Nucleotide Sequence of the Gene for the Hole-Forming Toxin Aerolysin of Aeromonas hydrophila

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The gene for the hole-forming toxin aerolysin from *Aeromonas hydrophila* was sequenced. Although most of the sequence seems unrelated to that of *Staphylococcus aureus* alpha-toxin, both proteins are very hydrophilic, and they each contain a nearly identical string of 10 amino acids.

Aerolysin is a cytolytic toxin exported by the gramnegative bacterium *Aeromonas hydrophila* (6–9, 14). There are two precursor forms of the toxin. It crosses the inner bacterial membrane as a preprotoxin containing a typical signal sequence, which is removed cotranslationally (8). The resulting protoxin is exported from the bacteria and activated by proteolytic removal of about 25 amino acids from the carboxy terminus (9). Like *Staphylococcus aureus* alpha-toxin (3), the mature protein binds to eucaryotic cells and aggregates to form holes approximately 3 nm in diameter, leading to destruction of the membrane permeability barrier and osmotic lysis (6). We have recently cloned the structural gene for preproaerolysin into *Escherichia coli* (10). Here we present its nucleotide sequence.

The plasmid pPH4 (10) was digested with the restriction enzymes shown in Fig. 1. The resulting fragments were inserted into the polylinker cloning regions of M13 mp18 and mp19 (15) and sequenced (19). The universal 17-base singlestranded primer was used to begin sequencing from the 3' end of the inserts. Sequencing was continued with 18-base oligonucleotide primers prepared by using a Sam One DNA synthesizer (Biosearch Inc., San Rafael, Calif.).

The sequence of the region containing the aerolysin gene is shown in Fig. 2. The longest open reading frame extends from base pairs (bp) 532 to 1989. This would produce a protein with a molecular weight of 53,800, very close to the observed molecular weight of preproaerolysin (8). Several features of the region preceding the gene, including the presence of sequences analogous to Shine-Dalgarno (20) and -10 and -35 regions, have been discussed earlier (10). The translation product of the first 69 bp beginning at 532 is a typical 23-amino-acid signal peptide (17). This is followed by a sequence corresponding to the amino terminus of purified aerolysin (10). In addition, the translated amino acid sequence matches the amino-terminal sequences of two of the cyanogen bromide fragments which were isolated and sequenced after cleavage of purified aerolysin with cyanogen bromide (5). There are no extended open reading frames in the 350-bp 5' untranslated sequence upstream of the gene, which appears to contain a promoter, and there is a palindromic sequence analogous to a Rho-independent termination signal just downstream of the termination codon (18). These observations, which indicate that aerolysin is translated from a monocistronic message, are inconsistent with the results of Chakraborty et al. (1). Those authors have recently cloned and mapped a fragment of A. hydrophila DNA containing a gene for a hemolytic toxin, as well as flanking regions downstream and upstream which they suggest modulate its expression and activity. Our earlier observation that hemolytic activity is not affected in recombinants containing plasmids with Tn5 insertions at or downstream of the BamHI site at bp 2176 (10) also contrasts with their report that hemolysis is changed by transposon insertions in comparable regions of the gene they have cloned. In addition, Chakraborty et al. (1) found that plasmid deletions obtained by digestion with PstI produced nonhemolytic recombinants, yet the sequence reported in Fig. 2 contains no PstI sites (we have confirmed this by restriction analysis of pPH4). Indeed, the restriction map of the aerolysin structural gene in Fig. 2 differs in several other respects from the map reported by Chakraborty et al. (1). This may mean either that the aerolysin genes of the two isolates of A. hydrophila have diverged enormously, or that two different hemolysins have been cloned. A. hydrophila is known to release a weakly hemolytic phospholipase (14), and it may produce another cytolysin (21).

The experimentally determined amino acid composition of purified aerolysin (Table 1) is very similar to the composition calculated assuming that the site of activation is between Arg-442 and Leu-443. This site was located by digesting the mature toxin with carboxypeptidase Y (12) and determining the amino acids released from the C terminus (not shown here). The nucleotide sequence predicts five cyanogen bromide fragments in both aerolysin and proaerolysin, including an unusually large fragment of 27,900 molecular weight, in good agreement with our experimental observations (9).

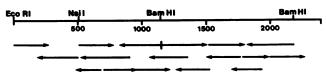


FIG. 1. Strategy for sequencing the aerolysin gene. Arrows represent sequences obtained by using fragments produced by restriction enzyme digestion at the sites indicated or sequences obtained using synthetic oligonucleotides as primers.

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GOG CCC GAG TCA GCT GOG GCC GTT CAC TGG CGA CGG GCA CAG GCC CCT TGC TTG CGG TGG CCG GTC ACT CGC TGC AAT TGC AGG GGT TGG GCA CAA TCA CCT TCG ATE CCE GCA CCC GCT CGC TCA ACG GCC GTC CCC ACC TGC AAC CGG GTC GCC AAC TGG TGC TGA GCC GCG ATG AAA CGG GTC GGG CAA CCG AGA TCC TGA TCC CCA ACC CCG AGG ATG AAC CCG AAT AAG GAT CAT GCA GCC AAA CGC TTA ATA TTT ATT TTG CTA AAT TAG AAA TAT CTT TTT TAT CTA TAT TCC AAA AGA TGA TTA AGT GAC GAA TAA AAT AAT AGA GOG AGT GOT CTG ATA TTA TAT CAA TCA ATA TTG AAT GAA GTT CAA TTT ATG ATT TTG TTA ATA TAT TGC GCA TAT TAA AAT GTG GEC TEG ATC GCA TAT TEA GAT TAA TET GAC TEA TAT TET OFT ACT CAC ATE CCA CCC GCT GAT ATA TAA GET TEG TEA ATE CAT ETC AAT ETT CAA TAT ATT GEG **↓**¹ TTG TTT TCA TTG GGC CAA GGG GTC TGT GGC GAC AAG TAT CGC CCC GTC AAT CGA GAA GAA GAC GCC CAA AGC GTT AAA AGC AAT ATT GTC GGC ATG ATG GGG CAA TGG Leu Phe Ser Leu Gly Gln Gly Val Cys Gly Asp Lys Tyr Arg Pro Val Asn Arg Glu Glu Ala Gln Ser Val Lys Ser Asn Ile Val Gly Met Met Gly Gln Trp CAA ATA AGC GGG CTG GCC AAC GGC TGG GTC ATT ATG GGG CCG GGT TAT AAC GGT GAA ATA AAA CCA GGG ACA GCG TCC AAT ACC TGG TGT TAT CCG ACC AAT CCT Gin Ile Ser Giy Leu Ala Asn Giy Trp Val Ile Met Gly Pro Gly Tyr Asn Gly Glu Ile Lys Pro Giy Thr Ala Ser Asn Thr Trp Cys Tyr Pro Thr Asn Pro 50 70 80 GTT ACC GGT GAA ATA CCG ACA CTG TCT GCC CTG GAT ATT CCA GAT GGT GAC GAA GTC GAT GTG CAG TGG CGA CTG GTA CAT GAC AGT GCG AAT TTC ATC AAA CCA Val Thr Gly Glu Ile Pro Thr Leu Ser Ala Leu Asp Ile Pro Asp Gly Asp Glu Val Asp Val Gln Trp Arg Leu Val His Asp Ser Ala Asn Phe Ile Lys Pro ACC AGC TAT CTC GCC CAT TAC CTC GGT TAT GCC TCG GTC GGC GGC AAT CAC AGC CAA TAT GTC GGC GAA GAC ATG GAT GTC ACC CGT GAT GGC CAC GGC TGG GTG Thr Ser Tyr Leu Ala His Tyr Leu Gly Tyr Ala Trp Val Gly Gly Asn His Ser Gln Tyr Val Gly Glu Asp Met Asp Val Thr Arg Asp Gly Asp Gly Trp Val 120 140 ATC COT GCC AAC AAT GAC GCC GCC TGT GAC GCC TAT CCC TGT GCT GAC AAG ACG GCC ATC AAG GTC AGC AAC TTC GCC TAT AAC CTG GAT CCC GAC AGC TTC AAG TAC GAC ACA GCC ACC AAC TGG TCC AAG ACC AAC ACC TAT GGC CTG AGC GAG AAG GTG ACC ACC AAG AAC AAG TTC AAG TGG CCA CTG GTG GGG GAA ACC CAA CTC Tyr Asp Thr Ala Thr Asn Trp Ser Lys Thr Asn Thr Tyr Gly Leu Ser Glu Lys Val Thr Thr Lys Asn Lys Phe Lys Trp Pro Leu Val Gly Glu Thr Gin Leu 250 TCC ATC GAG ATT CGT GCC AAT CAG TCC TGG GCG TCC CAG AAC GGG GGC TCG ACC ACC ACC TCC CTG TCT CAG TCC GTG CGA CCG ACT GTG CGC GCC CGC TCC AAG Ser Ile Glu Ile Arg Ala Asn Gln Ser Trp Ala Ser Gln Asn Gly Gly Ser Thr Thr Thr Ser Leu Ser Gln Ser Val Arg Pro Thr Val Pro Ala Arg Ser Lya 250 ATC CCG GTG AAG ATA GAG CTC TAC AAG GCC GAC ATC TCC TAT CCC TAT GAG TTC AAG GCC GAT GTC AGC TAT GAC CTG AGC GGC GCC TTC CTG CGC TGG GGC Ile Pro Val Lys Ile Glu Leu Tyr Lys Ala Asp Ile Ser Tyr Pro Tyr Glu Phe Lys Ala Asp Val Ser Tyr Asp Leu Thr Leu Ser Gly Phe Leu Arg Trp Gly 310 AAG CGT TAC ATC COG GGT GAA GTG AAG TGG TGG GAC TGG AAC TGG ACC ATA CAG CAG AAC GGT CTG TCT ACC ATG CAG AAC AAC CTG GCC AGA GTG CTG CGC CGG Lys Arg Tyr Ile Pro Gly Glu Val Lys Trp Trp Asp Trp Asn Trp Thr Ile Gln Gln Asn Gly Leu Ser Thr Met Gln Asn Asn Leu Ala Arg Val Leu Arg Pro 330 GTG CGG GCG GGG ATC ACC GGT GAT TTC AGT GCC GAG AGC CAG TTT GCC GGC AAC ATA GAG ATC GGT GCT CCC GTG CCG GTC GCG GCT GAC AGC AAG GTG GGT GGT GGT Val Arg Ala Gly Ile Thr Gly Asp Phe Ser Ala Glu Ser Gln Phe Ala Gly Asn Ile Glu Ile Gly Ala Pro Val Pro Leu Ala Ala Asp Ser Lys Val Arg Arg 430 GCT OGC AGT GTG GAC GGC GGT GGT CAA GGC CTG AGG CTG GAG ATC CGC GTC GAT CGC GAA GAG CTC TCC GGG CTT GGC TTC AAC AAG TCA GCC TCA GCC TCA GCG TGA CCC Ala Arg Ser Val Asp Gly Ala Gly Gln Gly Leu Arg Leu Glu Ile Pro Leu Asp Arg Glu Glu Leu Ser Gly Leu Gly Phe Asn Lys Ser Ala Ser Ala CTG CTG CCA ATC AAT AAC GGC AGC GCG TTG TAG TGA TGG AAC GGG GCC TCT GTG GCC CGG TTT TTG CTT GCA CTG GTC GGG CTT GTT AAA GGC TTG TGC TTT CCA 2175 BamHI TTT CCC CAC TTA TAC TGG OGC CAT CTT GTC GGA GTG CCA ACC GTC GAA OGA CGC GAG GCC GAG ACC GTT AAT TCG GGA TCC GTG GAA OCT GAT CAG GCT AGC ACC TEC GAA GGE AAA CAA GGE TAA CTT GCE GET TEC CEC GCE GGE GGA GGE ACA AGC CTC TCC CEC TCA TCA AGA GCA ATT CCT CEA TCA AGE GCE CAC AAG

AGG GAG TCT GTC CCG TCC GGT CTG CCC AGG AGG GGC

FIG. 2. DNA sequence of the aerolysin gene. Sequences which could be part of a promoter region and a Rho-independent terminator are underlined, as is the amino acid sequence of the signal peptide. The experimentally determined N-terminal amino acid sequences of two cyanogen bromide fragments are also underlined. Sites of proteolytic processing of preproaerolysin and proaerolysin are marked with arrows.

 TABLE 1. Comparison of the amino acid composition of aerolysin derived from the DNA sequence with that obtained by analysis of the purified protein

Amino acid(s)	Composition (mol/mol of protein) determined by:	
	Derivation ^a	Protein analysis ^b
Asp + Asn	59	63
Thr	29	29
Ser	34	33
Glu + Gln	35	39
Pro	23	23
Gly	44	41
Ala	28	26
Cys	4	ND
Val	34	33
Met	5	5
Ile	21	21
Leu	25	24
Tyr	21	21
Phe	10	10
His	6	6
Lys	22	23
Arg	24	20
Trp	18	ND

^a These calculations are based on the assumption that proaerolysin is processed at Arg-442.

^b Average of two determinations. ND, Not determined.

223 232 Aerolysin Thr.Ala.Thr.Asn.Trp.Ser.Lys.Thr.Asn.Thr. Alpha toxin Thr.Ser.Thr.Asn.Trp.Lys.Gly.Thr.Asn.Thr. 312 312

FIG. 3. Comparison of regions of potential sequence homology between aerolysin and alpha-toxin.

Analysis of the predicted amino acid sequence (2, 11)indicates that the protein is remarkably hydrophilic [average (H)19 hydropathy of -0.57] with no obvious hydrophobic stretches long enough to span a membrane. Because aerolysin forms holes in biological membranes, this is at first surprising; however, the pore-forming outer membrane porins also lack hydrophobic sequences (16). Like these proteins, aerolysin may aggregate to accomplish hole formation. It should also be noted that *S. aureus* alpha-toxin is even more hydrophilic than aerolysin (4).

A search of the National Biomedical Research Foundation protein sequence library (13) did not reveal any sequences homologous to aerolysin, but a direct comparison of aerolysin and alpha-toxin (4) uncovered similar strings of 10 amino acids in each protein. Seven of the 10 are identical, including a Trp in the middle of each sequence (Fig. 3). Another position is occupied by Ser in one protein and homologous Ala in the other. Because the two toxins are functionally so similar, it is tempting to speculate that this small area of similarity is significant. This research was supported by grants from the National Sciences and Engineering Research Council of Canada and the British Columbia Health Care Research Foundation.

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