MICROCALORIMETRIC STUDIES ON THE FORMATION OF MAGNESIUM COMPLEXES OF ADENINE NUCLEOTIDES

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Abstract.-Values for the thermodynamic quantities $(\Delta F, \Delta H, \Delta S)$ in reactions in which complexes of adenine nucleotides with magnesium ion $(ATPMg^{--}, ADPMg^{-}, AMPMg)$ are formed have been obtained by a microcalorimetric technique by using an isothermic Calvet's apparatus.

Experimental values measured at ionic strength $\mu = 0.2$ indicate that complex formation reactions are driven by the entropic factor and that stability of complexes increases with length of the phosphate chain.

Most important reactions in the field of bioenergetics involve the participation of magnesium complexes of adenine nucleotides. Much work has been devoted to the determination of the thermodynamic quantities of these complexes. Yet there is a great disparity in the results available, *vide*, in a recent review by R. C. Phillips et al.¹

We report the use of ^a microcalorimetric technique which permits direct determinations of the enthalpy of these complex formation reactions, as well as estimations of the apparent equilibrium constants.

Materials and Methods.--Microcalorimeter: The apparatus used was constructed by the "Institut de Microcalorimétrie et de thermogénèse" CNRS, Marseilles, France, according to the Calvet principle (cf. Calvet and Prat).² This microcalorimeter is an isothermic differential heat fluxmeter, the sensitivity of which is 2.9μ watt per millimeter recorder deflection. The experimental device is shown in Figure 1. Figure 2 exhibits the artifact observed when ¹ ml of buffer contained in the siphon is injected into the pyrex calorimetric cell containing 5 ml of the same buffer. The beginning of the recording shows the constant stirring heat flow.

Experimental procedure: Five ml of triethanolamine buffer containing appropriate concentrations of tetramethylammonium chloride and of magnesium ion are pipetted into the pyrex calorimetric cell. The empty siphon is held by a ground-glass stopper on this cell. The cell is put into the calorimeter for temperature stabilization for about 5 hr. After this, ¹ ml of adenine nucleotide solution which has been equilibrated with buffer of the same pH and ionic strength is injected into the siphon. The experiment can be started by pumping the nucleotide solution from the siphon into the calorimetric cell when the recording has come back to the base line (about one hr after filling the siphon). Pumping time is ten min.

All experiments were made at 30° C, pH 8.5, and at ionic strength 0.2 maintained by the electrolyte support (tetramethylammonium chloride), the concentration of which varies with the concentration of nucleotides and of magnesium ion. The concentration of TEA buffer is constant and equal to $0.15 M$.

Chemicals: The sodium salts of adenosine triphosphate (ATP) and of adenosine diphosphate (ADP) (chromatographically pure) were purchased from Schwarz BioResearch Inc., and the acid form of adenosine monophosphate "puriss grade" (AMP) from Fluka. Sodium ion was not eliminated because of its low affinity for adenine nucleotides³ and its low concentration with respect to the electrolyte support.

FIG. 1.-Experimental device used for the calorimetric determination, (A) Pyrex calori-(B) Helix stirrer in Teflon. (C) Siphon: (1) tube for reactant introduction; (2) tube for the equilibration of the pressure between the siphon and the exterior; (3) tube for the equilibration of the pressure between the calorimetric cell and the exterior; and (4) access through the siphon for the stirrer axis. (D) Thermocouples surrounding the calorimetric cell.

Tetramethylammonium chloride and TEA were purchased from Eastman Kodak and Merck, respectively. Magnesium sul $fate$ (RP) was supplied by Prolabo.

Calculation: All calculations were made with a small computer (Programma 101 Olivetti).

Results.-Principle of calculation: At pH 8.5 adenine nucleotides are almost entirely dissociated, i.e., present as ATP⁴⁻, ADP³⁻, and AMP²⁻, and we can consider that, as a first approximation, the quantity which is not dissociated is negligible. For simplicity we shall call A: adenine nucleotide, M: magnesium ion, and A.M: complex.

The reaction studied is:

$$
A + M \rightleftarrows A.M
$$

If a and b are the initial concentrations of adenine nucleotide and magnesium ion respectively, and x the concentration of complex at equilibrium, the apparent formation constant is:

$$
K_{\rm app}=\frac{x}{(a-x)(b-x)}\qquad (1)
$$

When this reaction occurs at constant temperature and pressure in a calorim eter of utilizable volume (V) the heat evolved during the reaction is:

$$
Q_1 = x \cdot \Delta H^\circ \cdot V \tag{2}
$$

where ΔH° is the enthalpy of the complex formation reaction. If a second experiment is done under the same conditions, but with a different initial concentration (c) of magnesium ion, the concentration of complex at equilibrium will be y , and

$$
K_{\rm app} = \frac{y}{(a-y)(c-y)}\tag{3}
$$

and

$$
Q_2 = y \cdot \Delta H^{\circ} \cdot V \tag{4}
$$

From the relationships 1, 2, 3, and 4, we obtain:

$$
\Delta H^{\circ} = \frac{(c-b) Q_1 Q_2 \pm \sqrt{\Delta}}{2(Q_1 ac - Q_2 ab) V}
$$
(5)

FIG. 2.---Artifacts produced by the stirring (AB), by the introduction of ¹ ml of buffer into the siphon (CD) and by the injection of ¹ ml of buffer into 5 ml of the same buffer from the siphon to the $\overline{50}$ B \overline{c} $\overline{10}$ calorimetric cell (EF).

with $\Delta = Q_1^2 \cdot Q_2^2 (b^2 - 2bc + c^2) - 4(Q_1ac - Q_2ab) (Q_1 Q_2^2 - Q_2Q_1^2).$

Experimental calorimetric determinations of Q_1 and Q_2 permit estimation of ΔH° , and the knowledge of ΔH° provides K_{app} from the relationships (1) and (2) or (3) and (4).

Experimental results.—(a) $ATPMg^{2-}$ complex: (1) ΔH° measurement. Assuming that, at the pH considered, ATP is entirely dissociated, the reaction of complex formation is: $\text{ATP}^{4-} + \text{Mg}^{2+} \rightleftharpoons \text{ATPMg}^{2-}$. In all experiments, we have chosen the adenosine triphosphate concentration (a) equal to $2.5 \times 10^{-3} M$; the magnesium ion concentration varies with the experiment; for Q_1 measurement we have $b = a$, and for Q_2 , $c = 2a$. Figure 3 shows a thermogram of the $ATPMg²$ complex formation reaction. The area between the time axis and the curve represents the quantity of heat evolved during the reaction. The Q_1 and Q_2 values shown in Table 1 were obtained by integration of thermograms. The ΔH° obtained from equation (5) is equal to 4.16 kcal/mole. In reality, this value of ΔH° is only an apparent value, termed ΔH° _{app} because, at pH 8.5, all ATP is not entirely dissociated and the true reaction scheme is:

$$
H^{+} + ATP^{4-} + Mg^{2+} \rightleftharpoons ATPMg^{2-} + H^{+}
$$

||
ATPH³⁻ + Mg²⁺ \rightleftharpoons ATPHMg⁻.

At ionic strength 0.2 and a temperature of 25° C. Phillips *et al.*¹ 4 </sup> give

FIG. 3.—Thermogram of the formation of ATPMg²⁻complex formation. The dotted line represents the injection and the stabilization of 1 ml of ATP solution in the siphon (in this case the stabilisation is endother mic). The pumping of ATP solution begins from point A.

$$
pK_{ATPH1} = 7.05
$$
\n
$$
pK_{ATPHMg} = 5.26.
$$
\n
$$
\frac{4}{3}f(w^{\text{watt}})
$$
\n
$$
= 5.26.
$$

 Q_1 and Q_2 experimental values of ATPMg²⁻ complex formation reaction. These values are corrected for the heat of dilution of the reactants and for the heat of injection artifact.

Correction of ΔH° takes into account the neutralization heat of protons which are liberated during the reaction and the ATPHMg- complex formation.

Let us call: a , the total concentration of nucleotides before the reaction; α , the concentration of ATP⁴⁻ at equilibrium; α' , the concentration of ATPH³⁻ at equilibrium; x, the concentration of ATPMg^{2-} at equilibrium; x', the concentration of ATPHMg⁻ at equilibrium; ΔH° ², the enthalpy of the complex ATPMg²⁻ formation reaction; $\Delta H^{\circ}{}_{A^-}$, the enthalpy of the complex ATPMg⁻ formation reaction; $(ATPH³⁻ + Mg²⁺ \rightleftharpoons ATPHMg⁻ + 2.2 kcal/mole accord$ ing to the method of Phillips et al.); ΔH° , the enthalpy of ionization of ATPH³⁻ $(-1.68 \text{ kcal/mole}:$ Phillips et al.); ΔH° _n, the enthalpy of ionization of the triethanolamine buffer 7.5 kcal/mole (our experimental determination).

The concentration of undissociated protons before reaction at any alkaline pH is equal to ATPH³⁻ concentration, hence

$$
\frac{[ATP^{4-}]}{[ATPH^{3-}]} = \frac{10^{-pK} \text{ATPH}^{-3}}{10^{-pH}}.
$$
 (6)

At pH 8.5 [ATPH³⁻] = [ATP⁴⁻]/28 and [ATPH³⁻] = $a/29$. When equilibrium is reached, equation (6) becomes:

$$
\frac{\alpha}{\alpha'} = 10^{-p} K_{ATPH^{-s}} / 10^{-p}^{\text{H}}
$$
 (7)

and

$$
x/x' = 10^{-pK} \text{ATPHMg}^{-}/10^{-pH}.
$$
 (8)

At pH 8.5, the indissociated proton concentration is

[ATPH³⁻] + [ATPHMg⁻] =
$$
\alpha' + x' = \frac{\alpha}{28} + \frac{x}{1740}
$$
.

The quantity (z) of neutralized protons all of which arise from ATPH^{3-} is

$$
z = \left(\frac{a}{29} - \frac{\alpha}{28} - \frac{x}{1740}\right) V = \left(\frac{x}{29} - \frac{0.016x}{29}\right) V.
$$

and the neutralization heat (Q_n) evolved during the reaction is

$$
Q_n = z(\Delta H^\circ{}_p - \Delta H^\circ{}_n)
$$

$$
Q_n = V\left(\frac{x}{29} - \frac{0.016x}{29}\right)(\Delta H^\circ{}_p - \Delta H^\circ{}_n).
$$

With this correction equation (2) becomes

$$
Q = V[x\Delta H^{\circ}{}_{A^{\circ}}{}^{-} + x'\Delta H^{\circ}{}_{A^{\circ}}{} + z(\Delta H^{\circ}{}_{p} - \Delta H^{\circ}{}_{n})]
$$

and

$$
Q = xV \bigg[\Delta H_{A^{+-}} + \frac{\Delta H^{\circ}_{A^{-}}}{1740} + \bigg(\frac{1}{29} - \frac{0.016}{29}\bigg) (\Delta H^{\circ}_{p} - \Delta H^{\circ}_{n}) \bigg]. \tag{9}
$$

Since $\Delta H^{\circ}{}_{A^{2-}}$, $\Delta H^{\circ}{}_{A^{-}}$, $\Delta H^{\circ}{}_{n}$, and $\Delta H^{\circ}{}_{n}$ are constant, the measured heat is always proportional to the concentration of the $ATPMg^{2-}$ complex.

We can now estimate the enthalpy of ATPMg²⁻ formation, i.e., the true ΔH° , using equation (9) (in this case only the positive root is meaningful)

$$
\Delta H^{\circ}{}_{\rm app} = \Delta H^{\circ}{}_{A^{\leftarrow}} + \frac{\Delta H^{\circ}{}_{A^{\leftarrow}}}{1740} + \left(\frac{1}{29} - \frac{0.016}{29}\right) (\Delta H^{\circ}{}_{p} - \Delta H^{\circ}{}_{n}) \tag{10}
$$

As $\Delta H^{\circ}{}_{A^-}$ is of the same order of magnitude as $\Delta H^{\circ}{}_{A^{\rightarrow}}$, the term $\Delta H^{\circ}{}_{A^-}/1740$ is negligible with respect to $\Delta H^{\circ}{}_{A^{2-}}$, and the $\Delta H^{\circ}{}_{A^{2-}}$ value, corrected for neutralization, is 4.47 kcal/mole. Yet we see in equation (10) that term 0.016/29 $(\Delta H^{\circ}{}_{n}-\Delta H^{\circ}{}_{n})$ is also negligible and equation (10) can be simplified to

$$
\Delta H^{\circ}{}_{\text{app}} = \Delta H^{\circ}{}_{A^{\text{2}}\text{}} + \frac{1}{29} \left(\Delta H^{\circ}{}_{p} - \Delta H^{\circ}{}_{n} \right) \tag{11}
$$

The $\Delta H^{\circ}{}_{A^{2-}}$ value in this equation is also equal to 4.47 kcal/mole, the difference between values of ΔH° at obtained from equation (10) or (11) being only of the order of 0.005 kcal/mole.

(2) Equilibrium constant measurement. For measurement of the apparent equilibrium constant at $\mu = 0.2$ we have to use ΔH° _{app} and the relationships 1 and 2, or 3 and 4. The result obtained by this means is K_{app} measured $(K_{\text{app}}_{\text{meas.}})$ = 47.100 M⁻¹. This value, like that of ΔH° , must be corrected for ATPH³⁻ and ATPHMg⁻ complex formation. For ionic strength 0.2, the true K_{app} is:

$$
K_{\rm app} = \frac{[ATPMg^{2-}]}{[ATP^{4-}] \cdot [Mg^{2+}]} \tag{12}
$$

If b_i is the total Mg concentration, at equilibrium we have:

$$
[\text{Mg}^{2+}] = b_t - x - x'
$$
 (13)

and

$$
[ATP^{4-}] = a - \alpha' - x - x'
$$
 (14)

Replacing α' and x' by their values as a function of α and x (equations (7) and (8)), we have from (12), (13), and (14)

$$
K_{\rm app} = \frac{x(1 + 10^{(\rm pK_{\rm ATPH} - \rm pH)})}{[b_t - x (1 + 10^{(\rm pK_{\rm ATPH} - \rm pH)})][a - x (1 + 10^{(\rm pK_{\rm ATPH} - \rm pH)})]} \quad (15)
$$

From equation (15) $K_{\text{app}} = 49.300 M^{-1}$

(b) $ADPMg^-$ complex: In this case $b = a, c = 2a$, and $a = 4.10^{-3} M$. Table shows values of Q_1 and Q_2 for ADPMg⁻ complex formation reactions. From 2 shows values of Q_1 and Q_2 for ADPMg⁻ complex formation reactions. these values we have $\Delta H^{\circ}{}_{\text{app}} = 2.99 \text{ kcal/mole}.$

Using $pK_{ADPHMg} = 5.31$ (ref. 1), $pK_{ADPH2} = 6.78$ (ref. 4), and ΔH° ionization of ADPH²⁻ = -1.37 kcal/mole (ref. 1) equations analogous to the preceding equations give the thermodynamic quantities of the $ADPMg$ ⁻ complex formation reaction. True $\Delta H^{\circ} = 3.15$ kcal/mole, K_{ann} mess = 4.720 M^{-1} , and K_{ann} = 4.950 M^{-1} .

(c) AMPMg complex: In this case $a = b$, $c = 8a$, and $a = 4.10^{-3}$ M.

The complex formation reaction is simpler than the two preceding reactions because only one species of complex has to be considered

$$
H^{+} + AMP^{2-} + Mg^{2+} \rightleftharpoons AMPMg + H^{+}
$$

||
AMPH-

and equation (15) in this case becomes

$$
K_{\rm app} = \frac{x(1 + 10^{(\rm pK_{\rm AMPH}-\rm pH)})}{(b_t - x)(a - x)}
$$
(16)

Table 3 shows values and means of Q_1 and Q_2 for the AMPMg complex formation reaction. With pK_{AMPH} ⁻ = 6.47 (ref. 4) and ΔH of AMPH⁻ ionization equal to -0.85 kcal/mole⁵ and using equations (11) and (16), we find that the thermodynamic quantities of the AMPMg complex formation reaction are: ΔH° _{app} = 1.70 kcal/mole; ΔH° = 1.78 kcal/mole; $K_{\text{app meas}}$ = 63 M^{-1} ; and $K_{\rm apo} = 64 M^{-1}.$

Discussion.-The main purpose in microcalorimetric experiments is to measure the true ΔH° of reactions. In our case we see that ΔH° increases with the length of phosphate chain (Fig. 4). These results do not agree with those of Burton⁶ (5 kcal/mole ATPMg²⁻ and 6 kcal/mole ADPMg⁻) and of Taqui Khan and Martel⁹ (3.4 kcal/mole for ATPMg) who obtained ΔH° by the Vant'Hoff

TABLE 2.

 Q_1 and Q_2 experimental values of ADPMg⁻ complex formation reaction. Values corrected as for Table I.

TABLE 3.

 Q_1 and Q_2 experimental values of AMPMg complex formation reaction. Values corrected as for Tables ¹ and 2.

TABLE 4

Thermodynamic data for the magnesium complex formation reactions of ATP⁴⁻, ADP³⁻, and AMP²⁻ at 30^oC and for an ionic strength of 0.2.

relationship but at an ionic strength of 0.1. increases with the length of phosphate chain is in good agreement with the work of Phillips et al.' The latter investigators, determining ΔH° of $ADPMg$ ⁻ and $ATPMg$ ²⁻ complex formation by the Vant'Hoff relationship, concluded that it was necessary to convert the apparent equilibrium constant (K_{app}) into the true equilibrium constant (K_0) , i.e., the constant at infinite dilution. Indeed, the Vant'Hoff relationship gives 3.6 kcal/mole and 3.3 kcal/mole, respectively, for the ΔH of ADP and ATP magnesium complex formation reactions at ionic strength 0.1, if Log K_{app} is plotted versus $1/T$, and 4.3 kcal/mole and 5.1 kcal/ mole, respectively, for the same reactions if Log K_0 is plotted versus $1/T$. However, these last values are about 1 kcal/mole higher than ours.

Secondly, equation (9) demonstrates that complex concentration is linearly proportional to the heat evolved during the reaction. This fact permits the determination of K_{app} for experimental ionic strength by enthalpic titration. The values obtained for ATPMg²⁻ and

However, the fact that ΔH°

FIG. 4.-Experimental data demonstrating that the ΔH° values (enthalpy of the complex formation reactions) are proportional to the length of the phosphate chain.

 $ADPMg$ ⁻ formation by our method are in good agreement with the most probable values in the literature^{1, $6-9$} for an ionic strength of about 0.2. Our $K_{\rm ann}$ of AMPMg is close to that of Taqui Khan et $al.^{10}$ and of Smith and Alberty.¹¹ With K_{ann} and ΔH° , the entropy and the energy variation of reactions studied can be estimated by:

$$
\Delta F = -RT \text{ Log } K_{\text{app}} \text{ and } \Delta S = \frac{\Delta H - \Delta F}{T}
$$

Table 4 shows the respective values of ΔF and ΔS , demonstrating that complex formation is driven by the entropic factor; it also shows a good correlation between AS and the length of phosphate chain. This fact can probably be correlated with the rearrangement of H₂O molecules during the complex formation reaction. Finally it should be pointed out that our K_{app} values can be corrected to K_0 according to the results of Phillips *et al.*⁴ These corrected K_{app} values will be published elsewhere.

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