



Published in final edited form as:

Tetrahedron Lett. 2007 September 17; 48(38): 6754–6758.

Rapid and efficient microwave-assisted synthesis of highly sulfated organic scaffolds

Arjun Raghuraman^a, Muhammad Riaz^a, Michael Hindle^b, and Umesh R. Desai^{a,*}

^aDepartment of Medicinal Chemistry and Institute for Structural Biology and Drug Discovery, Richmond, VA, USA

^bDepartment of Pharmaceutics, Virginia Commonwealth University, Richmond, VA, USA

Abstract

Sulfation of multiply hydroxylated small organic molecules is fraught with problems of poor yield, multitude of products and long reaction times. We have developed a rapid microwave-based method for synthesis of highly sulfated small organic molecules, which affords the per-sulfated product in moderate to excellent yields and high purity. The method is expected to be of value in the discovery of per-sulfated organic molecules as mimics of glycosaminoglycans, which are being increasingly recognized as modulators of key physiological functions.

Recent work in our laboratory shows that designed highly sulfated, aromatic, small organic molecules possess interesting physico-chemical and biological properties.¹⁻⁵ Biochemically, these molecules form multiple ionic as well as non-ionic interactions, which form the backbone of most protein-recognition elements. Structurally, these represent mimics of glycosaminoglycans (GAGs), which are increasingly being recognized as modulators of key physiological functions,^{6,7} while toxicologically, the sulfated structure represents a highly water-soluble, already-metabolized form that is expected to possess minimal toxicity. Despite these novel features, highly sulfated organic molecules remain largely unexplored.

A major limitation in exploring these novel structures is their challenging synthesis. Nearly all small organic sulfates reported in the literature are mono- or di-sulfated molecules,⁸⁻¹² typically prepared using sulfur trioxide complexes with amines in a highly polar solvent (DMF or DMA). Sulfation of such organic scaffolds may require as many as 13 hrs and temperatures as high as 95 °C in the presence of a large excess of the sulfating complexes,⁸⁻¹² while sugars, which contain multiple –OH groups, require reaction times in the range of 12 hrs to several days.¹³⁻¹⁹

Theoretically, this method could be extended for synthesis of highly sulfated drug-like molecules, yet practically it is a synthetic nightmare because these molecules possess significantly higher negative charge density.² The major challenge is driving the reaction to completion in order to sulfate all available hydroxyl groups (alcoholic or phenolic) on the substrate. As the number of –OH groups increase on a small scaffold, sulfation becomes progressively difficult because of anion crowding, resulting in numerous partially sulfated side-products.

*Corresponding author. Tel.: 001–804–828–7328; fax: 001–804–827–3664; e-mail: urdesai@vcu.edu

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

A further challenge is the isolation of the chemically pure per-sulfated product, which requires aqueous isolation techniques. Yields in the range of 11 and 100% have been reported, yet the presence of inorganic salts arising from the use of buffers and salts lead to significant inconsistencies and inaccuracies. Additionally, instability of highly anionic products introduce limitations on reaction times and temperature.¹³ This is likely to be especially true for highly sulfated, aromatic, small organic molecules, which are expected to be less stable than the saccharide scaffolds.

To avoid these problems with one-step sulfation, we recently synthesized some small aromatic per-sulfated structures using a two-step approach involving the 2,2,2-trichloroethyl protecting group.²⁰ The two-step protection-deprotection protocol resolved some of the problems of the direct sulfation approach, yet required careful real-time monitoring of the reaction by RP-HPLC to prevent product degradation and was not particularly applicable to substrates that were acid and/or metal sensitive. These limitations led us to seek an alternative sulfation approach, which can be rapid, efficient, and widely applicable to a number of poly-hydroxy scaffolds.

We hypothesized that significant rate enhancements are likely to be achieved using microwaves, especially because the ionic sulfated product may couple to microwaves through ionic conduction, e.g., in CH₃CN.²¹ CH₃CN was chosen as the solvent over the commonly used DMF because a) it can be evaporated at lower temperature (thus aiding isolation) and b) it was likely to solubilize the persulfated product with an amine counter-ion. We also hypothesized that introducing free base in the reaction mixture should promote the difficult per-sulfation reaction.

Sulfation of **1**²² with SO₃•Me₃N complex (6 equivalents per –OH group) at 100 °C in the absence of free base gave only 4.7 % of per-sulfated product **1s** in 20 minutes (Table 1, entry 1).²³ Inclusion of 1 equivalent of free Et₃N per –OH group resulted in 13.5 % conversion (entry 2), while 79.8 % of **1s** was formed with 5 equivalents of Et₃N (entry 3). Further increase to 10 equivalents Et₃N per –OH group had a negligible increase in the yield of **1s** (entry 4). Increasing the proportion of SO₃•Me₃N per –OH group from 1 to 9 molar equivalents (Table 1, entries 5–8) gradually increased the yield of the per-sulfated product from 14.5 to 79.5 %, while further increase to 12 equivalent was found to be not particularly advantageous (entry 9). For further studies, 6 and 10 molar equivalents of the sulfating complex and base, respectively, were chosen.

To assess the effect of solvent, we chose to evaluate nitromethane and DMF, both of which are solvents with high dielectric constant and known to be microwave-friendly. While only 23.2 and 17.2 % of per-sulfated product **1s** was formed from **1** in 10 minutes at 100 °C in CH₃NO₂ and DMF, respectively, 47.4% of the product was formed in CH₃CN (Table 1, entries 10 and 11). Thus, our initial choice of CH₃CN proved to be optimal. To assess the effect of temperature and reaction time, sulfation was performed for 10–30 minutes at 40 to 120 °C. While 30 minutes were required to yield 91.2 % of **1s** at 100 °C, only 10 minutes were needed for 90.8 % conversion at 120 °C. In striking contrast, no product was detected at 40 °C within 10 minutes. Finally, SO₃•py/py was found to give nearly twice as much per-sulfated product as SO₃•Me₃N/Et₃N (entries 7 and 12) in 10 minutes at 100 °C. Since pyridine is ~ 10,000-fold weaker base in comparison to Et₃N, this result suggests general base catalysis as the predominant mechanism of sulfation rather than a process involving deprotonation of the substrate followed by nucleophilic attack.

Appropriate control reactions in the absence of microwaves using two different substrates – **1** and **3** (entries 1 and 3 in Table 2) – at 60 °C in DMF with no free base showed poor product yields. For example, it took 24 hrs in the absence of microwaves to yield **1s** in 60% yield, while

3s was not detected even after 24 hrs (entry 3, Table 2). These results highlight the importance of microwaves in achieving rapid per-sulfation.

Having optimized the reaction conditions, we assessed whether the method works for a variety of different substrates. Per-sulfation of **2** proceeded smoothly in a manner identical to **1** (Table 2).²⁴ More importantly, persulfation of **3**, containing the crowded 3,4,5-trihydroxy moiety, was achieved under microwave conditions in an isolated yield of 54 %, while the conventional procedure completely failed to give **3s**. Finally, microwave-assisted per-sulfation also works extremely well for substrates **5** through **8** containing one to six –OH groups. Interestingly, **5** and **6** gave a mixture of products with SO₃•Me₃N, but yielded the per-sulfated products with SO₃•py.

Several points make the microwave-assisted synthetic protocol particularly attractive. A) The method appears to tolerate a range of functional groups including amide (Table 2, entries 1 through 4), ester (entry 4), aldehyde (entry 8) and double bond (entry 7). The relatively high isolated yields (~70–95%) in each case make the reaction especially suitable for library construction. B) The method works equally well for substrates containing one –OH group to those that contain six –OH groups. This is important because the small size of these molecules introduces considerable anion–anion repulsion as the number of sulfate groups increase. C) The method applies equally well to alcoholic and phenolic –OH groups, especially with SO₃•py complex. D) The method provides high purity per-sulfated product that is readily isolated using an aqueous G10 filtration column. Typically, the purity of these highly water soluble, per-sulfated, small, organic molecules was found to be more than 95% using reverse polarity capillary electrophoresis (see Supplementary Material). E) The method is particularly suitable for quantitative isolation of small amounts (<10 mg) of the per-sulfated products, however could be linearly scaled up at least 20-fold without affecting the yields to a significant extent.

In summary, we have developed a rapid and high yielding microwave-based synthesis of variably functionalized, persulfated organic molecules. The protocol is expected to greatly facilitate the construction of a library of persulfated, small organic molecules for screening as glycosaminoglycan mimetics.

Acknowledgments

This work was made possible by the financial support from the National Heart, Lung and Blood Institute (RO1 HL069975 and R41 HL081972) and the American Heart Association – National Center (EIA 0640053).

References and notes

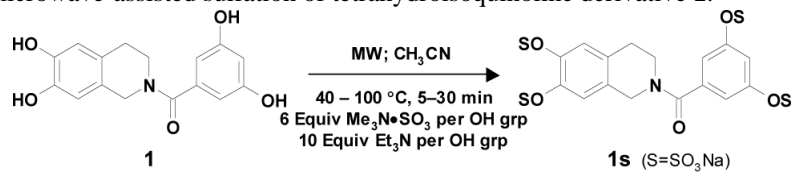
1. Monien BH, Henry BL, Raghuraman A, Hindle M, Desai UR. *Bioorg. Med. Chem* 2006;14:7988–98. [PubMed: 16914317]
2. Dantuluri M, Gunnarsson GT, Riaz M, Nguyen H, Desai UR. *Anal. Biochem* 2005;336:316–22. [PubMed: 15620899]
3. Gunnarsson GT, Desai UR. *Bioorg. Med. Chem. Lett* 2003;13:579–83.
4. Gunnarsson GT, Desai UR. *J. Med. Chem* 2002;45:4460–70. [PubMed: 12238925]
5. Gunnarsson GT, Desai UR. *J. Med. Chem* 2002;45:1233–43. [PubMed: 11881992]
6. Garg, HG.; Linhardt, RJ.; Hales, CA. *Chemistry and Biology of Heparin and Heparan Sulfate*. Elsevier Science; 2005.
7. Raman R, Sasisekharan V, Sasisekharan R. *Chem. Biol* 2005;12:267–77. [PubMed: 15797210]
8. Santos GA, Murray AP, Pujol CA, Damonte EB, Maier MS. *Steroids* 2003;68(2):125–32. [PubMed: 12606002]
9. Bogenstatter ML, A. Overman LE, Tomasi AL. *J. Am. Chem. Soc* 1999;121(51):12206–7.
10. Kawai N, Takao K, Kobayashi S. *Tetrahedron Lett* 1999;40(22):4193–6.

11. Liu Y, Lien IFF, Ruttgaizer S, Dove P, Taylor SD. *Org. Lett* 2004;6:209–12. [PubMed: 14723530]
12. Simpson LS, Widlanski TS. *J. Am. Chem. Soc* 2006;128:1605–10. [PubMed: 16448132]
13. Gama CI, Tully SE, Sotogaku N, Clark PM, Rawat M, Vaidehi N, Goddard WA 3rd, Nishi A, Hsieh-Wilson LC. *Nat. Chem. Biol* 2006;2:467–73. [PubMed: 16878128]
14. Lu LD, Shie CR, Kulkarni SS, Pan GR, Lu XA, Hung SC. *Org. Lett* 2006;8:5995–8. [PubMed: 17165913]
15. Fan RH, Achkar J, Hernandez-Torres JM, Wei A. *Org. Lett* 2005;7:5095–8. [PubMed: 16235966]
16. Orgueira HA, Bartolozzi A, Schell P, Litjens REJN, Palmacci ER, Seeberger PH. *Chem. Eur. J* 2003;9(1):140–69.
17. Codee JDC, van der Marel GA, van Boeckel CAA, van Boom JH. *Eur. J. Org. Chem* 2002:3954–65.
18. Haller M, Boons GJ. *J. Chem. Soc., Perkin Trans. 1* 2001;8:814–22.
19. Das SK, Mallet JM, Esnault J, Driguez PA, Duchaussoy P, Sizun P, Herault J-P, Herbert J-M, Petitou M, Sinay P. *Chem. Eur. J* 2001;7:4821–34.
20. Gunnarsson GT, Riaz M, Adams J, Desai UR. *Bioorg. Med. Chem* 2005;13:1783–9. [PubMed: 15698795]
21. Kappe OC, Doris Dallinger D. For a recent review on the impact of microwaves on drug discovery. *Nat. Rev. Drug Discov* 2006;5:51–63. [PubMed: 16374514]
22. See Supplementary Material for synthesis of starting materials.
23. RP-HPLC profile showed peaks from 4.3–6.0 min, in addition to one at 9.0 min. The peak at 4.3 was subsequently isolated after optimization of conditions and determined to be persulfated (**1s**). The peak at 9.0 min was identified as **1** by comparison with synthetically pure sample. Conversions (%) were determined by area normalization.
24. Representative procedure for per-sulfation: To a stirred solution of the poly-alcohol (20 mg, 0.066 mmol) in MeCN (1 mL) at rt, Et₃N (0.4 mL, 2.9 mmol) and Me₃N•SO₃ (220 mg, 1.6 mmol) was added. The reaction vessel was sealed and micro-waved (CEM-Discover microwave synthesizer) for 20 minutes at 100 °C. The reaction was repeated for 4 times and the reaction mixture was pooled for isolation of the product. The MeCN layer was decanted and pooled, while the residue was washed with MeCN (5 mL) and centrifuged. The combined MeCN layers were concentrated in vacuo. Water (5 mL) was added to the residue and stirred for 10 min. The water layer was concentrated to approximately 2 mL, loaded onto a Sephadex G10 column (~ 160 cm) and chromatographed using water as eluent. Fractions were combined based on RP-HPLC profiles, concentrated and reloaded onto a SP Sephadex C25 column for sodium exchange. Appropriate fractions were pooled, concentrated in vacuo and lyophilized to obtain a white powder. Spectral characteristics of the final sulfated compounds are as follows: **1s**: ¹H NMR (DMSO, 400 MHz) δ: 7.29–7.30 (m, 2H), 6.94–6.97 (m, 3H), 4.58 (s, 2H, isomer I), 4.48 (s, 2H, isomer II), 3.58 (s, 2H, isomer II), 3.50 (s, 2H, isomer I), 2.66 (br, 2H, isomer I & II); ESI (-ve) *m/z* calcd for C₁₆H₁₁NNa₄O₁₇S₄ [(MNa)⁻] 685.86, found 686.1; **2s**: ¹H NMR (DMSO, 400 MHz) δ: 7.65 (d, *J* = 2.4 Hz, 1H), 7.57 (d, *J* = 8.4 Hz, 1H), 7.29 (s, 2H), 6.99 (dd, *J* = 8.4, 1.6 Hz, 1H), 4.54 (s, 2H), 3.70 (br, 2H), 2.69 (t, *J* = 4.8 Hz, 2H); ESI (-ve) *m/z* calcd for C₁₆H₁₁NNa₄O₁₇S₄ [(M-Na)⁻] 685.86, found 686.0; **3s**: ¹H NMR (DMSO, 400 MHz) δ: 7.37 (s, 2H), 7.29 (s, 2H), 4.54 (s, 2H), 3.53 (s, 2H), 2.68 (s, 2H); ESI (-ve) *m/z* calcd for C₁₆H₁₀NNa₅O₂₁S₅ [(M-Na)⁻] 803.79, found 804.1; **4s**: ¹H NMR (DMSO, 400 MHz) δ: 6.96–7.70 (m, 5H), 4.83–5.16 (m, 1H isomers I - III), 4.18–4.44 (m, 2H, isomers I–III), 3.55–3.61 (m, 2H, isomers I–III), 2.99–3.13 (m, 2H, isomers I–III), 1.08–1.16 (m, 3H, isomers I–III); ESI (-ve) *m/z* calcd for C₁₉H₁₅NNa₄O₁₉S₄ [(M-Na)⁻] 757.88, found 757.5; **5s**: ¹H NMR (DMSO, 400 MHz) δ: 7.34 (d, 1H, *J*=8.0 Hz), 7.16 (t, 1H, *J*=8.0 Hz), 6.95 (t, 1H, *J*=8.0 Hz), 6.89 (d, 1H, *J*=8.0 Hz), 5.24 (d, 1H, *J* = 8.0 Hz), 4.86 (s, 2H), 4.62 (d, 2H, *J*=4.0 Hz), 4.35 (m, 1H), 4.21–4.27 (m, 1H), 3.93–3.99 (m, 1H), 3.75 (m, 1H); ESI (-ve) *m/z* calcd for C₁₆H₁₀NNa₅O₂₁S₅ [(M-Na)⁻] 772.81, found 772.7; **6s**: ¹H NMR (DMSO, 400 MHz) δ: 7.1 (d, *J* = 8.4 Hz, 1H), 6.81–6.83 (m, 2H), 4.02 (t, *J* = 8.0 Hz, 1H), 2.71–2.74 (m, 2H), 2.21–2.25 (m, 1H), 2.08–2.12 (m, 1H), 1.87–2.00 (m, 2H), 1.75–1.78 (m, 1H), 1.48–1.60 (m, 2H), 1.09–1.34 (m, 6H), 0.66 (s, 3H); ESI (-ve) *m/z* calcd for C₁₈H₂₂Na₂O₈S₂ [(M-Na)⁻] 453.07, found 453.1; **7s**: ¹H NMR (DMSO, 400 MHz) δ: 7.56 (s, 1H), 7.54 (s, 1H), 6.99–7.10 (m, 7H), 6.61 (s, 1H), 5.36 (s, 1H), 4.63 (s, 1H), 4.59 (s, 1H), 4.39 (s, 1H), 4.14–4.18 (m, 1H), 4.08 (d, *J* = 8.8 Hz, 1H) 3.81 (t, *J* = 10 Hz, 1H); ESI (-ve) *m/z* calcd for C₂₀H₁₆Na₆O₂₆S₆ [(M-Na)⁻] 978.77, found 979.0; **8s**: ¹H NMR (DMSO, 400 MHz) δ: 9.76 (s, 1H), 8.94–8.96 (m, 2H),

8.62–8.68 (m, 1H), 8.09–8.14 (m, 2H), 7.72–7.75 (m, 2H), 6.92–6.95 (m, 2H); ESI (-ve) m/z calcd for $C_{12}H_{11}NO_5S [(M-pyH^+)^-]$ 200.99, found 200.8

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

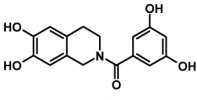
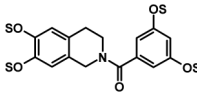
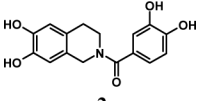
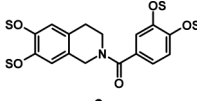
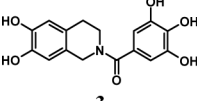
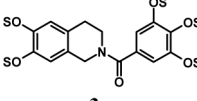
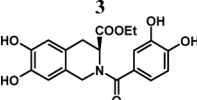
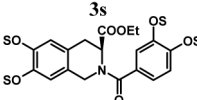
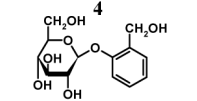
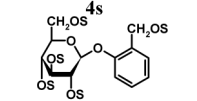
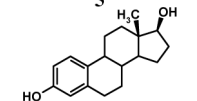
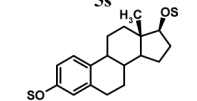
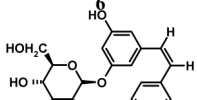
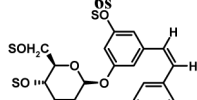
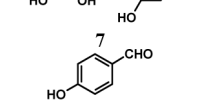
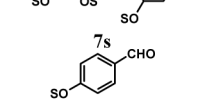
Table 1Optimization of microwave-assisted sulfation of tetrahydroisoquinoline derivative **1**.

	Time (min)	Temp. (°C)	Modifications to Conditions	HPLC Yield (%)
1	20	100	No base	4.7
2	20	100	1 equiv. Et ₃ N per OH grp.	13.5
3	20	100	5 equiv. Et ₃ N per OH grp.	79.8
4	20	100	— ^a	80.0
5	10	100	1 equiv. SO ₃ •Me ₃ N per OH grp.	14.5
6	10	100	3 equiv. SO ₃ •Me ₃ N per OH grp.	46.1
7	10	100	— ^a	47.4
8	10	100	9 equiv. SO ₃ •Me ₃ N per OH grp.	79.5
9	10	100	12 equiv. SO ₃ •Me ₃ N per OH grp.	80.7
10	10	100	DMF as solvent	17.2
11	10	100	CH ₃ NO ₂ as solvent	23.2
12	10	100	With 6 equiv SO ₃ •py/OH grp and 10 equiv py/OH grp as base.	82.3
13	10	40	— ^a	0
14	10	70	— ^a	18.2
15	10	120	— ^a	90.8
16	30	100	— ^a	91.2

^aReaction conditions here are as listed above with no additional modifications.

Table 2
Microwave-assisted sulfation of poly-hydroxyl substrates.

Substrate $\xrightarrow[\text{Et}_3\text{N (10 equiv/OH), SO}_3\bullet\text{Me}_3\text{N (6 equiv/OH), CH}_3\text{CN}]{\text{Microwaves, 100 }^\circ\text{C, 20-30 min}}$ Product

	Substrate	Product ^a	Isolated Yield (%)
1			85
2			87
3			54 ^b
4			74
5			84 ^c
6			94 ^c
7			72 ^d
8			97, ^{ce}

^aS: SO₃Na.

^bWith 9 equiv of SO₃•Me₃N/OH group.

^cWith SO₃•py (6–9 equiv/OH group) and pyridine as base.

^dReaction conditions: 120°, 10 min, SO₃•py (12 equiv/OH group) and pyridine as base.

^eS = SO₃•pyH⁺