

# Urinary schistosomiasis in Zimbabwean school children: predictors of morbidity

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## ABSTRACT

**Background:** The morbid effects of urinary bilharziasis are becoming more evident with the advent of sophisticated diagnostics such as ultrasound. However, such diagnosis of *Schistosoma haematobium* morbidity is often hampered by lack of funds, proper equipment, or training.

**Objective:** We performed a cross-sectional investigation of schoolchildren in a highly endemic area of east central Zimbabwe in order to assess the utility of a number of simple clinical indicators to predict *Schistosoma haematobium* morbidity.

**Methods:** Prevalence and intensity of *S. haematobium* infection was determined in 551 schoolchildren, with ultrasound examination of the kidneys and bladder performed on 222. The association of a number of demographic, parasitological, and clinical parameters with clinical outcome was evaluated.

**Results:** Overall, 60% of the children were infected with *S. haematobium*. Although lacking specificity, proteinuria and parasite eggs count best predicted bladder pathology. Presence of kidney dilation was associated with fatigue and pain upon urination, but these variables were not very sensitive.

**Conclusions:** None of the variables assessed were ideal predictors of morbidity. However, the results suggest that a combination of inexpensive, simple indicators may allow for improved targeting of *S. haematobium* treatment to those with severe morbidity and better monitoring of the progress of control campaigns when more expensive diagnostic methods are not available.

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## INTRODUCTION

An estimated 200 million people in 76 countries are infected with schistosomes. Of these, approximately 20 million suffer severe sequelae, with disease manifestations ranging from bladder carcinomas to liver fibrosis.<sup>1</sup> Current health policy on schistosomiasis aims at decreasing morbidity associated with infection.<sup>2,3</sup> Increasing the accessibility of methods to diagnose schistosomiasis induced pathology is instrumental in meeting this goal.

For *Schistosoma haematobium* morbidity assessment, ultrasound examination has achieved

widespread acceptance.<sup>4,5</sup> Field studies with portable ultrasound devices have shown that type and extent of schistosomiasis associated urinary tract pathology varies widely.<sup>6-9</sup> Being able to diagnose severe cases or define risk factors associated with pathology would be useful for targeting treatment and would also improve our understanding of the magnitude and etiology of such pathology. As ultrasound is a costly method of diagnosing urinary tract pathology, the current study assesses the utility of a number of less expensive clinical indicators, already extensively used in schistosomiasis endemic areas, to predict morbidity. We provide recommendations based on analysis of data from 222 Zimbabwean schoolchildren in a cross-sectional survey of *S. haematobium* associated pathology.

## METHODS

From 1998-1999, 551 9-16 year old primary schoolchildren from the Chikwaka Communal Lands of Zimbabwe, an area endemic for *S. haematobium*,<sup>6,10,11</sup> participated in this survey. Inclusion criteria comprised informed consent, minimum age of nine years with majority of the time spent in the immediate area, and no obvious indicator of

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ill health. During the preceding decade, no organized schistosomiasis treatment campaigns were conducted in the area.

Students were screened for *Schistosoma mansoni* and geohelminths using Kato-Katz thick smear method.<sup>12</sup> Additionally, 3 urine specimens per student, taken during the period for optimum egg excretion i.e. between 10:00 and 14:00 hrs on 3 different days, were examined for presence of *S. haematobium*. Eggs were visualized after staining with Lugol's iodine<sup>13</sup> A patient was considered infected if a 10 ml urine sample contained at least one egg. Reagent strips (Hemastix®, Bayer Diagnostics) were used on samples to determine the presence and extent of haematuria and proteinuria.

A subset of students ( $n=222$ ), from whom we had complete parasitological data, who were uninfected with *S. mansoni*, and who were present on the survey days, were examined using a portable ultrasound device (UF-5800A, Fukuda Denshi Co., with a 3.5 MHz convex probe). Thirty-three of these students were uninfected and served as endemic controls. Bladders were examined when full and the ultrasonographer was unaware of patient infection status. A transverse measurement of the bladder was taken and pathology was classified as **0**- No pathology, wall < 5mm; no masses, or polyps; **1**- Mild, wall < 5mm; Focal thickenings, no masses or polyps; or **2**- Severe, wall ≥ 5mm; and/or masses or polyps.<sup>6,14</sup> Kidneys were examined post voiding and dilation initially classified as follows: **0**- No dilation; **1**- < 5 mm; **2**- 5-15 mm; with > 2 cm parenchyma; **3**- > 15 mm with ≤ 2 cm parenchyma; or end-stage with absence of parenchyma.<sup>14</sup> Because one kidney can compensate for the other, for analysis we further categorised the data according to whether damage was uni- or bilateral: **0**- No pathology; **1**- Mild, one kidney with

grade 1 pathology; the other, 0 or 1; **2**- Moderate, one kidney, grade 2; the other, 0 or 1; **3**- Severe, both kidneys, grade 2; or one kidney, grade 3.

To those who underwent ultrasound examination, a questionnaire was administered eliciting information on education, demographic information, type and duration of water contact activities, past or present occurrence of symptoms, and treatment history. Univariate analyses, an odds ratio or chi-square for trend, were used to examine which questionnaire items were associated with morbidity.<sup>6</sup> Data analysis was performed using SPSS software (Version 10.0). For statistically significant indicators (two-sided  $P$ -values < 0.05), the sensitivity, specificity, and positive and negative predictive values were calculated. Combinations of clinical indicators were also compared. Results were stratified by gender when the gender-specific values varied more than ±5 percentage points from the combined value.

All study methods were approved by the Medical Research Council of Zimbabwe and the Institutional Review Board of the Johns Hopkins Bloomberg School of Public Health. Infected participants were given a single oral dose of 40mg/kg praziquantel (Biltricide®, Bayer LTD) at the end of the investigation at their school.

## RESULTS

*Schistosoma haematobium* infection was found in 329 of the 551 students surveyed (60%). In those who underwent ultrasound examination, morbidity was significantly linked with infection status (Table 1). Urinary tract pathology was present in 72% of those infected, with 49/184 (27%) having major wall thickenings of or masses or polyps on their bladder. Hydronephrosis was found in 67/188 (36%) of infected students, with 22% having bilateral dilations (Table 1). Dilation of the kidneys occurred alongside bladder pathology in 34/183 (19%) of cases. There was no correlation between presence or severity of bladder pathology and presence or extent of kidney pathology.

**Table 1. Urinary tract pathology and *Schistosoma haematobium* infection status**

Bladder Pathology	<i>S. haematobium</i> status		Total*
	Positive	Negative	
<b>0</b>	92	28	120
<b>1</b>	43	2	45
<b>2</b>	49	3 (3 <sup>†</sup> )	52
<b>Total</b>	184	33	217
	OR = 5.60 (2.07-15.14), $P < 0.01$ ‡		
Kidney Pathology			
<b>0</b>	95	23	118
<b>1</b>	26	6 (2 <sup>†</sup> )(3 <sup>§</sup> )	32
<b>2</b>	40	3 (2 <sup>†</sup> )	43
<b>3</b>	27	1	28
<b>Total</b>	188	33	221
	OR = 2.25 (1.02-4.99), $P = 0.04$ ‡		

\* Five students did not have a complete bladder examination and one student did not have a complete kidney evaluation, thus they were omitted from our analysis.

† Number with recent history of *S. haematobium* infection.

‡ Odds ratio (OR) is for presence of pathology (any grade) versus infection status.

§ Number with haematuria.

Previously, we found no association between haematuria, proteinuria, nor parasite egg count with kidney pathology in these infected schoolchildren.<sup>6</sup> Pain upon urination, however was associated with both presence and extent of hydronephrosis ( $P = 0.02$ ), while fatigue was associated with presence of kidney pathology ( $P = 0.03$ ).<sup>6</sup> Neither of these symptoms proved to be ideal tests for predicting kidney pathology (Table 2).

**Table 2 Diagnostic value of clinical indicators of bladder ( $n = 184$ ) and kidney ( $n = 188$ ) pathology in *Schistosoma haematobium* infected children**

	Frequency in population (%)	Sensitivity	Specificity	PPV*	NPV †
<b>BLADDER PATHOLOGY</b>					
<b>Proteinuria</b>					
Trace value = positive ‡	89	96	18	54	80
Trace value = negative §	52	65	60	62	63
<b>Egg count per 10 ml urine</b>					
> 25 <sup>th</sup> percentile ¶	75	94	17	53	73
> median	50	70	52	59	63
<b>Self-perceived haematuria</b>					
Overall	57	67	52	57	61
Males	65	75	47	57	67
Females	43	50	59	54	55
<b>KIDNEY PATHOLOGY</b>					
<b>Dysuria</b>					
Overall	41	48	66	56	58
Males	56	64	51	54	61
Females	15	21	91	67	56
<b>Fatigue</b>					
	28	35	79	61	58

\* Positive Predictive Value

† Negative Predictive Value

‡ Trace values on the dipstick were considered negative for proteinuria

§ Trace values on the dipstick were considered positive for proteinuria

¶ Test positive if parasite egg count was greater than the lowest quartile of egg count.

|| Test positive if parasite egg count was greater than median egg count.

## DISCUSSION

Urinary schistosomiasis is endemic in much of Africa, yet affected nations do not always treat it as a health priority. One reason for this is the perception that sequelae are mild, since infected people are often able to maintain an active lifestyle. The current investigation provides further evidence that severe urinary tract pathology can result even early in life and that the prevalence of pathology can be high, with half of the infected schoolchildren assessed in this study manifesting some form of bladder abnormality and over a third developing moderate to severe kidney dilation. As schistosomiasis control efforts currently focus on decreasing morbidity,<sup>2,3</sup>

tests that aid diagnosis of presence or degree of morbidity are essential for targeting treatment and tracking progress of control campaigns.

Intensity of infection has been correlated with extent of bladder lesions and hydronephrosis in a number of studies.<sup>8,15</sup> Bleeding and release of proteins into the urine is consequential of the damage caused by passage of eggs through the bladder wall, while high proteinuria levels have also been associated with severe renal damage in infected hosts.<sup>16</sup> In our study, proteinuria and intensity of infection were sensitive in identifying individuals with bladder pathology (Table 2). Self-perceived macrohaematuria was less sensitive, although it has the benefit of not requiring reagent strips. Its gender disparity, however, needs to be considered if using this test, with females

requiring additional testing. Although most variables tested were poorly specific, in that a high percentage of those without bladder pathology also had these symptoms, they provide some indication of which groups of patients should be targeted if morbidity control is the goal.

Fatigue and pain upon urination were more likely in patients with kidney pathology.<sup>6</sup> Although not everyone with morbidity suffered such symptoms (low sensitivity), fatigue and dysuria were not common among those without hydronephrosis (high specificity) (Table 2). Therefore, kidney pathology should be suspected when *S. haematobium* infected children exhibit these symptoms.

While the values obtained in this investigation are specific to the population observed, they do suggest trends that are likely to apply on a wider basis. In our study population, proteinuria, high parasite egg counts, and macrohaematuria were more likely in those who developed bladder pathology. Pain upon urination and fatigue were more often reported in those with kidney pathology than those without. We were, however, unable to identify an ideal diagnostic with both high specificity and sensitivity. Although imperfect, such criteria may be useful in identifying children with urinary tract pathology when more technically complex means to assess morbidity cannot be performed. Our findings also underscore the need for further research on other diagnostic candidates that may improve identification of children with urinary tract pathology and thus allow better monitoring of progress towards the goal of reducing *S. haematobium* related morbidity.

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## REFERENCES

1. WHO. Fact Sheet No. 115: Schistosomiasis. 1996;

- (Last accessed at <http://www.who.int/inf-fs/en/fact115.html> on 21 October 2003).
2. WHO. Report of the WHO informal consultation on schistosomiasis control. 1998.
  3. WHA. Schistosomiasis and soil-transmitted helminth infections. Resolution 54.19. 54th World Health Assembly 2001.
  4. Burki A, Tanner M, Burnier E, Schweizer W, Meudt R, Degremont A. Comparison of ultrasonography, intravenous pyelography and cystoscopy in detection of urinary tract lesions due to *Schistosoma haematobium*. *Acta Trop* 1986; **43**(2): 139-51.
  5. Hatz C, Jenkins JM, Meudt R, Abdel-Wahab MF, Tanner M. A review of the literature on the use of ultrasonography in schistosomiasis with special reference to its use in field studies. 1. *Schistosoma haematobium*. *Acta Trop* 1992; **51**(1): 1-14.
  6. Brouwer KC, Ndhlovu PD, Wagatsuma Y, Munatsi A, Shiff CJ. Epidemiological assessment of *Schistosoma haematobium*-induced kidney and bladder pathology in rural Zimbabwe. *Acta Trop* 2003; **85**(3): 339-47.
  7. Deleque P, Picquet M, Shaw DJ, Vercruyse J, Sambou B, Ly A. Morbidity induced by *Schistosoma haematobium* infections, as assessed by ultrasound before and after treatment with praziquantel, in a recently expanded focus (Senegal River basin). *Ann Trop Med Parasitol* 1998; **92**(7): 775-83.
  8. Vester U, Kardorff R, Traore M, et al. Urinary tract morbidity due to *Schistosoma haematobium* infection in Mali. *Kidney Int* 1997; **52**(2): 478-81.
  9. Wagatsuma Y, Aryeetey ME, Sack DA, Morrow RH, Hatz C, Kojima S. Resolution and resurgence of *Schistosoma haematobium*-induced pathology after community-based chemotherapy in Ghana, as detected by ultrasound. *J Infect Dis* 1999; **179**(6): 1515-22.
  10. Ndhlovu P, Cadman H, Gundersen S, et al. Circulating anodic antigen (CAA) levels in different age groups in a Zimbabwean rural community endemic for *Schistosoma haematobium* determined using the magnetic beads antigen-capture enzyme-linked immunoassay. *Am J Trop Med Hyg* 1996; **54**(5): 537-42.
  11. Taylor P, Makura O. Prevalence and distribution of schistosomiasis in Zimbabwe. *Ann Trop Med Parasitol* 1985; **79**(3): 287-99.
  12. Katz N, Chaves A, Pellegrino J. A simple device for quantitative stool thick-smear technique in *Schistosomiasis mansoni*. *Rev Inst Med Trop Sao Paulo* 1972; **14**(6): 397-400.
  13. Mott KE. A reusable polyamide filter for diagnosis of *S. haematobium* infection by urine filtration. *Bull Soc Pathol Exot Filiales* 1983; **76**(1): 101-4.
  14. WHO/TDR. Ultrasound in schistosomiasis. International workshop on the use of ultrasonography in relation to schistosomiasis. *C.E.R.M.E.S: Niamey, Niger* 1996.
  15. Traore M, Traore HA, Kardorff R, et al. The public health significance of urinary schistosomiasis as a cause of morbidity in two districts in Mali. *Am J Trop Med Hyg* 1998; **59**(3): 407-13.
  16. Sobh MA, Moustafa FE, Ramzy RM, Deelder AM, Ghoneim MA. *Schistosoma haematobium*-induced glomerular disease: an experimental study in the golden hamster. *Nephron* 1991; **57**(2): 216-24.