

# Mitochondrial Fusion in Yeast Requires the Transmembrane GTPase Fzo1p

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**Abstract.** Membrane fusion is required to establish the morphology and cellular distribution of the mitochondrial compartment. In *Drosophila*, mutations in the *fuzzy onions* (*fzo*) GTPase block a developmentally regulated mitochondrial fusion event during spermatogenesis. Here we report that the yeast orthologue of *fuzzy onions*, Fzo1p, plays a direct and conserved role in mitochondrial fusion. A conditional *fzo1* mutation causes the mitochondrial reticulum to fragment and blocks mitochondrial fusion during yeast mating. Fzo1p is a mitochondrial integral membrane protein with its GTPase domain exposed to the cytoplasm. Point mutations that alter conserved residues in the GTPase do-

main do not affect Fzo1p localization but disrupt mitochondrial fusion. Suborganellar fractionation suggests that Fzo1p spans the outer and is tightly associated with the inner mitochondrial membrane. This topology may be required to coordinate the behavior of the two mitochondrial membranes during the fusion reaction. We propose that the *fuzzy onions* family of transmembrane GTPases act as molecular switches to regulate a key step in mitochondrial membrane docking and/or fusion.

**Key words:** GTPase • membrane fusion • mitochondria • mtDNA • organelle morphology

THE homotypic, or self-fusion, of mitochondrial membranes plays an important role in controlling the copy number and cellular distribution of these essential organelles (Bereiter-Hahn and Voth, 1994). In budding yeast and some mammalian cells, frequent mitochondrial fusion events are balanced by mitochondrial fission, maintaining the compartment as a highly branched, tubular network (Hoffman and Avers, 1973; Stevens, 1981; Koning et al., 1993; Nunnari et al., 1997; Hermann, 1998). Mitochondrial fusion also functions to remodel mitochondrial morphology during differentiation. Individual mitochondria in rat skeletal muscle cells undergo postembryonic fusion to generate a branched mitochondrial reticulum (Bakeeva et al., 1981). During spermatogenesis in insects, fusion generates giant mitochondria that associate with the axoneme of the sperm flagella (Lindsley and Tokuyasu, 1980; Fuller, 1993). The generation of such large, interconnected mitochondrial compartments is thought to facilitate the distribution of energy and metab-

olites as well as chemical and electrical signals throughout these cells (Bakeeva et al., 1978; Ichas et al., 1997). The genetic and molecular mechanisms that control mitochondrial fusion have not been defined.

Molecules that regulate the heterotypic and homotypic fusion of membranes in the secretory and endocytic pathways have been studied in some detail. Both types of fusion require the cytosolic factors NSF and SNAPs (or homologues) as well as compartment-specific integral membrane proteins termed v- and t-SNAREs (Denesvre and Malhotra, 1996; Pfeffer, 1996; Rothman, 1996; Hay and Scheller, 1997; Edwardson, 1998; Götte and Fischer von Mollard, 1998; Patel et al., 1998; Rabouille et al., 1998). During heterotypic fusion, v/t-SNARE pairing promotes the stable association of the vesicle with the target membrane and may be sufficient to catalyze bilayer mixing (Weber et al., 1998). In contrast, the homotypic fusion of ER and Golgi membranes depends on self-interactions among resident t-SNAREs (Patel et al., 1998; Rabouille et al., 1998). Yeast vacuole fusion requires a v/t-SNARE pairing for optimal fusion but can occur (with lower efficiency) between vacuoles containing only the t-SNARE partner (Nichols et al., 1997). Thus, in some cases, the recognition of “like” membranes in homotypic fusion reactions may be mediated by homomeric interactions be-

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tween compartment-specific t-SNAREs. NSF and SNAPs appear to act generally during fusion to disrupt unproductive SNARE associations within a membrane before docking (Mayer et al., 1996; Otto et al., 1997) and may also regulate changes in SNARE conformation after docking but before membrane fusion (Söllner et al., 1993).

Although an isoform of the VAMP-1 SNARE has been reported to localize to mitochondria in transfected epithelial cells (Isenmann et al., 1998), molecules in the NSF, SNAP, and SNARE families have not been implicated in mitochondrial fusion. This is not entirely unexpected since mitochondrial fusion, unlike most other membrane fusion reactions in the cell, requires the sequential mixing of two distinct lipid bilayers. Moreover, the mitochondrial compartment is evolutionarily distinct from compartments of the secretory and endocytic pathways. Thus, the molecular machinery that mediates mitochondrial fusion is likely to be unique, even though the general mechanism of membrane fusion may be conserved. A novel gene required for mitochondrial fusion, *fuzzy onions* (*fzo*),<sup>1</sup> was recently identified through the analysis of *Drosophila* mutants defective in spermatogenesis (Hales and Fuller, 1997). During spermatid differentiation in flies, individual mitochondria coalesce, fuse into two giant organelles, and coil tightly around one another forming the Nebenkern, a spherical structure that resembles an onion in cross section (Lindsley and Tokuyasu, 1980; Fuller, 1993). Mutations in the *fzo* gene disrupt mitochondrial fusion and result in disorganized aggregates of individual mitochondria that do not associate properly with the elongating axoneme (Hales and Fuller, 1997). *fzo* encodes a predicted transmembrane GTPase that is detected on the mitochondrial compartment just as fusion begins and disappears soon after fusion is complete. Together, the *fzo* mutant phenotype and the expression pattern of the Fzo protein suggest that this molecule either regulates, or is a direct mediator of, mitochondrial fusion.

The discovery of *fzo* homologues in yeasts, nematodes, and mammals defined a new family of multiple domain, high molecular weight GTPases, and raised the possibility that these molecules act generally to control mitochondrial fusion events in different organisms and cell types (Hales and Fuller, 1997). Fzo family members contain an amino-terminal GTPase domain, two adjacent carboxy-terminal transmembrane domains, and multiple heptad repeats (see Fig. 1 A) (Hales and Fuller, 1997). Here we show that the *Saccharomyces cerevisiae* orthologue of *Drosophila* Fzo, Fzo1p, is a mitochondrial integral membrane protein required to maintain a tubular mitochondrial reticulum during mitotic growth and demonstrate a direct role for this protein in mitochondrial fusion during yeast mating. Mutations in two conserved *FZO1* GTPase motifs disrupt mitochondrial fusion but do not affect Fzo1p localization. Subcellular fractionation and protease protection experiments reveal that the amino terminus of the protein (containing the essential GTPase domain and

multiple heptad repeats) extends into the cytoplasm where it could participate in organelle docking/fusion and interact with molecules that regulate GTP binding or hydrolysis. In addition, the distribution of Fzo1p in submitochondrial fractionation studies suggests that the carboxy-terminus of this protein may interact with both mitochondrial membranes. This topology is similar to that of SNARE molecules and viral fusion proteins and suggests that Fzo1p could play a direct role in the docking and fusion of mitochondrial compartments.

## Materials and Methods

### Strains and Genetic Techniques

All yeast strains (Table I) are isogenic to FY10 (Winston et al., 1995). All media including YPDextrose, YPGlycerol, SDextrose, SGalactose, and SRaffinose were prepared as described by Sherman et al. (1986). Strains were grown at 30°C unless otherwise noted. Genetic and molecular cloning manipulations followed standard techniques (Maniatis et al., 1982; Sherman et al., 1986; Baudin et al., 1993). The *Escherichia coli* host strain JM109 (Promega Corp., Madison, WI) was used for all bacterial manipulation of plasmids.

### Plasmid Construction

For pRS416-*FZO1*, a genomic fragment containing the *FZO1* coding region plus 500 bp of 5' and 3' flanking sequence was PCR amplified from JSY999 using primers containing engineered EcoRI sites: P297 (5'-GGGG-GAATTCAGGTGACAGAATGTCTGGGTTGAAAG-3') and P298 (5'-GGGGGAATTCCTTGCTCCTTGTGTCTTTAAATGGAG-3'), and cloned into the unique EcoRI site in pRS416 (verified by sequencing; Stratagene, La Jolla, CA). An EcoRI fragment from pRS416-*FZO1* was subcloned into pRS414 (Stratagene) to generate pRS414-*FZO1*. The pRS414-*FZO1* and pRS416-*FZO1* plasmids complemented both the mitochondrial morphology defects and the loss of mtDNA in *fzo1Δ*. To generate a *GAL1* regulated form of *FZO1*, a PCR fragment containing a 3XMYC tag at the amino terminus of *FZO1* (N-3XMYC-*FZO1*) was amplified from JSY2028 (see below) using primers with engineered Sall, P379 (5'-GGGGTTCGACTATCTAATCGATGTCTAAATTTATTCTTC-3'), and XbaI, P380 (5'-GGGGTCTAGATTAACGATGTCTAG-GGAACAAAAGCTGGAG-3'), sites. The PCR fragment was cloned into pRS415-*GAL1* (Mumberg et al., 1994) (American Type Culture Collection, Rockville, MD) to generate p*GAL1*-N-3XMYC-*FZO1* (pRS415-*GAL1*-N-3XMYC-*FZO1*, verified by sequencing). Growth in SRaffinose provided sufficient expression of N-3XMYC-Fzo1p from the *GAL1* promoter to complement the mitochondrial morphology and growth defects of *fzo1Δ* cells. To generate mutations in the conserved *FZO1* GTPase domain, a 3.5-kb EcoRI fragment from pRS416-*FZO1* was cloned into the EcoRI site of pALTER-1 (Promega Corp.) to create pALTER-1-*FZO1*. Site-directed mutagenesis (Altered Sites II; Promega Corp.) was performed with the following mutagenic oligonucleotides: K200A (5'-GAT-GTAAATACTGGCGCCTCAGCTCTTTGCAAC-3', introduces an EheI site); S201N (5'-CAGGTGATGTAATACTGGTAAATATGCATTA-TGCAACTCTCTATTAAAGCAGCG-3', introduces an NsiI site); T221A (5'-GGATCAGCTACCATGCGCAAATGTATTTTCCGAA-3', introduces an Fsp I site); K371A (5'-GTTTTTTGTGTGAA-GCTTTTGACAAAATCAGGG-3', introduces a HindIII site). The mutants were identified by restriction digest and confirmed by sequence analysis. The mutagenized *fzo1* genes were subcloned into the EcoRI sites of pRS414 and pRS424.

### Generation and Characterization of *fzo1Δ::HIS3* Cells

The *fzo1Δ::HIS3* mutation was generated by transforming the diploid strain JSY1373 with a PCR fragment containing 50 bp of *FZO1* flanking sequence interrupted by the *HIS3* gene (Baudin et al., 1993). The *fzo1Δ::HIS3* disruption precisely removed the entire *FZO1* coding sequence and was verified by PCR analysis. In 39 tetrads from the sporulated heterozygous diploid (JSY1808), the *HIS3* marker segregated 2:2 with a slow growth defect on YPDextrose and an inability to grow on YPGlycerol. Mitochondrial morphology was visualized in four tetrads using a primary

1. Abbreviations used in this paper: 3-PGK, 3-phosphoglycerate kinase; 5-FOA, 5-fluoro-orotic acid; CoxIV, cytochrome oxidase subunit IV; DAPI, 4',6-diamidino-2-phenylindole; DiOC6, 3,3' dihexyloxocarbocyanine; *fzo*, *fuzzy onions* gene; GFP, green fluorescent protein; mtDNA, mitochondrial DNA.

Table I. Yeast Strains Used in This Study

Strain	Genotype + (Plasmid)
JSY999	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 rho<sup>+</sup></i>
JSY1373	<i>MAT<math>\alpha</math>/<math>\alpha</math> leu2<math>\Delta</math>1/leu2<math>\Delta</math>1 his3<math>\Delta</math>200/his3<math>\Delta</math>200 ura3-52/ura3-52 rho<sup>+</sup></i>
JSY1808	<i>MAT<math>\alpha</math>/<math>\alpha</math> leu2<math>\Delta</math>1/leu2<math>\Delta</math>1 his3<math>\Delta</math>200/his3<math>\Delta</math>200 ura3-52/ura3-52 <i>fzo1</i><math>\Delta</math>::<i>HIS3/FZO1</i> rho<sup>+</sup></i>
JSY1809	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>0</sup></i>
JSY1810	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>0</sup></i>
JSY1811	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>FZO1</i> rho<sup>+</sup></i>
JSY1812	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>FZO1</i> rho<sup>+</sup></i>
JSY2028	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>FZO1</i>::N-3XMYC rho<sup>+</sup></i>
JSY2034	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>FZO1</i>::N-3XMYC rho<sup>+</sup></i>
JSY2038	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>+</sup> + (pRS416-<i>FZO1</i>)</i>
JSY2270	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>FZO1</i> rho<sup>+</sup> + (pGAL1-N-3XMYC-<i>FZO1</i>)</i>
JSY2273	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>+</sup> + (pGAL1-N-3XMYC-<i>FZO1</i>)</i>
JSY2287	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>+</sup> + (pRS416-<i>FZO1</i>)</i>
JSY2288	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>FZO1</i> rho<sup>+</sup> + (pRS416-<i>FZO1</i>)</i>
JSY2354	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> + (pRS414)</i>
JSY2355	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> + (pRS414-<i>fzo1</i>[T221A])</i>
JSY2356	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> + (pRS414-<i>fzo1</i>[K371A])</i>
JSY2357	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> + (pRS414-<i>fzo1</i>[K200A])</i>
JSY2358	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> + (pRS414-<i>fzo1</i>[S201N])</i>
JSY2392	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> + (pRS414-<i>FZO1</i>)</i>
JSY2394	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>+</sup> + (pRS414-N-9XMYC-<i>FZO1</i>)</i>
JSY2555	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 rho<sup>0</sup></i>
JSY2579	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>0</sup> + (pRS416-<i>FZO1</i>)</i>
JSY2634	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>+</sup> + (pGAL1-N-3XMYC-<i>FZO1</i>) + (pRS416-ADH-COXIVpre-GFP)</i>
JSY2793	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>+</sup> + (pRS414-<i>FZO1</i>) + (pRS416-ADH-COXIVpre-GFP)</i>
JSY2802	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>lys2</i><math>\Delta</math>-202 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>+</sup> + (pRS414-<i>fzo1-1</i>) + (pRS416-ADH-COXIVpre-GFP)</i>
JSY2804	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>+</sup> + (pRS414-<i>fzo1-1</i>) + (pRS416-ADH-COXIVpre-GFP)</i>
JSY2926	<i>MAT<math>\alpha</math> leu2<math>\Delta</math>1 his3<math>\Delta</math>200 ura3-52 <i>trp1</i><math>\Delta</math>63 <i>fzo1</i><math>\Delta</math>::<i>HIS3</i> rho<sup>+</sup> + (pRS414-<i>FZO1</i>)</i>

anti-porin antiserum (1:200 dilution; Molecular Probes, Eugene OR) and a secondary goat anti-mouse FITC antibody (1:100 dilution) (Jackson ImmunoResearch Laboratories, West Grove, PA) (Pringle et al., 1991). DAPI (4',6-diamidino-2-phenylindole; 25 ng/ml) was included in the mounting medium to visualize mtDNA. The loss of mtDNA was confirmed by mating JSY1810 (*fzo1* $\Delta$ ) to a rho<sup>0</sup> tester strain JSY2555 (*FZO1*, rho<sup>0</sup>). pRS416-*FZO1* was transformed into a *fzo1* $\Delta$  strain (JSY1810) to generate JSY2579 (wild-type mitochondrial morphology, no detectable mtDNA). To generate the rho<sup>0</sup> strain JSY2555 (wild-type mitochondrial morphology, no detectable mtDNA), a rho<sup>+</sup> strain (JSY999) was grown twice to saturation in synthetic minimal medium containing 25  $\mu$ g/ml ethidium bromide (Fox et al., 1991).

Electron microscopy of wild-type (JSY1812) and *fzo1* $\Delta$  (JSY1810) cells was performed essentially as described with the following modifications (Yaffe, 1995). The strains were grown in YPDextrose before fixation, and two additional changes of anhydrous Spurr resin (Polysciences, Inc., Warrington, PA), followed by overnight incubation, were used to achieve maximum infiltration of the samples.

### Depletion of N-3XMYC-Fzo1p

To deplete the N-3XMYC-Fzo1p, the pGAL1-N-3XMYC-*FZO1* plasmid was transformed into JSY2038 (*fzo1* $\Delta$  rho<sup>+</sup> + pRS416-*FZO1*) and cells that had lost the pRS416-*FZO1* plasmid were selected on SRaffinose medium containing 5-FOA (5-fluoro-orotic acid) to yield JSY2273 (*fzo1* $\Delta$  rho<sup>+</sup> + pGAL1-N-3XMYC-*FZO1*). JSY2273 was grown for 24 h in SRaffinose medium lacking leucine to select for pGAL1-N-3XMYC-*FZO1*. To block expression of N-3XMYC-Fzo1p, the cells were collected, rinsed, and grown in SDextrose minus leucine to a density of 2.5  $\times$  10<sup>6</sup> cells/ml. Mitochondrial morphology and mtDNA nucleoid distribution were evaluated at the indicated time points by staining with DiOC<sub>6</sub> (3,3'-dihexyloxycarbocyanine) (Molecular Probes Inc.) (Hermann et al., 1997) or DAPI (Pringle et al., 1991), respectively. JSY2270 (*FZO1* rho<sup>+</sup> + pGAL1-N-3XMYC-*FZO1*), containing the wild-type *FZO1* gene, did not exhibit any changes in mitochondrial morphology or loss of mtDNA during the N-3XMYC-Fzo1p depletion (data not shown). N-3XMYC-Fzo1 protein levels in total cell extracts were analyzed by Western blotting (anti-MYC antibody) at the indicated time points (Harlow and Lane,

1988). Blots were stripped and reprobed with anti-3-PGK (3-phosphoglycerate kinase) (1:1,000 dilution) (Molecular Probes Inc.) to control for differences in protein loading. Mitochondrial morphologies were scored by GFP (green fluorescent protein) staining in JSY2273 cells containing the pRS416-ADH-COXIVpre-GFP plasmid (JSY2634; mito-GFP).

### Generation and Characterization of the *fzo1-1* Mutation

The *fzo1-1* temperature-sensitive allele was generated by low-fidelity PCR (Muhlrad et al., 1992). Mutagenized pRS414-*FZO1* plasmids were transformed into a strain in which *fzo1* $\Delta$ ::*HIS3* was covered with pRS416-*FZO1* (JSY2287). The loss of pRS416-*FZO1* from these cells was selected by growth on medium containing 5-FOA. Cells containing the pRS414-*FZO1* mutagenized plasmids were tested for growth at 25° and 37°C on SGlycerol medium. One strain that was inviable at 37°C was identified (*fzo1-1*). The plasmid containing the mutant version of *FZO1* was recovered, and when retransformed into a cell lacking *FZO1* caused temperature-sensitive growth on SGlycerol medium. Transformation of the pRS414-*fzo1-1* plasmid into a wild-type strain revealed that the *fzo1-1* temperature-sensitive growth defect was recessive. Mitochondrial morphology was examined in *fzo1-1* cells by staining with DiOC<sub>6</sub> (not shown) or mito-GFP.

To examine mitochondrial fusion during mating, *fzo1* $\Delta$  + pRS414-*fzo1-1* cells labeled with either mito-GFP (JSY2802) or Mitotracker red (JSY2804) (Molecular Probes Inc.), were mated at 25° or 37°C and analyzed as described by Nunnari et al. (1997).

### Construction and Analysis of MYC-tagged Fzo1p

A 3XMYC epitope was introduced immediately downstream of the initiating Met in *FZO1* (JSY2028) as described by Schneider et al. (1995). To add additional MYC epitope tags, the N-3XMYC-*FZO1* coding region plus 500 bp of 5' and 3' flanking sequence was amplified from JSY2028 using P297 and P298 and cloned into the EcoRI site of pRS426 generating pRS426-N-3XMYC-*FZO1* (verified by sequence analysis). N-3XMYC-*FZO1* was subcloned into the EcoRI site of pALTER-1 (lacking a NotI site) and digested with NotI to release the 3XMYC tag. NotI fragments

containing 3XMYC epitopes from the plasmid pMPY-3XMYC (Schneider et al., 1995) were inserted into the amino-terminal NotI site of *FZO1*. PCR screening and sequence analysis was used to identify a clone containing a 9XMYC insert (pALTER-1-N-9XMYC-*FZO1*). The N-9X-MYC-*FZO1* was cloned into the EcoRI site of pRS414 to generate pRS414-N-9XMYC-*FZO1*. *fzo1Δ* cells expressing only the 3XMYC-tagged (pRS426-N-3XMYC-*FZO1*; JSY2028) or the 9XMYC-tagged (pRS414-N-9XMYC-*FZO1*; JSY2394) Fzo1 protein grew normally on nonfermentable carbon sources, retained mtDNA, and had wild-type mitochondrial morphology, indicating that the fusion proteins were functional. All MYC-tagged forms of Fzo1p were detected by Western blotting of total protein extracts using an anti-MYC mouse monoclonal antibody (9E10; 1:1,000) (Berkeley Antibody Co., Richmond, CA) (Hermann et al., 1997). The N-9XMYC-Fzo1p was localized in JSY2392 (*fzo1Δ* + pRS414-*FZO1*) and JSY2394 by indirect immunofluorescence with a primary anti-MYC antibody (1:100 dilution) and a goat anti-mouse FITC secondary antibody (1:100 dilution) (Jackson ImmunoResearch) (Pringle et al., 1991).

### Biochemical Analysis of N-9XMYC-Fzo1p

A strain expressing N-9XMYC-Fzo1p (JSY2394) was grown in SGalactose medium lacking tryptophan to select for the pRS414-N-9XMYC-*FZO1* plasmid. Cell lysates (cytosol) were fractionated by differential sedimentation to generate a mitochondrial pellet and a postmitochondrial supernatant (Daum et al., 1982; Zinser and Daum, 1995). Samples (cell equivalents) from each fraction were analyzed by Western blotting. To determine the membrane association of N-9XMYC-Fzo1p, 100 μg of mitochondria purified from JSY2394 were pelleted at 12,000 g for 10 min, resuspended in 75 μl of 100 mM Na<sub>2</sub>CO<sub>3</sub>, pH 11.5, or 1% Triton X-100 containing 1 mM PMSF, 1 μg/ml aprotinin, and 1 μg/ml leupeptin, incubated on ice for 30 min, and then sedimented at 100,000 g for 30 min. Equal volumes of pellet and supernatant fractions were analyzed by Western blotting. Mitochondrial inner and outer membranes (Daum et al., 1982) were loaded onto 5 ml of 0.85–1.6 M sucrose step gradients (0.8, 1.1, 1.35, and 1.6 M) and then centrifuged for 16 h at 30,000 rpm in a Beckman SW50.1 rotor at 2°C (Fullerton, CA). The gradient was partitioned into 13 individual 400-μl fractions that were analyzed by Western blotting. Comparison of protein levels was performed using NIH Image 1.60 (National Institutes of Health, Bethesda, MD). The orientation of N-9XMYC-Fzo1p on mitochondria was determined by incubating 100 μg of purified mitochondria (from JSY2394) in breaking buffer (0.6 M mannitol, 20 mM Hepes-KOH, pH 7.4) containing 100 μg/ml trypsin (Sigma Chemical Co., St. Louis, MO) on ice. To disrupt the outer membrane, mitochondria were diluted with nine volumes of OS buffer (20 mM Hepes-KOH, pH 7.4) and trypsin was added to a final concentration of 100 μg/ml. After 20 min, the reaction was stopped by the addition of soybean trypsin inhibitor (2.5 mg/ml, Sigma Chemical Co.) and 1 mM PMSF. Samples were analyzed by Western blotting. Western blots were performed with the indicated antibodies at the following dilutions: anti-MYC (1:1,000), anti-porin (1:1,000), anti-3-PGK (1:1,000), anti-CoxIV (cytochrome oxidase subunit IV; 1:20,000; Molecular Probes Inc.), and anti-cytochrome *b*<sub>2</sub> (1:10,000).

### Analysis of FZO1 GTPase Point Mutants

pRS414, pRS414-*FZO1*, and pRS414 containing the *FZO1* GTPase point mutations were shuffled into JSY2287 (*fzo1Δ rho*<sup>+</sup> + pRS416-*FZO1*), to generate the following strains: (a) JSY2354 (*fzo1Δ* + pRS414); (b) JSY2392 (*fzo1Δ* + pRS414-*FZO1*); (c) JSY2355 (*fzo1Δ* + pRS414-*fzo1*[T221A]); (d) JSY2356 (*fzo1Δ* + pRS414-*fzo1*[K371A]); (e) JSY2357 (*fzo1Δ* + pRS414-*fzo1*[K200A]); and (f) JSY2358 (*fzo1Δ* + pRS414-*fzo1*[S201N]). None of the mutant Fzo1 proteins caused defects in mitochondrial morphology (anti-porin staining), mtDNA nucleoid retention (DAPI staining) or function (growth on glycerol) when introduced on low or high copy plasmids into a wild-type *FZO1* strain (JSY2288) (data not shown). Subcellular fractionation (see above) and Western blotting with an anti-Fzo1p antibody confirmed that the mutant proteins were expressed and localized to mitochondrial membranes in *fzo1Δ* cells (data not shown).

### Microscopic Techniques

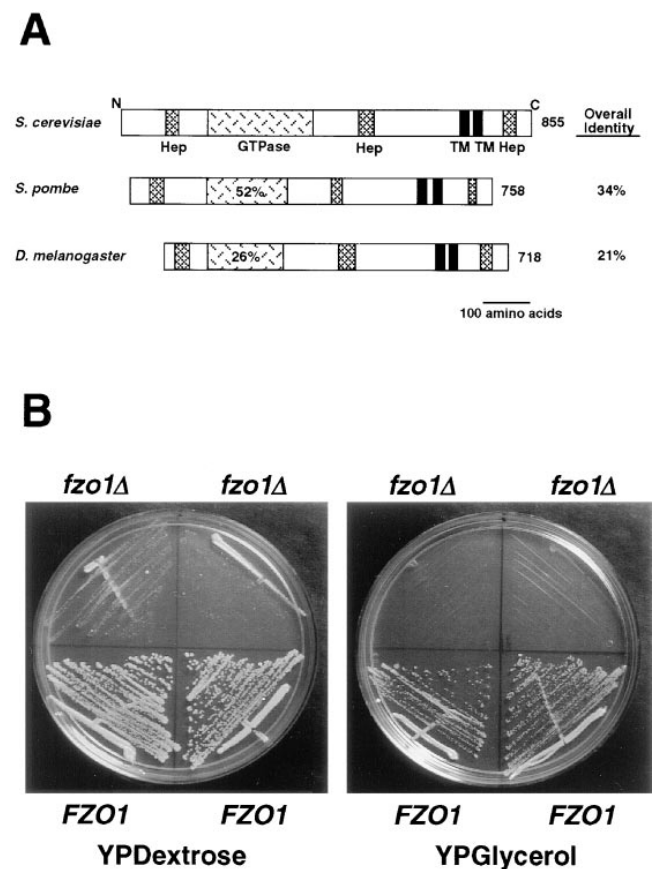
Cells were viewed on a Zeiss Axioplan microscope (1.25× optivar setting; Carl Zeiss Inc., Thornwood, NY) as described in Roeder et al. (1998). Images were captured using a Hamamatsu C5810 color-chilled 3CCD camera (Hamamatsu Photonics, Hamamatsu City, Japan) interfaced to a Macintosh Quadra 840AV computer. For three-dimensional fluorescence

microscopy, data collection was carried out using a Leica confocal microscope and a 100× 1.4 N.A. objective (Leica Inc., St. Gallen, Switzerland). All shutters, stage motion, and image acquisition were under computer control. Images were acquired by moving the stage in ~0.2-μm intervals. Thin sections were viewed with a Hitachi H-700 electron microscope (Tokyo, Japan) and images were captured using the Kodak 161 digital camera system v1.55b (Eastman Kodak Co., Rochester, NY). Digital images were assembled into figures and printed as described in Roeder and Shaw (1996).

## Results

### *S. cerevisiae FZO1* Is Required for Maintenance of Mitochondrial Morphology and Retention of Mitochondrial DNA

To determine whether Fzo family members act generally to control mitochondrial fusion, we disrupted one copy of

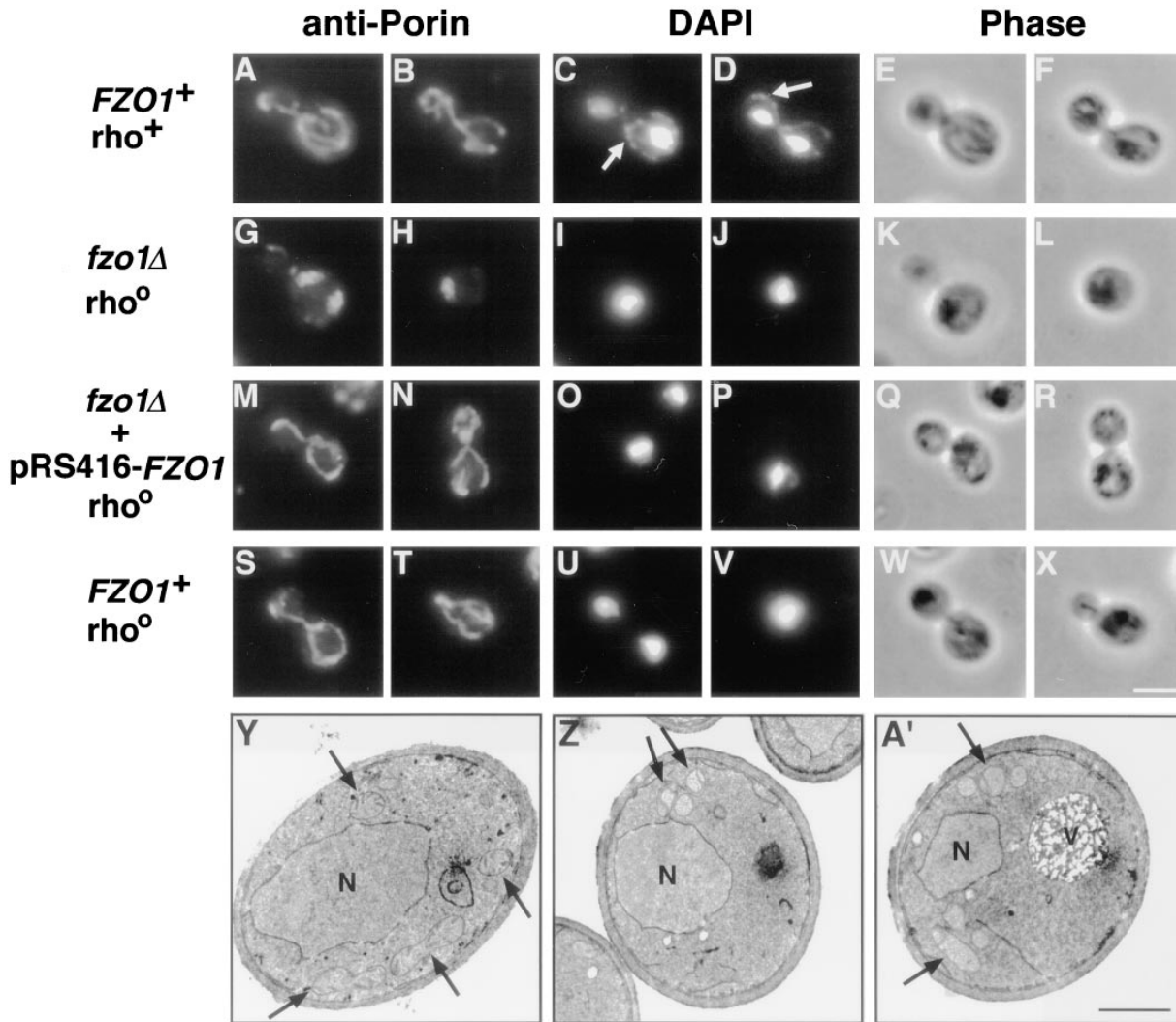


**Figure 1.** *S. cerevisiae FZO1* is required for mitochondrial function. (A) Domain structure of three Fzo family members showing the position of the predicted GTPase (GTPase), heptad repeat (Hep), and transmembrane (TM) domains. Amino acid identities between the entire protein sequences and the predicted GTPase domains of the *S. cerevisiae* Fzo1p, *S. pombe* Fzo1p (the newest family member), and *D. melanogaster* Fzo family members are indicated. GenBank/EMBL/DBJ accession numbers are as follows: *S. cerevisiae* (Z36048), *S. pombe* (ALO23533), and *D. melanogaster* (U95821). (B) *fzo1Δ* cells exhibit growth defects on fermentable and nonfermentable carbon sources. Two *fzo1Δ* (top, JSY1809 and JSY1810) and two wild-type (bottom, JSY1811 and JSY1812) strains from a single tetrad were grown on YPDextrose medium for 2 d at 30°C (left) or YPGlycerol medium for 5 d at 30°C (right).

*S. cerevisiae* *FZO1* in a diploid strain. After sporulation and dissection, *fzo1Δ* haploid cells exhibited a significant growth defect relative to wild type on medium containing the fermentable carbon source dextrose and were unable to grow on medium containing the nonfermentable carbon source glycerol (Fig. 1 *B*). This growth pattern, also referred to as a “petite” phenotype (Dujon, 1981), is characteristic of strains with defective mitochondrial respiration and indicates that the loss of *FZO1* disrupts normal mitochondrial function.

Indirect immunofluorescence revealed that Fzo1p is required for normal organization of the mitochondrial net-

work in vegetatively growing cells (Fig. 2). In a wild-type strain, porin-stained mitochondrial membranes appeared as branched, tubular networks distributed at the cell surface (Fig. 2, *A* and *B*). In contrast, less than 1% of *fzo1Δ* cells contained a wild-type mitochondrial network. Instead, *fzo1Δ* mutants contained between one and five spherical or slightly elongated mitochondrial structures that were localized to the cell cortex but were not distributed evenly around the cell periphery (Fig. 2, *G* and *H*). In addition, numerous small mitochondria were occasionally observed scattered throughout the cytoplasm in the mutant (data not shown). The mitochondrial morphology in



**Figure 2.** *fzo1Δ* cells exhibit defective mitochondrial morphology and lose their mtDNA. (*A–X*) Indirect immunofluorescence images of yeast cells stained with anti-porin antiserum to visualize mitochondrial membranes (*leftmost columns*), and DAPI to visualize nuclei and mtDNA (*middle columns*; *white arrows* in *C* and *D* indicate mtDNA nucleoids). Phase-contrast images of the same cells are shown in the *rightmost columns*. (*A–F*) Normal mitochondrial morphology and distribution of mtDNA nucleoids in a wild-type  $\rho^+$  strain (JSY1812). (*G–L*) Mutant *fzo1Δ rho<sup>0</sup>* cells (JSY1810) contained spherical or slightly elongated mitochondrial structures that lack DAPI-stained mtDNA. (*M–R*) Wild-type mitochondrial morphology, but not mtDNA, was restored in an *fzo1Δ rho<sup>0</sup>* mutant strain (JSY2579) upon reintroduction of the wild-type *FZO1* gene. (*S–X*) Mitochondrial morphology is normal in a wild-type  $\rho^0$  strain (JSY2555) that lacks mtDNA. (*Y–A'*) Transmission electron micrographs of wild-type and *fzo1Δ* cells. (*Y*) Mitochondrial profiles in wild-type cells (JSY1812) are dispersed equally throughout the cell cortex and display numerous cristae (*black arrows*). (*Z* and *A'*) Mitochondrial profiles in *fzo1Δ* cells (JSY1810) are clustered together near the cell periphery and contain fewer cristae. *N*, nuclei; *V*, vacuoles;  $\rho^+$ , containing mtDNA;  $\rho^0$ , lacking mtDNA. Bars: (*A–X*) 5  $\mu\text{m}$ ; (*Y–A'*) 1  $\mu\text{m}$ .

*fzo1Δ* cells resembles that in mutants with abnormal actin cytoskeletons (Drubin et al., 1993; Lazzarino et al., 1994). However, we did not observe defects in the organization of the actin and microtubule cytoskeletons in the *fzo1Δ* strain (data not shown). Although many of the yeast mitochondrial morphology mutants identified to date also have an associated mitochondrial inheritance defect (Burgess et al., 1994; Sogo and Yaffe, 1994; Berger et al., 1997; Hermann and Shaw, 1998), our analysis did not reveal a mitochondrial inheritance defect in *fzo1Δ* cells relative to wild type (data not shown). Thus, Fzo1p controls mitochondrial network organization in yeast but is not required for mitochondrial transport into daughter cells during division.

Transmission electron microscopy indicated that mitochondrial morphology and distribution were also abnormal at the ultrastructural level in *fzo1Δ*. In wild-type cells, mitochondrial profiles were distributed throughout the peripheral cytoplasm and contained elaborate cristae (invaginations of the inner mitochondrial membrane) (Fig. 2 *Y*). In contrast, mitochondrial profiles in *fzo1Δ* cells appeared to cluster together in one or two regions near the plasma membrane (Fig. 2, *Z* and *A'*), similar to the mitochondrial distribution observed by indirect immunofluorescence (Fig. 2, *G* and *H*). The clustered mitochondrial profiles in *fzo1Δ* could represent closely opposed tubules of a collapsed, but still interconnected, mitochondrial reticulum. Alternatively, the clusters could be composed of individual unfused mitochondrial fragments (see below). Cross sections also revealed that *fzo1Δ* mitochondria often contained fewer cristae or lacked cristae altogether (Fig. 2, *Z* and *A'*).

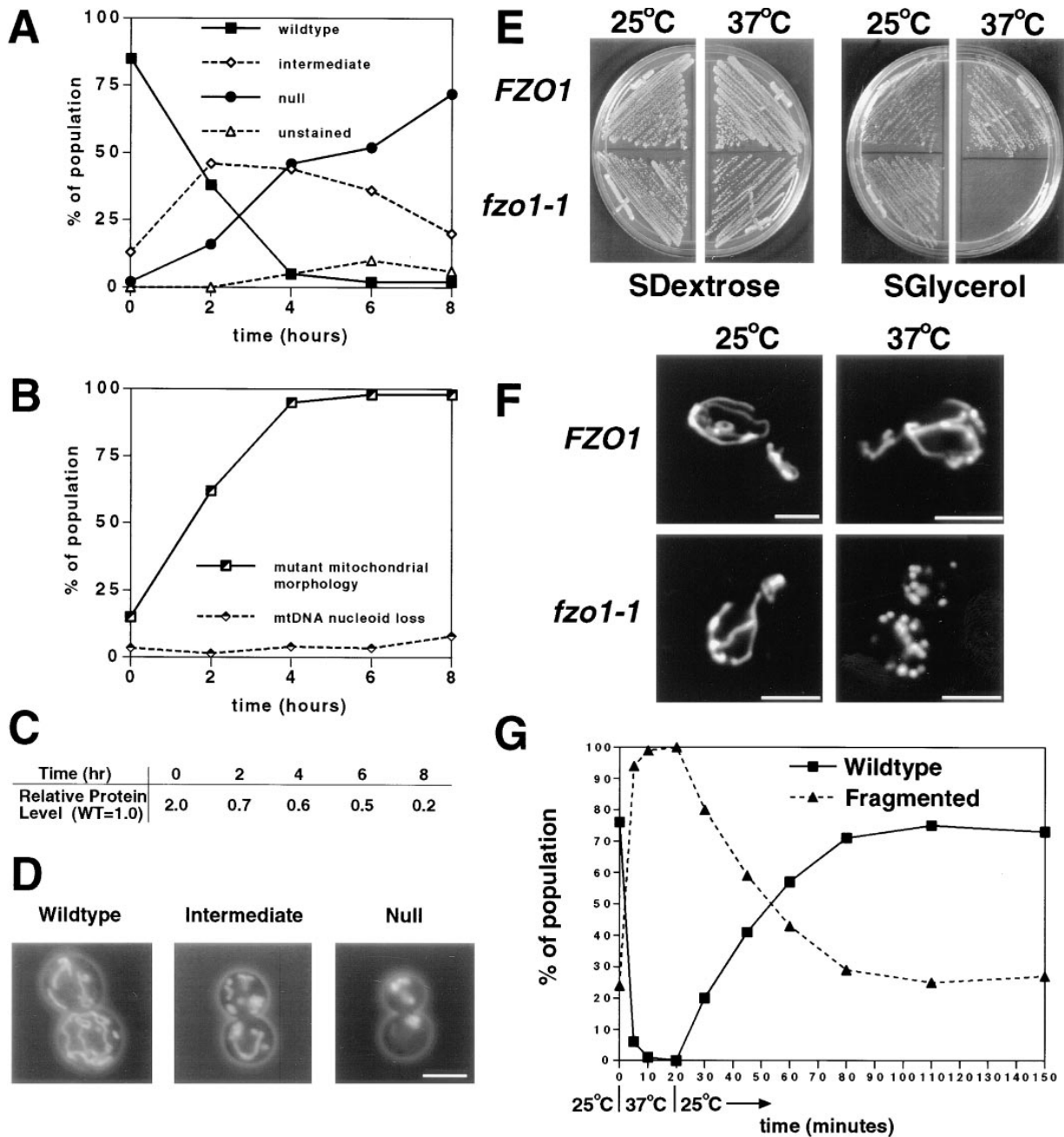
The wild-type function of Fzo1p is also required for the maintenance of mtDNA (mitochondrial DNA) nucleoids. Wild-type cells labeled with the DNA-specific dye DAPI always contained brightly stained nuclei as well as 25–50 punctate mtDNA nucleoids localized at the cell periphery (Fig. 2, *C* and *D*). In contrast, mitochondrial nucleoids were never detected in *fzo1Δ* cells (Fig. 2, *I* and *J*). Crosses of *fzo1Δ* with a known  $\rho^0$  strain (lacking mtDNA) confirmed that mitochondrial genomes were absent in *fzo1Δ* cells (data not shown). Since mtDNA encodes RNAs and proteins essential for mitochondrial respiratory function (Pon and Schatz, 1991), these results raised the possibility that *fzo1Δ* mitochondrial morphology defects were an indirect consequence of mtDNA loss. However, reintroduction of the wild-type *FZO1* gene was sufficient to restore elongated and branched mitochondrial networks in *fzo1Δ* cells (Fig. 2, *M* and *N*) in the absence of mtDNA (Fig. 2, *O* and *P*). The morphology of the restored mitochondrial networks was identical to that observed in an isogenic  $\rho^0$  strain containing a wild-type *FZO1* gene (Fig. 2, *S–V*). These results demonstrate that the loss of mtDNA, and the resulting defects in mitochondrial respiration, do not cause mitochondrial morphology defects in wild-type cells (Guan et al., 1993) or in *fzo1Δ* mutants and suggest that Fzo1p is directly required for the maintenance of normal mitochondrial morphology.

Fzo1p depletion studies revealed that changes in mitochondrial morphology preceded the loss of mtDNA nucleoids. A *GALI* regulated, N-3XMYC-tagged form of Fzo1p on a plasmid was introduced into the *fzo1Δ* mutant strain

using a plasmid shuffling technique to prevent mtDNA loss (refer to Materials and Methods). Growth in raffinose-containing medium (which neither induces nor represses transcription from the *GALI* promoter) resulted in sufficient expression of N-3XMYC-Fzo1p to fully support wild-type growth and restore wild-type mitochondrial morphology in *fzo1Δ* cells (Fig. 3, *A*, *solid squares*, *t* = 0, and *D*, *Wildtype*). After transfer to glucose-containing medium to block expression of N-3XMYC-Fzo1p from the *GALI* promoter, aliquots were harvested at the indicated times and mitochondrial morphology (DiOC<sub>6</sub> staining) and mtDNA distribution (DAPI staining) were examined. During the 8 h after transfer, wild-type mitochondrial networks broke down into smaller segments which eventually collapsed, forming between one and five spherical or slightly elongated membrane clusters per cell (Fig. 3, *A* and *D*). The number, shape, and size of these mitochondrial clusters at the 8-h time point were identical to those visualized in the *fzo1Δ* mutant by anti-porin staining (Fig. 2, *G* and *H*). The defects in mitochondrial morphology observed 8 h after the shift did not severely affect mitochondrial function, since these misshapen organelles were still able to accumulate the membrane potential-sensitive dye DiOC<sub>6</sub> (Chen, 1988). Western blotting with an anti-MYC antiserum showed a 10-fold reduction in N-3XMYC-Fzo1 protein levels during the course of the depletion experiment (Fig. 3 *C*). Cells lacking DAPI-stained mtDNA nucleoids were not detected, even after 8 h when the majority of cells exhibited defective mitochondrial morphology (Fig. 3 *B*). Moreover, 24 h after the initial N-3XMYC-Fzo1p expression block (11 doublings), only 25% of the cells lacked nucleoids, making it unlikely that Fzo1p is required for mtDNA partitioning into buds (data not shown). Together, these results indicate that the loss of Fzo1p leads first to defects in mitochondrial membrane morphology and only later to defects in mtDNA maintenance.

### **Mitochondrial Networks Fragment at 37°C in a Conditional *fzo1-1* Mutant**

Mitochondrial morphology is likely to be regulated, in part, by opposing membrane fusion and membrane fission reactions (Nunnari et al., 1997). If the primary effect of an *fzo1* mutation is to block membrane fusion, we predicted that ongoing mitochondrial fission would initially cause the mitochondrial network to fragment. To test this, a conditional *FZO1* allele (*fzo1-1*) was identified by replacing a wild-type *FZO1* plasmid with mutagenized *FZO1* plasmids in an *fzo1Δ* strain (refer to Materials and Methods). *fzo1-1* allowed growth of the *fzo1Δ* strain at 25° but not 37°C on SGlycerol medium (Fig. 3 *E*). The *fzo1-1* sequence contained three different nucleotide changes, resulting in amino acid substitutions K538I, N543I, and P553Q in the predicted polypeptide (refer to Materials and Methods). Staining with DiOC<sub>6</sub> (data not shown) or GFP (Fig. 3, *F* and *G*) demonstrated that mitochondrial network morphology was wild type in the majority (76%) of *fzo1-1* cells grown at the permissive temperature (Fig. 3, *F*, *fzo1-1*, 25°C, and *G*, *t* = 0 min). 10 min after shifting to 37°C, however, the networks in 99% of the *fzo1-1* cells had fragmented into many small, uniformly distributed, mito-



**Figure 3.** Depletion or loss of Fzo1p function leads to defects in mitochondrial morphology and mtDNA maintenance. (A–D) Depletion of Fzo1p causes defects in mitochondrial morphology that precede the loss of mtDNA nucleoids. *fzo1Δ* cells containing p*GALI*-N-3XMYC-*FZO1* (JSY2273) were shifted from raffinose medium, which allows nearly wild-type levels of Fzo1p expression, to dextrose medium, which represses expression from the *GALI* promoter. (A) Mitochondrial morphology scored by DiOC<sub>6</sub> staining after shift to depletion medium (*n* = 100): mitochondrial morphologies were classified as wild type (branched tubular network), intermediate (tubular condensed network), null (spherical or partially elongated structures), and unstained (nonrespiring mitochondria do not accumulate DiOC<sub>6</sub>). (B) Distribution of mtDNA nucleoids scored by DAPI staining during the Fzo1p depletion shown in A (*n* = 200). The mutant mitochondrial morphology class included the intermediate, null, and unstained categories scored in A. (C) N-3XMYC-Fzo1p levels during the depletion experiment. Protein extracts prepared from equivalent numbers of cells were analyzed by Western blotting with anti-MYC antiserum. N-3XMYC-Fzo1 protein levels during the depletion were normalized to the level of N-3XMYC-Fzo1p expressed from the wild-type *FZO1* promoter (WT = 1.0; JSY2034). (D) Representative wild-type, intermediate, and null mitochondrial morphologies visualized with a matrix-targeted form of the GFP (mito-GFP). (E and F) The temperature-sensitive *fzo1-1* mutation causes rapid and reversible fragmentation of the mitochondrial network. (E) *fzo1-1* cells cannot grow on a nonfermentable carbon source at 37°C. *fzo1Δ* strains containing pRS414-*FZO1* (JSY2926) or pRS414-*fzo1-1* (JSY2793) were grown on SDextrose media (left panels) for 2 d at 25° or 37°C and SGlycerol media (right panels) for 6 d at 25° or 37°C. (F) Mitochondrial morphology in *FZO1* (JSY2793) and *fzo1-1* (JSY2802) cells grown at 25°C and shifted to 37°C for 10 min. Mitochondrial compartments were visualized with mito-GFP. Representative cells are shown. (G) Log-phase *fzo1-1* cells (JSY2804) grown at 25°C (*t* = 0) were shifted to 37°C. After 20 min at 37°C, the cells were returned to 25°C. Mitochondrial morphology was quantified at the indicated times using mito-GFP (*n* ≥ 200). A representative experiment is shown. Bar, 2.5 μm.

chondrial compartments (Fig. 3, *F*, *fzo1-1*, 37°C, and *G*, *t* = 10 min). Upon extended incubation at 37°C, these mitochondrial fragments clustered together forming large membrane aggregates similar to those observed in the *fzo1Δ* strain (data not shown). If after 20 min at 37°C the temperature was reduced to 25°C, the mitochondrial network regained its wild-type morphology within 60 min (Fig. 3 *G*). In control experiments, mitochondrial fragmentation was never observed in wild-type strains grown at 25° or 37°C (Fig. 3 *F*, *FZO1*, 25°C and 37°C). The rapid fragmentation of the mitochondrial network observed in the conditional *fzo1-1* strain provides a direct demonstration that Fzo1p controls mitochondrial morphology in yeast and is consistent with a role for this protein in mitochondrial membrane fusion.

### *fzo1-1* Blocks Mitochondrial Fusion during Mating

Mitochondrial fusion and content mixing has been shown to occur in yeast zygotes soon after cell fusion (Azpiroz and Butow, 1993; Nunnari et al., 1997). To test directly the requirement of Fzo1p in mitochondrial fusion, we examined the effect of the *fzo1-1* mutation on mitochondrial content mixing during mating. Mitochondrial networks were visualized by labeling one haploid parent with the fluorescent vital dye Mitotracker red and the other haploid parent with a matrix targeted form of GFP (mito-GFP) (Nunnari et al., 1997). After mating and zygote formation, the distribution of the fluorophores was examined by fluorescence confocal microscopy. In matings between wild-type cells performed at 25° or 37°C, the two fluorescent markers rapidly and completely colocalized in zygotes, indicating that the parental mitochondrial mem-

Table II. Mitochondrial Fusion in Zygotes

Strains crossed	°C	Fused mitochondria	
		Unbudded zygotes	Budded zygotes
<i>FZO1</i> <sup>+</sup> × <i>FZO1</i> <sup>+</sup>	25	40/50 (80%)	50/50 (100%)
	37	18/50 (36%)	51/51 (100%)
<i>FZO1</i> <sup>+</sup> × <i>fzo1-1</i>	25	40/55 (73%)	53/56 (95%)
	37	1/49 (2%)	32/60 (53%)
<i>fzo1-1</i> × <i>fzo1-1</i>	25	27/50 (54%)	51/55 (93%)
	37	0/51 (0%)	4/53 (8%)

branes had fused and mitochondrial contents had mixed (Nunnari et al., 1997) (Table II). Mitochondrial fusion also occurred efficiently in *fzo1-1* × *fzo1-1* zygotes formed at 25°C (Fig. 4, *A–D*; Table II). In contrast, mitochondrial networks fragmented and failed to fuse in *fzo1-1* × *fzo1-1* zygotes formed at 37°C (Fig. 4, *E–L*; Table II). When *fzo1-1* zygotes were optically sectioned using the confocal microscope, mitochondrial membranes containing the haploid-derived green and red fluorescent markers never colocalized (Fig. 4, *E–L*). Moreover, *fzo1-1* × *fzo1-1* zygotes formed at the restrictive temperature contained unfused mitochondria even after they had completed karyogamy and formed a new bud (Fig. 4, *I–L*). Thus, *fzo1-1* causes a block, and not simply a delay, in mitochondrial docking and/or fusion. Finally, mitochondrial fusion was also reduced in matings between *fzo1-1* and wild-type parents at 37°C (Table II), suggesting that the function of Fzo1p is required in both haploid parents for efficient mitochondrial fusion.

We think it is unlikely that the *fzo1-1* mutation prevents

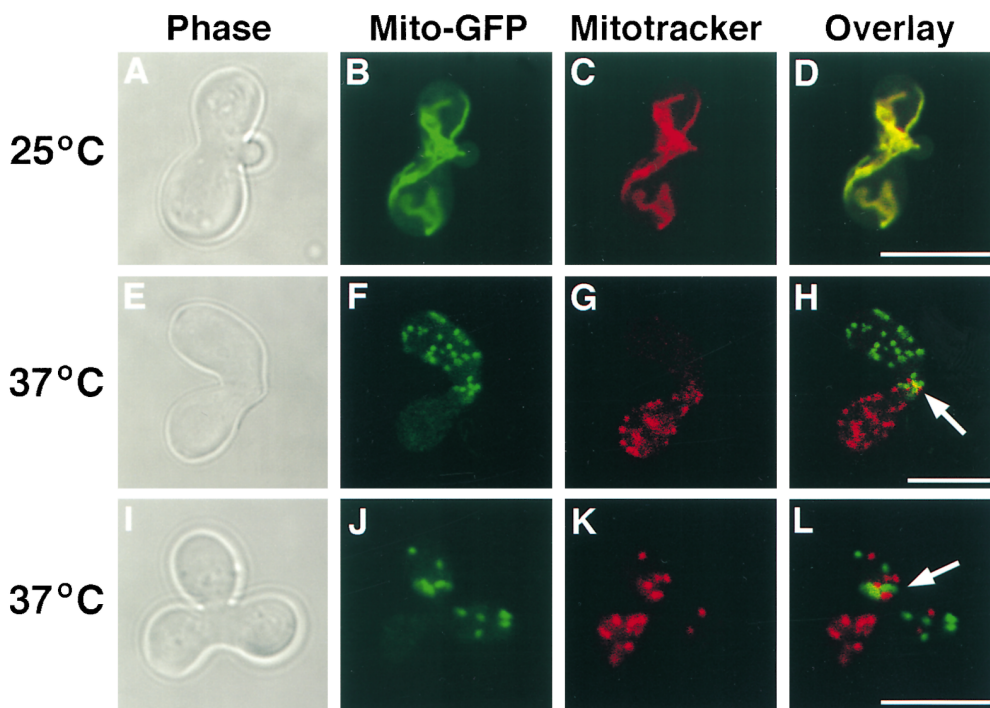


Figure 4. *FZO1* is required for mitochondrial fusion during mating. *fzo1-1* cells of opposite mating type (JSY2802 and JSY2804) were labeled with mito-GFP or Mitotracker red and mated at 25° (*A–D*) and 37°C (*E–H* and *I–L*). Confocal microscopy was used to score the distribution of mito-GFP (green in *B*, *F*, and *J*) and Mitotracker (red in *C*, *G*, and *K*) in serial optical sections (representative single optical section are shown). Fusion and mixing of mitochondrial contents (yellow in *D*) was evaluated in merged mito-GFP and Mitotracker red images (*D*, *H*, and *L*). Zygote morphology was visualized by phase-contrast microscopy (*A*, *E*, and *I*). The zygote bud is on the right in *A* and on the top in *I*. White arrows, regions where parental mitochondrial membranes have intermixed. Bar, 5 μm.



mitochondrial fusion indirectly by interfering with mitochondrial motility and/or distribution. First, although mitochondrial networks fragment at 37°C in *fzo1-1* × *fzo1-1* zygotes, these membrane fragments were segregated normally into daughter cells, indicating that mitochondrial motility was not severely impaired (Fig. 4, *I-L*, arrow). Second, the fragmentation of mitochondria in *fzo1-1* × *fzo1-1* zygotes does not appear to block their association. We often observed red and green mitochondrial compartments in close apposition near the zygote neck or in the diploid daughter cell (Fig. 4, *H* and *L*, arrows). Together, these results provide compelling evidence that Fzo1p plays an essential and direct role in mitochondrial fusion.

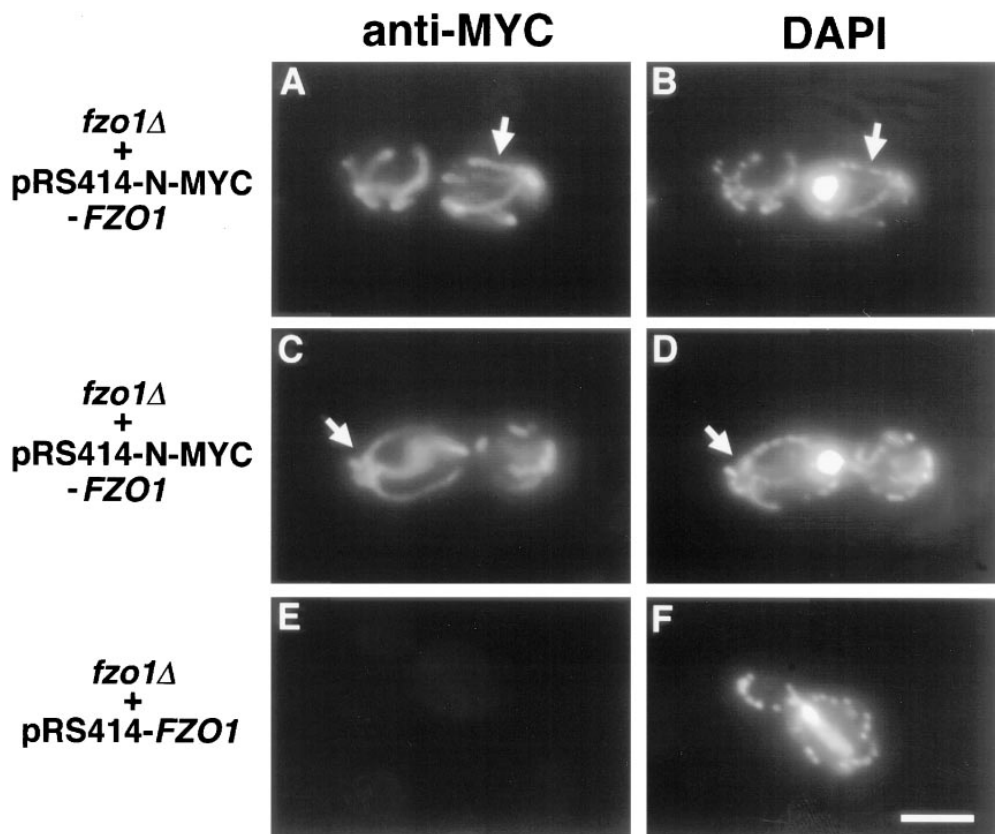
### *Fzo1p Is a Mitochondrial Outer Membrane Protein with Its GTPase Domain Facing the Cytoplasm*

The Fzo1 protein is localized on the mitochondrial network in vegetatively growing yeast cells (Fig. 5). A plasmid encoding Fzo1p tagged near the amino terminus with nine MYC epitopes (N-9XMYC-Fzo1p) was constructed and shown to rescue the mitochondrial morphology defect, the mtDNA loss phenotype, and the glycerol growth defect in the *fzo1Δ* strain (data not shown). Anti-MYC antibodies recognized a 116-kD protein in extracts prepared from wild-type cells expressing the N-9XMYC-Fzo1p (Fig. 6 *A*, lane 2) but not the native Fzo1 protein (Fig. 6 *A*, lane 1). Indirect immunofluorescence with anti-MYC antibodies revealed that N-9XMYC-Fzo1p localized to the mitochondrial network in wild-type cells and was uniformly distributed on this compartment (Fig. 5, *A-D*). The pat-

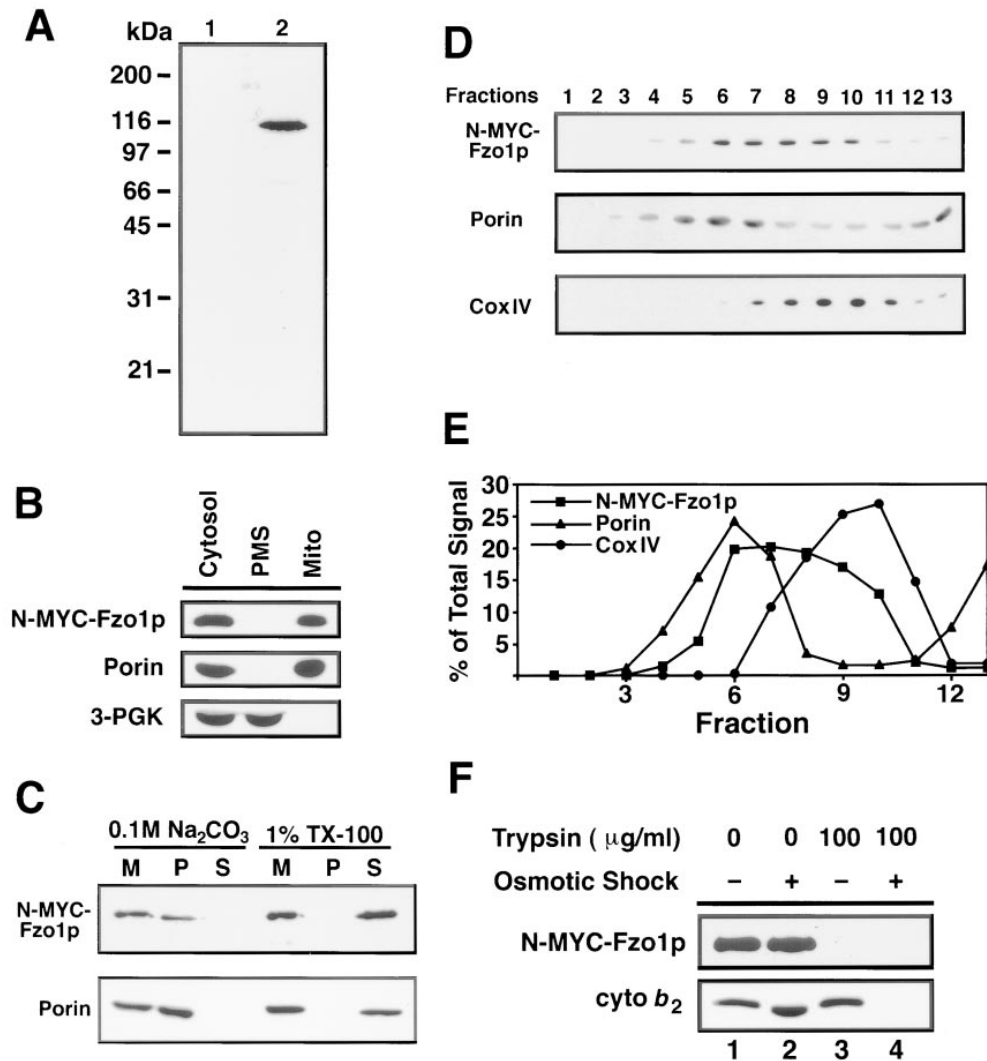
tern of DAPI-stained mtDNA nucleoids in these cells overlapped with the fluorescent signal confirming that N-9XMYC-Fzo1p was located on the mitochondrial network (Fig. 5, *B* and *D*; compare *white arrows* in *A* and *C* with *B* and *D*). Similar results were obtained when cells were stained with polyclonal antibodies generated against the native Fzo1 protein (data not shown). In control experiments, no signal was detected in cells expressing only the wild-type Fzo1p (Fig. 5 *E*).

Subcellular fractionation confirmed the mitochondrial localization of Fzo1p (Fig. 6 *B*). N-9XMYC-Fzo1p cofractionated with the mitochondrial membrane protein porin, during differential centrifugation of a postnuclear cell extract (Fig. 6 *B*, *Mito*). No N-9XMYC-Fzo1p was detected in the post-mitochondrial supernatant fraction that contained the cytoplasmic protein 3-PGK (Fig. 6 *B*, *PMS*). Similar results were obtained from wild-type cells using anti-Fzo1p antiserum to follow fractionation of the native Fzo1 protein (data not shown).

Fzo1p behaved like an integral membrane protein, as predicted based on the two closely spaced and conserved hydrophobic domains near its carboxy terminus (refer to Fig. 1 *A*) (Hales and Fuller, 1997). When mitochondria containing N-9XMYC-Fzo1p were extracted with 100 mM Na<sub>2</sub>CO<sub>3</sub>, pH 11.5, to release peripheral membrane proteins, both the N-9XMYC-Fzo1p and the integral outer-membrane protein porin were resistant to sodium carbonate extraction and remained associated with the membrane pellet (Fig. 6 *C*). In contrast, Fzo1p and porin were released into the supernatant when mitochondrial mem-



**Figure 5.** Fzo1p localizes to the mitochondrial network. (*A-D*) *fzo1Δ* cells (JSY2394) expressing the N-9XMYC-Fzo1 protein were stained with anti-MYC antiserum (*A* and *C*) and DAPI (*B* and *D*). The N-9XMYC-Fzo1p staining was completely coincident with mitochondria as marked by mtDNA nucleoids (compare *white arrows* in *A* and *C* with *B* and *D*). (*E* and *F*) Anti-MYC serum did not stain *fzo1Δ* cells (JSY2392) expressing the Fzo1p lacking the MYC tag. Bar, 5 μm.



**Figure 6.** Fzo1p is a mitochondrial integral membrane protein with its GTPase domain exposed to the cytoplasm. (A) Total cell extracts from *fzo1Δ* cells containing either the wild-type pRS414-*FZO1* (lane 1; JSY-2392) or pRS414-N-9XMYC-*FZO1* plasmids (lane 2; JSY2394) analyzed by Western blotting with an anti-MYC antiserum. The same *fzo1Δ* strain harboring pRS414-N-9XMYC-*FZO1* (JSY2394) was used for all of the experiments described in 6B-F. (B) N-9XMYC-Fzo1p cofractionated with the mitochondrial protein porin. Protein extracts from equivalent numbers of cells were fractionated by differential centrifugation, separated by SDS-PAGE, and analyzed by Western blotting with anti-MYC, anti-porin, and anti-3-PGK serum. *Cytosol*, postnuclear cytoplasmic extract; *Mito*, crude mitochondrial pellet; *PMS*, supernatant depleted of mitochondria. (C) N-9XMYC-Fzo1p is an integral membrane protein. Purified mitochondria (*M*) were treated to solubilize peripheral membrane proteins (0.1 M Na<sub>2</sub>CO<sub>3</sub>) or integral membrane proteins (1% Triton X-100), separated into pellet (*P*) and supernatant (*S*) fractions,

and analyzed by SDS-PAGE and Western blotting. The release of soluble cytochrome *b*<sub>2</sub> and the peripheral F<sub>1</sub>F<sub>0</sub> ATPase subunit into the supernatant fraction after 0.1M Na<sub>2</sub>CO<sub>3</sub> treatment was confirmed by Western blotting (data not shown). (D and E) N-9XMYC-Fzo1p fractionates in an intermediate density mitochondrial membrane fraction. Fractions from a 30–50% sucrose step gradient (*top*, fraction 1) analyzed by Western blotting with anti-MYC, anti-porin, and anti-CoxIV serum. (E) The percentage of total N-MYC-Fzo1p, porin, and CoxIV per fraction. (F) The GTPase domain of Fzo1p is exposed on the cytoplasmic surface of mitochondria. Untreated (lane 1), trypsin-treated (lanes 3 and 4, 100 μg/ml), and osmotically shocked (lanes 2 and 4) mitochondria were analyzed by SDS-PAGE and Western blotting with anti-MYC and anti-cytochrome *b*<sub>2</sub> serum.

branes were solubilized with 1% Triton X-100 (Fig. 6 C). Fzo1p also remained associated with mitochondrial membranes after the organelles were disrupted by osmotic and mechanical methods to release soluble intermembrane space and matrix proteins (Fig. 6 D and data not shown).

Analysis of submitochondrial membrane fractions separated on sucrose density gradients indicated that Fzo1p associates with both the inner and outer mitochondrial membranes. N-9XMYC-Fzo1p fractionated at an intermediate density that overlapped with, but was distinct from, the distribution of both inner membrane vesicles containing the integral membrane protein CoxIV and outer membrane vesicles containing porin (Fig. 6, D and E). Although this result does not prove that Fzo1p crosses both mitochondrial membranes, this fractionation pattern is

characteristic of “trapped” translocation intermediates that span both mitochondrial membranes at contact sites formed by the translocation pore (Pon et al., 1989). Proteins that fractionate with both inner and outer mitochondrial membranes and are enriched at contact sites have been observed previously (Pon et al., 1989).

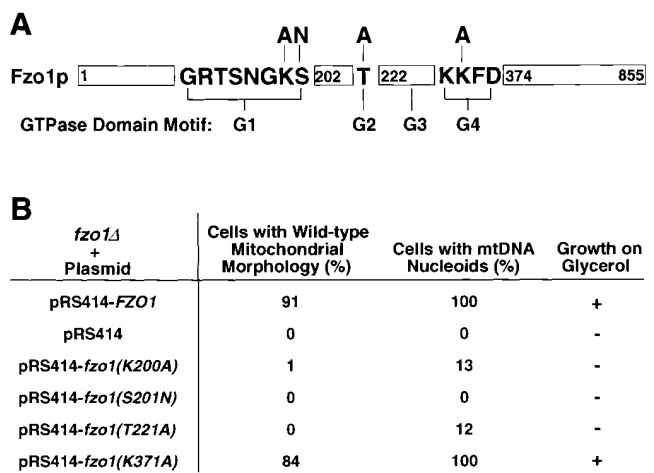
Fzo1p is oriented with its amino-terminal GTPase domain and adjacent heptad repeats exposed on the cytoplasmic face of the mitochondrial compartment. Treatment of isolated, intact mitochondria with trypsin resulted in the complete digestion of the MYC tag on the amino terminus of Fzo1p (Fig. 6 F, lane 3). In contrast, cytochrome *b*<sub>2</sub>, a protein in the intermembrane space, was resistant to proteolysis, indicating that the outer mitochondrial membrane remained intact (Fig. 6 F, lane 4).

Cytochrome *b*<sub>2</sub> could be digested, however, if the mitochondrial outer membrane was disrupted by osmotic shock (Fig. 6 F, lane 4). Since Fzo1p behaves like an integral membrane protein, these data also indicate that at least one of the hydrophobic domains at the carboxy-terminus of Fzo1p is embedded in the mitochondrial outer membrane.

The signals and machinery that target Fzo proteins to mitochondria may be conserved. When a cDNA encoding the *Drosophila melanogaster* Fzo protein was introduced into wild-type cells on a *GAL1* low copy plasmid, the *Drosophila* protein cofractionated with mitochondrial membranes as assayed by Western blotting with antibodies specific for the *D. melanogaster* homologue (data not shown). However, the *Drosophila* Fzo protein did not rescue mitochondrial morphology or mtDNA loss phenotypes in the *S. cerevisiae fzo1Δ* strain.

### *Fzo1p* Function Requires the Conserved GTPase Domain

The Fzo1p GTPase domain contains four conserved motifs designated G1–G4 (Fig. 7 A). In most GTPases, these domains are required for GTP binding and hydrolysis as well as conformational changes elicited by nucleotide binding (Bourne et al., 1991). Conserved residues in three of the four G motifs in Fzo1p (*K200A* and *S201N* in G1, *T221A* in G2, and *K371A* in G4) were altered by site-directed mutagenesis (Fig. 7 A). All of these amino acid substitutions are known to disrupt either nucleotide binding or interactions with effector proteins in other GTPases (Sigal et al., 1986; Adari et al., 1988; Cales et al., 1988; Feig and Cooper, 1988; Farnsworth and Feig, 1991; Vojtek et al., 1993; Murphy et al., 1997). When low copy plasmids containing the mutated *fzo1* genes were introduced into *fzo1Δ* cells, the mutant Fzo1 proteins were expressed at wild-type levels and were targeted to the mitochondrial compartment as assayed by differential centrifugation and Western blotting with anti-Fzo1p antiserum (data not shown). However, the *fzo1(K200A)*, *fzo1(S201N)*, and *fzo1(T221A)* mutant genes failed to rescue the glycerol growth defect or the mitochondrial morphology defects in the *fzo1Δ* strain (Fig. 7 B). Interestingly, a significant percentage (13 and 12%, respectively) of cells containing the *fzo1(K200A)* and *fzo1(T221A)* mutant genes contained detectable mtDNA nucleoids (Fig. 7 B), although the total number of nucleoids was reduced relative to wild type (1–5 instead of 25–50) (data not shown). It is possible that these mutant Fzo1 proteins retain some residual functions required for mtDNA maintenance. Alternatively, cells containing these mutant proteins may simply lose their mitochondrial genomes more slowly than *fzo1*-null cells. Mutation of a conserved residue in the G4 domain (*K371A*) did not disrupt the function of *FZO1* (Fig. 7 B). This result is somewhat surprising, since lysine 371 is conserved throughout the GTPase superfamily and is known to be required for high-affinity GTP binding by Ras (Der et al., 1988; Bourne et al., 1991). In addition, the same mutation in *Drosophila fzo* had a modest but significant effect on the function of the protein (Hales and Fuller, 1997). Finally, wild-type cells overexpressing the mutant Fzo1 proteins (10–20 fold from a 2- $\mu$  vector) did not ex-



**Figure 7.** GTPase domain mutations block Fzo1p function. (A) Schematic representation of Fzo1p illustrating the location and sequence of the GTPase domain motifs (G1–G4; not to scale) with mutations changing conserved residues indicated. The mutant amino acids are depicted above the original residues. (B) Mutations in the G1 and G2 motifs disrupt the function of Fzo1p. Glycerol growth, mitochondrial morphology (anti-porin, *n* = 400), and mtDNA nucleoid distribution (DAPI staining, *n* ≥ 100) analyzed in *fzo1Δ* cells (JSY2354) containing wild-type *FZO1* (JSY2392) or mutated *fzo1* genes (JSY2355–2358) carried on low copy plasmids (pRS414).

hibit dominant growth or mitochondrial morphology defects when compared with cells overexpressing the wild-type Fzo1p (10–20 fold from a 2- $\mu$  vector) (data not shown). These observations indicate that GTP binding and/or hydrolysis is essential for Fzo1p function.

## Discussion

### *Fzo1p* Regulates Mitochondrial Fusion

We have shown that Fzo1p is a transmembrane GTPase required for mitochondrial fusion in yeast. Mitochondrial membranes rapidly fragment at the nonpermissive temperature in a conditional *fzo1-1* strain. Since the opposing processes of mitochondrial fusion and fission are responsible for the reticular structure of the mitochondrial network in mitotically dividing cells (Nunnari et al., 1997), the simplest explanation for this result is that *fzo1-1* causes a selective block in fusion and that fragmentation occurs as a result of continuing mitochondrial fission. *fzo1-1* is the only yeast mitochondrial morphology mutant with a fragmentation phenotype, consistent with its novel role in mitochondrial fusion. Mitochondrial fusion and content mixing were also blocked in homozygous *fzo1-1* × *fzo1-1* zygotes and reduced in heterozygous *fzo1-1* × *FZO1*<sup>+</sup> matings. Functional Fzo1 protein could be required on opposing mitochondrial membranes for efficient fusion. If this is the case, then the residual mitochondrial fusion observed in *fzo1-1* × *FZO1*<sup>+</sup> matings could result from new protein synthesis and complementation in the heterozygous zygote. Alternatively, functional Fzo1 protein on only one of the fusion partners may allow fusion between

Fzo1p<sup>+</sup> and Fzo1p<sup>-</sup> mitochondrial membranes at lower efficiency.

Disruption of the *FZO1* gene, depletion of the Fzo1 protein, or prolonged incubation of the *fzo1-1* strain at 37°C lead to the severe clumping and aggregation of mitochondrial membranes. It seems likely that a constitutive block in fusion is responsible for the dramatic defects in mitochondrial morphology we observed under all of these conditions. Although mitochondrial inheritance is often defective in yeast mutants with abnormal mitochondrial morphology (Burgess et al., 1994; Sogo and Yaffe, 1994; Berger et al., 1997; Hermann, 1998), the loss of Fzo1p function did not affect the motility or transport of mitochondrial membranes during mitotic division and the abnormal mitochondrial compartments in *fzo1-1* and *fzo1Δ* cells were efficiently transmitted to daughter buds (refer to Fig. 2 G). Defects in Fzo1p function also lead to the loss of mitochondrial genomes. Although we cannot rule out that Fzo1p participates directly in mtDNA replication and/or segregation, several lines of evidence suggest that the loss of mtDNA in *fzo1Δ* cells is a secondary consequence of changes in mitochondrial morphology. First, when Fzo1p is depleted from wild-type cells, defects in mitochondrial membrane structure are observed many hours before DAPI-stained mtDNA nucleoids are lost. Second, a number of studies indicate that mutations affecting mitochondrial morphology result in decreased mtDNA stability (Guan et al., 1993; Burgess et al., 1994; Sogo and Yaffe, 1994; Berger et al., 1997; Hermann, 1998). Further experiments are required to determine the mechanism by which mtDNA is lost from *fzo1* mutant strains.

### ***Fzo1 Mitochondrial Membrane Association and Topology***

Immunolocalization, fractionation, and protease digestion studies indicated that Fzo1p is an integral membrane protein with its amino terminus displayed on the mitochondrial surface. This topology positions Fzo1p's GTPase domain and adjacent heptad repeats in the cytoplasm where they could interact with binding partners and/or regulatory molecules that might regulate Fzo1p function. Given the conservation of the domain structure and overall charge distribution of the different Fzo family members (Fig. 1 A; Hales and Fuller, 1997), we predict that Fzo homologues from other organisms will display a similar mitochondrial distribution and topology.

Mitochondrial fusion has been reported to initiate at stable contact sites between the inner and outer mitochondrial membranes (Bereiter-Hahn and Voth, 1994). The carboxy terminus of Fzo proteins could play an important role in coordinating the behavior of the two lipid bilayers at these sites during the fusion reaction. Protease digestion and protein solubilization studies suggested that at least one of Fzo1p's carboxy-terminal hydrophobic domains is embedded in the outer mitochondrial membrane. In addition, Fzo1p migrated in sucrose gradients with an intermediate density fraction that partially overlapped both inner and outer mitochondrial membrane markers. This fractionation pattern has been observed for proteins that are physically associated with mitochondrial contact sites (Pon et al., 1989), suggesting that Fzo1p is located in these struc-

tures. There are a number of topologies that could account for the ability of Fzo1p to fractionate with both mitochondrial membranes. It is possible that Fzo1p spans both membranes with its carboxy terminus in the matrix as originally proposed by Hales and Fuller (1997). Alternatively, the carboxy-terminal tail of Fzo1p could extend into the intermembrane space and interact with proteins in the inner membrane. Finally, Fzo1p might be exclusively associated with the outer membrane at contact sites formed by other proteins. Localization of the carboxy terminus of Fzo1p will help to distinguish between these models.

Although the *Drosophila* and yeast Fzo proteins are both required for mitochondrial fusion, several observations suggest that they may be regulated differently. First, our studies indicate that the *Drosophila* Fzo protein is efficiently targeted to mitochondrial membranes in wild-type yeast but cannot rescue mitochondrial phenotypes in the *fzo1Δ* strain (data not shown). Second, the *Drosophila* Fzo and yeast Fzo1 proteins exhibit distinct expression patterns in the two organisms. *Drosophila* Fzo was only detected on sperm mitochondria during a short period of time when mitochondrial fusion was occurring (Hales and Fuller, 1997), suggesting that the timing of mitochondrial fusion could be developmentally regulated by controlling Fzo expression, localization, and/or degradation. In contrast, yeast Fzo1p protein levels and mitochondrial localization did not change during mitotic growth, mating, or meiosis (data not shown), consistent with the observation that mitochondrial fusion occurs during all stages of the yeast life cycle (Pon and Schatz, 1991; Nunnari et al., 1997; Hermann and Shaw, 1998). In time-lapse studies, yeast mitochondrial fusion is observed when the tip of a mitochondrial tubule encounters the tip or side of another mitochondrial tubule (Nunnari et al., 1997). This has led to the suggestion that key fusion components are localized or specifically activated at the tips of mitochondrial tubules (Nunnari et al., 1997). Our observation that Fzo1p is uniformly distributed on the mitochondrial compartment suggests that the fusion machinery is not localized at tips. Instead, Fzo1p could be activated locally at sites of membrane contact.

### ***Fzo1 Function Requires the Amino-terminal GTPase Domain***

The GTPase domain of Fzo1p is essential for its function and could act as a molecular switch to regulate mitochondrial docking and/or fusion. The *fzo1(K200A)* and *fzo1(S201N)* mutations in the G1 motif are equivalent to mutations in the Ras GTPase that reduce guanine nucleotide binding (Sigal et al., 1986; Feig and Cooper, 1988; Farnsworth and Feig, 1991). The *fzo1(T221A)* substitution in the G2 motif is based on a Ras mutation which eliminates interactions with effector molecules (Adari et al., 1988; Cales et al., 1988; Vojtek et al., 1993; Murphy et al., 1997). None of these mutations disrupted Fzo1p localization suggesting that GTP binding and/or hydrolysis is not required to target Fzo1p to mitochondria.

In contrast, the *fzo1(K371A)* mutation in the G4 motif retained wild-type function and was able to rescue defects in mitochondrial fusion, mtDNA maintenance, and glycerol growth in *fzo1* mutant cells. This lysine is highly con-

served among the superfamily of GTPases (Bourne et al., 1991) and is required for efficient nucleotide binding and stabilization of the GTP binding pocket in Ras (Lys117) (Der et al., 1988; Pai et al., 1990). Although mutating Lys117 in Ras significantly reduces its affinity for GTP, it does not disrupt its oncogenic potential (Clanton et al., 1986; Der et al., 1988). In addition, the analogous mutation in the G4 motif of *Drosophila* Fzo did not completely disrupt mitochondrial fusion (Hales and Fuller, 1997). These results suggest that the yeast and fly Fzo proteins may have a high intrinsic affinity for GTP that is not completely compromised by alterations in the G4 lysine. Alternatively, this lysine may be completely (yeast Fzo1p) or partially (fly Fzo) dispensable with respect to nucleotide binding.

Mutant forms of the yeast dynamin-like proteins Vps1p and Dnm1p (Vater et al., 1992; Otsuga et al., 1998), mammalian dynamin (Herskovits et al., 1993; van der Bliek et al., 1993), and Ras (Sigal et al., 1986; Feig and Cooper, 1988; Farnsworth and Feig, 1991) induce dominant-interfering phenotypes when overexpressed in wild-type cells. These dominant phenotypes are thought to result because the mutant forms of the proteins either titrate out or block the activities of binding partners required for their function. In contrast, none of the disabled forms of Fzo1p we tested induced dominant mitochondrial phenotypes in wild-type cells (data not shown). The simplest interpretation of these results is that Fzo1p acts alone or that mutations in the GTPase domain of Fzo1p completely disrupt its ability to interact with itself or other proteins.

### Models for the Mechanism of Fzo1p Action

Given what is known regarding the role of other integral membrane proteins in fusion, we propose that Fzo GTPases act as molecular switches to directly mediate mitochondrial docking and/or membrane fusion. Both the domain structure of the Fzo family members and the topology we have determined for Fzo1p are consistent with this model. Fzo molecules share structural features with two classes of integral membrane proteins that mediate membrane fusion events. The SNAREs regulate membrane docking and fusion during vesicle transport and during the homotypic fusion of organelle membranes (Denesvre and Malhotra, 1996; Hay and Scheller, 1997; Edwardson, 1998; Götte and Fischer von Mollard, 1998; Weber et al., 1998). The viral-type fusion proteins regulate extracellular membrane docking and fusion events (Hernandez et al., 1996; Huovila et al., 1996). Both SNAREs and viral fusion proteins contain multiple heptad repeats in their amino-terminal domains. These repeats form parallel coiled coils that are proposed to mediate the direct association of v- and t-SNAREs on opposing membranes and the oligomerization of viral fusion proteins within a membrane (Kee et al., 1995; Hernandez et al., 1996; Hanson et al., 1997; Hay and Scheller, 1997; Hughson, 1997). Our studies indicate that the two amino-terminal heptad repeats in Fzo1p extend into the cytoplasm. If Fzo1p serves a SNARE-like function, these repeats could mediate the direct or indirect association of Fzo1p with itself or with another protein binding partner on an opposing mitochondrial membrane. The cytoplasmic GTPase domain of Fzo1p might function

to control the rate or fidelity of such Fzo1p interactions, similar to the manner in which Rab GTPases regulate SNARE-SNARE associations (Lupashin and Waters, 1997). The idea that Fzo1p functions as a novel type of mitochondrial SNARE is attractive, given that none of the v- or t-SNAREs encoded by the *S. cerevisiae* genome localize to the mitochondrial compartment (H.R.B. Pelham, personal communication).

Alternatively, Fzo1p may operate more like viral fusion proteins (White, 1990, 1992; Hernandez et al., 1996; Hughson, 1997; Qiao et al., 1998). These integral membrane proteins contain additional hydrophobic fusion peptides that normally remain masked. When activated, the fusion peptide inserts into the target membrane generating a docked state. Conformational changes in the protein pull the membranes together and promote fusion. Fzo1p contains two hydrophobic sequences in its cytoplasmic domain that resemble fusion peptides (Hermann, 1998). It is possible that Fzo1p uses its GTPase activity as a switch to stimulate the insertion of these putative fusion peptides into a neighboring mitochondrial membrane. The GTPase domain might also regulate conformational changes in Fzo1p that pull the two docked mitochondrial membranes together after peptide insertion. The observation that Fzo1p is tightly associated with both mitochondrial membranes suggests that it might regulate the fusion of the inner membrane as well. Whether the predicted transmembrane domain and heptad repeat closest to the carboxy terminus of Fzo1p are in a position to influence inner membrane behavior remains to be seen.

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