

## **A bacteriocin produced by certain M-type 49 *Streptococcus pyogenes* strains when incubated anaerobically**

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### SUMMARY

Bacteriocin production (P)-typing of 75 M-type 49 group-A streptococci obtained from a variety of epidemiological incidents in different countries gave no evidence of production under the usual aerobic test conditions. However, with anaerobic incubation, 28% of the strains gave a pattern of inhibitory activity against the indicator strains which was indistinguishable from that previously attributed to the bacteriocin, streptococcin A-FF22 (SA-FF22). Isolation and partial purification of the M type 49 bacteriocin (SA-M49) by freeze-thaw elution from anaerobically grown lawn cultures, followed by ammonium sulphate precipitation and Sephadex chromatography, showed the activity to be associated with a heat-stable proteinaceous molecule of molecular weight approximately 8000 - properties similar to those of SA-FF22. SA-FF22 and SA-M49 were found to have identical inhibitory spectra including immunity of the producer strains to the inhibitory activity of both the homologous and heterologous bacteriocin preparations. SA-M49 production occurred in some strains of phage subtypes II, III and provisional VI and, since it was a consistent property for all isolates from single outbreaks of infection, it provides a means of discriminating between strains of each of these three phage subtypes. There was no evidence of any increased incidence of SA-M49 production in M-type 49 strains associated with nephritic sequelae.

### INTRODUCTION

Application of an inhibitor 'fingerprinting' scheme based upon a deferred antagonism test on blood agar has shown that many group-A streptococcus (*Streptococcus pyogenes*) strains produce antibacterial agents (Tagg & Bannister, 1979). By use of this scheme, all of 12 M-type 4 streptococci tested produced an identical range of activity against nine standard indicator strains. In code form this inhibitory pattern is referred to as production (P)-type 655, a type which appears to be uniquely associated with M-type 4 strains. Similarly, only the six M-type 57 strains tested were found to produce P-type 614 inhibitory activity (Tagg & Bannister, 1979). Proteinaceous antibiotics isolated from prototype

strains representative of these two serotypes have subsequently been characterized as bacteriocins (Johnson, Tagg & Wannamaker, 1979; Simpson & Tagg, 1983).

The first reported group-A streptococcus bacteriocin was streptococcin A-FF22 (SA-FF22) (Tagg, Read & McGiven, 1973), produced by group A streptococcus strain FF22 (M type 52) and found in many respects similar to nisin, an antibiotic produced by *Streptococcus lactis* (Tagg & Wannamaker, 1978). As part of the inhibitor 'fingerprinting' scheme (Tagg & Bannister, 1979) an assessment is made of the sensitivity (S)-type of test strains, using a set of nine standard inhibitory streptococci. Strain FF22 is included as one of these standard inhibitor producers and has been found widely active against other group A streptococci, all of 54 prototype strains representing different M serotypes being sensitive (Tagg & Bannister, 1979). Other relevant characteristics include the immunity of strain FF22 to its own bacteriocin, an essential feature of any bacteriocinogenic strain (Tagg, Dajani & Wannamaker, 1976) and the enhanced production of SA-FF22 under conditions of anaerobic incubation (Tagg & Bannister, 1979). M type 49 streptococci have frequently been implicated in the development of pyoderma, sometimes complicated by ensuing acute glomerulonephritis (Maxted, Fraser & Parker, 1967). Discrimination between different M49 strains has been aided by the use of a phage-typing scheme, five different phage subtypes initially being reported amongst 72 strains representing a variety of epidemiological incidents from ten worldwide sources (Skjold & Wannamaker, 1976). In our preliminary studies (Tagg & Skjold, unpublished) using the standard inhibitor 'fingerprinting' scheme there was no evidence of bacteriocin-like inhibitor production by any of the available M type 49 streptococci; however, some strains were distinctive in that they were insensitive to SA-FF22 when S-typed. This apparent immunity to SA-FF22 action was the first indication of the possible production by these strains of an antibiotic substance which might be closely similar to SA-FF22. In the present study we have examined a large collection of M49 streptococci for inhibitor production and for its relationship to the phage subtype and the nephritis association of the strains.

#### MATERIALS AND METHODS

The nine standard indicator stains (II-I9) and the basic P-typing procedure used in this study have been described previously (Tagg & Bannister, 1979). For purposes of the present study calcium carbonate (0.5%, w/v) was added to the typing medium to minimize inhibition due to the accumulation of acidic metabolites (Tagg & Martin, 1984). Most of the M-type 49 group-A streptococci tested for inhibitor production were selected from those used in previous phage-typing studies (Skjold & Wannamaker, 1976; Skjold *et al.* 1983). Four strains provided by courtesy of Dr W. R. Maxted (Colindale) had been isolated in South Oxfordshire (Mayon-White & Perks, 1982). Anaerobic incubation was at 32°C for 18 h in an atmosphere of 85% nitrogen, 10% hydrogen and 5% carbon dioxide (Forma Scientific anaerobic glovebox). For production of partially purified SA-M49 and SA-FF22 the methods for inoculation of lawn cultures, freeze-thaw extraction of the culture liquor, precipitation with 80% saturated ammonium sulphate and chromatography on a calibrated column of Sephadex G100 were as previously described (Tagg, Read & McGiven, 1973*b*; Tagg *et al.* 1973*a*). Assay of preparations

Table 1. *Sensitivity to SA-FF22 and SA-FF22-like inhibitor production by M type 49 streptococci*

Strain designation	Strain characteristics	Sensitivity to SA-FF22*	Inhibitor production* (P-type)	
			Aerobically	Anaerobically
FF22	M52, SA-FF22 producer	—	476	476
EB1	SA-FF22 negative derivative of FF22	+	000	000
GT-9653	M49, phage subtype II	—	000	476
GT-9538	M49, phage subtype III	—	000	476
81-086	M49, phage subtype VI	—	000	476
GT-8760	M49, phage subtype I	+	000	000
GT-6481	M49, phage subtype V	+	000	000
GT-7903	M49, phage subtype IV	+	000	000

\* Tested by deferred antagonism with producer growth at 32°C on blood agar plus 0.5% (w/v) calcium carbonate.

for inhibitory activity was by a well-diffusion method on Columbia blood agar base medium (Tagg *et al.*, 1973*a*). Tests for protease and heat susceptibility were by conventional techniques (Tagg *et al.* 1973*a*). Phage typing was by the method of Skjold & Wannamaker (1976). The types defined were the originally described types I–V (Skjold & Wannamaker, 1976), provisional VI and provisional VII, the latter representing sensitivity to phage 5 (Skjold *et al.* 1983).

## RESULTS

In view of our preliminary observation that certain *M* type 49 streptococci were insensitive to SA-FF22, several of these strains were re-examined for inhibitor production under anaerobic conditions (Table 1). With anaerobic incubation all of the SA-FF22-resistant strains were found to be P-type 476, a P-type identical to that given by strain FF22. Neither the SA-FF22-sensitive M49 strains nor strain EB1, a SA-FF22-negative derivative of strain FF22, produced detectable inhibitors under these incubation conditions.

Clusters of epidemiologically related M49 strains were tested for P-type 476 inhibitor production both aerobically and anaerobically (Table 2). Approximately 28% of the strains, including representatives from three of the seven phage subtype categories, had inhibitor-positive representatives. All of the strains within a defined infection cluster were found to be either inhibitor-positive or inhibitor-negative. Four of the seven clusters of inhibitor-positive strains and 12 of the 16 inhibitor-negative clusters were considered to have had a nephritic association. Inhibitor-positive strains of phage subtype III were present in nephritis-associated outbreaks in Trinidad in 1965–6 and in 1975–6. Also associated with the latter outbreak were two inhibitor-positive isolates of the provisional phage subtype VI.

Table 2. *P*-type 476 inhibitor production by clusters\* of *M* type 49 strains of various phage subtypes and epidemiological sources

Phage subtype	Source†	Nephritis Assoc-iation	Inhibitor production		No. of isolates tested
			Aerobically	Anaerobically	
I	Red Lake 1966	+	—	—	4
	Alabama 1964-5	+	—	—	4
II	The Netherlands 1960-2	+	—	—	4
	Great Britain, 1966?	—	—	+	4
	Chile (year unknown)	+	—	+	4
	Oxford‡, 1980-1	—	—	—	4
	Kent‡ 1980	—	—	+	2
III	Alabama, 1964-5	+	—	—	2
	Trinidad, 1965-6	+	—	+	4
	Trinidad‡, 1975-6	+	—	+	4
IV	Minneapolis/ Red Lake, 1953	+	—	—	3
	Alaska‡, 1975-9	+	—	—	1
V	Alabama, 1964-5	+	—	—	4
	Alaska, ‡ 1975-9	+	—	—	4
VI (prov)‡	South Oxford- shire§ 1977	—	—	—	4
	Alabama 1964-5	+	—	—	2
	Czechoslovakia, 1961-2	+	—	—	5
	Lincolnshire, ‡ 1980	—	—	—	6
	Cambridgeshire, ‡ 1980	—	—	—	4
	Trinidad, ‡ 1975-6	+	—	+	2
	Kent, ‡ 1980	—	—	+	1
	VII (prov)	Alaska, ‡ 1975-9	+	—	—
Alabama, 1964-5	+	—	—	1	

\* Each cluster of isolates was from a similar geographical source and date and had the same phage subtype.

† The strains were selected from those previously described (Skjold & Wannamaker, 1976) unless otherwise designated.

‡ Skjold *et al.* (1983).

§ Mayon-White & Perks (1982).

Although the observed cross-resistance in deferred antagonism tests between strain FF22 and the M49 inhibitor producers suggested that the M49 inhibitor was closely similar to SA-FF22, isolation and characterization of the inhibitor was required to substantiate this. Strain GT-9653 was selected as the prototype producer of SA-M49 activity for use in these studies.

Inhibitor production in liquid media was unreliable, only trace amounts being detected in pre-reduced, anaerobically incubated cultures in tryptic soy broth, Todd Hewitt broth or brain-heart infusion. Addition of 5% (v/v) blood and incubation at various temperatures (25, 32, 37°C) was tried, but found unhelpful in promoting consistent production. Low yields (titre 2) of SA-M49 activity were obtained by freeze-thaw extraction of lawn cultures grown anaerobically (24 h/32°C) on the P-typing medium. Greater recoveries (titre 4-8) were obtained with use of a sheep blood rather than human blood supplement to the medium.

Table 3. *Inhibitory activity of partially-purified preparations of SA-M49 and SA-FF22*

Test strain	Inhibitory activity* of partially purified	
	SA-M49	SA-FF22
Group A streptococcus strain GT 9653	—	—
Group A streptococcus strain EB1	++	++
Group A streptococcus strain GT8760	++	++
Standard indicator strains		
<i>Micrococcus luteus</i> T18(I1)	+++	+++
Group A streptococcus FF22(I2)	—	—
Group F streptococcus T29(I3)	—	—
Group E streptococcus T6(I4)	+	+
Group A streptococcus 71-679(I5)	++	++
Group N streptococcus T21(I6)	+++	+++
Group A streptococcus 71-698(I7)	+++	+++
Group A streptococcus W1(I8)	++	++
Group C streptococcus T148(I9)	—	—

\* Tested by the well-diffusion method. Zone diameter: +++ (> 14 mm), ++ (10–14 mm), + (6–10 mm), — (no inhibition), well diameter 6 mm.

Further studies showed that it was the plasma component of the blood that was important in promoting inhibitor production and heating the plasma at 56°C for 30 min did not interfere with this effect. Subsequently, crude preparations of SA-M49 were prepared by freeze-thaw elution from batches of lawn cultures incubated 24 h/37°C anaerobically on Columbia blood agar base plus 5% (v/v) sheep plasma. Partial purification of SA-M49 by precipitation of the inhibitor with 80% saturated ammonium sulphate followed by chromatography on a calibrated Sephadex G100 column allowed an estimation of the molecular weight of approximately 8000. The spectrum of activity of the partially purified SA-M49 against the standard indicators was the same as found in P-typing tests and was identical to that of a similarly purified preparation of SA-FF22 (Table 3). Both SA-M49 and SA-FF22 were stable to heating at 80°C for 30 min and loss of activity followed treatment for 60 min with 1 mg/ml trypsin.

#### DISCUSSION

The production by certain M-type 49 isolates of SA-M49, a bacteriocin apparently identical to SA-FF22, provides a useful additional strain marker within this widely occurring nephritogenic serotype of group A streptococci. SA-M49 production did not appear to correlate with any increased association with nephritis. However, it was found to be a consistent property within clusters of epidemiologically-related isolates and since it seems to occur independently of the phage subtype of the strain, it offers a means of further subdividing M49 strains when used in conjunction with phage typing. Previously (Skjold *et al.* 1983) it had been suggested on the basis of phage typing data that the same strain may have persisted over a 9-year period to be responsible for two outbreaks of nephritis in Trinidad. Isolates from both of these outbreaks were found to be SA-M49 positive,

affording additional evidence for the close-relatedness of the strains involved in these two episodes.

None of the M49 strains of phage subtypes IV, V or PR VII obtained in 1975–9 in association with a high incidence of nephritis in an Alaskan Eskimo population were SA-M49 positive. Previously it has been reported (Skjold *et al.* 1983) that in association with this same outbreak a number of group A streptococci were isolated having T14 antigen, as did the M type 49 strains, but differing from the latter in being serum opacity factor-negative and non-typable with available phages. These strains, which appear to represent a new M type, have also been tested for inhibitor production (Tagg, unpublished) and all are P-type 226, an inhibitory pattern quite unlike that detected in any of the M type 49 strains.

An unusual feature of SA-M49 production was its apparent dependence upon anaerobic incubation of the producer strain. Recovery from liquid cultures was poor, extraction from lawn cultures on sheep-plasma-containing media being required to provide sufficient activity for purification and characterization studies. Partially purified SA-M49 was closely similar to a SA-FF22 in its molecular weight, heat-stability, sensitivity to trypsin and activity spectrum.

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