. Gorbalenya, S. A. Kotova, T. V. Rotanova, and E. V. Shimbarevich. 1991. Site-directed mutagenesis of La protease: a catalytically active serine residue. FEBS Lett. 287:211-214.
- Amerik, A. Y., L. G. Chistyakova, N. I. Ostroumova, A. I. Gurevich, and V. K. Antonov. 1988. Cloning, expression and structure of the shortened, functionally active *lon* gene of *Escherichia coli*. Bioorg. Khim. 14:408–411.
- Casadaban, M. J., J. Chou, and S. N. Cohen. 1980. In vitro gene fusions that join an enzymatically active β-galactosidase segment to amino-terminal fragments of exogenous proteins: *Escherichia coli* plasmid vectors for the detection and cloning of translational initiation signals. J. Bacteriol. 143:971–980.
- Chin, D. T., S. A. Goff, T. Webster, T. Smith, and A. L. Goldberg. 1988. Sequence of the lon gene in Escherichia coli. J. Biol. Chem. 263:11718-11728.
- 5. Furuichi, T., M. Inouye, and S. Inouye. 1985. Novel one-step cloning vector with a transposable element: application to the *Myxococcus xanthus* genome. J. Bacteriol. 164:270-275.
- Gill, R. E., and M. G. Cull. 1986. Control of developmental gene expression by cell-to-cell interactions in *Myxococcus xanthus*. J. Bacteriol. 168:341-347.
- 7. Gill, R. E., M. Karlok, and D. Benton. 1993. Myxococcus xanthus encodes an ATP-dependent protease which is required for developmental gene transcription and intercellular signaling. J. Bacteriol. 175:4538-4544.
- 8. Hagen, D. C., A. P. Bretscher, and D. Kaiser. 1978. Synergism between morphogenetic mutants of *Myxococcus xanthus*. Dev. Biol. 64:284–296.
- Hagen, T. J., and L. J. Shimkets. 1990. Nucleotide sequence and transcriptional products of the csg locus of Myxococcus xanthus. J. Bacteriol. 172:15-23.
- 10. Ito, K., S. Udaka, and H. Yamagata. 1992. Cloning, character-

ization, and inactivation of the Bacillus brevis lon gene. J. Bacteriol. 174:2281-2287.

- Komano, T., T. Furuichi, M. Teintze, M. Inouye, and S. Inouye. 1984. Effects of deletion of the gene for the developmentspecific protein S on differentiation in *Myxococcus xanthus*. J. Bacteriol. 158:1195–1197.
- Kroos, L., and D. Kaiser. 1987. Expression of many developmentally regulated genes in *Myxococcus xanthus* depends on a sequence of cell interactions. Genes Dev. 1:840–854.
- Kroos, L., A. Kuspa, and D. Kaiser. 1986. A global analysis of developmentally regulated genes in *Myxococcus xanthus*. Dev. Biol. 177:252-266.
- LaRossa, R., J. Kuner, D. Hagen, C. Manoil, and D. Kaiser. 1983. Developmental cell interactions of *Myxococcus xanthus*: analysis of mutants. J. Bacteriol. 153:1394–1404.
- O'Connor, K. A., and D. R. Zusman. 1983. Coliphage P1mediated transduction of cloned DNA from *Escherichia coli* to *Myxococcus xanthus*: use for complementation and recombinational analyses. J. Bacteriol. 155:317–329.
- 16. Rosenberg, E. 1984. Myxobacteria: development and cell interactions. Springer-Verlag, New York.
- Sanger, F., S. Nicklen, and A. R. Coulson. 1977. DNA sequencing with chain-terminating inhibitors. Proc. Natl. Acad. Sci. USA 74:5463-5467.
- Shimkets, L. J., R. E. Gill, and D. Kaiser. 1983. Developmental cell interactions in *Myxococcus xanthus* and the *spoC* locus. Proc. Natl. Acad. Sci. USA 80:1406–1410.
- Tojo, N., S. Inouye, and T. Komano. 1993. Cloning and nucleotide sequence of the Myxococcus xanthus lon gene: indispensability of lon for vegetative growth. J. Bacteriol. 175:2271-2277.
- Yanisch-Perron, C., J. Vieira, and J. Messing. 1985. Improved M13 phage cloning vectors and host strains: nucleotide sequences of the M13mp18 and pUC19 vectors. Gene 33:103–119.