SHORT REPORT

Sensory nerve excitability and neuropathy in end stage kidney disease

A V Krishnan, R K S Phoon, B A Pussell, J A Charlesworth, M C Kiernan

.....

J Neurol Neurosurg Psychiatry 2006;77:548-551. doi: 10.1136/jnnp.2005.079988

Background: Peripheral neuropathy is present in 65% of patients with end stage kidney disease (ESKD) starting dialysis. Studies of membrane potential and axonal ion channel function may help explain the pathophysiology.

Objectives: To follow changes in median sensory axon excitability in patients with ESKD treated with haemodialysis, and correlate them with clinical rating scales and serum levels of potential neurotoxins.

Methods: Sensory nerve action potentials were recorded from the second digit following stimulation of the median nerve in 12 ESKD patients. Stimulus-response behaviour using two stimulus durations, threshold electrotonus to 100 ms polarising currents, a current-threshold relation, and recovery of excitability following supramaximal stimulation were recorded before, during, and after haemodialysis. Serum concentrations of potential neurotoxins were measured.

Results: Before dialysis, there were changes in nerve excitability consistent with axonal depolarisation: refractoriness was increased; superexcitability and depolarising threshold electrotonus were reduced. Following dialysis there were improvements in all indices, with correlations between excitability abnormalities and serum potassium measurements. Neuropathic symptoms correlated with excitability changes.

Conclusions: Nerves are depolarised before haemodialysis in ESKD patients. The correlation of excitability abnormalities with potassium indicates that the achievement of normokalaemia should be a priority in treating such patients.

Neuropathy in end stage kidney disease (ESKD) presents as a symmetrical length dependent process with predominant sensory involvement.¹ The pathophysiology of uraemic neuropathy has not been established, although its development has been attributed to retention of "middle molecules" (300–12 000 Daltons).² Although this hypothesis has not been proven, the concept of a dialysable neurotoxin remains prevalent.³

Assessment of nerve excitability provides information about the biophysical properties of nodal and internodal axonal function.⁴⁻⁶ Previous studies of motor nerve excitability in ESKD^{7 8} have shown pre-dialysis abnormalities consistent with axonal depolarisation, with resolution of these changes following dialysis. Given the predominantly sensory nature of uraemic neuropathy, our aim in the present study was to explore further the pathophysiology of uraemic neuropathy by recording the sensory nerve excitability properties in ESKD patients before, during, and after dialysis, and to correlate excitability changes with clinical features and levels of potential neurotoxins.

METHODS

Consecutive patients with ESKD were studied (eight men and four women, aged 17 to 70 years, mean age 53.5 years). All were receiving haemodialysis. None of the patients had a history of other illnesses known to cause neuropathy. Patients gave informed consent to all procedures, which were approved by our regional and institutional review boards.

Neurological history and examination were undertaken before dialysis, and neuropathic symptoms were graded using subsets IA, IIA, and IIB of the neuropathy symptom score (NSS).[°] Each symptom received a score of 1, and the number of symptoms present in each subset was summed to give a total neuropathy symptom score (T-NSS). The maximum possible T-NSS was 9.

Routine nerve conduction studies (NCS) were undertaken in all patients on the sural, tibial, common peroneal, and superficial radial nerves using a Medelec Synergy system (Oxford Instruments, High Wycombe, UK) and standard techniques.¹⁰ The severity of neuropathy in the present study was staged using a modified form of a previously devised system¹¹:

- stage 0, no neuropathy (T-NSS <2 with normal NCS);
- stage 1, asymptomatic neuropathy (T-NSS = 0 with abnormalities on NCS);
- stage 2, symptomatic neuropathy (T-NSS >2 with normal NCS or T-NSS ≥1 with abnormal NCS; neuropathic symptoms non-disabling);
- stage 3, disabling neuropathy (T-NSS ≥2 with normal NCS or T-NSS ≥1 with abnormal NCS; neuropathic symptoms reported to be disabling.

Disability was defined as a curtailment in recreational activities, absence of feeling in the hand or inability to walk independently.¹¹

Excitability properties of median nerve sensory axons were measured before, during, and one hour after a standard five hour haemodialysis session. Serum electrolytes, urea, creatinine, parathyroid hormone, and β 2-microglobulin were measured at the time of the excitability studies. Kt/V, a urea based measure of dialysis adequacy which relates the clearance of urea to its distribution volume,¹² was also calculated.

Median nerve sensory excitability studies were carried out using a previously described technique.⁵ The following excitability parameters were recorded: stimulus–response behaviour using two stimulus durations; threshold electrotonus

Abbreviations: ESKD, end stage kidney disease; Kt/V, ratio of clearance of urea to distribution volume; NCS, nerve conduction study; NSS, neuropathy symptom score; RRP, relative refractory period; SNAP, sensory nerve action potential; TEd, depolarising threshold electrotonus; TEh, hyperpolarising threshold electrotonus; T-NSS, total neuropathy symptom score

	Pre-dialysis	During dialysis	Post-dialysis	Normal controls
TEd 90–100 ms	41.1 (2.6)%***	49.3 (2.6)%***	50.7 (2.1)%	50.6 (0.9)%
TEh 90–100 ms	-82.7 (11.5)%***	-110.4 (11.5)%***	-111.4 (9.5)%	-131.9 (3.2)%
TEd 40–60 ms	43.6 (3.2)%***	50.9 (2.1)%***	48.5 (1.3)%	51.4 (0.7)%
TEd 10–20 ms	55.0 (3.7)%***	60.8 (1.2)%***	65.2 (1.5)%	65.9 (0.5)%
TEh 10–20 ms	-51.3 (6.4)%***	-56.4 (7.6)%***	-72.4 (5.0)%	-84.6 (1.2)%
RRP	4.2 (0.4) ms***	4.5 (0.8) ms***	2.7 (0.1) ms	3.6 (0.1) ms
Superexcitability	-13.1 (2.3)%***	-16.2 (2.3)%***	-18.2 (2.3)%	-17.2 (0.7)%
Late subexcitability	7.4 (1.5%)***	6.9 (2.0%)**	6.1 (1.2)%	12.0 (0.5)%
electrotonus and reco	very cycle parameters. 1	ter dialysis, with normal Ed and TEh are represe rsis and during dialysis r	ented for different ti	me intervals as state

to 100 ms polarising currents (a marker of internodal axonal membrane function); a current–threshold relation (a measure of an inwardly rectifying K^+ conductance); refractoriness (owing to inactivation of nodal transient Na⁺ channels); superexcitability (dependent on paranodal fast K^+ channels); and late subexcitability (determined in part by nodal slow K^+ channels).

Abnormalities of nerve conduction and excitability were established by comparing the results with normative data from our unit⁵ ¹³ and other centres.^{14–17} In the case of excitability studies, individual measurements were corrected for age and temperature before statistical tests were applied, using relations established by normative data.⁵ ¹⁸ Single comparisons of excitability parameters were analysed using Student's unpaired *t* tests for comparisons with normative data (n = 50, age range 18 to 61, mean 36 years) and paired *t* tests for comparisons before and after dialysis. Correlations were analysed using Pearson's correlation coefficient. A probability (p) value of <0.05 was considered statistically significant. Results are expressed as mean (SEM).

RESULTS

The complete excitability protocol could not be completed in two of the 12 ESKD patients because sensory nerve action potentials (SNAP) amplitudes were too small for threshold tracking. All 12 patients reported symptoms of neuropathy (mean T-NSS, 1.9 (0.2)). Sensory symptoms were present in all patients, predominantly paraesthesiae and numbness, while six experienced motor symptoms. Four patients described pain, a symptom of small fibre dysfunction. According to the neuropathy staging system (above), 8% had no evidence of neuropathy, 58% had stage 2, and 33% had stage 3. In keeping with sensory predisposition of uraemic neuropathy, sural SNAP amplitude¹³ was the most commonly affected NCS parameter, reduced in 75% of patients (to a mean of 6.0 (1.2) μ V), with relative preservation of conduction velocity (mean velocity, $42.6 (0.9) \text{ ms}^{-1}$), consistent with axonal neuropathy.

Excitability recordings in patients before dialysis showed significant abnormalities. Maximum SNAP amplitude, measured peak to peak,⁵ was significantly reduced (p<0.00005)—accompanied by a shift to the left of stimulus–response curves, consistent with axonal depolarisation.¹⁹ Depolarising threshold electrotonus (TEd) at the 90–100 ms interval (TEd 90–100) was lower in ESKD patients than in controls (p<0.0005), as was hyperpolarising threshold electrotonus (TEh) at the same time interval (p<0.00005), leading to a "fanned in" appearance of the threshold curve (fig 1A).¹⁹⁻²¹ Changes in threshold electrotonus were also noted in TEd 10–20 ms (p<0.0005), TEd 40–60 ms (p<0.0005), and TEh 10–20 ms (p<0.0005). During the

recovery cycle (fig 1B), refractoriness was significantly greater in ESKD patients (p<0.005) and this was accompanied by a prolongation of the relative refractory period (p<0.05). Superexcitability and late subexcitability were both reduced in pre-dialysis recordings (p<0.05 and 0.005, respectively). There was no significant difference in strength–duration properties between EKSD patients and controls.

Excitability recordings undertaken one hour after completion of dialysis showed significant improvement in excitability parameters (table 1; fig 1 panels C and D). Persistent abnormalities were, however, noted in TEd 40–60 ms (p = 0.06), TEh 10–20 ms (p < 0.005), TEh 90–100 ms (p < 0.05), and late subexcitability (p < 0.005), indicating that haemodialysis failed to restore nerve excitability to normal completely.

A significant correlation was noted between T-NSS and both pre-dialysis subexcitability (r = 0.79, p<0.01) and TEh 90–100 ms (r = 0.73, p<0.01), a sensitive indicator of membrane potential (fig 1E), suggesting that patients with greater neuropathic symptoms showed more pronounced changes in membrane excitability before dialysis. Correlation analysis was also undertaken to investigate the relation between neurophysiological variables and the levels of potential neurotoxins. There was close correlation in predialysis recordings between serum K⁺ and both threshold electrotonus and late subexcitability. The correlation with TEd 90-100 ms was further strengthened when post-dialysis data were included in the analysis (r = 0.73, p<0.01). When sensory data from the present study were pooled with results obtained previously on pre-dialysis changes in lower limb motor nerves,⁸ there remained a close correlation between K⁺ and both TEh 90–100 ms (r = 0.81, p<0.05; fig 1F) and TEd 90–100 ms (r = 0.76, p<0.05). There was no correlation between parameters from standard NCS with symptom severity or serum K⁺.

The only other potential toxin which correlated well with pre-dialysis excitability parameters was urea (TEd 40–60 ms, r = 0.86, p<0.01; TEd 90–100 ms, r = 0.76, p<0.05). Urea concentration, however, also correlated well with pre-dialysis K⁺ (r = 0.90, p<0.01). No significant correlation was noted between Kt/V, a urea based measure of dialysis adequacy, and excitability parameters. Critically, all patients had Kt/V greater than 1.2, in keeping with current guidelines of dialysis adequacy.¹²

Sensory excitability parameters from the present study were compared with results of a previous study of median motor nerve excitability.⁷ While no differences were noted between sensory and motor axons with respect to parameters of threshold electrotonus, refractoriness, or superexcitability, late subexcitability was significantly lower in motor axons

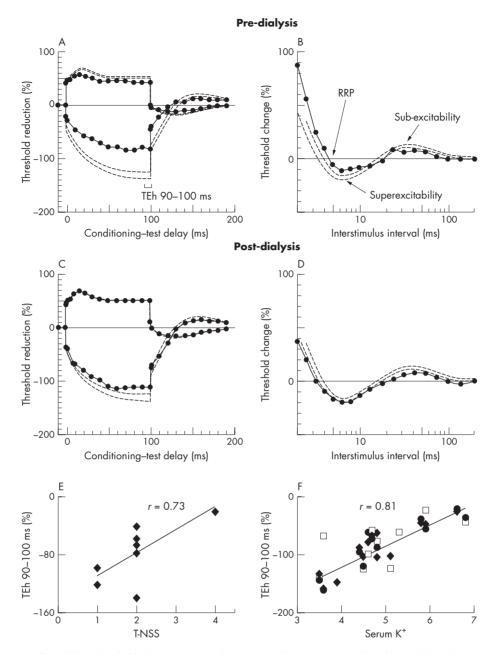


Figure 1 Comparison of pre-dialysis threshold electrotonus (A) and recovery cycle (B) in patients with end stage kidney disease (ESKD) (continuous lines with circles) with post-dialysis recordings (C, D). Broken lines represent 95% confidence intervals for normal controls. The conditioning-test interval corresponding to hyperpolarising threshold electrotonus at 90–100 ms (TEH 90–100 ms) is depicted in (A). Pre-dialysis traces demonstrate "fanning in" of threshold electrotonus, a prolonged relative refractory period, and reduced superexcitability and late subexcitability. Abnormalities had largely resolved by one hour post-dialysis. (E) Relation between pre-dialysis TEH 90–100 ms and total neuropathy symptom score for sensory nerves in ESKD patients. (F) Pre-dialysis TEH 90–100 ms and K⁺ for pooled data from median sensory studies (empty square) and lower limb motor studies⁸ (recorded

than sensory axons (motor studies, 4.5 (1.8)%; sensory studies, 9.1 (1.2)%, p<0.05), although the mean pre-dialysis serum K⁺ was almost identical in the two groups (motor studies, 5.0 (0.2) mmol/l; sensory studies, 4.9 (0.3) mmol/l)).

DISCUSSION

The present study confirms that sensory axons are depolarised in ESKD patients before dialysis. Specifically, there was fanning in of threshold electrotonus with increases in refractoriness and the duration of the relative refractory period, and attenuation of superexcitability, all consistent with membrane depolarisation.²¹ Correlation was noted between clinical symptoms and excitability parameters. Patients with a greater number of neuropathic symptoms showed more severe changes in these excitability parameters. These findings suggest a direct relation between excitability abnormalities and neuropathic symptoms, and mirror the results for lower limb motor nerves.⁸

Of the potential neurotoxins, K^+ and urea correlated most closely with excitability parameters. The correlations with threshold electrotonus indices were further strengthened following inclusion of pre-dialysis excitability results from lower limb motor nerves.⁸ Given the dependence of membrane potential on the concentration gradient for $K^{+,3}$ alterations in K^+ concentration would be likely to produce abnormalities of membrane potential. In contrast, there is no evidence to suggest that urea exerts any effect on membrane potential.

While the serum K⁺ abnormalities noted in the present study improved following haemodialysis, such improvement is likely to be transient. Following dialysis, there is rebound of K⁺ in ESKD patients, which typically occurs within six hours of dialysis owing to re-equilibration between the intracellular and the extracellular compartments.²² Such prolonged hyperkalaemia may cause disruption of normal ionic gradients essential for axonal survival, thereby leading to accumulation of Ca⁺ within the cell and subsequent axonal degeneration.²³

The paradoxical reduction in late subexcitability rather than the expected increase with membrane depolarisation¹⁹ mirrors the findings noted in median motor axons in ESKD patients.⁷ Late subexcitability, which is determined by activation of nodal slow K⁺ channels and the difference between membrane potential (E_r) and potassium equilibrium potential (E_k) , increases with axonal depolarisation if extracellular K⁺ remains unchanged.¹⁹ Raised extracellular K⁺, as may occur in ESKD patients, will lead to a reduction in $(E_r - E_k)$ and thereby decreased subexcitability. There was a significantly greater reduction in late subexcitability in median nerve motor axons7 than in sensory axons, despite similar pre-dialysis mean K⁺ concentrations. This suggests that raised K⁺ has greater effects on motor axons than on sensory axons, a difference that has been noted in previous in vitro studies.^{24 25} This apparently paradoxical reduction in subexcitability suggests that hyperkalaemia is driving axonal depolarisation. Overall, the present study on sensory axons in ESKD, combined with previous studies in motor axons,78 provides clear evidence that the achievement of normokalaemia3 rather than just the avoidance of hyperkalaemia should become a priority in the treatment of patients with ESKD to avoid derangement in nerve excitability, especially in those patients with incipient neuropathy.

ACKNOWLEDGEMENTS

AK was supported by the Australian Association of Neurologists Research Fellowship and the National Health and Medical Research Council of Australia. Grant support from the Brain Foundation, the Sylvia and Charles Viertel Charitable Foundation, and the Ramaciotti Foundation is gratefully acknowledged.

Authors' affiliations

A V Krishnan, M C Kiernan, Prince of Wales Medical Research Institute and Prince of Wales Clinical School, University of New South Wales, Sydney, Australia

R K S Phoon, B A Pussell, J A Charlesworth, Department of Nephrology, Prince of Wales Hospital

Competing interests: none declared

Correspondence to: Dr Matthew Kiernan, Prince of Wales Medical Research Institute, Barker Street, Randwick, Sydney, NSW 2031, Australia; M.kiernan@unsw.edu.au

Received 10 September 2005 In revised form 12 December 2005 Accepted 13 December 2005

REFERENCES

- 1 Bolton CF. Peripheral neuropathies associated with chronic renal failure. Can J Neurol Sci 1980;7:89-96.
- Babb AL, Popovich RP, Christopher TG, et al. The genesis of the square meterhour hypothesis. Trans Am Soc Artif Intern Org 1971;17:81-91
- 3 Bostock H, Walters RJ, Andersen KV, et al. Has potassium been prematurely Viscarded as a contributing factor to the development of uraemic neuropathy? Nephrol Dial Transplant 2004;19:1054–7.
- 4 Krishnan AV, Lin CS-Y, Kiernan MC. Nerve excitability properties in lower limb motor axons: evidence for a length-dependent gradient. Muscle Nerve 2004;29:645-55
- 5 Kiernan MC, Lin CS, Andersen KV, et al. Clinical evaluation of excitability measures in sensory nerve. Muscle Nerve 2001;24:883–92.
- 6 Kiernan MC, Burke D, Andersen KV, et al. Multiple measures of axonal excitability: a new approach in clinical testing. Muscle Nerve 2000;23:399-409.
- 7 Kiernan MC, Walters RJ, Andersen KV, et al. Nerve excitability changes in chronic renal failure indicate membrane depolarization due to hyperkalaemia. *Brain* 2002;**125**:1366–78.
- 8 Krishnan AV, Phoon RK, Pussell BA, et al. Altered motor nerve excitability in end-stage kidney disease. Brain 2005;128:2164-74.
- 9 Dyck PJ, Sherman WR, Hallcher LM, et al. Human diabetic endoneurial sorbitol, fructose, and myo-inositol related to sural nerve morphometry. Ann Neurol 1980;8:590-96.
- 10 Kimura J. Electrodiagnosis in diseases of nerve and muscle. Philadelphia: FA Davis, 1983.
- 11 Dyck PJ. Detection, characterization, and staging of polyneuropathy: assessed in diabetics. Muscle Nerve 1988:11:21-32
- 12 National Kidney Foundation. NKF-DOQI clinical practice guidelines for hemodialysis adequacy. Am J Kidney Dis 1997;**30**(suppl 2):S15–66. 13 **Burke D**, Skuse NF, Lethlean AK. Sensory conduction of the sural nerve in polymourser, the sural New York Sensory conduction of the sural nerve in
- polyneuropathy. J Neurol Neurosurg Psychiatry 1974;37:647-52.
- 14 Buschbacher RM. Tibial nerve motor conduction to the abductor hallucis. Am J Phys Med Rehabil 1999;**78**:S15–20.
- 15 Puksa L, Stalberg E, Falck B. Reference values of F wave parameters in healthy subjects. Clin Neurophysiol 2003;114:1079-90.
- 16 Ma DM, Kim SH, Spielholz N, et al. Sensory conduction study of distal radial nerve. Arch Phys Med Rehabil 1981;62:562-64.
- 17 Ma DM, Liveson JA. Nerve conduction handbook. Philadelphia: FA Davis, 1983
- 18 Kiernan MC, Cikurel K, Bostock H. Effects of temperature on the excitability properties of human motor axons. Brain 2001;124:816-25.
- 19 Kiernan MC, Bostock H. Effects of membrane polarization and ischaemia on the excitability properties of human motor axons. Brain 2000;123:2542-51. 20 Bostock H, Baker M. Evidence for two types of potassium channel in human
- motor axons in vivo. Brain Res 1988;462:354-58. 21 Kaji R. Physiology of conduction block in multifocal motor neuropathy and
- other demyelinating neuropathies. Muscle Nerve 2003;27:285-96.
- 22 Ahmed J, Weisberg LS. Hyperkalemia in dialysis patients. Semin Dialysis 2001;14:348-56
- 23 Stys PK. General mechanisms of axonal damage and its prevention. J Neurol Sci 2005;233:3-13.
- 24 Neumcke B, Schwarz W, Stampfli R. Slow actions of hyperpolarization on sodium channels in the membrane of myelinated nerve. Biochim Biophys Acta 1979;558:113-18.
- 25 Palti Y, Moran N, Stampfli R. Potassium currents and conductance. Comparison between motor and sensory myelinated fibers. Biophys J 1980;32:955-66.