A **CHROMATOGRAPHIC AND SPECTROPHOTOMETRIC STUDY OF THE PRODUCTS OF THE REACTION OF** OSMIUM **TETROXIDE WITH UNSATURATED LIPIDS**

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ABSTRACT

Thin films of methyl oleate, oleic acid, and di-octadecenoyl phosphatidylcholine were reacted with a 2% solution of OsO₄ in water for 1 hr at 0°. As controls, methyl 9, 10-dihydroxystearate and 9,10-dihydroxystearic acid were reacted with $OsO₄$ in 0.25 N NaOH in methanol for 1 hr at room temperature. The reaction products were isolated, purified, and analyzed by thin-layer chromatography, gas-liquid chromatography, and infrared and visible spectroscopy. In all cases, the products were identified as diesters of osmic acid in which two molecules of fatty acids are linked through 1 molecule of osmic acid.

INTRODUCTION

I have recently presented evidence (1) that the product of reaction of a thin film of methyl oleate and aqueous osmium tetroxide is bis(methyl 9,10-dihydroxystearate)osmate. On the basis of C, H, and O analyses, infrared, visible, and nuclear magnetic resonance spectroscopy, and molecular weight determination, the following structure was suggested:

An apparently identical compound was isolated by transmethylation of the lipids of *Acanthamoeba* after fixation in Millonig's osmium tetroxide (2). The studies have now been extended to include the reactions of oleic acid and di-ll-octadecenoyl phosphatidylcholine with aqueous osmium tetroxide. The products of these reactions have been compared chromatographically and by infrared and visible spectroscopy to each other and to the products of reaction of methyl 9,10-dihydroxystearate and 9, 10-dihydroxystearic acid with osmium tetroxide in methanolic NaOH. The data indicate that, in all cases, the olefins and the glycols are converted into diesters which probably have the common structure:

This formulation differs from that proposed previously in possessing a hexavalent rather than a tetravalent Os and is in keeping with the structures proposed by Criegee (3, 4) for diesters formed from glycols and tetramethyl dipotassium osmate.

EXPERIMENTAL

MATERIALS: Methyl oleate and oleic acid were purchased from Applied Science Laboratories, Inc., State College, Pa. Oleic acid-1-¹⁴C was purchased from New England Nuclear Corp., Boston, and purified of any contaminating short chain fatty acids and neutral impurities by partitioning procedures previously described (5). Methyl oleate-1-¹⁴C was synthesized by reaction of oleic acid- $1-^{14}$ C with diazomethane in ether. Threo-9, 10-dihydroxystearic acid was purchased from K & K Laboratories, Inc., Plainview, N. Y., and converted to its methyl ester by reaction with diazomethane. The methyl 9, 10-dihydroxystearate was then purified by fractional crystallization from heptane. Purified 9,10 dihydroxystearic acid was prepared by saponification of the purified methyl 9,10-dihydroxystearate. The purity of all fatty acids and methyl esters was checked by thin-layer chromatography on silicic acid and by gas-liquid chromatography of the methyl esters on ethylene glycolsuccinate and SE-30.

Phosphatidylcholine was isolated from large cultures of *Agrobacterium tumefaciens (6).* The packed cells (750 g) were extracted with 14 liters of chloroform: methanol (2:1) overnight, and the filtrate was evaporated to dryness. The dried residue was extracted with chloroform which yielded 27 g of lipid. One-half of the lipids was fractionated on a column of 500 g of silicic acid eluting sequentially with 10 liters each of chloroform, 25% methanol in chloroform, 50% methanol in chloroform, and 100% methanol. The eluted fractions were analyzed by thin-layer chromatography, and the 50% methanol fraction was found to contain most of the phosphatidylcholine. This fraction was purified by chromatography on a smaller column of silicic acid. The fraction that was eluted by 40% methanol in chloroform but not by 35% methanol was found to be pure phosphatidylcholine by thin-layer chromatography. Fatty acid analyses (see Results) showed the material to be essentially di-ll-octadecenoyl phosphatidylcholine.

CHROMATOGRAPHY: Thin-layer chromatography of the reaction products of osmium tetroxide and methyl oleate, oleic acid, methyl 9,10-dihydroxystearate, and 9, 10-dihydroxystearic acid was carried out on silicic acid with a solvent system of heptane: diethyl ether: glacial acetic acid, 50:50:1. The soluble reaction products of osmium tetroxide and phosphatidylcholine were chromatographed on silicic acid with a large number of solvents in none of which did the osmium derivative move from the origin. Phosphatidylcholine was, therefore, readily separable from its osmium derivative. The osmium derivative of the phospholipid did migrate on thin-layer plates of cellulose with chloroform or methanol as the developing solvent, but not when heptane or diethyl ether were used. Compounds were usually eluted from the silicic acid or cellulose with anhydrous methanol. Spots that contained osmium were brown and gave a positive reaction when sprayed with the benzidine reagent described by Riemersma (7). Colorless lipids were visualized under ultraviolet light after the plates were sprayed with 0.1% dichlorofluorescein in 50% ethanol. In the experiments with 14 C-compounds, all areas of the plates were eluted with methanol and counted.

Fatty acids were analyzed as their methyl esters by gas-liquid chromatography on both 17% ethylene glycol succinate at 190° and on SE-30 at 205°. The columns measured 6 ft \times 5 mm, the inlet pressure was maintained at 10 lb/in2, and an argon ionization detector was used. The detector was calibrated with standards at known concentrations. Details of these procedures as applied to the compounds of interest in this paper have been published (1, 2).

ANALYSES: Phosphorus was analyzed by a modification (8) of the procedure of Fiske and SubbaRow (9). For determination of infrared spectra, the compounds were dissolved in CC14, or made into mulls in Nujol. A Perkin-Elmer 21 spectrophotometer and a Beckman IR 7 spectrophotometer were used. Visible spectra were obtained with a Cary recording spectrophotometer. Radioactivity was measured in a scintillation spectrometer with the use of 0.4% diphenyloxazole in toluene as scintillator-solvent. Quench corrections were made by the channels ratio method.

RESULTS

Synthesis of Osmium Derivatives

Tetramethyl dipotassium osmate was synthesized exactly as described by Criegee et al. (4) . Osmium tetroxide (1.02 g) was dissolved in 10 ml of methanol, added to 25 ml of I N KOH in methanol, and the solution warmed for a few minutes. The deep green crystals that formed after cooling the solution were collected, washed, and dried.

*Methyl oleate-1-*¹⁴C (1 mmole, 3.6×10^6 cpm) was reacted with 1.05 mmole of OsO₄ (as a 2% solution in water) for 1 hr at 0° . The black tarry oil that formed was washed with water and extracted into heptane, leaving behind a black residue. The insoluble material (presumably OsO2) weighed 167.5 mg. The dark purple-brown heptane solution contained 85% of the initial radioactivity. The soluble product was chromatographed on silicic acid plates, 94% of the radioactivity was recovered in a brown, osmium-positive spot with an Rf' identical to that previously found (1) for bis(methyl 9, 10-dihydroxystearate)osmate. No radioactivity was present on the thin-layer plate in the areas in which methyl oleate and methyl 9,10-dihydroxystearate would have been. Gas-liquid chromatographic analysis (1) revealed only bis(methyl 9,10-dihydroxystearate)osmate. Both unreacted methyl oleate and methyl 9,10 dihydroxystearate are easily detected by this method. The product was converted quantitatively to methyl 9, 10-dihydroxystearate when heated at 80 $^{\circ}$ for 1 hr in 1 N HCl in 50 $\%$ methanol (1). The specific radioactivity of the osmium derivative after purification by thin-layer chromatography was 8.3 \times 10³ cpm/mg. The theoretical specific radioactivity for bis(methyl 9, 10-dihydroxystearate)osmate synthesized from methyl oleate $(3.6 \times 10^6 \text{ cm/mole})$ is 8.2 $\times 10^3 \text{ cm/mg}$. This agrees with the analytical data published previously (1) which indicated that the product contains 2 moles of fatty acid and 1 mole of osmium. The product purified by thin-layer chromatography was used for the spectral analyses.

Methyl 9,10-dihydroxystearate (0.2 moles) was reacted at room temperature with 0.15 mmoles of $OsO₄$ for 1 hr in 4 ml of 0.25 N NaOH. The deep yellow-orange solution was acidified, whereupon a dark brown precipitate formed. An equal volume of water was added to the mixture, and it was extracted with heptane. The heptane was evaporated to give 85 mg of a dark purple-brown oil. This product was identical to bis(methyl 9,10-dihydroxystearate)osmate synthesized from methyl oleate by gas-liquid and thin-layer chromatographic analyses. No unreacted methyl 9,10 dihydroxystearate was detected. The osmium derivative could be converted to methyl 9,10 dihydroxystearate by hydrolysis. The product was used for spectral analyses after purification by thin-layer chromatography.

Oleic acid-1-¹⁴C (1 mmole, 4×10^6 cpm) was reacted with 1.05 mmole of $OsO₄$ (2% water) for I hr at 0°. The black precipitate that formed was washed with water and was then extracted with heptane:ether, 1:1. The extract contained 95% of the radioactivity. A nonradioactive black residue (113 mg), presumably $OsO₂$, remained. The extract was analyzed by thin-layer chromatography, which revealed one dark brown, osmium-

positive spot with an Rf of 0.39. This material contained 55% of the radioactivity that was applied to the plate. The remainder of the radioactivity was present in the area corresponding to unreacted oleic acid. No radioactivity was detected in the area that would have contained 9,10 dihydroxystearic acid if it were present. The product was converted quantitatively to 9,10 dihydroxystearic acid by hydrolysis (1). The specific radioactivity of the purified osmium derivative was 8.9 \times 10³ cpm/mg, which is in reasonable agreement with the theoretical specific radioactivity for bis(9,10 - dihydroxystearic acid)osmate synthesized from oleic acid (4 \times 10⁶ cpm/mmole), i.e., 9.6×10^3 cpm/mg. The purified material was used for the spectral analyses.

9,10-Dihydroxystearic acid (0.09 mmoles) was reacted with 0.07 mmoles OsO4 in 4 ml of 0.25 N NaOH at room temperature for 1 hr. The color of the solution progressed from yellow-orange through brown-yellow, green-brown, light purple, and, finally, to a deep violet after 25 min. The color changes are related to the reduction of $OsO₄$ to tetramethyl dipotassium osmate (3, 4). The solution was cooled and acidified which resulted in a brown solution to which an equal volume of water was added. The product was extracted into heptane: ether, 1:1. The solvent was evaporated, and 34.2 mg of a dark brown oil was recovered. When chromatographed on a thin-layer of silicic acid, one major brown, osmium-positive spot was found with the same Rf as the product formed from oleic acid-1-¹⁴C and $OsO₄$. Nothing was present in the area on the chromatogram corresponding to 9, 10-dihydroxystearic acid. The osmium derivative was converted quantitatively to the glycol acid by hydrolysis in 1 N HC1. Spectral analyses were performed on material purified by silicic acid chromatography.

Phosphatidylcholine (136 mg) was reacted with 25 ml of 2% OsO₄ in water for 1 hr at 0^o. The black precipitate that formed was washed with water and then extracted several times with methanol. The red-brown methanol solution was evaporated which yielded 101 mg of the methanol-soluble product. The methanol-insoluble material was washed with ether and air-dried to give 83 mg of product. The starting material contained 160 μ eq of phosphorus, the methanol-soluble product 83 μ eq of phosphorus and the methanol-insoluble fraction 75μ eq of phosphorus. The minimal molec-

¹ Rf = relative to the solvent front.

ular weight of the methanol-soluble product calculated for one equivalent of phosphorus is 1 150 (101 $mg/0.083$ mmoles = 1150 mg/mmole). This value agrees very closely with the theoretical molecular

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FIGURE 1 Thin-layer chromatogram of the methanolsoluble osmium derivative of di-ll-octadecenoyl phosphatidyleholine. The osmium derivative and a sample of the unreacted phospholipid were applied in narrow bands. The plate was developed first with diethyl ether, then air-dried, and redeveloped with chloroform. The unreacted phospholipid migrated with the ether front, while the osmium derivative remained at the origin. In chloroform the osmium derivative migrated with the front, leaving a faint gray, phosphorus-negative band at the origin. The brown band at the chloroform front was phosphorus-positive and osmium-positive.

weight for the addition product containing 1 mole of phosphatidylcholine and 1 mole of $O₈O₄$ $(850 + 254 = 1104)$. By analogy to the reactions with methyl oleate and oleic acid, it may be assumed that the methanol-insoluble material contained $OsO₂$ in addition to the compound that contained the phosphorus.

The methanol-soluble osmium derivative did not move from the origin of silicic acid thin-layer plates in any of the solvents used. Unreacted phospholipid was not present. When chromatography on thin-layers of cellulose was done, phosphatidylcholine was found to move with the solvent front in diethyl ether (Fig. 1), but the methanol-soluble osmium derivative remained at the origin. The osmium derivative migrated with the front when the same plate was redeveloped with chloroform (Fig. 1). The spot was osmiumpositive. The only other material detected was a faint gray band at the origin.

The fatty acids of the phosphatidylcholine and its methanol-soluble osmium derivative were converted to methyl esters by methanolysis in 0.5 N sodium methoxide and analyzed by gas-liquid chromatography (1, 2). The original phosphatidylcholine had the composition: 4% hexadecanoate; 2% 9-hexadecenoate; 94% 11-octadecenoate; 2% 11,12-methylene octadecanoate, and can, therefore, be considered to be essentially a di-l 1 octadecenoyl phosphatidylcholine. The methanolsoluble osmium derivative had the composition: 9.3% hexadecanoate; 87% bis(methyl 11,12 dihydroxystearate)osmate; and 3% 11,12-methylene octadecanoate. It contained no dihydroxyste-

FIGURE 2 Infrared spectrum of $OsO₄$ in CCl₄

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FIGURE 3 Infrared spectra of CCl₄ solutions of methyl oleate, methyl 9, 10-dihydroxystearate, and the purified products of the reactions of S0O4 with methyl oleate and methyl 9,10-dihydroxystearate. The spectra of the two osmium derivatives are essentially identical and differ markedly from the spectrum of methyl 9, 10-dihydroxystearate.

arate or olefinic fatty acids. When the osmium derivative was hydrolyzed in 1 N HCl at 80° for 1 hr, the fatty acid analysis revealed: 9.1 % hexadecanoate; 14.7% bis(methyl 11,12-dihydroxystearate)osmate; and 76% 11,12-dihydroxystearate.

Infrared Spectral Data

The infrared spectrum of $OsO₄$ is shown in Fig. 2. The spectrum has one sharp absorption band at 10.53 μ . No absorption bands were found for $OsO₂$, and tetramethyl dipotassium osmate had no absorption in the region of 10.5 μ . It, of course, does not have an Os=O group. In Fig. 3, the infrared spectra of the osmium derivatives of methyl oleate and methyl 9, 10-dihydroxystearate dissolved in CCI4 are compared to the spectra of authentic methyl oleate and methyl 9,10-dihydroxystearate. The spectra of the two osmium derivatives are nearly identical and show only two significant differences from the spectrum of methyl oleate. One difference is the absence of the absorption band at 3.3 μ attributable to the *cis*double bond of methyl oleate; the other is the presence of a new absorption band at 10.1 μ . This

FIGURE 4 The infrared spectra of CC14 solutions of oleic acid and the purified products of the reactions of Os04 with oleic acid and 9, 10-dihydroxystearic acid. The spectrum of 9, 10-dihydroxystearic acid was obtained as a mull in Nujol because of its insolubility in CC14. The spectra of the two osmium derivatives are essentially identical and differ markedly from the spectrum of 9,10-dihydroxystearic acid.

new band is undoubtedly due to the Os=O function, as suggested also by Stoeckenius and Mahr (10), although its position is somewhat different from the corresponding absorption band of $OsO₄$. Neither the osmium derivative prepared from methyl oleate nor the osmium derivative of methyl 9,10-dihydroxystearate has an absorption band at 2.9 μ , that is characteristic of hydroxyl groups. That band is, of course, present in the spectrum of authentic methyl 9,10-dihydroxystearate. This confirms the absence of methyl 9, 10-dihydroxystearate as indicated by thin-layer and gas-liquid chromatography.

Similar spectra are shown in Fig. 4 for the osmium derivatives of oleic acid and 9,10 dihydroxystearic acid as compared to authentic

oleic acid and 9,10-dihydroxystearic acid. Again, the spectra of the two osmium derivatives are nearly identical. The band at 10.1 μ attributable to the Os=O group is present; the band at 3.3 μ due to the *cis-olefin* is missing; and there is no absorption in the region of 3 μ where hydroxyl groups would absorb if they were present. The hydroxyl absorption band is clearly evident in the spectrum of authentic 9, 10-dihydroxystearic acid. The broad absorption between $3-4$ μ underlying the sharp bands is a result of the COOH group and is present in the spectra of all four acids. The product of reaction of oleic acid and OsO4 shows a small band at 6.5 μ similar to the band Stoeckenius and Mahr (10) attributed to heavy metal soaps. In the present experiments, it seems unlikely that much

FIGURE 5 Infrared spectra of CC14 solutions of di-ll-octadecenoyl phosphatidylcholine and its methanolsoluble osmium derivative. The spectrum of the methanol-insoluble osmium derivative was obtained as a mull in Nujol. The osmium derivatives show no increase in absorption in the hydroxyl region (3μ) (See Table I). The phospholipids were dried in vacuo over P_2O_5 to remove water of hydration.

of the product could be in this form because of its solubility and chromatographic properties, but the possibility cannot be entirely eliminated.

The spectra of the methanol-soluble and

methanol-insoluble osmium derivatives of di-octadecenoyl phosphatidylcholine are compared with the spectrum of the original phospholipid in Fig. 5. The major differences between the spectra of

TABLE I

Relative Absorption in the Hydroxyl Region of Phosphatidylcholine, Methanol-Soluble Osmium Derivative of Phosphatidylcholine and Methyl 9, 10-Dihydroxystearate

	Phosphatidyl- choline	Osmium Phosphatidyl- choline	derivative of Methyl 9, 10- Dihydroxy- stearate
$D_{3.1} \mu/D_{3.45} \mu$	0.099	0.094	0.196
$D_{3,1} \mu/D_{5,8} \mu$	0.13	0.12	0.23
$D_{5,8} \mu / D_{3,45} \mu$	0.76	0.78	0.85

 $D_{3,1}$ μ is due to hydroxyl groups; $D_{3,45}$ μ is due to methylene groups; $D_{5.8} \mu$ is due to ester carbonyl groups.

the methanol-soluble osmium derivative and the starting material are the absence of the absorption band at 3.3 μ *(cis-double bond)* and the presence of a new band at 10.1 μ (Os=O). There is no increase in the absorption in the hydroxyl region of the spectrum. This is more clearly shown in the expanded spectra of that region. One can compare quantitatively (11) the absorption of the hydroxyl groups (3.1 μ) in the two compounds by relating these absorptions to those of the methylene groups (3.45 μ) and to those of the carbonyl groups (5.8 μ). As shown in Table I, the methanol-soluble osmium derivative has relatively less absorption in the hydroxyl region than does the original phosphatidylcholine. This absorption may be due to residual water, which is very difficult to remove

FIGURE 6 Visible spectra in acidic and alkaline methanol of tetramethyl dipotassium osmate and the osmium derivatives of methyl oleate, methyl 9,10-dihydroxystearate, and di-11-octadecenoyl phosphatidylcholine.

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FIGURE 7 Proposed mechanism for the synthesis of his (methyl 9, 10-dihydroxystearate)osinate or bis (9,10-dihydroxystearic acid)osmate from $0sO₄$ and methyl 9,10-dihydroxystearate or 9,10-dihydroxystearic acid. This mechanism is based on the work of Criegee (3, 4).

FIGURE 8 Proposed mechanism for the synthesis of his (methyl 9,10-dihydroxystearate)osmate or bis (9,10-dihydroxystearic acid)osmate from 0s04 and methyl oleate or oleic acid. This mechanism is very similar to one proposed by Becker (12) for the analogous reaction with acenapthylene.

from phospholipids, to $-P$ -OH groups, or to hydroxyl groups neutralizing the quaternary nitrogen.

The infrared spectrum of the methanol-insoluble osmium derivative of di-octadecenoyl phosphatidylcholine could not be obtained in solution in CC14. As a mull in Mujol, its spectrum is very similar to that of the soluble osmium derivative with the characteristic $10.1-\mu$ band from Os=O.

Visible Spectral Data

The visible spectra of the osmium derivatives of methyl oleate, methyl 9, 10-dihydroxystearate, and

di-octadecenoyl phosphatidylcholine are very similar (Fig. 6). All show a maximum at about 700 $m\mu$ when the compounds are dissolved in 0.1 μ NaOH in methanol, and all show a change to 2 maxima at about 465 and 565 m μ when the compounds are dissolved in 0.1 N H_2SO_4 in methanol. Similar spectra were obtained for tetramethyl dipotassium osmate, except that only 1 maximum was obtained in acidic methanol. Similar results were also found for the osmium derivatives of oleic acid and 9, 10-dihydroxystearic acid, but because of their low solubility these compounds give opalescent solutions and, therefore, poor spectra.

FIGURE 9 Proposed mechanism for the effect of changes in pH on the visible spectra of diesters of osmic acid.

DISCUSSION

All of the data in this and the previous papers (1, 2) are in agreement with the conclusion that the product of the reaction between thin films of oleic acid, or its methyl ester, and aqueous solutions of OsO₄ is a stable diester containing one molecule of osmic acid and two molecules of the glycol formed by oxidation of the olefin. This conclusion is supported by elemental analyses, molecular weight determination, nuclear magnetic resonance spectra, gas-liquid and thin-layer chromatography, the specific radioactivity of the products synthesized from radioactive precursors, and infrared and visible spectral data. Additional support for this identification is given by the fact that indistinguishable compounds are formed from the reaction of 9, 10-dihydroxystearic acid, or its methyl ester, with OsO₄ in alkaline methanol. These model reactions are analogous to those that were studied by Criegee $(3, 4)$. Criegee showed that $OsO₄$ is reduced by alkaline methanol to tetramethyl dipotassium osmate and that this compound reacts with two molecules of glycol in a transesterification reaction as illustrated in Fig. 7.

A plausible mechanism for the synthesis of these products starting from the olefinic acid (or its methyl ester) is shown in Fig. 8. In this case, the reaction is envisaged as occurring in several steps, although intermediates may not accumulate in detectable amounts. Oleic acid (or its methyl ester) would react with $OsO₄$ in an oxidationreduction reaction to form the osmic acid ester of 9,10-dihydroxystearic acid. Reactions of this kind have been well characterized by Criegee (3, 4) for other olefins. Two molecules of the monoester could then react forming the diester and **Os03** which could be expected to be unstable and, therefore, dismutate to $OsO₂$ and $OsO₄$. This proposed reaction scheme accounts for all the products that have been found: Bis(9,10 - dihydroxystearic acid)osmate and $OsO₂$.

In Criegee's research, only monoesters were found as the products of reaction of olefins and OsO4 in anhydrous organic solvents, but recently Becker (12) has found that acenapthylene forms a diester with osmic acid when reacted with OsO4 in aqueous dioxane. The reaction mechanism suggested by Becker is almost identical to the scheme shown in Fig. 8, except that he proposes the free glycol as an intermediate. The glycol is not included in the reaction sequence in Fig. 8 because its presence could not be detected by the sensitive methods of gas-liquid chromatography, thin-layer chromatography, and infrared spectroscopy. Riemersma (7) has presented indirect evidence for the formation of glycols during the reaction of phospholipids and $OsO₄$ in aqueous *t*-butanol. Stoeckenius and Mahr (10) noted an increase in the absorption in the hydroxyl region of infrared spectra of unsaturated lipids after reaction with OsO4 in anhydrous chloroform. This observation is difficult to evaluate because, under anhydrous conditions, there is no source for hydroxyl groups. It may be that the absorption band was due to water present in the KBr pellets used in the infrared spectroscopy. All of the compounds synthesized in the present study showed an artifactual "hydroxyl" absorption band when the spectra were obtained as KBr pellets.

The structure of the diester in Fig. 8 differs slightly from that proposed in the earlier papers (1, 2). There are two reasons for suggesting that the product contains a hexavalent osmium atom. First, the same product is formed from the olefin and the glycol, and the most plausible explanation are reaction mechanisms that lead to a hexavalent osmium. Second, the acid-base shift in the visible spectra of the osmium derivatives is evidence for a

titratable group attached to the osmium such as a "carbonyl" type oxygen that can undergo "ketol" transformation (Fig. 9).

The fatty acid and phosphorus analyses and the infrared and visible spectra leave little doubt that the products of the reaction between $OsO₄$ and di-ll-octadecenoyl phosphatidylcholine are directly analogous to the products formed from oleic acid and methyl oleate. A reasonable structure for, and mechanism of synthesis of, the methanol-

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H_{2}C - 0 - C - (CH_{2})_{9}CH = CH(CH_{2})_{5}CH_{3}
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H_{2}C - 0 - C - (CH_{2})_{9}CH = CH(CH_{2})_{5}CH_{3}
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H_{2}C - 0 - P^{50}_{1}O - CH_{2}CH_{2}N(CH_{3})_{3}
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O = C
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H_{2}C - 0 - C - (CH_{2})_{9}CH - CH(CH_{2})_{5}CH_{3}
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H_{2}C - 0 - C - (CH_{2})_{9}CH - CH(CH_{2})_{5}CH_{3}
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H_{2}C - 0 - P^{50}_{1}O - CH_{2}CH_{2}N(CH_{3})_{3}
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H_{2}C - 0 - P^{50}_{1}O - CH_{2}CH_{2}N(CH_{3})_{3}
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FIGURE 10 Proposed mechanism of synthesis, and structure, of the methanol-soluble osmium derivative of di-11-octadecenoyl phosphatidylcholine.

soluble osmium derivative is shown in Fig. 10. In a reaction identical to that proposed in Fig. 8 for oleic acid, OsO4 would react with two molecules of a monounsaturated fatty acid to form a diester of osmic acid. In this instance, however, the two monounsaturated fatty acids are on the same phospholipid molecule, giving an "internal" diester. The methanol-insoluble osmium derivative of the phospholipid might result when the osmic acid diester links fatty acids of different phospholipid molecules, thus leading to the formation of a polymer. One possible configuration of such a polymer is shown in Fig. 11.

The data presented in this paper, therefore, give some direct experimental support to the hypotheses of Wigglesworth (13), Baker (14), and Stoeckenius and Mahr (10) that the fixation of biological material by $OsO₄$ may, in part, be due to the linkage of the double bonds of fatty acids through stable diesters. If more than one unsaturated fatty acid, or one polyunsaturated fatty acid, occurs in the lipids, polymerization is possible. Stoeckenius has interpreted electron micrographs of lipid-water systems fixed with $OsO₄$ (15, 16) and the infrared spectra of the products of reaction of olefinic lipids and $OsO₄$ in chloroform (10) to indicate the accumulation of osmium at the polar end of lipid molecules. He suggests that, after an initial stoichiometric reaction of Os04 with the double band, the osmic ester might break down, releasing the osmium which could then migrate to the polar

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(CH_{3})_{3}NCH_{2}CH_{2} - \frac{P}{P} - O - CH_{2} \underset{Q}{\overset{P}{\underset{Q}{\bigcup}}} O \underset{H_{2}C}{\overset{P}{\underset{Q}{\bigcup}}} O \underset{Q \leq \underset{Q}{\bigcup} O}{\overset{P}{\underset{Q}{\bigcup}}} O
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$$
H_{2}C - O - \underset{G}{C}CH_{2})_{9}CH - \underset{Q}{C}H \underset{Q}{\underset{Q}{\bigcup}} O \underset{Q}{\overset{Q}{\underset{Q}{\bigcup}}} O}
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CH_{3}(CH_{2})_{5}CH - \underset{Q}{C}H \underset{Q}{\overset{Q}{\bigcup}} O
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CH_{3}(CH_{2})_{5}CH - \underset{Q}{C}H \underset{Q}{\underset{Q}{\bigcup}} O \underset{Q}{\overset{Q}{\bigcup}} O
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CH_{3}(CH_{2})_{5}CH - \underset{Q}{C}H \underset{Q}{\underset{Q}{\bigcup}} O - O - CH_{2}
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CH_{3}(CH_{2})_{5}CH - \underset{Q}{\underset{Q}{\bigcup}} O \underset{Q}{\overset{Q}{\bigcup}} O
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H_{2}C - O - \underset{Q}{\overset{Q}{\bigcup}} O \underset{Q}{\overset{Q}{\bigcup}} O
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H_{2}C - O - \underset{C}{C}CH_{2})_{9}CH - \underset{C}{C}H \underset{C}{\bigcup} H \underset{Q}{\overset{Q}{\bigcup}} O
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H_{2}C - O - \underset{Q}{\underset{Q}{\bigcup}} O \underset{Q}{\overset{Q}{\bigcup
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FIGURE 11 One possible structure for the methanol-insoluble osmium derivative of di-11-octadecenoyl phosphatidyleholine.

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groups. The reactions proposed in Fig. 8 provide a mechanism whereby osmium could be released while the fatty acyl chains are still linked as diesters of osmic acid. Under the reaction conditions described in the present paper, however, little if any osmium appears to be bound to anionic groups. The solubility of most of the products, the specific radioactivity data, the chromatographic prop-

erties, and the phosphorus analyses all indicate that the osmium derivatives contain only the one molecule of esterified osmium for every two molecules of fatty acid. Whether a reaction between osmium and the polar groups of lipids occurs during biological fixations remains for future research to determine.

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REFERENCES

- 1. KORN, E. D. 1966. *Biochim. Biophys. Acta.* 116:317.
- 2. KORN, E. D. 1966. *Biochim. Biophys. Acta.* 116:325.
- 3. CRIEGEE, R. 1936. *Ann. Chem.* 522:75.
- 4. CRIEGEE, R., B. MARCHAND, and H. WAN-NOWIUS. 1942. *Ann. Chem.* 550:99.
- 5. McBRIDE, O. W., and E. D. KORN. 1964. *J. Lipid Res.* 5:448.
- 6. KANESHIRO, T., and A. G. MARR. *J. Lipid Res.* 3:148.
- 7. RIEMERSMA, J. C. 1963. *J. Histochem. Cytochem.* 11:436.
- 8. DAVIDOFF, F., and E. D. KORN. 1963. *J. Biol. Chem.* 238: 3199.
- 9. **FISKE,** C. H., and Y. SUBBARow. 1925. *J. Biol. Chem.* 66:375.
- 10. STOECKENIUS, W., and S. C. MAHR. 1966. *Lab. Invest.* 14: 1196.
- 11. Official and Tentative Methods of the American Oil Chemists' Society. 1964. American Oil Chemists' Society, Chicago. 2nd edition. Cd 14-16.
- 12. BECKER, R. Diplom. Arbeit Germany, Technische Hochschule Karlsruhe. 1959.
- 13. WIGGLESWORTH, V. B. 1947. *Proc. Roy. Soc. (London), Ser. B.* 147:185.
- 14. BAKER, J. R., 1958. *J. Histochem. Cytochem.* 6:303.
- 15. STOECKENIUS, W. 1960. *Proc. European Regional Conf. Electron Micr., Delft.* 2:176.
- 16. STOECKENIUS, W. 1962. *J. Cell Biol.* 12:221.