Increase in Transduction Efficiency of Tn551 Mediated by the Methicillin Resistance Marker

BRIGITTE BERGER-BÄCHI

Institute of Medical Microbiology, University of Zurich, Zurich, Switzerland

Received 2 June 1982/Accepted 26 January 1983

The transduction efficiency of Tn551, a staphylococcal transposon coding for erythromycin resistance (*ermB*), was increased by a factor of about five in acceptor strains carrying the gene for methicillin resistance (*mec*), as compared with methicillin-sensitive strains. This enhancement was independent of the generalized transducing phage used and of the chromosomal insertion site of the Tn551 transposon.

The methicillin resistance gene (mec) in staphylococci has been reported to be chromosomally located (3), probably residing on an extra piece of DNA not present in methicillin-sensitive (Mec^s) strains (7). However, based on its genetic behavior, it might be integrated into a transposable element (2). The methicillin resistance shows an unexpected interaction in promoting the transduction efficiency of the Tn551 transposon. This activity might indicate some recombinational functions encoded or activated by the methicillin resistance gene.

The strains used in this study are described in Table 1. A set of isogenic strains, all derivatives of strain NCTC8325 and varying in just one factor, were constructed to see whether the genetic background of the strain carrying the Tn551 insertion had any influence on transduction efficiency of the transposon. As mediators for erythromycin resistance, the transducing phages (29 and 80α) were grown on strains BB285, BB291, BB293, and BB296. Strains BB285, BB291, and BB293 had the same Tn551 insert. In strain BB296, Tn551 is located at another site. This had been verified by Southern blots (6) with radioactively labeled Tn551 DNA to EcoRI digests of the chromosomal DNA (data not shown). Strain BB291 is a Mec^r derivative of strain BB285; both strains carry the three prophages ϕ 11, ϕ 12, and ϕ 13, whereas strains BB293 and BB296 are phage-free. Three Mec^s strains were used as acceptor strains for the transduction, viz., RN450, a prophage-free strain, FK268, a strain with the three prophages and the penicillinase plasmid pI524, and BB255, which is a plasmid-free derivative of FK268. The two Mec^r acceptor strains, BB262 and BB270, were derivatives of strain BB255.

The transduction frequencies for erythromy-

cin resistance obtained after UV irradiation of the different lysates are shown in Fig. 1. The irradiation time of 10 min corresponded to a reduction of the plaque-forming ability of the transducing lysates by approximately 90% (data not shown). UV irradiation of the transducing lysates increases the frequency of stable transductants for chromosomal genes (1), and a maximum is observed after a 90% reduction of the PFU. This effect was observed also in this model system. There was no decrease of the erythromycin resistance transduction efficiency after 10 min of UV irradiation of the transducing lysate, which reduced the phage viability by 90%. But as a rule, erythromycin-resistant (Em^r) transductants were obtained in higher proportions when the acceptor strain carried a mec determinant. The transducing frequencies with phage 29 were somewhat lower than those with phage 80α , but the effect was the same. The increase in transduction efficiency was independent of the genetic background of the donor. Two strains with different mec determinants and isogenic except for the region of DNA carried along with the *mec* determinant in the transducing phage showed the same increase in transduction efficiency. Where tested, the Mec^s strains RN450, FK268, and BB255 gave similar transducing efficiencies for erythromycin resistance, with no marked increase after the UV irradiation of the phage. The influence of prophages or of the plasmid content of an acceptor strain could therefore be ruled out. The increase of the transduction efficiencies in Mec^r strains was 3to 13-fold, with a mean of about 5-fold (Table 2). The low value for strain BB262 with phage 80α grown on strain BB293 was an exception. The transduction of whole plasmids seemed not to be affected by the mec determinant. Plasmid pBB2,



J. BACTERIOL.



FIG. 1. Transduction frequencies of Tn551. The transducing phages were grown and propagated by the agar overlay method. UV irradiation for 10 min reduced the plaque-forming ability of the transducing lysate by 90%. A 0.3-ml portion of an overnight culture of an acceptor strain was exposed to various dilutions of the irradiated phages for 15 min at 37°C, and the transductants were scored on brain heart infusion plates containing 20 μ g of erythromycin per ml. The phages used were 80 α (A) and 29 (B) grown on strains BB285 (a), BB291 (b), BB293 (c), and BB296 (d). The acceptor strains were the Mec^s strains RN450 (\bigcirc), FK268 (\triangle), and BB255 (\blacksquare) and the Mec^r strains BB270 (\textcircledleft) and BB262 (\blacktriangle).

a 14-kilobase, Tn551-carrying plasmid was transduced with the same efficiency into strain BB270 as into strain BB255 (0.9×10^{-4} and 1.3×10^{-4} , respectively). The increased transduction efficiency was restricted to the transposon Tn551 when chromosomally located. The transduction frequencies of *purA*, a chromosomal marker, into a Mec^r (BB295) or Mec^s (BB359) recipient were identical $(3.1 \times 10^{-6} \text{ and } 3.8 \times 10^{-6})$, respectively). The increased efficiency might be due to an increase in recombination functions either provided by or directed by the *mec* determinant. Further studies will be needed to elucidate the various functions of the methicillin resistance determinant.

TO A D I	T 4	• ••	•	
TABL	JE 1	. Stra	ains	used

Strain	Genotype	Origin		
RN450	Same as for NCTC 8325-4	R. P. Novick (5)		
FK268	Same as for NCTC 8325 (pI524)	S. Cohen (4)		
BB255	Same as for NCTC 8325	This laboratory; FK268 cured from pI524 by ethidium bromide		
BB262	Same as for NCTC 8325, mec	This laboratory, by transduction of strain BB255 with phage 83α -grown on strain E691 <i>mec</i> originally isolated from a patient		
BB270	Same as for NCTC 8325, mec	This laboratory, by transduction of strain BB255 with phage ϕ 11 grown on strain E142 mec originally isolated from a patient		
BB285	Same as for NCTC 8325, Ω2000 (chr::Tn551)	This laboratory; Em ^r Mec ^s derivative of strain BB270, obtained by heat inactivation of a temperature-sensitive plasmid carry- ing Tn551 (pRN3208; R. P. Novick) previously introduced into strain BB270; loss of Mec ^r not yet further characterized		
BB291	Same as for NCTC 8325, Ω2000 (chr::Tn551) mec	This laboratory, by transduction of strain BB270 with phage 80α grown on strain BB285		
BB293	Same as for NCTC 8325-4, Ω2000 (chr::Tn551)	This laboratory, by transduction of strain RN450 with phage 80α grown on strain BB285		
BB296	Same as for NCTC 8325-4, Ω2002 (chr::Tn551)	This laboratory, by heat inactivation of a temperature-sensitive plasmid carrying Tn551 (pRN3208; R. P. Novick) previously introduced in strain RN450		
BB295	Same as for NCTC 8325, mec purA102 nov-142	This laboratory, by transduction of strain BB270 with phage 80α grown on strain ISP86 (NCTC 8325 nov-142 uraA141 hisG15 purA102 pig-131; P. Pattee)		
BB359	Same as for NCTC 8325, purA102 nov-142	This laboratory, by transduction of strain BB255 with phage 80α grown on strain BB295		
BB303	Same as for NCTC 8325-4(pBB2)	This laboratory, by transformation of strain RN450 with the largest <i>Eco</i> RI fragment of φ11de which harbors Tn551		

TABLE 2. Enhancement of the transduction efficiency of Tn551 in Mec^r strains as compared with Mec^s strains

Transducing phage	Acceptor strains (Mec ^r /Mec ^s)	Enhancement valve ^a with the following donor strain for Tn551:			
		BB285	BB291	BB293	BB296
80α	BB270/BB255	3.1	5.2	4.4	3.2
80α	BB262/BB255	5.3	5.4	1.0	4.1
29	BB270/BB255	3.6	4.3	3.5	8.3
29	BB262/BB255	4.5	7.1	4.2	13.8

^a The enhancement value was calculated by dividing the highest transduction frequencies obtained for erythromycin resistance in the Mec^r strains BB270 and BB262 by the frequency obtained in the isogenic Mec^s strain BB255.

ACKNOWLEDGMENTS

I thank F. H. Kayser for comments on the manuscript and stimulating discussions.

This work was supported partially by Swiss National Foundation grant 3.546-0.79.

LITERATURE CITED

- 1. Arber, W. 1960. Transduction of chromosomal genes and episomes in *Escherichia coli*. Virology 11:273-288.
- Cohen, S., H. M. Sweeney, and S. K. Basu. 1977. Mutations of prophage φ11 that impair the transducibility of their *Staphylococcus aureus* lysogens for methicillin resistance. J. Bacteriol. 129:237-245.
- 3. Kuhl, S. A., P. A. Pattee, and J. N. Baldwin. 1978.

Chromosomal map location of the methicillin resistance determinant in *Staphylococcus aureus*. J. Bacteriol. 135: 460-465.

- Murphy, E., and R. P. Novick. 1979. Physical mapping of Staphylococcus aureus penicillinase plasmid pI524: characterization of an invertible region. Mol. Gen. Genet. 175:19-30.
- Novick, R. P. 1967. Properties of a cryptic high frequency transducing phage in *Staphylococcus aureus*. Virology 33:155-166.
- Southern, E. M. 1975. Detection of specific sequences among DNA fragments separated by gel electrophoresis. J. Mol. Biol. 98:503-517.
- Stewart, G. C., and E. D. Rosenblum. 1980. Genetic behavior of the methicillin resistance determinant in *Staph*ylococcus aureus. J. Bacteriol. 144:1200-1202.