

Clonal Spread of Imipenem-Resistant *Acinetobacter baumannii* among Different Cities of China[∇]

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A total of 342 imipenem-resistant *Acinetobacter baumannii* isolates (IRABs) were collected from 16 Chinese cities. Six predominant clones had spread widely, and four clones were detected in distant hospitals. The majority of the IRABs contain *bla*_{OXA-23}, with *ISAbal* upstream of the gene. These results suggested that clonal spread played an important role in the outbreak of IRABs in China.

A. baumannii is recognized as an increasingly important opportunistic gram-negative pathogen that is frequently associated with nosocomial outbreaks worldwide (21). Carbapenems are useful agents that could combat many severe *Acinetobacter* infections. Resistance to carbapenems is now accumulating, largely through clonal spread (13). A few lineages even achieve an epidemic status, reaching multiple hospitals or countries (6). One mechanism of resistance to carbapenems is due to the carbapenem-hydrolyzing β -lactamases of molecular classes B and D (12). Four of the eight clusters of OXA-type carbapenemases were identified in *A. baumannii*, including clusters OXA-23, OXA-24, OXA-51, and OXA-58 (22). *ISAbal* may have an important role in the expression and transfer of OXA-type carbapenemase genes (17, 20).

In this study, we investigated the prevalence of imipenem-resistant *A. baumannii* isolates (IRABs) in China and characterized the genes of the carbapenemases in IRABs. Three hundred forty-two nonduplicate IRABs were recovered from 16 cities through January 2005 to December 2005 in China (Table 1; Fig. 1). All isolates were identified by the Vitek GNI⁺ card (BioMérieux, Marcy-l'Étoile, France) as members of the *Acinetobacter calcoaceticus*-*A. baumannii* complex. Species identification was confirmed by sequence analyses of the 16S-23S rRNA gene spacer region (4).

The MICs of 10 antimicrobial agents were determined by the agar dilution technique by following the standards of Clinical and Laboratory Standards Institute (5); these agents included imipenem (Merck KGaA, Darmstadt, Germany), meropenem (Sumitomo Pharmaceuticals Co., Ltd., Osaka, Japan), piperacillin (Sigma, Deisenhofen, Germany), ceftazidime (Sigma, Deisenhofen, Germany), cefepime (Sino-American Shanghai Squibb Pharmaceuticals Ltd., Shanghai, China), aztreonam (Sigma, Deisenhofen, Germany), amikacin (Sigma, Deisenhofen, Germany), gentamicin (Sigma, Deisenhofen, Germany), ciprofloxacin (Bayer, Leverkusen, Germany), and minocycline (Sigma, Deisenhofen, Germany). The MICs of

four other agents, ampicillin-sulbactam, cefoperazone-sulbactam, piperacillin-tazobactam, and polymyxin E, were determined by Etest (AB Biodisk, Solna, Sweden) according to the manufacturer's instructions. The results were interpreted according to CLSI 2006 guidelines (5). *Pseudomonas aeruginosa* ATCC 27853 was used as a control. All isolates were found to be resistant to imipenem and meropenem. The rates of resistance to ampicillin-sulbactam, cefoperazone-sulbactam, and minocycline were 68.0%, 54.2%, and 75.9%, respectively. The rate of resistance to polymyxin E was 10.8%, the lowest among the tested agents. The rates of resistance to all other tested antimicrobial agents were more than 90%.

To characterize these isolates genetically, the genomic DNA of all the isolates was digested with *Apa*I and the resultant DNA fragments were subjected to pulsed-field gel electrophoresis (PFGE) as described previously (18). According to the datum-interpreting criteria described by Tenover et al. (19), these *A. baumannii* isolates belonged to 29 distinct clones (Fig. 2). Among them, six clones were dominant, consisting of 303 strains in total (Table 1).

Crude β -lactamase preparations were extracted by the sonication method (25). pI values were determined by the Phast-System electrophoresis system (Amersham Pharmacia Biotech, Piscataway, NJ) according to the manufacturer's instructions. pI reference proteins were stained with Coomassie brilliant blue R-250. The pattern was analyzed by Curve Expert software (version 1.3) developed by Daniel Hyams (free software). OXA-23-producing *A. baumannii* (a collection of our laboratory) was used as the positive control. Metallo- β -lactamase-producing isolates were screened by the imipenem-EDTA double-disk synergy test (10). IMP-4-producing *A. baumannii* and VIM-2-producing *P. aeruginosa* were used as the positive controls. All tested isolates had one band at pI 6.64 and were negative by the imipenem-EDTA double-disk synergy test.

*bla*_{IMP-type}, *bla*_{VIM-type}, *bla*_{SIM-1}, *bla*_{OXA-23-like}, *bla*_{OXA-24-like}, *bla*_{OXA-58-like}, and *bla*_{OXA-51-like} were analyzed by PCR using primers as described previously (9, 11, 20). PCR mapping was performed using the *ISAbal*-OXA-23-like primers or the *ISAbal*-OXA-51-like primers as reported previously (26). The results showed that all the isolates contained *bla*_{OXA-51-like} but were negative for the OXA-24-like, OXA-58-like, SIM-1, IMP-type, or VIM-type gene. Three hundred twenty-two isolates

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TABLE 1. The PFGE clone distribution of 342 clinical isolates among 16 cities

City	Total no. of isolates	No. of isolates of PFGE clone						
		A	B	C	D	E	F	Other
Beijing	5	2						3
Shanghai	30		17	10				3
Guangzhou	17					10		7
Shenyang	15		10					5
Chongqing	4	1						3
Hangzhou	149	17	5	97	24			6
Ningbo	23	21		1				1
Zhoushan	1	1						
Jianxing	7			6				1
Huzhou	8			6				2
Quzhou	8			6				2
Jinhua	17	1			7		8	1
Taizhou	13			11				2
Lishui	21			20				1
Shaoxing	15	15						
Wenzhou	9	4		3				2
Total	342	62	32	160	31	10	8	39

contained the *bla*_{OXA-23-like} gene. PCR with the *ISAbal*-OXA-23-like primers generated a PCR product in 314 IRABs, and sequencing analysis showed that the PCR product contained the OXA-23 gene (the GenBank accession number AJ132105), suggesting that a majority of the clones contain the OXA-23 gene with *ISAbal* upstream of it. PCR with the *ISAbal*-OXA-51-like primers generated a PCR product, which contained the OXA-66 gene (the GenBank accession number AY750909) in 13 strains, as revealed by sequencing analysis. There was a 34-bp nucleotide sequence between *ISAbal* and *bla*_{OXA-23} and a 7-bp nucleotide sequence between *ISAbal* and *bla*_{OXA-66}.

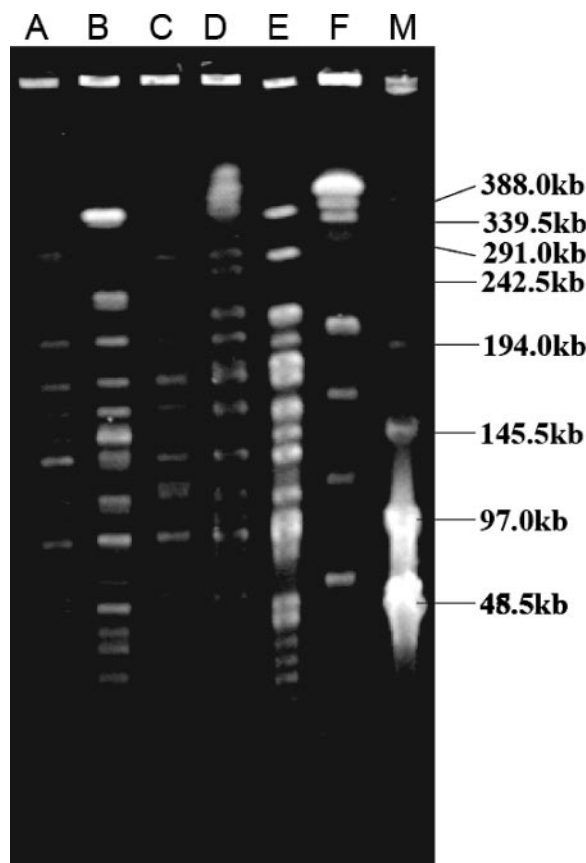


FIG. 2. PFGE analysis of the six major clones of *A. baumannii*. Genomic DNA isolated from the six major *A. baumannii* clones was digested with *Apa*I before PFGE analysis. Lane M, λ ladder used as molecular size marker; lanes A to F, clones A to F, respectively, of *A. baumannii*.



FIG. 1. The geographic distribution of 16 cities in China. On the left, a map of China shows three autonomous cities (Beijing, Chongqing, and Shanghai) and three provincial cities (Hangzhou [Zhejiang], Guangzhou [Guangdong], and Shenyang [Liaoning]), in which *Acinetobacter baumannii* isolates were collected. On the right, a map shows the 11 cities, including Hangzhou, in Zhejiang Province.

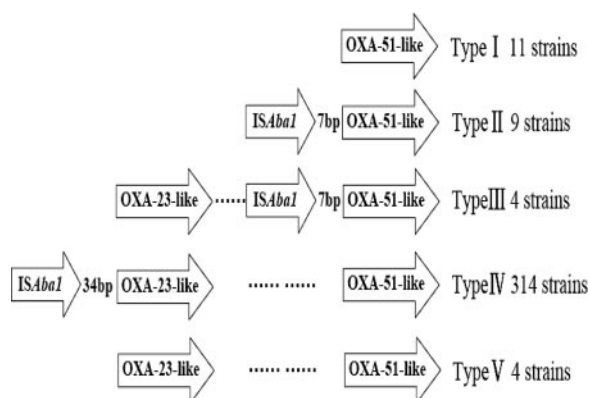


FIG. 3. PCR mapping of $bla_{OXA-51-like}$, $bla_{OXA-23-like}$, and $ISAbal$ in IRABs. According to the PCR mapping results, 342 strains were divided into five types: type I, consisting of 11 strains, contains $bla_{OXA-51-like}$ without the upstream $ISAbal$; type II, consisting of 9 strains, contains $bla_{OXA-51-like}$ with the upstream $ISAbal$; type III, consisting of 4 strains, contains both $bla_{OXA-51-like}$ with the upstream $ISAbal$ and $bla_{OXA-23-like}$ without the upstream $ISAbal$; type IV, consisting of 314 strains, contains $bla_{OXA-51-like}$ without $ISAbal$ and $bla_{OXA-23-like}$ with $ISAbal$; and type V, consisting of 4 strains, contains $bla_{OXA-51-like}$ and $bla_{OXA-23-like}$ without $ISAbal$ upstream of either gene.

According to the PCR mapping, the 342 strains belonged to five types (Fig. 3).

Attempts to transfer bla_{OXA-23} by conjugation using the rifampin-resistant *Escherichia coli* 600 as the recipient failed. Repeated attempts to detect the presence of plasmids capable of hybridizing with the bla_{OXA-23} and bla_{OXA-66} probes also failed. ApaI-digested PFGE DNAs were transferred to nylon membranes and were hybridized with a v-DCTP (DuPont Corporation)-labeled bla_{OXA-23} gene probe. The fragments of the ApaI-digested PFGE DNA of pulsotypes A, B, and C showed hybridization bands at about >220 kb in clone A, about >300 kb in clone B, and about 220 kb in clone C, respectively (data not shown). These results are indicative of a chromosomal location for the bla_{OXA-23} gene.

Our study revealed that IRABs have resistance to most antimicrobial agents. Some studies have indicated that colistin may be useful for treating infections caused by carbapenem-resistant pathogens (14). We confirmed that in vitro, the most active agent against these resistant isolates was polymyxin E.

This is the first report to investigate IRABs in China on such a large scale. We found that clones A, B, C, and D were the dominant isolates and had spread widely among hospitals, cities, and even remote areas. Some isolates with the same genetic basis spread in many hospitals in China, suggesting that the spread of isolates plays an important role in the increase of IRABs. So, it is important to monitor and control the spread of *A. baumannii* having resistance to most β -lactams.

In China, bla_{OXA-23} and $bla_{OXA-51-like}$ were the most popular carbapenemase genotypes. OXA-23-producing IRABs spread widely in the world (1, 2, 7, 9, 16, 23, 24). $bla_{OXA-51-like}$ genes are the endogenous genes of *A. baumannii*, and their protein products have weak carbapenem-hydrolyzing activities (3, 8). An insertion of $ISAbal$ may provide a promoter for the OXA-type carbapenemase genes to increase their expression (20). There were only 15 isolates (4.39%) that lacked $ISAbal$ up-

TABLE 2. The distribution of carbapenemase genes in the major PFGE clones of IRABs

Type	No. of strains of clone							No. of strains
	A	B	C	D	E	F	Other	
I	1		2				8	11
II	3		4				2	9
III							4	4
IV	57	32	152	31	10	8	24	314
V	1		2				1	4
Total (no. of affected cities)	62 (8)	32 (3)	160 (9)	31 (2)	10 (1)	8 (1)	39 (14)	342 (16)

stream of OXA-type carbapenemase coding genes, suggesting that an insertion of $ISAbal$ may be a major factor responsible for the carbapenem resistance.

All isolates belonging to clones B, D, E, and F had a PCR mapping of type IV. In contrast, isolates of clones A and C and sporadic isolates had complex PCR mapping results (Table 2). In the process of the dissemination, the strains may have acquired or lost different gene elements under different antimicrobial pressures (15).

Nucleotide sequence accession numbers. Sequences for the 34 bp between $ISAbal$ and bla_{OXA-23} and the 7 bp between $ISAbal$ and bla_{OXA-66} have been submitted to GenBank under accession numbers DQ923478 (26) and DQ923479, respectively.

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REFERENCES

- Akalin, H., C. Ozakin, and S. Gedikoglu. 2006. Epidemiology of *Acinetobacter baumannii* in a university hospital in Turkey. *Infect. Control Hosp. Epidemiol.* **27**:404–408.
- Boo, T. W., F. Walsh, and B. Crowley. 2006. First report of OXA-23 carbapenemase in clinical isolates of *Acinetobacter* species in the Irish Republic. *J. Antimicrob. Chemother.* **58**:1101–1102.
- Brown, S., H. K. Young, and S. G. Amyes. 2005. Characterisation of OXA-51, a novel class D carbapenemase found in genetically unrelated clinical strains of *Acinetobacter baumannii* from Argentina. *Clin. Microbiol. Infect.* **11**:15–23.
- Chang, H. C., Y. F. Wei, L. Dijkshoorn, M. Vaneechoutte, C. T. Tang, and T. C. Chang. 2005. Species-level identification of isolates of the *Acinetobacter calcoaceticus*-*Acinetobacter baumannii* complex by sequence analysis of the 16S-23S rRNA gene spacer region. *J. Clin. Microbiol.* **43**:1632–1639.
- Clinical and Laboratory Standards Institute. 2006. Performance standards for antimicrobial susceptibility testing; 16th informational supplement. CLSI document M100-S16. Clinical and Laboratory Standards Institute, Wayne, PA.
- Coelho, J. M., J. F. Turton, M. E. Kaufmann, J. Glover, N. Woodford, M. Warner, M. F. Palepou, R. Pike, T. L. Pitt, B. C. Patel, and D. M. Livermore. 2006. Occurrence of carbapenem-resistant *Acinetobacter baumannii* clones at multiple hospitals in London and southeast England. *J. Clin. Microbiol.* **44**:3623–3627.
- Dalla-Costa, L. M., J. M. Coelho, H. A. Souza, M. E. Castro, C. J. Stier, K. L. Bragagnolo, A. Rea-Neto, S. R. Penteado-Filho, D. M. Livermore, and N. Woodford. 2003. Outbreak of carbapenem-resistant *Acinetobacter baumannii* producing the OXA-23 enzyme in Curitiba, Brazil. *J. Clin. Microbiol.* **41**:3403–3406.
- Héritier, C., L. Poirel, P. E. Fournier, J. M. Claverie, D. Raoult, and P. Nordmann. 2005. Characterization of the naturally occurring oxacillinase of *Acinetobacter baumannii*. *Antimicrob. Agents Chemother.* **49**:4174–4179.
- Jeon, B. C., S. H. Jeong, I. K. Bae, S. B. Kwon, K. Lee, D. Young, J. H. Lee, J. S. Song, and S. H. Lee. 2005. Investigation of a nosocomial outbreak of imipenem-resistant *Acinetobacter baumannii* producing the OXA-23 beta-lactamase in Korea. *J. Clin. Microbiol.* **43**:2241–2245.
- Lee, K., Y. S. Lim, D. Yong, J. H. Yum, and Y. Chong. 2003. Evaluation of

- the Hodge test and the imipenem-EDTA double-disk synergy test for differentiating metallo- β -lactamase-producing isolates of *Pseudomonas* spp. and *Acinetobacter* spp. *J. Clin. Microbiol.* **41**:4623–4629.
11. Lee, K., J. H. Yum, D. Yong, H. M. Lee, H. D. Kim, J. D. Docquier, G. M. Rossolini, and Y. Chong. 2005. Novel acquired metallo- β -lactamase gene, *bla*_{SIM-1}, in a class 1 integron from *Acinetobacter baumannii* clinical isolates from Korea. *Antimicrob. Agents Chemother.* **49**:4485–4491.
 12. Livermore, D. M. 2002. The impact of carbapenemases on antimicrobial development and therapy. *Curr. Opin. Investig. Drugs* **3**:218–224.
 13. Livermore, D. M., and N. Woodford. 2006. The beta-lactamase threat in *Enterobacteriaceae*, *Pseudomonas* and *Acinetobacter*. *Trends Microbiol.* **14**: 413–420.
 14. Nordmann, P., and L. Poirel. 2002. Emerging carbapenemases in gram-negative aerobes. *Clin. Microbiol. Infect.* **8**:321–331.
 15. Peleg, A. Y., C. Franklin, J. M. Bell, and D. W. Spelman. 2006. Emergence of carbapenem resistance in *Acinetobacter baumannii* recovered from blood cultures in Australia. *Infect. Control Hosp. Epidemiol.* **27**:759–761.
 16. Poirel, L., and P. Nordmann. 2006. Carbapenem resistance in *Acinetobacter baumannii*: mechanisms and epidemiology. *Clin. Microbiol. Infect.* **12**:826–836.
 17. Poirel, L., and P. Nordmann. 2006. Genetic structures at the origin of acquisition and expression of the carbapenem-hydrolyzing oxacillinase gene *bla*_{OXA-58} in *Acinetobacter baumannii*. *Antimicrob. Agents Chemother.* **50**: 1442–1448.
 18. Seifert, H., and P. Gerner-Smidt. 1995. Comparison of ribotyping and pulsed-field gel electrophoresis for molecular typing of *Acinetobacter* isolates. *J. Clin. Microbiol.* **33**:1402–1407.
 19. Tenover, F. C., R. D. Arbeit, R. V. Goering, P. A. Mickelsen, B. E. Murray, D. H. Persing, and B. Swaminathan. 1995. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J. Clin. Microbiol.* **33**:2233–2239.
 20. Turton, J. F., M. E. Ward, N. Woodford, M. E. Kaufmann, R. Pike, D. M. Livermore, and T. L. Pitt. 2006. The role of ISAbal in expression of OXA carbapenemase genes in *Acinetobacter baumannii*. *FEMS Microbiol. Lett.* **258**:72–77.
 21. Villegas, M. V., and A. I. Hartstein. 2003. *Acinetobacter* outbreaks, 1977–2000. *Infect. Control Hosp. Epidemiol.* **24**:284–295.
 22. Walther-Rasmussen, J., and N. Hoiby. 2006. OXA-type carbapenemases. *J. Antimicrob. Chemother.* **57**:373–383.
 23. Wilks, M., A. Wilson, S. Warwick, E. Price, D. Kennedy, A. Ely, and M. R. Millar. 2006. Control of an outbreak of multidrug-resistant *Acinetobacter baumannii-calcoaceticus* colonization and infection in an intensive care unit (ICU) without closing the ICU or placing patients in isolation. *Infect. Control Hosp. Epidemiol.* **27**:654–658.
 24. Ying, C. M., T. K. Ling, C. C. Lee, and J. M. Ling. 2006. Characterization of carbapenem-resistant *Acinetobacter baumannii* in Shanghai and Hong Kong. *J. Med. Microbiol.* **55**:799–802.
 25. Yu, Y. S., Q. Yang, X. W. Xu, H. S. Kong, G. Y. Xu, and B. Y. Zhong. 2004. Typing and characterization of carbapenem-resistant *Acinetobacter calcoaceticus-baumannii* complex in a Chinese hospital. *J. Med. Microbiol.* **53**:653–656.
 26. Zhou, H., B. R. Pi, Q. Yang, Y. S. Yu, Y. G. Chen, L. J. Li, and S. S. Zheng. 2007. Dissemination of imipenem-resistant *Acinetobacter baumannii* strains carrying the ISAbal-*bla*_{OXA-23} genes in a Chinese hospital. *J. Med. Microbiol.* **56**:1076–1080.