## Folic Acid and Pteroylpolyglutamate Contents of Archaebacteria

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Cell extracts of methanogens and the thermoacidophile Sulfolobus solfataricus contained little or no folic acid (pteroylglutamate) or pteroylpolyglutamate activity  $(<0.1$  nmol/g [dry weight]). However, the halophile Halobacterium salinarum contained pteroylmono- or pteroyldiglutamates, and Halobacterium volcanii and Halobacterium halobium contained pteroyltriglutamates at levels equivalent to those in eubacteria  $(>1 \text{ nmol/g})$ [drv weight]).

The archaebacteria, eubacteria, and eucaryotes differ from each other in numerous biological properties (16-18, 35), including coenzyme biochemistry. The methanogenic archaebacteria employ novel cofactors in the pathway of methane biosynthesis (36), and novel pterins and cofactorlike compounds are present in halophilic and thermoacidophilic archaebacteria (22-24, 30, 34). However, the familiar growth factors thiamine, riboflavin, nicotinic acid, pantothenate, pyridoxine, and biotin are found in methanogens (21); and the first five of these have been detected in halophiles and thermoacidophiles (K. M. Noll, personal communication). Methanogens contain extremely low levels of folic acid activity and tetrahydrofolate-dependent enzymes (9, 21). Traces of folate activity have been attributed to a hydrolytic product of methanopterin (a folate analog) which functions as a one-carbon carrier (8, 12, 20, 21, 33).

Folate is a generic term for forms of pteroylglutamic acid (PteGlu<sub>n</sub>;  $1 \le n \le 7$ ) which are central to one-carbon metabolism in eubacteria and eucaryotes (2, 3, 11, 19, 26, 31). A given organism mainly contains <sup>a</sup> single chain length (2, 11). The specificities of the bioassay strains and changes in folate content after treatment with conjugase (which releases the PteGlu nucleus from PteGlu<sub>n</sub>) permit the identification of the folate moieties (10, 26, 31).

Here we provide evidence for pteroylpolyglutamates in halobacteria.

(A preliminary report of this study was presented previously [D. P. Nagle, Jr., V. E. Worrell, C. W. Jones, and R. Teal, Abstr. Annu. Meet. Am. Soc. Microbiol. 1987, 1130, p. 194].)

Bacterial strains and media. The bacterial strains used in this study are listed in Table 1. The methanogens were grown essentially as described previously (1) in folate-free media from which vitamins, yeast extract, Casitone (Difco Laboratories, Detroit, Mich.), and Trypticase (BBL Microbiology Systems, Cockeysville, Md.) were omitted. Methanococcus voltae was supplemented with 0.5 g of leucine, 1.0 g of isoleucine, and 50 mg of calcium pantothenate per liter; 40  $\mu$ M NiCl<sub>2</sub>; and 1  $\mu$ M sodium selenate. Methanosarcina barkeri was grown on methanol (medium 1043 [5]) with <sup>1</sup> g of vitamin-free Casamino Acids (Difco) per liter. Sulfolobus solfataricus was grown at  $60^{\circ}$  C in medium containing, per liter, 1.3 g of  $(NH_4)_2SO_4$ , 0.2 g of  $KH_2PO_4$ , 0.25 g of  $MgSO_4 \cdot 7H_2O$ , 76 mg of  $CaCl_2 \cdot 2H_2O$ , and 1.0 g of Casamino Acids (Difco) adjusted to pH 3.0. Halobacteria were grown at 43°C in medium containing, per liter, 7.6 g of

vitamin-free Casamino Acids (Difco), 3 g of sodium citrate, 250 g of NaCl, 2 g of KCl, 20 g of  $MgCl_2 \cdot 7H_2O$ , 0.4 g of FeSO<sub>4</sub>. 7H<sub>2</sub>O, and 0.2 mg of MnSO<sub>4</sub>. H<sub>2</sub>O (from medium 97 [4]). Eubacteria were grown in folic acid assay medium without folate (Difco). Pteroic acid was a gift from L. D'Ari (University of California, Berkeley), and methanopterin was from J. C. Escalante-Semerena and W. J. Jones (University of Illinois, Urbana). Acid hydrolysis of methanopterin was in <sup>6</sup> N HCl (1 h, 100°C).

Bioassay for folic acid. Cells were centrifuged and, except for the halophiles, washed by suspension in 0.9% NaCl and centrifuged. Pellets were heated  $(20 \text{ min}, 100^{\circ}\text{C})$  in H<sub>2</sub>O  $(1 \text{ to}$ 2 ml per g of cells) and centrifuged. The folate content was reproducible among different extracts and stable on storage  $(-20^{\circ}C)$ . Extreme care was taken to avoid contamination of glassware and media with exogenous folate. Most media components were tested for folate content with Streptococcusfaecalis, and in particular, vitamin-free Casamino Acids (Difco) contained less than 2 pmol of folic acid per g. Streptococcus faecalis and Lactobacillus casei were grown in folic acid assay medium and folic acid Casei medium (Difco), respectively (7, 10). All assays were done in triplicate. The  $A_{540}$  of cultures was read after 24 to 48 h. Unknown values were estimated from the linear portion of the folate standard curves, between 0.1 and <sup>1</sup> ng/ml for

TABLE 1. Bacterial strains used in this study

Organism	Strain no. <sup><i>a</i></sup>	Source"
Archaebacteria		
<b>Methanobacterium</b> formicicum	<b>ATCC 33274</b>	W. J. Jones
Methanobacterium thermo- <i>autotrophicum</i> Marburg	<b>DSM 2133</b>	DSM, H. Hippe
Methanococcus voltae	<b>DSM</b> 1537	W. J. Jones
Methanosarcina barkeri	<b>DSM 800</b>	M. J. McInerney
Sulfolobus solfataricus	<b>ATCC 35091</b>	<b>ATCC</b>
Halobacterium volcanii	<b>ATCC 29605</b>	R. Gupta
Halobacterium salinarum	<b>ATCC 19700</b>	<b>ATCC</b>
Eubacteria		
<b>Bacillus subtilis</b>	OU 10093	K. Kealy
Lactobacillus casei	<b>ATCC 7469</b>	<b>ATCC</b>
Pseudomonas fluorescens	<b>OU 10025</b>	K. Kealy
Streptococcus faecalis (faecium)	<b>ATCC 8043</b>	<b>ATCC</b>

<sup>a</sup> Abbreviations: ATCC, American Type Culture Collection, Rockville, Md.; DSM, Deutsche Sammlung von Mikroorganismen, Gottingen, Federal Republic of Germany; OU, Department of Botany and Microbiology Culture Collection, University of Oklahoma, Norman, Okla.

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Streptococcus faecalis  $(A_{540}, 0.09 \text{ to } 0.97)$  and 0.01 and 0.4 ng/ml for Lactobacillus casei  $(A_{540}, 0.04$  to 0.567). Unknowns with the lowest levels of folate yielded  $A_{540}$ s of less than 0.05, with considerable scatter among replicates. For every condition assayed, uninoculated sterile controls did not exhibit growth after 96 h. All unknowns were tested for the inhibition of folate-dependent growth of the assay strains. Tubes with folate (0.4 to 0.5 ng/ml) were challenged with 0.05 ml of extracts and read at 24 h. Conjugase was prepared from desiccated hog kidney (Sigma Chemical Co., St. Louis, Mo.), and treatment at pH 4.5 was done essentially as described previously (10). Correction for folate in the enzyme and controls which lacked enzyme (sham conjugase) were done. Protein determinations were done (25) after cells were heated in a base. Folate levels were expressed on a dry weight basis (assuming 0.55 g of protein per g [dry weight] [13]).

Growth of indicator strains. Streptococcus faecalis grew on folate, pteroic acid, and thymidine (since all other products of folate metabolism were present in the medium). Folate-dependent growth was inhibited by the antifolate compound trimethoprim, whereas thymidine-dependent growth was not (Table 2) (28). The organism is known to grow on Pte $Glu_1$ , Pte $Glu_2$ , and reduced derivatives (26, 31). Lactobacillus casei was supported by folate and methanopterin and utilizes PteGlu<sub>2</sub> and PteGlu<sub>3</sub> (26, 31). Both *Lacto*bacillus casei and Streptococcus faecalis subsisted on acidhydrolyzed methanopterin (Table 2), but the methanopterin compounds were at least 3 orders of magnitude less potent than folate. These compounds did not inhibit growth of Streptococcus faecalis.

Folate and pteroylpolyglutamate content. Levels of folate activity in representative species grown in folate-free media are presented in Table 3. Values for free folate activity in more than 35 strains of eubacteria have been reported (11, 14, 21, 29, 32); these values range from 0.68 (Bacillus sphaericus) to 261 (Pseudomonas convexa) nmol/g (dry weight), with most being between 1 and 10 nmol/g (dry weight). Very high levels of  $H_4$ PteGlu<sub>3</sub> have been found in the purinolytic Clostridium acid-urici (3,900 nmol/g [dry weight]) (6). The results for Bacillus subtilis and Pseudomonas fluorescens are in good agreement with those obtained with different strains and assay systems (11, 14, 21, 32). The halophilic archaebacteria contained levels of folate activity similar to those of the eubacteria, whereas the methanogens and the thermoacidophile Sulfolobus solfataricus contained considerably less. The value for folate in Methanococcus voltae was similar to that reported previously (21), while that in Methanobacterium thermoautotrophicum Marburg was higher than in strain  $\Delta H$ , possibly because of strain, growth, or assay differences (21).

When extracts were challenged with trimethoprim in the Streptococcus faecalis assay, absorbances decreased by more than 70% (to  $\leq$  0.02) for methanogens and by more than 95% (to  $\leq$ 0.01) for halophiles (data not shown), indicating that little of the folate activity was due to the presence of thymidine. Folate was stable in Sulfolobus solfataricus extracts, as more than 80% of the authentic folic acid that was mixed with cells was recovered. All extracts were tested for possible inhibition of the folate-dependent growth of the assay organisms. One Methanococcus voltae preparation inhibited both strains (decreasing growth by >80%). The organism did not grow well in folate-free medium, and this extract was high in FeS, which may explain the inhibition.

Extracts were tested for pteroylpolyglutamates by treatment with conjugase (Table 3). The results of trimethoprim challenge of conjugase-treated extracts indicated that additional growth was due to folate activity (data not shown). The eubacteria Bacillus subtilis and Pseudomonas fluorescens contained additional folate activity (about 4- and 10-





<sup>a</sup> Results of triplicate determinations repeated 2 to 6 times, expressed as  $\pm$ standard deviation of the mean.

**b** NM, Not measured.

-, Conjugase-treated extract inhibited growth of *Lactobacillus casei* (five repetitions).

fold, respectively) in both assays, indicating the presence of PteGlu<sub>n</sub>, with  $n > 3$ . It has been reported that conjugase treatment causes increases in folate activity of 45-fold for Bacillus subtilis (11) and 15-fold for Streptococcus pneumoniae (29).

Conjugase-treated extracts of Halobacterium halobium and Halobacterium volcanii contained additional growth factor for Streptococcus faecalis, but not that for Lactoba $c$ illus casei, which indicates that PteGlu<sub>3</sub> was present. However, since the folate activity of Halobacterium salinarum extracts did not increase significantly after digestion, the predominant forms present were  $PreGlu<sub>1</sub>$  and  $PreGlu<sub>2</sub>$ . It is not likely that the dimeric pterin sulfohalopterin II (from Halobacterium marismortui) would be active if it was present in these halophiles (22). Sulfolobus solfataricus extracts contained more folate activity in both assays after incubation at pH 4.5 (sham treatment). After conjugase treatment, the extracts contained even slightly more activity in the Streptococcus faecalis system but became inhibitory for the folate-dependent growth of Lactobacillus casei, causing a decrease in the  $A_{540}$  of more than 65%, which may reflect the presence of novel natural products in this archaebacterium. One thermoacidophile contains tetrahydromethanopterin (30), and Sulfolobus solfataricus contains a novel pterin (23), suggesting that such pterins may be the source of folate activity.

Methanogen extracts contained significantly more folate activity after conditions of conjugase treatment (pH 4.5, heating), which is consistent with the results for acidhydrolyzed methanopterin (Table 1). With one exception, methanogens contain very high levels of tetrahydromethanopterin (1,300 to 9,600 nmol/g [dry weight]) (15). Since methanopterin is far less potent than folate, the results suggest that it is the source of folate activity in methanogens.

Halobacterium volcanii contains folates and was the only archaebacterium listed in Table <sup>3</sup> that was susceptible to the antifolates methotrexate and trimethoprim (unpublished data). This is consistent with the report that it is the only trimethoprim-sensitive halobacterium and that it appears to contain a trimethoprim-sensitive dihydrofolate reductase (27).

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