Actin cytoskeleton dynamics linked to synovial fibroblast activation as a novel pathogenic principle in TNF-driven arthritis

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Accepted 25 July 2007 Rheumatoid arthritis is a chronic inflammatory disorder whose origin of defect has been the subject of extensive research during the past few decades. While a number of immune and non-immune cell types participate in the development of chronic destructive inflammation in the arthritic joint, synovial fibroblasts have emerged as key effector cells capable of modulating both joint destruction and propagation of inflammation. Ample evidence of aberrant changes in the morphology and biochemical behaviour of rheumatoid arthritis synovial fibroblasts have established the tissue evading and ''transformed'' character of this cell type. We have recently demonstrated that actin cytoskeletal rearrangements determine the pathogenic activation of synovial fibroblasts in modelled TNF-mediated arthritis, a finding correlating with similar gene expression changes which we observed in human rheumatoid arthritis synovial fibroblasts. Here, we show that pharmacological inhibition of actin cytoskeleton dynamics alters potential pathogenic properties of the arthritogenic synovial fibroblast, such as proliferation, migration and resistance to apoptosis, indicating novel opportunities for therapeutic intervention in arthritis. Recent advances in this field of research are reviewed and discussed.

R neumatoid arthritis (KA) represents one of the most

common chronic inflammatory disorders, affecting

approximately 1% of the population worldwide, thus

creating a substantial personal social and economical burden heumatoid arthritis (RA) represents one of the most common chronic inflammatory disorders, affecting creating a substantial personal, social and economical burden.¹ It is a chronic destructive arthropathy that manifests primarily as a painful and prolonged inflammation of the synovial membrane, a thin tissue that lines the diarthrodial joints. Inflammation is accompanied by a gradual increase in the thickness of the synovial lining, the surface layer of the synovial tissue, which is located adjacent to and in direct contact with the intra-articular cavity of the joint. The thickening of the synovial layer, mediated by the intrinsic hyperproliferation of synovial cells and inflammatory infiltrates, gives rise to a pathogenic mass called pannus, comprised primarily of synovial fibroblasts and macrophages, as well as T cells and B cells, which propels the inflammatory machinery to the destruction of bone and cartilage,² leading to irreversible destruction of joint structure and function.³

It is widely accepted that RA is a systemic disease, with a variety of combinatory determinants being attributed to its aetiopathogenesis. These include individual genetic susceptibility, environmental stimuli, physical stress and defective immune responses.⁴ Traditional mechanistic concepts have strongly implicated T cell and B cell-dependent pathways driving development of the disease. Following the success of anticytokine rather than antilymphocyte-targeted therapies in human RA,⁵ a role for chronic innate immune perturbations has also gained momentum, with the macrophages as major contributors of the inflammatory cytokine milieu.⁶ Dissecting the causative molecular mechanisms underlying RA development, the unchallengeable fact is that tumour necrosis factor (TNF) is indeed capable of orchestrating the pathogenic inflammatory cascade. A key translational experiment demonstrating the pathogenic potential of TNF in arthritis was originally provided by the generation of transgenic mice overexpressing human TNF (hTNF-Tg) and developing chronic, erosive and symmetric polyarthritis with histological characteristics resembling the pathogenic process in human RA.7 This model was also instrumental in predicting the significant efficacy of anti-TNF therapy in clinical trials.⁷ In accordance with the phenotype of this transgenic mouse, another murine model of RA, generated by genetic deletion of the AU-rich regulatory region of the murine TNF gene (TNF^{AARE}) further established the pivotal significance of TNF in RA as well as in Crohn's-like inflammatory bowel disease.⁸ RA development in the absence of the lymphocytic compartment in the two TNF transgenic models⁸ ⁹ challenged the indispensable role of adaptive immunity in RA and indicated the important role of non-immune cells, such as synovial fibroblasts in the development of this pathology.

Despite these advances, what exactly keeps TNF fuelling the inflammatory response is still unclear, while the increasing number of non-responders to anti-TNF therapy¹⁰ points to additional mechanisms existing in RA. As anticytokine therapies target a specific agent or pathway, research has been shifted towards cellular targets and processes that are central to the deleterious events initiated by inflammatory cytokines, such as TNFa.

THE SYNOVIAL FIBROBLAST: A ''TRANSFORMED'' CELL WITH A LEADING ROLE IN ARTHRITIS

Chronic persistent inflammation, wound healing and cancer exhibit significant common features: new extracellullar matrix (ECM) deposition and expansion of neighbouring mesenchymal cells such as fibroblasts or myofibroblasts;¹¹ stromal fibroblasts being suggested as potential ''inducers'' of certain carcinomas;¹² experimental evidence in genetically modified mice lacking TGF-βRII in fibroblasts revealed that deregulated

Abbreviations: ECM, extracellular matrix; EMT, epithelial–mesenchymal transition; Lat-A, latrunculin A; MMPs, matrix metalloproteinases; RA, rheumatoid arthritis; RASFs, rheumatoid arthritis synovial fibroblasts; SF, synovial fibroblast; SCID, severe combined immunodeficient; SRF, serum response factor; $TNFx$, tumour necrosis factor α ; Wt, wild type

expression of fibroblast products (eg, hepatocyte growth factor) influences epithelial function and leads to tumorigenesis, indicating the importance of the homeostatic responses of the stromal compartment in modulating/maintaining the tissue integrity.¹³

In chronic inflammatory arthritis, a stromal cell type that has emerged with a dominant role in both joint damage and the propagation of inflammation is the synovial fibroblast (SF) .¹⁴ SFs or fibroblast-like type B synoviocytes along with macrophages or macrophage-like type A synoviocytes, predominate the cellular population of the normal synovium. Although the origin of SFs is still debatable, they represent a heterogeneous population of cells in terms of tissue localisation, physiology (intimal and subintimal) and derivation (non-epithelial, mesenchymal cells) that display differential activation and differentiation properties.¹⁵ While the primary physiological role of SFs is to provide a nourishing environment for the cartilage and proteoglycans that lubricate the articular surfaces, they also anchor to the ECM through the expression of cell adhesion molecules, thus regulating the cellular population that enters the synovial fluid space. In the context of RA, an immune or environmental trigger may lead to the activation of SFs. This in turn results in inappropriate production of chemokines, adhesion molecules and matrix-degrading components, mainly matrix metalloproteinases (MMPs) and cathepsins, contributing to the subsequent cartilage and joint destruction.¹⁶ Indeed, synovial hyperplasia, a hallmark of RA, has been shown to precede the accumulation of inflammatory cells, suggesting a primary role for SFs in RA aetiology.17 18

Notwithstanding the notion that cytokines like TNFa can trigger SF activation and proliferation,¹⁹ ²⁰ it seems that SFs can maintain their activation status without the need of continuous stimulation from the pro-inflammatory microenvironment. This capability of SFs was originally documented by studies on the severe combined immunodeficient (SCID) mouse coimplantation model of RA.²¹ In that model it was elegantly shown that co-implantation of isolated human rheumatoid arthritis synovial fibroblasts (RASFs) and normal human cartilage in the knee joint of healthy SCID mice resulted in the induction of an RA-like phenotype through the production of cartilage-destroying enzymes from the proliferating RASFs. Consistent with the latter finding, we have observed a similar effect by transferring immortalised SFs from our well-established T cell/B cell-independent transgenic animal model, 22 to the knee joint of histocompatible normal recipients. Interestingly, the injected SFs were also able to migrate to other parts of the body, such as the ankle joint, and cause the development of an arthritis-like disease similar to that of the donor.23 The ability of RASFs to migrate, invade and destroy neighbouring articular cartilage resembles to a great extent that of cancer cells and the process of metastasis.24

Despite the tumour-like behaviour of the activated RASF, there is little evidence so far in favour of its function as a malignant cell.²⁵ Presently, no clear evidence exists regarding gene rearrangements and microsatellite instability,²⁶ which is well reported for cancer cells.²⁷ ²⁸ The identification of abnormal gene expression of a number of proto-oncogenes, such as ras, raf, sis, myb, c-myc, c-fos and c-jun in the RA synovium²⁹⁻³¹ and the association of RA and somatic mutations in the tumoursuppressor gene $p53$,³² do not provide conclusive evidence for its characterisation as a malignant cell. Moreover, mutations in PTEN, a tumour suppressor gene, have been described in several human cancers, and have been associated with the invasiveness and metastatic properties of malignant tumours.^{33 34} Interestingly, non-genetically related lack of expression of PTEN at sites of SF invasion further corroborated the argument that defective apoptosis may largely account for the synovial

hyperplasia in RA.³⁵ Apart from the hyperproliferation status of RASFs, shown by their fast growth in vitro and the increased expression of factors such as c -myc and NF- κB ,^{30 36} resistance to apoptosis provides an alternative scenario for SF hyperplasia. Defective apoptosis was well documented by the site-specific upregulation of several anti-apoptotic factors, including Sentrin-1, Bcl-2, and Flip, where synovial invasive growth and destruction is evident.³⁷⁻³⁹

Another intriguing characteristic of the ''transformed'' RASF stems from its ability to produce a variety of pro-angiogenic factors like FGF, VEGF, TGF- β , GM-CSF that are necessary for the generation of blood vessels fuelling the destructive growth of the pannus and RA.40 Consistent with the latter is the previously reported finding of anchorage-independent growth and loss of contact cell inhibition, suggesting that RASFs acquire characteristics found in transformed cells with a tumorigenic potential.41 Moreover, the role of epithelial– mesenchymal transition (EMT) that has been widely discussed lately in terms of stromal-associated carcinogenesis,^{42 43} is presently mentioned as a potential mechanism for the development of several stromal-related rheumatic diseases, such as RA.⁴⁴ The upregulation of TGF- β and activation of its pathway in arthritic synovium,^{45 46} the stress fibre formation and the myofibroblastoid α smooth muscle actin expression⁴⁷⁻⁵⁰ as well as the MMP secretion in RASFs, point to the reprogramming occuring during EMT.⁵¹ However, the absence of epithelial characteristics in the synovium is puzzling. Consequently, the term ''transformed'' has most commonly been attributed to the activated RASF in order to summarise its unique characteristics that contribute to RA pathogenesis.²⁴

FUNCTIONAL GENOMICS POINTS TOWARDS THE ACTIN CYTOSKELETON MACHINERY

The distinct alteration in the cellular physiology of RASFs can also be viewed in the context of their interaction with the ECM. The latter, coupled with components of the cytoskeleton, provides the necessary hydraulic resistance to the mechanical pressure opposed to the growing pannus and the synovial layer, thereby holding the synovial fluid within the joint cavity while it may trap inflammatory molecules.⁵² Furthermore, loss of ECM–SF adhesion has been shown to affect focal adhesion kinases, causing a potential resistance to apoptosis, otherwise conferred by these factors.⁵³ Indeed, the capability of cells like the SFs to adhere to the ECM reflects their status of cytoskeleton organisation and cellular morphology,⁵⁴ while it may also be related to changes in their migration and proliferation potential.⁵⁵ Interestingly, our recent high-throughput expression profiling analysis on SFs derived from the hTNF-Tg animal model, revealed a number of deregulated genes, including gsn, aqp1, cdc42hom, eef1a1, tuba1, rab14, lsp1, lst1, mylc2b, pitpnm and pstpip1, which are known to be involved in actin filament and cytoskeleton organisation.⁵⁶ More importantly, the expression status of a vast majority of these genes is being further replicated and validated in an independent cohort of patients with RA (unpublished data), hinting that there is a simultaneous alteration in RASFs' specific gene expression status and actin cytoskeleton rearrangements.

Notably visualisation of F-actin in vitro on both wild type (Wt) and hTNF-Tg SFs (TgSFs), shows a substantial increase in stress fibre formation in TgSFs compared to WtSFs, indicating altered actin dynamics in TgSFs (fig 1A, C).⁵⁶ A similar outcome is deduced by the treatment of WtSFs with recombinant human TNF α (fig 1B), suggesting that the "transformed" morphology of the SFs can be triggered by TNFa. This complements the previously reported finding that recombinant TNFa can induce morphological changes and increased proliferation of SFs.¹⁹ Most importantly, we have previously shown that TgSFs have

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an aberrant phenotype, compared with those of Wt mice. We have observed TgSFs to be more adhesive and to show greater motility and proliferation potential compared to Wt.⁵⁶ In vitro analysis on the effect of latrunculin A (Lat-A), an inhibitor of Factin polymerisation,⁵⁷ has indicated the potential of actinmodulating drugs in altering the cytoskeletal properties of TgSFs. More specifically, treatment with Lat-A results in decreased migration of TgSFs, while adhesion seems to be unaffected compared to WtSFs (fig 2A, B). This is in keeping with previous findings of Lat-A treatment and myofibrils.⁵⁸ At the same time, TgSFs treated with anti-TNF α do not show altered migration properties, suggesting that this effect might not be TNFa-driven. Alternatively, this could indicate that there is an imprinted phenotype in the TgSF cells due to the chronic overexpression of TNFa, which cannot be reversed by a short treatment with anti-TNFa antibody. In terms of other properties, treatment with Lat-A seems to have a dramatic effect on the proliferation levels of TgSFs which decline even lower than the levels of WtSFs, while DMSO and TNF α do not have any effect on the rate of proliferation of TgSFs (fig 2D). Most importantly, apoptosis assays using staurosporine as an inducer of cell death revealed that Lat-A disruption of the TgSFs cytoskeleton causes decreased resistance to apoptosis at the levels of the WtSFs (fig 2C). This result is consistent with previous findings showing that treatment with cytochalasin D (another inhibitor of actin polymerisation) enhances apoptosis in HeLa cells, while phalloidin (an inducer of actin polymerisation) does not affect apoptosis.⁵⁹ Notably, defective migration potential and abnormal cytoskeleton organisation are unique characteristics that define the "transformed" state of a cell.⁶⁰ Since an actin cytoskeleton-disrupting drug, like Lat-A, can cause profound changes in the properties of the ''transformed'' SF and act independently of TNF α , it is only rational to suggest that treating patients with RA with a combination of anti-TNF and a drug that targets the actin cytoskeleton may be more effective than anti-TNFa treatment alone.

THE ACTIN CYTOSKELETON NETWORK IN THE CONTEXT OF RA

As already discussed, the actin cytoskeleton plays an essential role in the homeostasis and normal function of the cell. Processes such as cell shape modulation, differentiation, migration, development, wound healing and maintenance of polarity are greatly dependent on the actin cytoskeleton. To perform these functions properly, the cell has to be able to respond efficiently to all extracellular and intracellular stimuli and modulate its actin cytoskeleton accordingly. For instance, it

has to be able to modulate the amount of filamentous F-actin versus monomeric G-actin and the number of stress fibres in order to be able to move. For that purpose, a complex intracellular protein network has been established, enabling the cell to modulate actin polymerisation, depolymerisation, nucleation as well as actin organisation depending on the cell's needs. Since these processes are crucial for the cell homeostasis and survival, it is not surprising that more than one protein regulates each step. For example, profilin, thymosin β , α actinin and VASP $^{62-64}$ mediate actin polymerisation.⁶¹ while cofilin/ADF,⁶¹ gelsolin,⁶¹ and caspase 11^{65} are critical for actin depolymerisation, and filamin,⁶⁶ and Arp2/3⁶¹ modulate the nucleation of actin bundles (fig 3). Apart from the control of actin polymerisation, modulation of actin organisation is also of utmost importance. Thus, the actin cytoskeleton structure can be modified by a number of signalling and/or actin-binding proteins. This results in stress fibre, focal adhesion, lamellipodia or filopodia formation depending on the specific requirements of the cell at the specific time. The assembly of projections in the membrane surrounding the cell is regulated mainly by small GTPases of the Rho family of proteins, Rac1, Rho and $Cdc42.^{67-69}$ The Rho family of proteins belongs to the Ras protein superfamily and its members are found in different isoforms. Each of these proteins has the capacity to bind to GTP and constantly be converted from an active GTP-bound form to an inactive GDP-bound form. In the active state they can interact with a number of effector molecules in order to transmit a specific signal and have a specific outcome. For instance, two members of this family, Rac1 and Cdc42, when activated by growth factors or by integrins, the receptors that connect the ECM with the inner compartment of the cell, mediate the lamellipodia and filopodia formation, respectively, while Rho is responsible for the stress fibre formation. 6770

In the context of RA, it was recently shown that in human cultured RASFs, TNFa-induced activation of NF-kB and cytokine secretion is dependent on the activation of RhoA, suggesting a central role of this GTPase in the arthritic inflammatory response.71 As RhoA induces stress fibre formation, it is remarkable that the p65/RelA subunit of NF-kB colocalises and binds to F-actin, while re-establishment of the actin cytoskeleton organisation seems to bring back to normal the distribution of p65 in the cytoplasm.72 Moreover, our studies have shown that depletion of gelsolin, an actin-binding protein known to be involved in actin depolymerisation by inhibiting Factin formation, from mice with RA resulted in exacerbation of the disease.56 This suggests a critical role for gelsolin in modulating the actin cytoskeleton in RASFs and thus in the

Figure 1 Immunofluorescence of stress fibre formation on wild type, wild type treated with hTNFa and hTNF-Tg primary SFs, respectively. (Details on primary SF isolation, F-actin and DAPI staining procedures have been shown previously.)⁵

Figure 2 Application of latrunculin A on the ''transformed'' synovial fibroblast. Disruption of actin cytoskeleton was induced with 4 μg/ml latrunculin A (Sigma) for 2 h at 37°C as previously described.⁹² As control of $TNF\alpha$ involvement in the actin cytoskeleton organisation 1 µg/ml anti-TNF (infliximab) was added on TgSFs for the same period of time. (A, B, D) Adhesion, migration and proliferation assays were performed as previously described.⁵⁶ (C) Apoptosis assays were performed by treating cells with 0.002– 0.05 μ M staurosporine, for 24 h at 37°C overnight. Cell survival was determined using the crystal violet assay as before.⁸ All results are expressed as the means + SE of one representative experiment performed in triplicate. Statistical analysis was performed using Student t test, with a level of significance set at $p<0.05$.

progression of RA. The importance of tissue remodelling in RA was further reinforced by recent experiments on cadherin-11 deficient mice, which showed diminished adhesion, migration and invasion properties of RASFs, while they seem to confer resistance to $K/B\times N$ serum transfer model of arthritis.⁷³ Interestingly, the effect of the actin cytoskeleton on both cell morphology and gene transcription has been the subject of an elegant study, involving Mal, a regulator of gene expression. It was shown that upon F-actin formation, Mal dissociates from Gactin and activates the serum response factor (SRF), whose target genes range from genes regulating cell growth, proliferation

and differentiation to genes that regulate the actin cytoskeleton machinery.74 Not surprisingly, ablation of the SRF in murine embryonic stem cells resulted in low F-actin levels and reduced adhesion and migration capabilities, thus documenting a dynamic link between SRF and remodelling of the actin cytoskeleton.75

In light of the evidence discussed so far, we propose a model whereby an immune or environmental stimulus could trigger TNF α to induce the reorganisation of the actin cytoskeleton, through activation of small GTPases, which may lead to morphological changes in the SFs, ultimately resulting in the

Figure 3 A schematic representation of the proposed interaction between actin cytoskeleton rearrangements and the maintenance of rheumatoid arthritis pathogenesis.

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''transformed'' phenotype of SFs characterised by increased migration, adhesion and proliferation properties (fig 3). Furthermore, increased F-actin formation could lead to increased gene expression mediated by SRF and NF-kB, resulting in sustaining the disorganised actin cytoskeleton and then in the production of pro-inflammatory cytokines, thereby maintaining the detrimental effects of the arthritogenic response (fig 3).

The essential role of the actin cytoskeleton in maintaining cell homeostasis and survival, as well as the role of the multitude of proteins that are crucial for fine-tuning its organisation, is also manifested very clearly in several disease states, other than RA. A recessively inherited genetic disorder known as neutrophil actin dysfunction has been linked to a severe defect in actin assembly and polymerisation in the polymorphonuclear leucocytes, showing the significance of actin polymerisation for neutrophil motility.⁷⁶ In line with neutrophil actin dysfunction, it has been established that a missense point mutation in the gene encoding β -actin that alters depolymerisation dynamics is associated with an autosomal dominant disease characterised by developmental malformations, deafness and dystonia syndrome, where cocontraction of agonistic and antagonistic muscles occurs.⁷⁷ Furthermore, in recent studies in which the neuropathology of brains from identical twins with dystonia was examined, cofilin and actin aggregates were found within the neocortex.⁷⁸ Notably, some of these structures showed typical rod-like morphology and sizes similar to those found in the brains of patients with Alzheimer's disease.79 Also of interest is the fact that the dominantly inherited familial amyloidosis Finnish type is attributed to the accumulation of a 71-amino acid amyloidogenic fragment of mutant gelsolin.⁸⁰ Interestingly, serum amyloid A was found to be elevated in patients with RA,⁸¹ with recent evidence suggesting an induction of synovial hyperplasia and angiogenesis as an outcome of its elevation.⁸²

Not surprisingly, actin and/or actin-binding protein-related disorders have been associated with cancer progression and metastasis. Just like in RA, many of the changes occurring during cancer progression as well as during the metastatic process are linked to deregulations in actin cytoskeleton and the actin-binding proteins at cell–matrix or cell–cell adhesion sites.^{83 84} Reduction in the expression level of the cell-cell adhesion molecule E-cadherin, for example, is known to be associated with cancer progression and poor prognosis.^{85 86} Moreover, deregulation in several ECM adhesion proteins that directly or indirectly associate with actin have been shown to be critical for cancer progression and metastasis, either by affecting cancer cell proliferation and apoptosis rates or by inducing anoikis, a type of apoptosis induced by loss of contact and adhesion to the ECM.⁸⁷⁻⁹⁰ Some of them have been proposed as potential targets for future therapeutical anticancer interventions.⁹¹

CONCLUSIONS

Accumulating evidence suggests that there is a direct but complex interaction between the actin cytoskeleton and the arthritic SF, of a yet unknown nature. Gaining a better understanding of the role of the actin cytoskeleton in modulating the SF function to initiate and maintain the arthritogenic response may provide new insights into novel therapeutic modalities. A rational strategy to achieve that goal would be the inhibition of signalling players that induce actin polymerisation by small peptide molecules or by interfering directly with their gene expression. Functional and in vivo studies will hopefully uncover such targets, whose impact on RA therapy remains to be elucidated.

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REFERENCES

- 1 Silman AJ, Pearson JE. Epidemiology and genetics of rheumatoid arthritis. Arthritis Res 2002;4(Suppl 3):S265–72.
- 2 Tak PP. Examination of the synovium and synovial fluid. In: Firestein GS, Panayi GS, Wollheim FA, eds. Rheumatoid arthritis. Frontiers in pathogenesis and treatment. New York: Oxford University Press, 2000:55–68.
- 3 Firestein GS. Rheumatoid synovitis and pannus. In: Klippel JH, Dieppe PA, eds. Rheumatology. London: Mosby, 1998:5.13.1–24.
- 4 Firestein GS. Evolving concepts of rheumatoid arthritis. Nature 2003;423:356–61.
- 5 Strand V, Kimberly R, Isaacs JD. Biologic therapies in rheumatology: lessons learned, future directions. Nat Rev Drug Discov 2007;6:75–92.
- 6 **Feldmann M**, Brennan FM, Maini RN. Role ot cytokines in rheumatoid arthritis. Annu Rev Immunol 1996;14:397–440.
- 7 Elliott MJ, Maini RN, Feldmann M, Long-Fox A, Charles P, Katsikis P, et al. Treatment of rheumatoid arthritis with chimeric monoclonal antibodies to tumor necrosis factor alpha. Arthritis Rheum 1993;36:1681-90.
- 8 Kontoyiannis D, Pasparakis M, Pizarro TT, Cominelli F, Kollias G. Impaired on/ off regulation of TNF biosynthesis in mice lacking TNF AU-rich elements: implications for joint and gut-associated immunopathologies. Immunity 1999;10:387–98.
- 9 Kollias G, Douni E, Kassiotis G, Kontoyiannis D. On the role of tumor necrosis factor and receptors in models of multiorgan failure, rheumatoid arthritis, multiple sclerosis and inflammatory bowel disease. Immunol Rev 1999;169:175–94.
- 10 Lipsky PE, van der Heijde DM, St Clair EW, Furst DE, Breedveld FC, Kalden JR, et al. Infliximab and methotrexate in the treatment of rheumatoid arthritis. Anti-Tumor Necrosis Factor Trial in Rheumatoid Arthritis with Concomitant Therapy Study Group. N Engl J Med 2000;343:1594–602.
- 11 **Dvorak HF**. Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. N Engl J Med 1986;315:1650-9.
- 12 **Bhowmick NA**, Neilson EG, Moses HL. Stromal fibroblasts in cancer initiation and progression. Nature 2004;432:332–7.
- 13 Bhowmick NA, Chytil A, Plieth D, Gorska AE, Dumont N, Shappell S, et al. TGFbeta signaling in fibroblasts modulates the oncogenic potential of adjacent epithelia. Science 2004;303:848–51.
- 14 Kontoyiannis D, Kollias G. Fibroblast biology: synovial fibroblasts in rheumatoid arthritis – leading role or chorus line? Arthritis Res 2000;2:342–3.
- 15 **Chang HY**, Chi JT, Dudoit S, Bondre C, van de Rijn M, Botstein D, et al. Diversity, topographic differentiation, and positional memory in human fibroblasts. Proc Natl Acad Sci USA 2002;99:12877–82.
- 16 Ritchlin C. Fibroblast biology: effector signals released by the synovial fibroblast in arthritis. Arthritis Res 2000;2:356-60.
- 17 Lopez-Bote JP, Bernabeu C, Marquet A, Fernandez JM, Larraga V. Adjuvantinduced polyarthritis. Synovial cell activation prior to polyarthritis onset. Arthritis Rheum 1988;31:769–75.
- 18 Marinova-Mutafchieva L, Williams RO, Funa K, Maini RN, Zvaifler NJ. Inflammation is preceded by tumor necrosis factor-dependent infiltration of mesenchymal cells in experimental arthritis. Arthritis Rheum 2002;46:507–13.
- Butler DM, Piccoli DS, Hart PH, Hamilton JA. Stimulation of human synovial fibroblast DNA synthesis by recombinant human cytokines. J Rheumatol 1988;15:1463–70.
- 20 Dayer JM, Krane SM, Russell RG, Robinson DR. Production of collagenase and prostaglandins by isolated adherent rheumatoid synovial cells. Proc Natl Acad Sci USA 1976;73:945-9.
- 21 Muller-Ladner U, Kriegsmann J, Franklin BN, Matsumoto S, Geiler T, Gay RE, et al. Synovial fibroblasts of patients with rheumatoid arthritis attach to and invade normal human cartilage when engrafted into SCID mice. Am J Pathol 1996;149:1607–15.
- 22 Keffer J, Probert L, Cazlaris H, Georgopoulos S, Kaslaris E, Kioussis D, et al. Transgenic mice expressing human tumour necrosis factor: a predictive genetic model of arthritis. Embo J 1991;10:4025–31.
- 23 Aidinis V, Plows D, Haralambous S, Armaka M, Papadopoulos P, Kanaki MZ, et al. Functional analysis of an arthritogenic synovial fibroblast. Arthritis Res 2003;5:R140–57.
- 24 Firestein GS. Invasive fibroblast-like synoviocytes in rheumatoid arthritis. Passive responders or transformed aggressors? Arthritis Rheum 1996;39:1781–90.
- 25 Taniguchi K, Kohsaka H, Inoue N, Terada Y, Ito H, Hirokawa K, et al. Induction of the p16INK4a senescence gene as a new therapeutic strategy for the treatment of rheumatoid arthritis. Nat Med 1999;5:760–7.
- 26 Kullmann F, Widmann T, Kirner A, Justen HP, Wessinghage D, Dietmaier W, et al. Microsatellite analysis in rheumatoid arthritis synovial fibroblasts. Ann Rheum Dis 2000;59:386–9.
- 27 Woerner SM, Kloor M, von Knebel Doeberitz M, Gebert JF. Microsatellite instability in the development of DNA mismatch repair deficient tumors. Cancer Biomark 2006;2:69–86.
- 28 Aplan PD. Causes of oncogenic chromosomal translocation. Trends Genet 2006;22:46–55.
- 29 Muller-Ladner U, Gay RE, Gay S. Activation of synoviocytes. Curr Opin Rheumatol 2000;12:186–94.
- 30 Qu Z, Garcia CH, O'Rourke LM, Planck SR, Kohli M, Rosenbaum JT. Local proliferation of fibroblast-like synoviocytes contributes to synovial hyperplasia. Results of proliferating cell nuclear antigen/cyclin, c-myc, and nucleolar organizer region staining. Arthritis Rheum 1994;37:212–20.
- 31 Xue C, Takahashi M, Hasunuma T, Aono H, Yamamoto K, Yoshino S, et al. Characterisation of fibroblast-like cells in pannus lesions of patients with rheumatoid arthritis sharing properties of fibroblasts and chondrocytes. Ann Rheum Dis 1997;56:262–7.
- 32 Yamanishi Y, Boyle DL, Rosengren S, Green DR, Zvaifler NJ, Firestein GS. Regional analysis of p53 mutations in rheumatoid arthritis synovium. Proc Natl Acad Sci USA 2002;99:10025–30.
- 33 Li J, Yen C, Liaw D, Podsypanina K, Bose S, Wang SI, et al. PTEN, a putative protein tyrosine phosphatase gene mutated in human brain, breast, and prostate
cancer. *Science* 1997;**275**:1943–7.
- 34 Steck PA, Pershouse MA, Jasser SA, Yung WK, Lin H, Ligon AH, et al. Identification of a candidate tumour suppressor gene, MMAC1, at chromosome 10q23.3 that is mutated in multiple advanced cancers. Nat Genet 1997;15:356–62.
- 35 Pap T, Franz JK, Hummel KM, Jeisy E, Gay R, Gay S. Activation of synovial fibroblasts in rheumatoid arthritis: lack of expression of the tumour suppressor
- PTEN at sites of invasive growth and destruction. Arthritis Res 2000;2:59–64. 36 Muller-Ladner U, Gay RE, Gay S. Role of nuclear factor kappaB in synovial inflammation. Curr Rheumatol Rep 2002;4:201–7.
- 37 Franz JK, Pap T, Hummel KM, Nawrath M, Aicher WK, Shigeyama Y, et al. Expression of sentrin, a novel antiapoptotic molecule, at sites of synovial invasion in rheumatoid arthritis. Arthritis Rheum 2000;43:599–607.
- 38 Pope RM. Apoptosis as a therapeutic tool in rheumatoid arthritis. Nat Rev Immunol 2002;2:527–35.
- 39 Catrina AI, Ulfgren AK, Lindblad S, Grondal L, Klareskog L. Low levels of apoptosis and high FLIP expression in early rheumatoid arthritis synovium. Ann Rheum Dis 2002;61:934–6.
- 40 Koch AE. Angiogenesis as a target in rheumatoid arthritis. Ann Rheum Dis 2003;62(Suppl 2):ii60–7.
- 41 Lafyatis R, Remmers EF, Roberts AB, Yocum DE, Sporn MB, Wilder RL. Anchorage-independent growth of synoviocytes from arthritic and normal joints. Stimulation by exogenous platelet-derived growth factor and inhibition by transforming growth factor-beta and retinoids. J Clin Invest 1989;83:1267–76.
- 42 Kang Y, Massague J. Epithelial-mesenchymal transitions: twist in development and metastasis. Cell 2004;118:277–9.
- 43 Thiery JP. Epithelial-mesenchymal transitions in tumour progression. Nat Rev Cancer 2002;2:442–54.
- 44 Zvaifler NJ. Relevance of the stroma and epithelial-mesenchymal transition (EMT) tor the rheumatic diseases. Arthritis Res Ther 2006;8:210.
45 Taketazu F, Kato M, Gobl A, Ichijo H, ten Dijke P, Itoh J, *et al*. Enhanced
- expression of transforming growth factor-beta s and transforming growth factorbeta type II receptor in the synovial tissues of patients with rheumatoid arthritis. Lab Invest 1994;70:620–30.
- 46 Pohlers D, Beyer A, Koczan D, Wilhelm T, Thiesen HJ, Kinne RW. Constitutive upregulation of the TGF-b pathway in rheumatoid arthritis synovial fibroblasts. Arthritis Res Ther 2007;9:R59.
- 47 Kasperkovitz PV, Timmer TC, Smeets TJ, Verbeet NL, Tak PP, van Baarsen LG, et al. Fibroblast-like synoviocytes derived from patients with rheumatoid arthritis show the imprint of synovial tissue heterogeneity: evidence of a link between an increased myofibroblast-like phenotype and high-inflammation synovitis.
Arthritis Rheum 2005;**52**:430–41.
- 48 Mattey DL, Dawes PT, Nixon NB, Slater H. Transforming growth factor beta 1 and interleukin 4 induced alpha smooth muscle actin expression and myofibroblast-like differentiation in human synovial fibroblasts in vitro: modulation by basic fibroblast growth factor. Ann Rheum Dis 1997;56:426–31.
- 49 Bauer S, Jendro MC, Wadle A, Kleber S, Stenner F, Dinser R, et al. Fibroblast activation protein is expressed by rheumatoid myofibroblast-like synoviocytes. Arthritis Res Ther 2006;8:R171.
- 50 Steenvoorden MM, Tolboom TC, van der Pluijm G, Lowik C, Visser CP, DeGroot J, et al. Transition of healthy to diseased synovial tissue in rheumatoid arthritis is associated with gain of mesenchymal/fibrotic characteristics. Arthritis Res Ther 2006;8:R165.
- 51 Radisky DC, Levy DD, Littlepage LE, Liu H, Nelson CM, Fata JE, et al. Rac1b and reactive oxygen species mediate MMP-3-induced EMT and genomic instability.
Nature 2005;**436**:123–7.
- 52 Konttinen YT, Li T-F, Hukkanen M, Ma J, Xu J-W, Virtanen I. Fibroblast biology: signals targeting the synovial fibroblast in arthritis. Arthritis Res 2000;2:348–55.
- 53 Frisch SM, Vuori K, Ruoslahti E, Chan-Hui PY. Control of adhesion-dependent cell survival by focal adhesion kinase. J Cell Biol 1996;134:793-9 54 Gumbiner BM. Cell adhesion: the molecular basis of tissue architecture and
- morphogenesis. Cell 1996;84:345–57. 55 Lelievre SA, Bissell MJ, Pujuguet P. Cell nucleus in context. Crit Rev Eukaryot
- Gene Expr 2000;10:13–20. 56 Aidinis V, Carninci P, Armaka M, Witke W, Harokopos V, Pavelka N, et al.
- Cytoskeletal rearrangements in synovial fibroblasts as a novel pathophysiological determinant of modeled rheumatoid arthritis. PLoS Genet 2005;1:e48.
- 57 Giganti A, Friederich E. The actin cytoskeleton as a therapeutic target: state of the art and future directions. Prog Cell Cycle Res 2003;5:511–25.
- 58 Wang J, Sanger JM, Sanger JW. Differential effects of Latrunculin-A on myofibrils in cultures of skeletal muscle cells: insights into mechanisms of myofibrillogenesis. Cell Motil Cytoskeleton 2005;62:35–47.
- 59 Kulms D, Dussmann H, Poppelmann B, Stander S, Schwarz A, Schwarz T. Apoptosis induced by disruption of the actin cytoskeleton is mediated via activation of CD95 (Fas/APO-1). Cell Death Differ 2002;9:598–608.
- 60 Puck TT, Krystosek A. Role of the cytoskeleton in genome regulation and cancer. Int Rev Cytol 1992;132:75–108.
- 61 **Disanza A**, Steffenc A, Hertzoga M, Frittoli E, Rottnerd K, Scitaa G. Actin
polymerization machinery: the finish line of signaling networks, the starting point
of cellular movement. C*ell Mol Life Sci* 2005;**62**:955–70
- 62 Castellano F, Le Clainche C, Patin D, Carlier M, Chavrier P. A WASP–VASP complex regulates actin polymerization at the plasma membrane. *Embo J*
2001;**20**:5603–14.
- 63 Krause M, Bear J, Loureiro J, Gertler F. The Ena/VASP enigma. J Cell Sci 2002;115:4721–6.
- 64 Kwiatkowski AV, Gertler FB, Loureiro JJ. Function and regulation of Ena/VASP proteins. Trends Cell Biol 2003;13:386–92.
- 65 Li J, Brieher WM, Scimone ML, Kang SJ, Zhu H, Yin H, et al. Caspase-11 regulates cell migration by promoting Aip1–Cofilin-mediated actin depolymerization. Nat Cell Biol 2007;9:276–86.
- 66 Stossel TP, Condeelis J, Cooley L, Hartwig JH, Noegel A, Schleicher M, et al. Filamins as integrators of cell mechanics and signalling. Nat Rev Mol Cell Biol 2001;2:138–45.
- Bishop A, Hall A. Rho GTPases and their effector proteins. Biochem J 2000;348:241–55.
- 68 Hall A. Rho GTPases and the actin cytoskeleton. Science 1998;279:509–14.
- 69 **Ridley AJ**. Rho tamily proteins: coordinating cell responses. Trends Cell Biol
2001;11:471–7.
- 70 Small JV, Stradal T, Vignal E, Rottner K. The lamellipodium: where motility begins. Trends Cell Biol 2002;12:112–20.
- 71 Xu H, Liu P, Liang L, Danesh FR, Yang X, Ye Y, et al. RhoA-mediated, tumor necrosis factor alpha-induced activation of NF-kappaB in rheumatoid synoviocytes: inhibitory effect of simvastatin. Arthritis Rheum 2006;54:3441-51.
- 72 Are AF, Galkin VE, Pospelova TV, Pinaev GP. The p65/RelA subunit of NFkappaB interacts with actin-containing structures. Exp Cell Res 2000;256:533–44.
- 73 Lee DM, Kiener HP, Agarwal SK, Noss EH, Watts GF, Chisaka O, et al. Cadherin-11 in synovial lining formation and pathology in arthritis. Science 2007;315:1006–10.
- 74 Vartiainen MK, Guettler S, Larijani B, Treisman R. Nuclear actin regulates dynamic subcellular localization and activity of the SRF cofactor MAL. Science 2007;316:1749–52.
- 75 Schratt G, Philippar U, Berger J, Schwarz H, Heidenreich O, Nordheim A. Serum response factor is crucial for actin cytoskeletal organization and focal adhesion assembly in embryonic stem cells. J[']Cell Biol 2002;156:737–50.
- 76 Southwick FS, Dabiri GA, Stossel TP. Neutrophil actin dysfunction is a genetic disorder associated with partial impairment of neutrophil actin assembly in three family members. J Clin Invest 1988;82:1525–31.
- 77 Procaccio V, Salazar G, Ono S, Styers ML, Gearing M, Davila A, et al. A mutation of β -actin that alters depolymerization dynamics is associated with autosomal dominant developmental malformations, deafness, and dystonia. Am J Hum Genet 2006;78:947-60.
- 78 Gearing M, Juncos JL, Procaccio V, Gutekunst CA, Marino-Rodriguez EM, Gyure KA, et al. Aggregation of actin and cofilin in identical twins with juvenile-onset dystonia. Ann Neurol 2002;52:465–76.
- 79 Bamburg JR, Wiggan OP. ADF/cofilin and actin dynamics in disease. Trends Cell Biol 2002;12:598–605.
- 80 de la Chapelle A, Tolvanen R, Boysen G, Santavy J, Bleeker-Wagemakers L, Maury CPJ, et al. Gelsolin-derived familial amyloidosis caused by asparagine or yrosine substitution for aspartic acid at residue 187. Nat Genet 1992;**2**:157–60.
- 81 Vallon R, Freuler F, Desta-Tsedu N, Robeva A, Dawson J, Wenner P, et al. Serum amyloid A (apoSAA) expression is up-regulated in rheumatoid arthritis and induces transcription of matrix metalloproteinases. J Immunol 2001;166:2801–7.
- 82 Lee MS, Yoo SA, Cho CS, Suh PG, Kim WU, Ryu SH. Serum amyloid A binding to formyl peptide receptor-like 1 induces synovial hyperplasia and angiogenesis. J Immunol 2006;177:5585–94.
- 83 Akiyama T, Kawasaki Y. Wnt signalling and the actin cytoskeleton. Oncogene 2006;25:7538–44.
- 84 Vasiliev J. Cytoskeletal mechanisms responsible for invasive migration of neoplastic cells. Int J Dev Biol 2004;48:425–39.
- 85 Avizienyte E, Wyke A, Jones R, McLean G, Westho VM, Brunton V, et al. Srcinduced de-regulation of E-cadherin in colon cancer cells requires integrin signaling. Nat Cell Biol 2002;4:632–8.
- 86 Bryant D, Stow JL. The ins and outs of E-cadherin trafficking. Trends Cell Biol 2004;14:427–34.
- 87 Attwell S, Roskelley C, Dedhar S. The integrin-linked kinase (ILK) suppresses anoikis. Oncogene 2000;19:3811–5.
- 88 Dai DL, Makretsov N, Campos EI, Huang C, Zhou Y, Huntsman D, et al. Increased expression of integrin-linked kinase is correlated with melanoma progression and poor patient survival. Clin Cancer Res 2003;9:4409–14.
- 89 Hannigan G, Troussard AA, Dedhar S. Integrin-linked kinase: a cancer therapeutic target unique among its ILK. Nat Rev Cancer 2005;5:51–63.
- 90 Kato K, Shiozawa T, Mitsushita J, Toda A, Horiuchi A, Nikaido T, et al. Expression of the Mitogen-Inducible Gene-2 (mig-2) is elevated in human uterine leiomyomas but not in leiomyosarcomas. Hum Pathol 2004;35:55–60.
- 91 Yoganathan TN, Costello P, Chen X, Jabali M, Yan J, Leung D, et al. Integrin-linked kinase (ILK): a ''hot'' therapeutic target. Biochem Pharmacol 2000;60:1115–9.
- 92 Spector I, Shochet NR, Blasberger D, Kashman Y. Latrunculins-novel marine macrolides that disrupt microfilament organization and affect cell growth: I. Comparison with cytochalasin D. Cell Motil Cytoskeleton 1989;13:127–44.