

# Reduced Loading of Intracellular $\text{Ca}^{2+}$ Stores and Downregulation of Capacitative $\text{Ca}^{2+}$ Influx In Bcl-2–Overexpressing Cells

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**Abstract.** The mechanism of action of the oncogene *bcl-2*, a key regulator of the apoptotic process, is still debated. We have employed organelle-targeted chimeras of the  $\text{Ca}^{2+}$ -sensitive photoprotein, aequorin, to investigate in detail the effect of Bcl-2 overexpression on intracellular  $\text{Ca}^{2+}$  homeostasis. In the ER and the Golgi apparatus, Bcl-2 overexpression increases the  $\text{Ca}^{2+}$  leak (while leaving  $\text{Ca}^{2+}$  accumulation unaffected), hence reducing the steady-state  $[\text{Ca}^{2+}]$  levels. As a direct consequence, the  $[\text{Ca}^{2+}]$  increases caused by inositol 1,4,5 trisphosphate (IP3)-generating agonists were reduced

in amplitude in both the cytosol and the mitochondria. Bcl-2 overexpression also reduced the rate of  $\text{Ca}^{2+}$  influx activated by  $\text{Ca}^{2+}$  store depletion, possibly by an adaptive downregulation of this pathway. By interfering with  $\text{Ca}^{2+}$ -dependent events at multiple intracellular sites, these effects of Bcl-2 on intracellular  $\text{Ca}^{2+}$  homeostasis may contribute to the protective role of this oncogene against programmed cell death.

**Key words:** calcium • organelles • oncogene • aequorin • cell signaling

## Introduction

In the past years, Bcl-2, the mammalian homologue of CED-9, the antiapoptotic gene initially described in *Caenorhabditis elegans*, has emerged as a major checkpoint controlling the escape of cells from their death fate (Kroemer, 1997). In mammals, Bcl-2 is a member of a broad family of related gene products, with various degrees of sequence identity and different functions. While Bcl-2 and some members of the family (e.g., Bcl-x) protect the cells from apoptosis, others (e.g., Bax) have the opposite effect. However, while there is a general consensus on the role of Bcl-2, its mechanism of action remains elusive. On one hand, the various members of the family have been shown to dimerize and to interact with cofactors of caspases, key executors of the apoptotic machinery (Li and Yuan, 1999); on the other, the specific localization of the protein on the ER and outer mitochondrial membrane (Lithgow et al., 1994) suggests that these proteins may interfere with some

specific functions localized in these organelles. In particular, Bcl-2 has been shown to prevent the release of cytochrome c and AIF from mitochondria (Kluck et al., 1997; Yang et al., 1997), events that are considered essential in triggering apoptosis, whereas expression of Bax has opposite effects (Jurgensmeier et al., 1998).

Among the various functions occurring on ER and mitochondrial membranes, ion fluxes are recognized to play major roles. Thus, the intriguing possibility has emerged that members of the Bcl-2 family affect the apoptotic process by modulating ion fluxes in these organelles (Minn et al., 1997; Schendel et al., 1998). In particular, recent work in cell clones stably expressing the oncogene suggests that Bcl-2 causes an overload of  $\text{Ca}^{2+}$  within the ER (Lam et al., 1994; He et al., 1997; Kuo et al., 1998). However, this effect appears in contrast with the observation that Bcl-2 insertion in artificial lipid bilayers results in the formation of ionic channels of low selectivity (Minn et al., 1997). Such property should in fact cause a reduction in the ER  $\text{Ca}^{2+}$  content (due to an increased leak) rather than excess accumulation. In this report, we have undertaken a detailed investigation of  $\text{Ca}^{2+}$  homeostasis at the

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subcellular level using the recombinant aequorin (AEQ)<sup>1</sup> approach developed in our laboratory (Rizzuto et al., 1992; De Giorgi et al., 1996). We conclude that Bcl-2 overexpression affects Ca<sup>2+</sup> handling by reducing the state of filling of the intracellular Ca<sup>2+</sup> stores. The implications of these results for the protective action of Bcl-2 on apoptosis are discussed.

## Materials and Methods

### Cell Culture and Transfection

HeLa cells were grown in DME supplemented with 10% FCS in 75-cm<sup>2</sup> Falcon flasks. Before transfection, cells were seeded onto 13-mm glass coverslips and allowed to grow to 50% confluence. At this stage, transfection with 4 μg of DNA (3 μg Bcl-2 + 1 μg AEQ) was carried out as previously described (Rizzuto et al., 1995a) and AEQ measurements or immunocytochemistry were performed 36 h after transfection. For fura-2 measurements, the cells were seeded onto 24-mm coverslips and transfection was carried out with 8 μg plasmid DNA (6 μg Bcl-2 + 2 μg mtGFP).

### Aequorin Measurements

For cytosolic AEQ (cytAEQ) and mitochondrial AEQ (mtAEQ), the coverslip with the cells was incubated with 5 μM coelenterazine for 1–2 h in DME supplemented with 1% FCS, and then transferred to the perfusion chamber. For reconstituting with high efficiency, the AEQ chimeras targeted to the Golgi apparatus and the ER (GoAEQ and erAEQ, respectively), the luminal [Ca<sup>2+</sup>] of these compartments first must be reduced. This was obtained by incubating the cells for 1 h at 4°C in KRB (Krebs-Ringer modified buffer: 125 mM NaCl, 5 mM KCl, 1 mM Na<sub>3</sub>PO<sub>4</sub>, 1 mM MgSO<sub>4</sub>, 5.5 mM glucose, and 20 mM Hepes, pH 7.4, at 37°C) supplemented with coelenterazine 5 μM, the Ca<sup>2+</sup> ionophore ionomycin, and 600 μM EGTA. After this incubation, the cells were extensively washed with KRB supplemented with 2% BSA and 1 mM EGTA, and then AEQ reconstitution was carried out as described.

All AEQ measurements were carried out in KRB. Agonists and other drugs were added to the same medium. In experiments with permeabilized cells, a buffer mimicking the cytosolic ionic composition (intracellular buffer, IB: 140 mM KCl, 10 mM NaCl, 1 mM K<sub>3</sub>PO<sub>4</sub>, 5.5 mM glucose, 2 mM MgSO<sub>4</sub>, 1 mM ATP, 2 mM sodium succinate, 20 mM Hepes, pH 7.05, at 37°C) was employed. This solution was supplemented with either 100 μM EGTA or a fixed [Ca<sup>2+</sup>] (buffered with 2 mM EGTA). The experiments were terminated by lysing the cells with 100 μM digitonin in a hypotonic Ca<sup>2+</sup>-rich solution (10 mM CaCl<sub>2</sub> in H<sub>2</sub>O), thus discharging the remaining AEQ pool. The light signal was collected and calibrated into [Ca<sup>2+</sup>] values as previously described (Brini et al., 1995; Rizzuto et al., 1995a). In brief, a 13-mm round coverslip with the transfected cells was placed in a perfused, thermostated chamber located in the close proximity of a low-noise photomultiplier, with built-in amplifier discriminator. The output of the discriminator was captured by a Thorn-EMI photon counting board and stored in an IBM-compatible computer for further analyses. The AEQ luminescence data were calibrated off-line into [Ca<sup>2+</sup>] values, using a computer algorithm based on the Ca<sup>2+</sup> response curve of wild-type and mutant AEQs, as previously described (Brini et al., 1995; Barrero et al., 1997).

In some experiments using erAEQ, after reaching the plateau level, an artifactual slow decrease in [Ca<sup>2+</sup>] was observed. Such an artifact is due to a small fraction of missorted AEQ in a low [Ca<sup>2+</sup>] compartment and a simple practical cure for it has been described by Maechler et al. (1999).

In the experiment of Fig. 4, cells transfected with erAEQ were used to determine the steady [Ca<sup>2+</sup>]<sub>er</sub> obtained by incubating the cells in low Ca<sup>2+</sup> medium. Two protocols were adopted that gave identical results. In the first, cells incubated in normal medium were depleted according to the

standard protocol, but the Ca<sup>2+</sup> concentration during the refilling was reduced to 100 μM. In the second protocol, the cells were preincubated in 100 μM CaCl<sub>2</sub> for 16 h, depleted, and finally refilled with an extracellular Ca<sup>2+</sup> concentration of 100 μM.

### Fura-2 Measurements

The coverslip with the cells was incubated with 5 μM fura-2 (added to DME + 1% FCS) at 37°C for 30 min, and after a brief washout was then transferred to the thermostated stage of a Zeiss Axiovert inverted microscope equipped with a Sutter filterwheel and 340/380 excitation filters. The fluorescence data were collected with a Princeton Instruments back-illuminated camera and expressed as emission ratios, using Metafluor software (Universal Imaging). Within the microscope field, transfected and untransfected cells were identified before carrying out the fura-2 monitoring by revealing mitochondrial-targeted green fluorescent protein (mtGFP) fluorescence (excitation 480 nm, emission 510 nm).

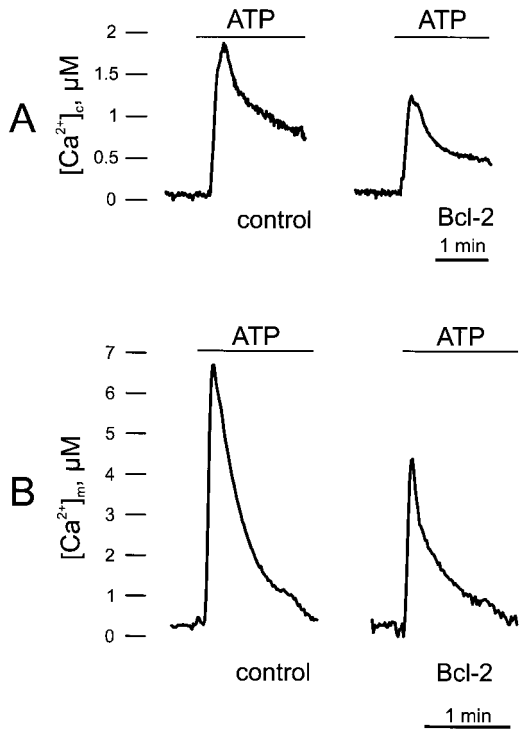
## Results and Discussion

To directly investigate the role of Bcl-2 in Ca<sup>2+</sup> homeostasis, we have selectively measured the [Ca<sup>2+</sup>] in different cell compartments, i.e., the cytoplasm and the organelles acting as sources (ER and Golgi apparatus) or targets (mitochondria) of the Ca<sup>2+</sup> signal. Targeted AEQ chimeras developed in our lab (Rizzuto et al., 1992; Brini et al., 1995; Montero et al., 1995; Pinton et al., 1998) were employed. This approach has two major advantages. The first is the high subcellular specificity of the probes, which allows to clearly identify the effects of Bcl-2 expression on the signaling properties of the various compartments; the second is that the Ca<sup>2+</sup> probe can be cotransfected with Bcl-2. We and others have previously shown that in transient cotransfection, the two recombinant proteins are expressed by the same subset of cells, thus avoiding the risks associated with the use of stable clones or single cells (Brini et al., 1995).

The effect of Bcl-2 expression on the cytosolic Ca<sup>2+</sup> signal elicited by an agonist, ATP, acting on receptors coupled, through Gq proteins, to the production of inositol 1,4,5 trisphosphate (IP<sub>3</sub>) was first investigated. In the experiment shown in Fig. 1, HeLa cells, either coexpressing Bcl-2 and cytAEQ (Bcl-2-transfected) or expressing only cytAEQ (controls) were challenged with ATP, which acts on a Gq-coupled P<sub>2</sub>Y receptor. Both in control and Bcl-2-transfected cells, ATP stimulation causes a rapid rise in cytoplasmic Ca<sup>2+</sup> concentration ([Ca<sup>2+</sup>]<sub>c</sub>), followed by a gradually declining sustained plateau. In Bcl-2-transfected cells, the [Ca<sup>2+</sup>] increases evoked by stimulation with ATP are significantly smaller than in controls (peak amplitude 1.53 ± 0.22 vs. 2.06 ± 0.13 μM, *n* = 10; Fig. 1 A).

Since Bcl-2 is localized in the mitochondria also, and thus could, at least in principle, enhance Ca<sup>2+</sup> uptake in this organelle, a simple explanation for this result would be that cytosolic Ca<sup>2+</sup> is more rapidly cleared by mitochondria, acting as an immobile buffer, placed in closed proximity to IP<sub>3</sub>-gated channels. Fig. 1 B, however, shows that this is not the case. Indeed, Bcl-2 overexpression does not increase, but rather reduces, the mitochondrial [Ca<sup>2+</sup>] rise induced by ATP (peak amplitude 6.56 ± 0.72 in controls vs. 4.42 ± 0.46 μM in Bcl-2-expressing cells; *n* = 10). Interestingly, the reduction caused by Bcl-2 overexpression appears larger in the mitochondria than in the cytosol, both in absolute terms and as percent (26 vs. 33% reduction, respectively). While not responsible for the alteration

<sup>1</sup>Abbreviations used in this paper: AEQ, aequorin; cytAEQ, cytosolic AEQ; erAEQ, AEQ chimera targeted to the ER; GFP, green fluorescent protein; GoAEQ, AEQ chimera targeted to the Golgi apparatus; IB, intracellular buffer; IP<sub>3</sub>, inositol 1,4,5 trisphosphate; KRB, Krebs-Ringer modified buffer; mtAEQ, mitochondrial AEQ; mtGFP, mitochondrially targeted GFP; SERCA, sarcoendoplasmic reticulum Ca<sup>2+</sup> ATPases; tBuBHQ, 2,5-di-(*tert*-butyl)-1,4-benzohydroquinone.



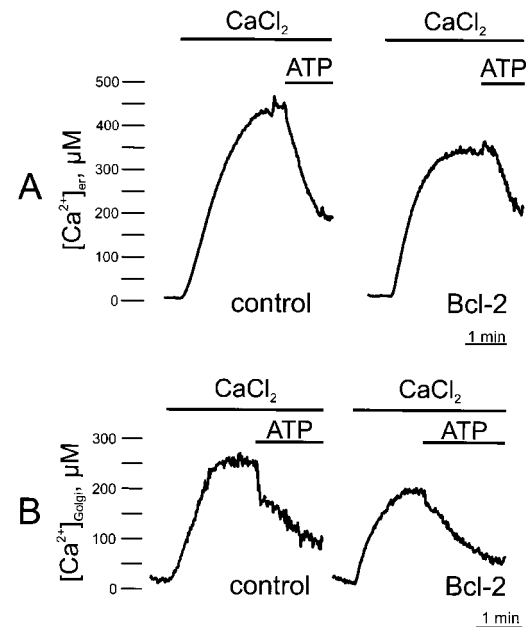
**Figure 1** Cytosolic (A) and mitochondrial (B)  $\text{Ca}^{2+}$  homeostasis in control and Bcl-2-overexpressing HeLa cells. Parallel batches of HeLa cells were either cotransfected with the appropriate AEQ chimera (cytAEQ and mtAEQ for monitoring the cytoplasm and the mitochondria, respectively) and Bcl-2 (Bcl-2), or transfected with the AEQ chimera alone (control). 36 h after transfection, the measurement of AEQ luminescence was carried out and calibrated into  $[\text{Ca}^{2+}]$  values, as described in Materials and Methods. Where indicated, the cells, perfused with KRB, were challenged with  $100 \mu\text{M}$  ATP added to the same buffer. These and the following traces are representative of ten experiments that gave similar results.

of the cytosolic  $\text{Ca}^{2+}$  signal, the effect of Bcl-2 on mitochondrial  $\text{Ca}^{2+}$  homeostasis appears of major functional relevance, given that key events occurring in the mitochondrial matrix, such as the stimulation of ATP production (Jouaville et al., 1999) and possibly the opening of the permeability transition pore, PTP (Bernardi, 1999), are regulated by  $[\text{Ca}^{2+}]_m$ .

A second possible explanation for the reduction in the agonist-dependent  $[\text{Ca}^{2+}]_c$  (and  $[\text{Ca}^{2+}]_m$ ) increases was next considered, i.e., a decrease in the amount of  $\text{Ca}^{2+}$  released by the agonist-sensitive  $\text{Ca}^{2+}$  stores. To investigate this possibility, the  $[\text{Ca}^{2+}]$  in the lumen of the ER was measured in cells cotransfected with Bcl-2 and an erAEQ chimera (Montero et al., 1995). In addition, since we recently have demonstrated that the Golgi apparatus shares many of the  $\text{Ca}^{2+}$  homeostatic properties of the ER (Pinton et al., 1998), this latter organelle was also investigated. Fig. 2 shows the calibrated  $[\text{Ca}^{2+}]$  values in the two compartments. To obtain reliable quantitative estimates of the  $[\text{Ca}^{2+}]$  in the lumen of these two organelles, their  $[\text{Ca}^{2+}]$  needs to be decreased during both the reconstitution of AEQ with coelenterazine and the subsequent initial phase of perfusion with KRB/EGTA in the luminometer (see

Materials and Methods). Under those conditions, the  $[\text{Ca}^{2+}]$  was  $<10 \mu\text{M}$  in both organelles. When the  $[\text{Ca}^{2+}]$  in the perfusion medium was switched to  $1 \text{ mM}$ , the  $[\text{Ca}^{2+}]$  in the lumen of the two compartments gradually increased, reaching in control cells plateau values of  $452 \pm 46 \mu\text{M}$  ( $n = 7$ ) in the ER and  $262 \pm 47 \mu\text{M}$  ( $n = 7$ ) in the Golgi apparatus, respectively. In Bcl-2 transfected cells, lower steady state values were attained in both compartments ( $304 \pm 38 \mu\text{M}$ ,  $n = 7$  for the ER, and  $186 \pm 34 \mu\text{M}$ ,  $n = 7$  for the Golgi apparatus, respectively). When the cells were treated with ATP, rapid decreases in the  $[\text{Ca}^{2+}]$  of the two compartments were observed, both in control and Bcl-2-expressing cells. In both organelles, the decrease of  $[\text{Ca}^{2+}]$  was larger and faster in controls compared with Bcl-2-expressing cells, reflecting the higher filling state and thus the more rapid flow through the IP<sub>3</sub>-gated channels.

The data reported above differ substantially from those reported in WEHI7.2 clones, where Bcl-2 overexpression results in an augmentation of the ER  $\text{Ca}^{2+}$  content (Lam et al., 1994; He et al., 1997). This discrepancy may be due to the use of transient vs. stable transfections, in which the results might reflect unique properties of the selected clone and/or the long-term expression of a foreign gene may result in adaptation phenomena that are only indirect consequences of the expressed protein. This interpretation might also explain the results obtained by Kuo and coworkers in clones of MCF10A cells overexpressing



**Figure 2.**  $\text{Ca}^{2+}$  homeostasis in the lumen of the ER (A) and of the Golgi apparatus (B) in control and Bcl-2-overexpressing HeLa cells. Parallel batches of HeLa cells were either cotransfected with the appropriate AEQ chimera (erAEQ and GoAEQ for monitoring the lumen of the ER and of the Golgi apparatus, respectively) and Bcl-2 (Bcl-2), or transfected with the AEQ chimera alone (control). 36 h after transfection, the organelles were depleted of  $\text{Ca}^{2+}$  to optimize AEQ reconstitution. After reconstitution, the cells were transferred to the luminometer chamber and AEQ luminescence was collected and calibrated into  $[\text{Ca}^{2+}]$  values. Where indicated, the cells were stimulated with  $100 \mu\text{M}$  ATP. All other conditions are as in Fig. 1.

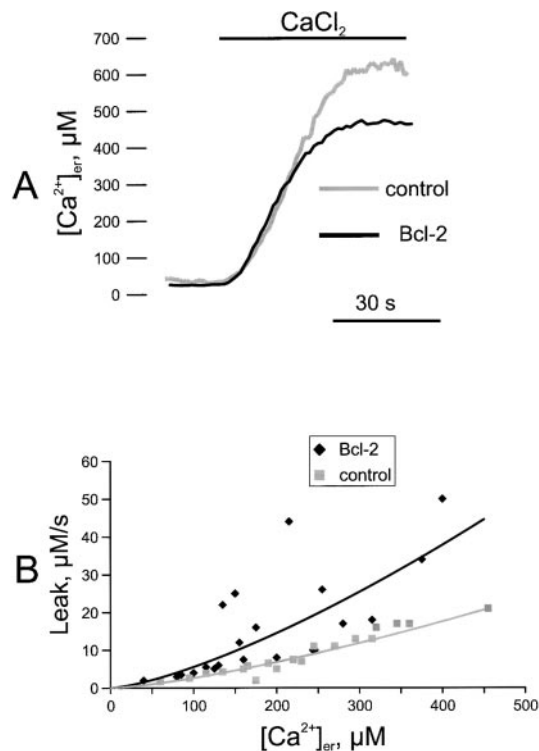
Bcl-2, in which an increase in SERCA2 (sarcoendoplasmic reticulum  $\text{Ca}^{2+}$  ATPases 2) protein leads to accelerated  $\text{Ca}^{2+}$  uptake and enhanced  $\text{Ca}^{2+}$  loading (Kuo et al., 1998). On the other hand, it is possible that, although the duration of these experiments is compatible with the time course of the apoptotic process, the events observed in transient experiments could include only some of the relevant effects of Bcl-2 on the mechanisms controlling programmed cell death.

But how does Bcl-2 affect ER and Golgi  $\text{Ca}^{2+}$  handling? Given that the steady-state  $[\text{Ca}^{2+}]$  in both organelles depends on the equilibrium between active accumulation and passive leak, the effect of Bcl-2 could be on either process. On one hand, Bcl-2 could reduce  $\text{Ca}^{2+}$  uptake by the SERCA, either by a direct effect on the pump or by reducing the resting cytosolic  $[\text{Ca}^{2+}]$ . On the other, it could increase the passive diffusion of  $\text{Ca}^{2+}$  from the ER. To clarify this critical issue, we investigated these possibilities independently.

At first, the possibility that Bcl-2 affected primarily cytosolic  $[\text{Ca}^{2+}]$  (and hence  $[\text{Ca}^{2+}]_{\text{er}}$  only indirectly) was verified. Given that AEQ is not accurate enough to reveal small differences of  $[\text{Ca}^{2+}] \sim 10^{-7}$  M, the resting  $[\text{Ca}^{2+}]_{\text{c}}$  was measured with the fluorescent indicator fura-2 (Grynkiewicz et al., 1985). In this experiment, to identify Bcl-2-overexpressing cells in single-cell imaging experiments, the cells were cotransfected with mitochondrially targeted GFP, mtGFP(S65T) (Rizzuto et al., 1995b). GFP-positive cells were distinguished from controls by the typical fluorescence emitted upon illumination with blue light. No difference in the resting cytoplasmic  $[\text{Ca}^{2+}]$  between Bcl-2-transfected and control cells was observed (data not shown).

Then, we investigated whether Bcl-2 overexpression had any direct effect on the activity of the SERCA pump. For this purpose, the kinetics of  $\text{Ca}^{2+}$  accumulation were studied in permeabilized cells (Fig. 3 A). After reconstituting the photoprotein as in the experiment of Fig. 2, Bcl-2-transfected and control cells were transferred to the luminometer chamber and perfused with a buffer (IB, see Materials and Methods) mimicking intracellular ionic conditions, supplemented with 100  $\mu\text{M}$  EGTA (IB/EGTA). After permeabilization with 100  $\mu\text{M}$  digitonin added to the same medium and wash in IB/EGTA, the medium was switched to IB, containing 200 nM  $[\text{Ca}^{2+}]$  (IB/ $\text{Ca}^{2+}$ ). Under those conditions,  $[\text{Ca}^{2+}]_{\text{er}}$  gradually increased, reaching, in control cells, a plateau value of  $728 \pm 60$ ,  $n = 5$ . In Bcl-2-transfected cells, the maximal rate of  $\text{Ca}^{2+}$  accumulation was the same as in control cells ( $13 \pm 2$  vs.  $12 \pm 3$   $\mu\text{M}/\text{s}$ ,  $n = 5$ ), arguing that the  $\text{Ca}^{2+}$  pumping activity is not modified. However, the steady-state value eventually attained was distinctly lower ( $504 \pm 52$   $\mu\text{M}$ ,  $n = 5$ ). Therefore, these data exclude that the alteration of ER  $\text{Ca}^{2+}$  homeostasis is due to a reduction in the activity of the SERCAs or to soluble cytosolic factors (that are released into the medium after permeabilization with digitonin). Rather, by demonstrating a difference only when ER refilling approaches the high  $[\text{Ca}^{2+}]_{\text{er}}$  values of intact resting cells, i.e., in conditions in which passive leak becomes significant, they suggest that Bcl-2 acts on the latter process.

We sought direct evidence for this interpretation by measuring the  $\text{Ca}^{2+}$  leak rate from the ER of Bcl-2-express-

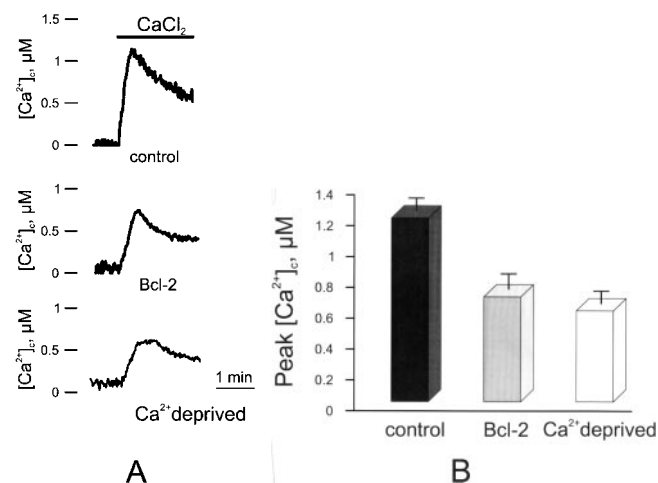


**Figure 3.** Kinetics of ER  $\text{Ca}^{2+}$  uptake and release in control and Bcl-2-transfected cells. **A**, ER  $\text{Ca}^{2+}$  refilling in permeabilized cells. Parallel batches of HeLa cells were either cotransfected with erAEQmut and Bcl-2 (Bcl-2), or transfected with erAEQmut alone (control). 36 h after transfection, depletion of  $\text{Ca}^{2+}$  stores and AEQ were carried out as in Fig. 2. The coverslip with the cells was then transferred to the luminometer chamber. The perfusion medium was changed to IB/EGTA, and permeabilization was carried out by perfusing 100  $\mu\text{M}$  digitonin (added to the same medium) for 1 min. After a 2-min wash with IB/EGTA,  $\text{Ca}^{2+}$  accumulation in the ER was initiated by switching the medium to IB containing a buffered  $\text{Ca}^{2+}$  concentration of 0.2  $\mu\text{M}$  (see text for details). AEQ calibration was carried out as described in Materials and Methods. **B**, Dependence on  $[\text{Ca}^{2+}]_{\text{er}}$  of the  $\text{Ca}^{2+}$  leak rate from the ER. Transfection, depletion of  $\text{Ca}^{2+}$  stores, and AEQ reconstitution were carried out as in A, then the coverslip with the cells was transferred to the luminometer and perfused with KRB/ $\text{Ca}^{2+}$  until the steady-state  $[\text{Ca}^{2+}]_{\text{er}}$  was reached.  $\text{Ca}^{2+}$  release was initiated by treating the cells with 50  $\mu\text{M}$  tBuBHQ. Based on the experimental trace, the maximal rates of  $\text{Ca}^{2+}$  release (measured from the first derivative) at different values of  $[\text{Ca}^{2+}]_{\text{er}}$  were calculated and plotted for Bcl-2-transfected and control cells. The plot contains data obtained from ten independent experiments. In each experiment, duplicate samples of control and Bcl-2-overexpressing cells were measured. Due to the mixing time in the luminometer chamber, the kinetics of  $[\text{Ca}^{2+}]_{\text{er}}$  decrease are sigmoidal and the maximal rate is obtained 2–3 s after addition of tBuBHQ. Accordingly, we considered the maximal rates the best approximation for the initial rate of  $[\text{Ca}^{2+}]_{\text{er}}$  decrease. The fitting of the curve shown in B was performed using Microsoft Excel software.

ing and control cells. After the depletion protocol, the ER was first refilled by exposing the cells to extracellular  $[\text{Ca}^{2+}]$  ranging from 0.2–3 mM, thus resulting in different levels of steady-state  $[\text{Ca}^{2+}]_{\text{er}}$ . The SERCA blocker, 2,5-di-(*tert*-butyl)-1,4-benzohydroquinone (tBuBHQ), was

then added that initiates the release of stored  $\text{Ca}^{2+}$ . Given that, by definition, the rates of  $\text{Ca}^{2+}$  uptake and leak in steady state are equal, the rate of  $[\text{Ca}^{2+}]_{\text{er}}$  decrease upon blockade of the SERCA must reflect the rate of  $\text{Ca}^{2+}$  cycling across the ER membrane and, thus, of the leak rate at any given steady-state  $[\text{Ca}^{2+}]_{\text{er}}$ . Fig. 3 B shows the relationship between  $[\text{Ca}^{2+}]_{\text{er}}$  and leak rate, which can be fitted by a power equation. In Bcl-2-overexpressing cells, the data are more scattered, but for each  $[\text{Ca}^{2+}]_{\text{er}}$  value, the rate of  $\text{Ca}^{2+}$  efflux is faster than in control cells. This result is consistent with the evidence that Bcl-2 can induce cation channels of low selectivity in artificial lipid bilayers (Minn et al., 1997).

Previously, we have demonstrated that a reduction of  $\sim 30\%$  in steady-state  $[\text{Ca}^{2+}]_{\text{er}}$  results in a substantial activation ( $>50\%$ ) of the so-called capacitative  $\text{Ca}^{2+}$  influx (Hofer et al., 1998). The reduction in steady-state  $[\text{Ca}^{2+}]_{\text{er}}$  caused by Bcl-2 overexpression is thus expected to cause an activation of this pathway. However, indirect evidence (namely, the observation that the resting cytosolic  $\text{Ca}^{2+}$  level is indistinguishable from controls) suggests that this is not the case. To further investigate the problem, we measured the capacitative  $\text{Ca}^{2+}$  influx in Bcl-2-transfected and control HeLa cells expressing cytAEQ (Fig. 4). The cells were treated with the SERCA blocker ( $10 \mu\text{M}$  tBuBHQ) while perfused with  $\text{Ca}^{2+}$ -free medium (KRB/EGTA). This procedure first evoked a small transient increase in  $[\text{Ca}^{2+}]_{\text{c}}$  due to release of  $\text{Ca}^{2+}$  from the stores. When the release of stored  $\text{Ca}^{2+}$  was complete ( $\sim 3$  min),  $\text{Ca}^{2+}$  was readded to the medium. This maneuver evoked a second, larger  $[\text{Ca}^{2+}]_{\text{c}}$  increase via the capacitative pathway, which was markedly lower in Bcl-2-transfected than in controls cells ( $1.19 \pm 0.11$  in controls vs.  $0.68 \pm 0.13 \mu\text{M}$  in Bcl-2-expressing cells,  $n = 6$ ). The question then arises as to the mechanism(s) through which Bcl-2 downregulates capacitative  $\text{Ca}^{2+}$  influx. Two possibilities were considered: one was that the reduction of this  $\text{Ca}^{2+}$  influx pathway is an adaptive mechanism to the long lasting reduction in steady-state  $[\text{Ca}^{2+}]_{\text{er}}$ ; the second was that the reduction is a direct consequence of Bcl-2 overexpression. To distinguish between these two possibilities, we verified whether a long-term reduction of  $[\text{Ca}^{2+}]_{\text{er}}$ , obtained independently of Bcl-2, could cause a comparable downregulation of the capacitative  $\text{Ca}^{2+}$  influx. Capacitative  $\text{Ca}^{2+}$  influx was thus compared in controls and in cells whose  $[\text{Ca}^{2+}]_{\text{er}}$  was reduced to values similar to those of Bcl-2-overexpressing cells by prolonged incubation with a low ( $100 \mu\text{M}$ ) extracellular  $[\text{Ca}^{2+}]$ . In cells that had been maintained at low external  $[\text{Ca}^{2+}]$ , and thus experienced a long-term reduction in the resting  $[\text{Ca}^{2+}]_{\text{er}}$  levels, the  $[\text{Ca}^{2+}]_{\text{c}}$  peak evoked by activation of the capacitative  $\text{Ca}^{2+}$  influx was markedly lower than in controls ( $0.59 \pm 0.11 \mu\text{M}$ ,  $n = 5$ ). Of interest, the level of steady-state  $[\text{Ca}^{2+}]_{\text{er}}$  in cells incubated in low extracellular  $\text{Ca}^{2+}$  was very similar ( $269 \pm 46 \mu\text{M}$ ,  $n = 4$ ) to that measured in Bcl-2-overexpressing cells incubated in normal ( $1 \text{ mM}$ ) extracellular  $\text{Ca}^{2+}$  concentration. The simplest explanation of these results is that the decrease in capacitative  $\text{Ca}^{2+}$  influx is an adaptive consequence to the prolonged reduction in steady-state  $[\text{Ca}^{2+}]_{\text{er}}$ , though, at the moment, it cannot be excluded that other factors (e.g., a reduction in plasma membrane potential) contribute to this effect. This reduction in capacitative



**Figure 4.** Capacitative  $\text{Ca}^{2+}$  influx in control, Bcl-2-transfected, and  $\text{Ca}^{2+}$ -deprived cells. A, Representative trace. B, Average  $[\text{Ca}^{2+}]_{\text{c}}$  peak. Parallel batches of HeLa cells were either cotransfected with cytAEQ and Bcl-2 (Bcl-2), or transfected with cytAEQ alone (control and  $\text{Ca}^{2+}$ -deprived). In the  $\text{Ca}^{2+}$ -deprived cells, the cells were maintained in the 18 h preceding the experiment in KRB containing a lower ( $0.1 \text{ mM}$ )  $\text{CaCl}_2$  concentration (KRB/low $\text{Ca}^{2+}$ ). After transferring the coverslip with the cells to the luminometer chamber, the cells, perfused with KRB/EGTA, were challenged with  $10 \mu\text{M}$  tBuBHQ (a specific blocker of the  $\text{Ca}^{2+}$  ATPase present in the ER and Golgi apparatus) added to the same medium. After 2 min, capacitative  $\text{Ca}^{2+}$  entry was initiated by changing the medium to KRB/ $\text{Ca}^{2+}$  ( $\text{CaCl}_2$ ). All other conditions as in Fig. 1.

$\text{Ca}^{2+}$  influx, on one hand, appears critical to maintain a resting  $[\text{Ca}^{2+}]_{\text{c}}$  indistinguishable from that of control cells, despite a major reduction of  $[\text{Ca}^{2+}]$  in the ER; on the other hand, it could account, at least in part, for a higher resistance of cells expressing Bcl-2 to apoptotic cell death, as the reduction of this ubiquitous  $\text{Ca}^{2+}$  influx pathway may prevent the overload of  $\text{Ca}^{2+}$ , considered one of the key events in apoptosis. In agreement with this view, Ma and coworkers recently demonstrated that SERCA overexpression, by causing ER  $\text{Ca}^{2+}$  overload, increases spontaneous apoptosis (Ma et al., 1999). Finally, our data are not in contrast with previous results showing that the SERCA inhibitor, thapsigargin, activates programmed cell death (Bian et al., 1997). In this latter case, in fact, the  $\text{Ca}^{2+}$  depletion is acute (1–2 min) and complete, whereas in Bcl-2-transfected cells, the drop in  $[\text{Ca}^{2+}]_{\text{er}}$  is modest and develops slowly. Thus, thapsigargin not only causes a massive activation of the capacitative  $\text{Ca}^{2+}$  influx pathway, but also such a drastic reduction in the level of  $[\text{Ca}^{2+}]_{\text{er}}$  to interfere with the basic activity of ER chaperonins.

We have demonstrated that overexpression of Bcl-2 causes a reduction of the steady-state  $[\text{Ca}^{2+}]$  levels, both in the ER and in the Golgi apparatus, the two main agonist-sensitive  $\text{Ca}^{2+}$  stores. We also showed that this alteration affects the  $\text{Ca}^{2+}$  signal to which the effector systems are exposed, in particular those located in local environments highly dependent on the filling state of the intracellular stores (such as the mitochondria). The chronic reduction in  $[\text{Ca}^{2+}]$  of the ER results in a downregulation of the ca-

capacitative  $\text{Ca}^{2+}$  influx. The alterations in  $\text{Ca}^{2+}$  handling provoked by Bcl-2 overexpression could be part of the mechanisms through which Bcl-2 exerts its role in the control of cell growth. Targets of Bcl-2 effects on  $\text{Ca}^{2+}$  homeostasis could be: the ER itself, where sorting and trafficking of proteins, e.g., enzymes or membrane receptors, are modulated by luminal  $[\text{Ca}^{2+}]$  (Booth and Koch, 1989); the mitochondria, where  $\text{Ca}^{2+}$  accumulation regulates the production of ATP (RobbGaspers et al., 1998; the concentration of which is a key determinant of the apoptosis vs. necrosis choice, Leist et al., 1997), and, possibly, the release of the apoptotic cofactor cytochrome c (Krajewski et al., 1999); or the cytoplasm, where bulk or local  $[\text{Ca}^{2+}]$  increases activate enzymatic effectors, such as the  $\text{Ca}^{2+}$ -dependent protease, calpain (Carafoli and Molinari, 1998; Squier et al., 1999), or the phosphatase, calcineurin (Wang et al., 1999). Whether, and how, these routes interact, and how they are affected by the alteration in  $\text{Ca}^{2+}$  homeostasis caused by Bcl-2, awaits further investigation.

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