

Amantadine-induced conformational and dynamical changes of the influenza M2 transmembrane proton channel

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The M2 protein of influenza A virus forms a transmembrane proton channel important for viral infection and replication. Amantadine blocks this channel, thus inhibiting viral replication. Elucidating the high-resolution structure of the M2 protein and its change upon amantadine binding is crucial for designing antiviral drugs to combat the growing resistance of influenza A viruses against amantadine. We used magic-angle-spinning solid-state NMR to determine the conformation and dynamics of the transmembrane domain of the protein M2TMP in the apo- and amantadine-bound states in lipid bilayers. ¹³C chemical shifts and torsion angles of the protein in 1,2-dilauroyl-*sn*-glycero-3-phosphatidylcholine (DLPC) bilayers indicate that M2TMP is α -helical in both states, but the average conformation differs subtly, especially at the G34–I35 linkage and V27 side chain. In the liquid-crystalline membrane, the complexed M2TMP shows dramatically narrower lines than the apo peptide. Analysis of the homogeneous and inhomogeneous line widths indicates that the apo-M2TMP undergoes significant microsecond-time scale motion, and amantadine binding alters the motional rates, causing line-narrowing. Amantadine also reduces the conformational heterogeneity of specific residues, including the G34/I35 pair and several side chains. Finally, amantadine causes the helical segment N-terminal to G34 to increase its tilt angle by 3°, and the G34–I35 torsion angles cause a kink of 5° in the amantadine-bound helix. These data indicate that amantadine affects the M2 proton channel mainly by changing the distribution and exchange rates among multiple low-energy conformations and only subtly alters the average conformation and orientation. Amantadine-resistant mutations thus may arise from binding-incompetent changes in the conformational equilibrium.

high-resolution structure | membrane protein | solid-state NMR | conformational heterogeneity | chemical-shift perturbation

The M2 protein of the influenza A virus forms a membrane-bound proton channel that acidifies the endosomally trapped virus, which triggers the release of the viral RNA into the infected cell, initiating viral replication (1, 2). The cationic amine amantadine inhibits viral replication by blocking this proton channel and thus has been used for the prophylaxis and treatment of influenza A infections (3, 4). However, in the last few years, amantadine resistance has skyrocketed among influenza A viruses in Asia and North America (5), making it imperative to develop alternative antiviral drugs.

The M2 protein contains a transmembrane α -helical domain (6) that has the essential amantadine-sensitive proton channel activity of the intact protein (7). Mutagenesis and electrophysiological experiments showed that the residues important for proton conduction and amantadine interaction lie on one face of the helix, namely, V27, A30, S31, and G34 (8, 9). Neutron diffraction data indicated that the amantadine ring is localized at ≈ 6 Å from the center of dioleoylphosphatidylcholine (DOPC) bilayers, close to V27 (10). Fourier analysis of the periodic oscillations in the channel reversal potential, pH-sensitive current, and amantadine resistance of cysteine mutants of the M2

protein yielded a functional structure of the channel (11). The active form of the channel is a tetramer for the intact protein (12, 13) and the transmembrane peptide (M2TMP) (14), as shown by ¹⁹F solid-state NMR (SSNMR) of the membrane-bound peptide.

The most extensive molecular-level structural information of M2TMP came from static ¹⁵N SSNMR data of Cross and coworkers (15, 16). Using uniaxially aligned lipid membranes, they determined the orientation of M2TMP in the apo (15) and complexed (16) states from ¹⁵N chemical shift and N–H dipolar couplings. The apo peptide is tilted by 38° from the bilayer normal (15), whereas the amantadine-complexed peptide exhibits a kink with 31° and 20° tilt angles (16). However, sample preparation conditions such as solvents, membrane composition, and peptide concentration varied greatly in these studies, which may contribute to the observed orientation difference. From the ¹⁵N orientational data, no direct information on the backbone and side chain conformations can be extracted. The side chain conformation may be especially sensitive to amantadine binding, yet so far only one ¹³C–¹⁵N distance (17) and four ¹⁹F–¹⁹F distances (14, 18) have been reported. Recently, amantadine was found to cause substantial narrowing of the ¹⁵N NMR spectra (19, 20), suggesting that it either reduces the conformational heterogeneity or changes the dynamics of the protein, but which factor dominates is unknown.

To elucidate the atomic-resolution conformation and dynamics of the backbone and side chains of this important proton channel with and without amantadine, we have used magic-angle-spinning (MAS) ¹³C and ¹⁵N NMR techniques on M2TMP bound to 1,2-dilauroyl-*sn*-glycero-3-phosphatidylcholine (DLPC) bilayers. To identify sites of structural perturbation, we measured and compared the ¹³C and ¹⁵N isotropic shifts and torsion angles of eight residues spread throughout the peptide. The data indicate that amantadine most significantly perturbs the backbone of G34 and I35 and the side chain of V27. Analysis of the homogeneous and inhomogeneous ¹³C line widths in the gel and liquid-crystalline (LC) phases of the membrane indicates that the apo peptide backbone undergoes significant motion on the microsecond time scale. Amantadine binding alters the motional rates and reduces the conformational distribution. Thus, the central feature of M2TMP structure appears to be the presence of multiple low-energy conformations, which are readily modified and selected by amantadine.

Results and Discussion

M2TMP Conformation With and Without Amantadine. We chose eight residues in M2TMP for ¹³C- and ¹⁵N-labeling. Based on the

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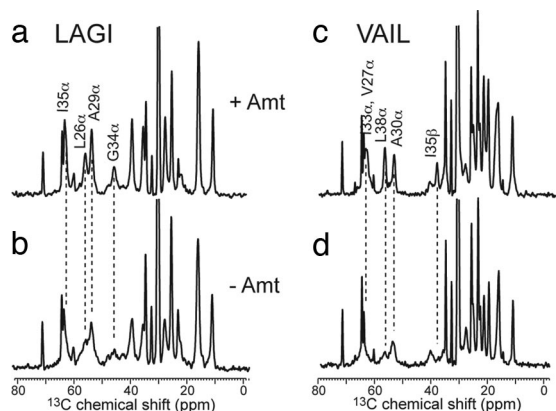


Fig. 1. ^{13}C CP-MAS spectra of M2TMP at 303 K with (a and c) and without (b and d) amantadine. (a and b) LAGI. (c and d) VAIL. Note the significant line-narrowing and intensity increase in the presence of amantadine.

approximate seven-residue periodicity of the protein (11), these sites cover channel-lining positions (V27, A30, and G34), helix-helix interfaces (L26, I33, and L38), and lipid-facing positions (A29 and I35). In this way, we assess the impact of amantadine binding to M2TMP structure from all regions of the tetrameric bundle. Two peptides were synthesized that each contained four uniformly ^{13}C , ^{15}N -labeled residues. The “LAGI” sample contained labeled L26, A29, G34, and I35, and the “VAIL” sample contained labeled V27, A30, I33, and L38. Fig. 1 shows representative ^{13}C cross-polarization (CP) MAS spectra of the peptide in DLPC bilayers with (red) and without (black) amantadine at 303 K. The ^{13}C isotropic line widths narrow substantially upon amantadine binding. In the apo peptide, many backbone signals such as G34 C^α are broad and poorly defined, whereas with amantadine, all C^α resonances narrow and increase in intensity. Side chain signals also are narrowed but less dramatically. This ^{13}C line-narrowing is similar to that seen in ^{15}N spectra of the protein (16, 19). In the next section, we investigate the origin of this line-narrowing.

To determine the M2TMP conformation and its perturbation by amantadine, we measured the ^{13}C and ^{15}N isotropic chemical shifts of the peptide without and with amantadine. ^{13}C — ^{13}C 2D double-quantum (DQ)-filtered correlation spectra and ^{15}N — ^{13}C correlation spectra were measured at 243 K where the peptide motion is frozen. Both spectra remove all lipid natural-abundance ^{13}C signals, thus simplifying resonance assignment. Fig. 2 shows the 2D ^{13}C — ^{13}C correlation spectra of LAGI and VAIL with (red) and without (black) amantadine. The spectra are readily assigned based on the connectivity patterns. Various chemical-shift changes are observed, for example, at V27 α , A30 β , I35 γ 1, and L38 β . The largest C^α shift change occurs at V27, which exhibits a 1.0-ppm upfield shift in the complex. G34 C^α is not detected in the 2D ^{13}C — ^{13}C correlation spectrum because of the DQ excitation condition, but its signal is visible in the 2D ^{15}N — ^{13}C spectra [supporting information (SI) Fig. 7] and shows a downfield ^{15}N chemical-shift change of 2.5 ppm. Interestingly, the two Ile residues flanking G34 also exhibit ^{15}N chemical-shift changes but in opposite directions, causing their amantadine-bound ^{15}N shifts to differ by 5.8 ppm. SI Table 2 lists the isotropic shifts of the apo and complexed M2TMP.

Despite the chemical-shift perturbations by amantadine, no change is large enough to indicate a nonhelical structure (21), which is reflected by the positive C^α and C' secondary shifts and negative C^β secondary shifts (Fig. 3 a–c) for all eight labeled residues. Fig. 3d plots the amantadine-induced average absolute chemical-shift changes of each residue. The maximum perturbation occurs at the channel-lining G34, followed by its adjacent

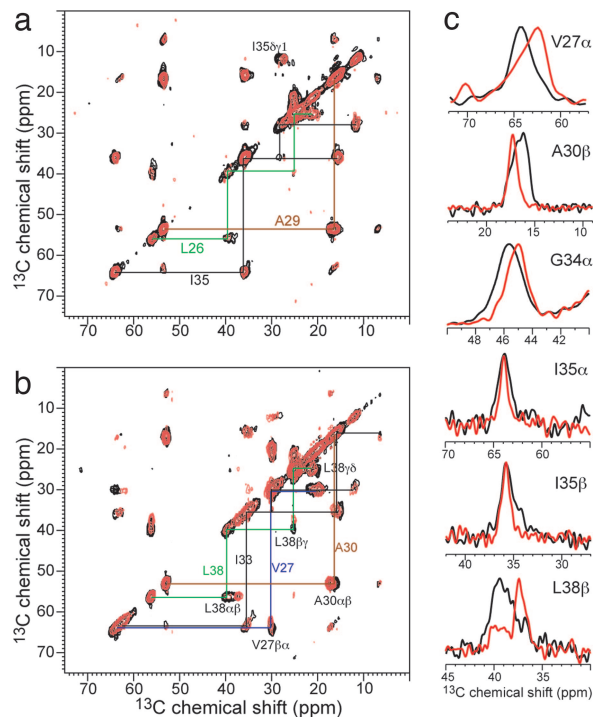


Fig. 2. 2D ^{13}C — ^{13}C DQ-filtered spectra of M2TMP in DLPC bilayers without (black) and with (red) amantadine at 243 K. Intraresidue connectivities and cross-peaks with chemical-shift changes are indicated. (a) LAGI. (b) VAIL. (c) Selected 1D cross-sections that exhibit line-narrowing and chemical-shift changes upon amantadine binding. The G34 α trace was extracted from 1D CP spectra.

I35 and I33. A second local maximum is seen at V27, consistent with its proximity to amantadine (10). In terms of residue location, the channel-lining residues, the interfacial residues, and the lipid-facing residues except for the G34-neighboring I35 have

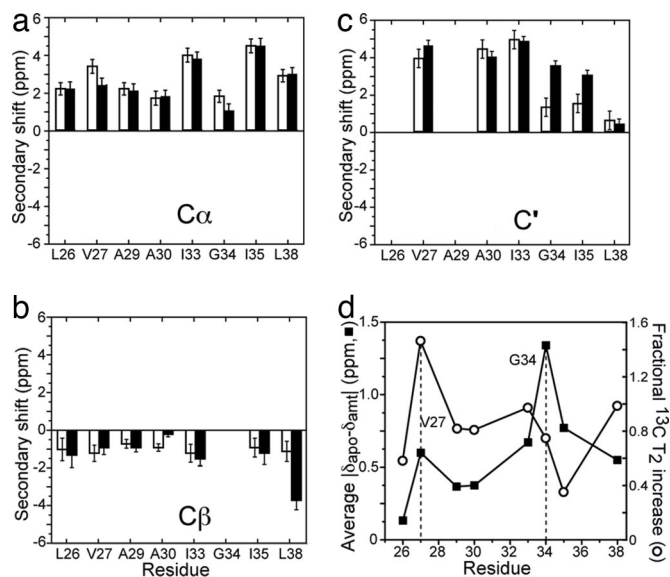


Fig. 3. Amantadine-induced isotropic shift and T_2 changes of M2TMP. (a–c) Secondary shifts are plotted for C^α (a), C^β (b), and C' (c). Open and filled bars correspond to the apo and complexed M2TMP, respectively. The average chemical-shift uncertainty is 0.35 ppm, estimated from the intrinsic line widths of the spectra. (d) Average absolute chemical-shift changes (filled squares) and fractional ^{13}C T_2 increase at 303 K (open circles). Local maxima of chemical shift and T_2 perturbation occur at V27 and G34.

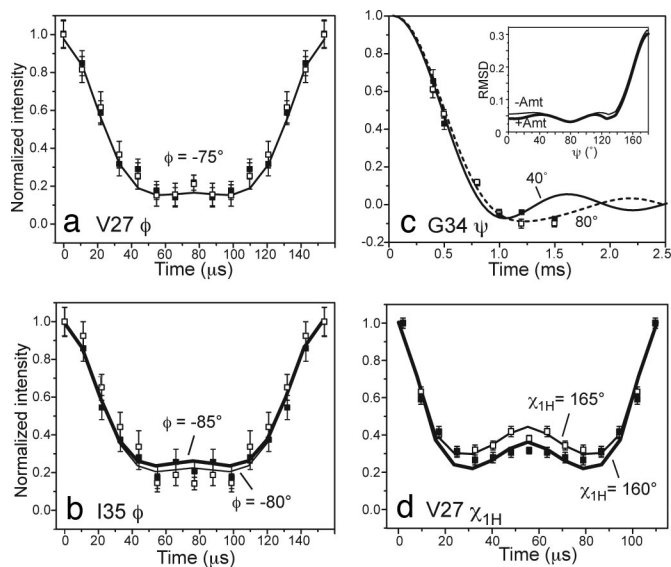


Fig. 4. Selected torsion angle data of M2TMP without (open squares) and with (filled squares) amantadine, along with their rmsd-quantified best-fit simulations (lines). Thick lines indicate best-fit curves for the complexed M2TMP when different from the apo peptide. (a) V27 ϕ . (b) I35 ϕ . (c) G34 ψ . (Inset) The rmsd between the simulation and the experiment. (d) V27 χ_{1H} .

average chemical-shift changes of 0.79 ppm, 0.46 ppm, and 0.33 ppm, respectively.

To obtain more quantitative conformational constraints, we measured ϕ , ψ , and χ_{1H} torsion angles by using dipolar correlation techniques. The ϕ angles of all labeled residues except for G34 were measured by using the HNCH technique (22), which correlates the N—H and C^α — H^α bond orientations of each residue. Most ϕ angles fall between -45° and -85° , with no large difference between the apo and complexed peptide within the angular resolution of the technique (Fig. 4a, Table 1). When taking into account the significant ^{15}N , $^{13}\text{C}^\alpha$, and $^{13}\text{C}'$ shift changes at G34–I35, the observed I35 ϕ angles of -80° for the apo-M2TMP and -85° for the complex (Fig. 4b) may reflect a real torsion angle difference. The ψ angle of G34 was measured by using the NCCN technique (23). The data (Fig. 4c) yielded a best-fit ψ angle of -80° at 243 K with and without amantadine. However, the angular resolution of the technique is limited in the α -helical region (23), as shown by the shallow rmsd minimum. Thus, we used the TALOS program (24) to predict the G34

Table 1. (ϕ , ψ , χ_1) torsion angles of eight residues in M2TMP in DLPC bilayers with and without amantadine

Residue	Torsion angle	– Amantadine	+ Amantadine
L26	ϕ	$-80 \pm 5^\circ$	$-80 \pm 5^\circ$
V27	ϕ	$-75 \pm 5^\circ$	$-75 \pm 5^\circ$
	χ_{1H}	$165 \pm 3^\circ$	$160 \pm 3^\circ$
A29	ϕ	$-75 \pm 15^\circ$	$-65 \pm 10^\circ$
A30	ϕ	$-83 \pm 10^\circ$	$-87 \pm 10^\circ$
I33	ϕ	$-74 \pm 10^\circ$	$-74 \pm 10^\circ$
	χ_{1H}	$157 \pm 3^\circ$	$157 \pm 3^\circ$
G34	ϕ^*	$-66 \pm 7^\circ$	$-67 \pm 7^\circ$
	ψ^*	$-37 \pm 6^\circ$	$-42 \pm 5^\circ$
I35	ϕ	$-80 \pm 5^\circ$	$-85 \pm 10^\circ$
	χ_{1H}	$163 \pm 3^\circ$	$165 \pm 3^\circ$
L38	ϕ	$-45 \pm 10^\circ$	$-45 \pm 10^\circ$

Angles that change between the two states are in bold italic type.

*These torsion angles were obtained from TALOS calculations.

torsion angles based on the experimental chemical shifts of the I33/G34/I35 triplet, yielding G34 (ϕ , ψ) of $(-66^\circ, -37^\circ)$ and $(-67^\circ, -42^\circ)$ for the apo and complexed peptide, respectively (Table 1). Indeed, the calculated -40° NCCN curve agrees well with the experimental data.

For the β -branched Val and Ile residues, the χ_{1H} torsion angle were obtained by correlating the C^α — H^α and C^β — H^β bond orientations (25). Here, the angular resolution is much higher, $\pm 3^\circ$, because of the nearly *trans* nature of the measured conformation. I33 and I35 show no χ_{1H} change, consistent with their lack of C^α/C^β chemical-shift perturbation. In contrast, the channel-lining V27 exhibits a significant χ_{1H} difference of 5° (Fig. 4d), consistent with the 1.0-ppm C^α and $C^\gamma 1$ chemical-shift change at this residue (SI Table 2).

Overall, the chemical shifts and torsion angle data indicate that amantadine induces only small conformational changes in M2TMP, with the main sites of perturbation being the G34–I35 pair and V27. This small conformational change contrasts with the large and extensive dynamic changes shown below.

M2TMP Dynamics and Conformational Heterogeneity. The 1D ^{13}C MAS spectra show dramatic differences in the line widths of the apo- and amantadine-complexed M2TMP. In general, NMR line widths have two contributions: inhomogeneous line widths mainly attributable to conformational heterogeneity and homogeneous line widths attributable to relaxation induced by stochastically fluctuating local fields on the microsecond time scale and residual dipolar couplings (26). Homogeneous broadening is not refocused in a spin-echo experiment, whereas inhomogeneous broadening is. Thus, to distinguish conformational heterogeneity from microsecond-time scale dynamics and to compare them between the apo- and complexed M2TMP, we measured the ^{13}C T_2 relaxation times by using a Hahn-echo experiment (27). The echo-derived T_2 is related to the homogeneous line width Δ by $T_2 = 1/\pi\Delta$ (28). Because residual dipolar couplings resulting from imperfect ^1H decoupling or insufficiently fast MAS are the same between the apo and complexed peptide, any T_2 or Δ differences should mainly result from dynamic differences of the two states. SI Fig. 8 shows representative T_2 decay curves at 303 K. All resolved sites exhibit longer T_2 curves with amantadine than without; this is true for both the backbone and side chains and for both channel-lining residues and other residues (SI Table 3). For backbone C^α , side chain carbons, and methyl carbons, the average T_2 increases by 1.5 ms, 2.1 ms, and 2.5 ms, respectively. The T_2 increase also varies with the residue location with respect to the channel: the three channel-lining residues experience the largest average T_2 increase of 2.0 ms, followed by T_2 increases of 1.9 ms and 1.4 ms for the interfacial and lipid-facing residues, respectively. Fig. 3d plots the fractional ^{13}C T_2 increase of the complexed peptide over the apo-M2TMP as a function of residues, calculated as $\frac{1}{\Sigma_{i=1}^n} (T_{2,+Amt}^i - T_{2,-Amt}^i) / T_{2,-Amt}^i$ where n is the number of sites measured in each residue. The largest T_2 increase is seen at V27, which again correlates with its purported close distance to the fused ring of amantadine (10, 29).

In ^1H -decoupled solids, ^{13}C T_2 relaxation times increase when motional correlation times either decrease or increase beyond the microsecond time window (26). Thus, both fast motions in the extreme narrowing limit and slow motions in rigid solids give rise to long T_2 curves. Comparison of the ^{13}C T_2 at 303 K, 243 K, and other temperatures (SI Table 4) suggests that the motional rates of C^α sites in the apo peptide are near the characteristic frequency of the T_2 minimum ($\approx 2\pi \cdot 70$ kHz) in the LC phase, and that amantadine binding increases the motional rates. This finding suggests that amantadine may widen the pore slightly, reducing steric hindrance and facilitating motion (30). Indeed, most side chain methyl groups show an increase in the motional rates in the amantadine-bound state based on the

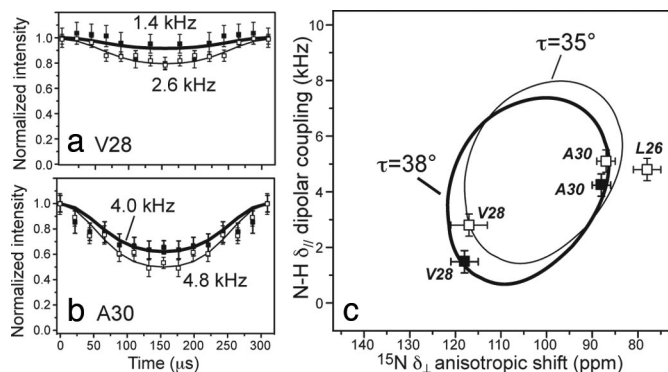


Fig. 5. Orientation of amantadine-bound M2TMP. (a and b) ^{15}N – ^1H dipolar coupling of unoriented M2TMP in DLPC bilayers with amantadine (filled squares, thick line). For comparison, the apo peptide data published recently are superimposed (open squares, thin line) (31). (a) V28. (b) A30. (c) PISA wheels of M2TMP constructed from the δ_{\parallel} N–H dipolar couplings and δ_{\perp} ^{15}N anisotropic shifts. The data fit to a wheel with a tilt angle τ of 38° (thick line). The apo peptide shows a $\tau = 35^\circ$ (open symbols, thin line) (31).

temperature-dependent T_2 curves, consistent with their motion being facilitated by a widening of the pore.

To assess the conformational heterogeneity of the protein, we compare the homogeneous line widths Δ derived from the T_2 with the apparent line widths Δ^* measured from the spectra. In the LC phase, the C^α line width of apo-M2TMP is almost completely homogeneously broadened by motion, as seen by the similar Δ and Δ^* (SI Fig. 9). Amantadine binding reduces Δ by a factor of two. For the side chains, conformational heterogeneity is detectable in both the apo and complexed peptide because of narrower intrinsic line widths. To evaluate the conformational heterogeneity without different homogeneous line widths between the apo and complexed peptide, we froze the DLPC-bound M2TMP to 243 K, where the homogeneous line widths become similar between the two states (SI Table 5). Under this condition, most sites show similar Δ^* and hence similar conformational heterogeneity between the apo and complexed peptide. The exceptions are G34, I35, and the side chains of L26, A30, and L38, where the complexed peptide has significantly narrower lines, indicating reduced conformational heterogeneity; this also is seen in the cross-sections of the ^{13}C 2D spectra (Fig. 2c).

In summary, in the LC phase of the lipid bilayer, the apo-M2TMP backbone undergoes large-amplitude microsecond-time scale motion that causes significant homogeneous broadening of the ^{13}C spectra and consequent loss of intensity. Amantadine binding increases the T_2 relaxation times for all sites by changing the motional rates, thus narrowing the intrinsic line widths. When the motion is frozen, the conformational distribution of the peptide is revealed to be reduced by amantadine at specific residues, including the G34–I35 junction and several methyl-rich side chains.

Amantadine-Induced M2TMP Orientation Change. We recently measured the orientation of the apo-M2TMP by using a powder-sample approach that exploits fast rigid-body uniaxial diffusion of the peptide backbone around the bilayer normal (31). Under this condition, motionally averaged powder spectra are obtained that indicate the peptide orientation from the bilayer normal (32). We now use this approach to determine the orientation of M2TMP in complex with amantadine. ^{15}N – ^1H dipolar couplings and ^{15}N chemical-shift anisotropies (CSA), which are extremely sensitive to the helix orientation, were measured. Fig. 5 a and b shows the N–H dipolar-shift (DIPSHIFT) curves of V28 and A30 at 313 K where the peptide is uniaxially mobile.

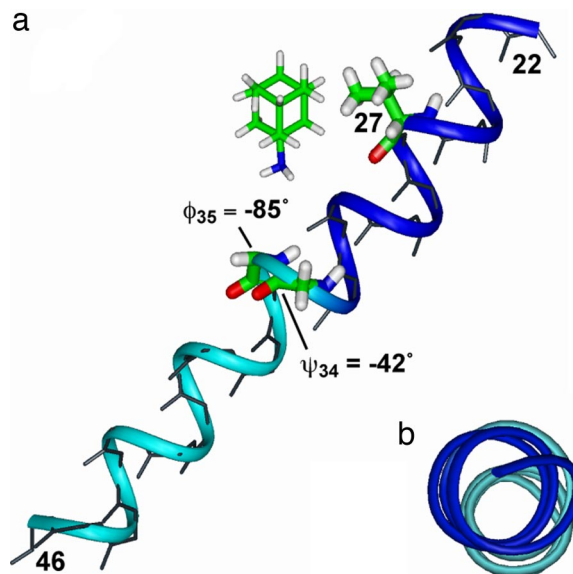


Fig. 6. Chemical shift and torsion-angle restrained backbone and partial side chain structure of amantadine-bound M2TMP. (a) Side view. (b) Top view. The exact position and orientation of amantadine is unknown and is shown here only as a reference to the peptide. The G34 ψ and I35 ϕ angles create a helix kink of 5° , highlighted by the blue N-terminal and the cyan C-terminal segments.

Amantadine binding decreases the N–H dipolar coupling of both residues. Correlating the motionally averaged δ_{\parallel} N–H dipolar coupling with the δ_{\perp} edge of the ^{15}N CSA obtained from static 1D spectra (data not shown), we obtain 2D “PISA wheels” (33, 34) (Fig. 5c). For the apo peptide in DLPC bilayers, previous data yielded a tilt angle τ of 35° (31), whereas the current amantadine-bound M2TMP has a slightly larger τ of 38° . The rotation angle of the wheel is unchanged. The 3° increase, although small, is consistent with amantadine binding at the N terminus of the helix, pushing it open slightly. The orientation of the segment C-terminal to G34 is not probed here because no ^{15}N labels are used in that region.

Fig. 6 shows the chemical shift and torsion-angle constrained structure of M2TMP in the presence of amantadine, refined from the ^{15}N NMR-derived model 1NYJ (15). At the G34–I35 junction, a G34 ψ angle of -42° and I35 ϕ of -85° were used. The resulting helix shows a small kink of 5° between the segments N-terminal and C-terminal to G34, visible in the top view (Fig. 6b). The kink is defined as the angle between the average N–H bond orientation for residues 27–33 and for residues 37–43. This kink is reminiscent of the recent ^{15}N NMR data of dimyristoylphosphatidylcholine (DMPC)-bound M2TMP, which showed a bend of 11° at G34 (16). We found that the exact value of the kink is sensitive to the G34/I35 torsion angles. With $\psi_{34} = -60^\circ$, the kink increases to $\approx 16^\circ$, whereas with a more ideal ϕ_{35} of -60° , the kink is almost completely removed.

Conclusion

The NMR data here provide an extensive set of high-resolution conformational and dynamical constraints of the backbone and side chains of M2TMP in lipid bilayers without and with amantadine and elucidate the nature of the spectral line-narrowing caused by amantadine. The data indicate that amantadine binding to M2TMP exerts the largest effect on the dynamics and conformational heterogeneity of the protein, affects to a lesser extent the average backbone and side chain conformations, and only subtly affects the helix orientation. The apo peptide exhibits large-amplitude microsecond-time scale

motion that homogeneously broadens the NMR lines. Amantadine increases the motional rates of most backbone C^α sites, causing substantial line-narrowing. It also reduces the conformational heterogeneity of certain residues, including G34, I35, and the side chains of L26, A30, and L38. Perturbation of the average conformation occurs mainly at G34–I35 and at the V27 side chain. Combined, the data strongly suggest that conformational plasticity is essential to proton conduction and gating of the apo channel, and at least part of amantadine's mechanism of action is to modify and select among the multiple low-energy conformations of M2TMP. This finding is consistent with energy surface mapping (35) and analytical ultracentrifugation data of M2TMP and its mutants (36, 37). It is possible, then, that amantadine resistance can arise from mutations that alter the protein conformational distribution and dynamics, thus preventing amantadine binding.

The observed large V27 chemical-shift and T_2 changes are in excellent agreement with mutagenesis data indicating strong interaction of this residue with amantadine. Mutation of V27 to Ala, Ser, Ile, and Thr caused either complete or partial resistance to amantadine in various viral strains (3, 9). Thus, the interaction of amantadine with the channel is exquisitely sensitive to the size and hydrophobicity of the side chain at this position.

Complexed with amantadine, the M2TMP helix shows a small degree of nonideality in the backbone torsion angles. In particular, the deviation of the G34–I35 conformation from the ideal helix geometry causes a helix kink of 5° , which may have an effect on the interhelical interaction of H37 imidazole rings downstream (19). The exact value of the kink and the exact orientation of the helix in the DLPC membrane differ slightly from those found in the DMPC membrane. Given the differences in sample preparation conditions, such as membrane thickness and the state of alignment (19, 20), these differences further underscore the structural plasticity of the peptide. The spectra of the amantadine-bound M2TMP show a single signal for each label; thus, the four helices of the tetramer are rotationally symmetric and chemically identical. This conclusion implies that, in the LC phase, not only does amantadine have the same uniaxial mobility as the tetrameric bundle, but it also exchanges among the four helices on a time scale faster than the nuclear spin interactions ($<10^{-5}$ s) (19).

Materials and Methods

Peptides and Lipids. Fmoc-protected uniformly ^{13}C , ^{15}N -labeled amino acids were either prepared in-house (38) or purchased from Sigma and Cambridge Isotope Laboratories. The transmembrane peptide of the M2 protein of the Udorn strain (residues 22–46) (39) was synthesized by PrimmBiotech (Cambridge, MA) and purified to $>95\%$ purity. The amino acid sequence is SSD-PLVVAASIIILHLILWILDRL. In addition to peptides containing multiple uniformly ^{13}C , ^{15}N -labeled residues, two peptides each containing a single ^{15}N label at V28 and A30 were synthesized for orientation measurements.

Membrane Sample Preparation. M2TMP was reconstituted into lipid vesicles by detergent dialysis (18). DLPC lipids were chosen because of the favorable dynamics of the protein in this membrane (31) and the similar phase transition temperature (-2°C) of this bilayer to biological membranes. The vesicle solution was prepared by suspending dry DLPC powder (Avanti Polar Lipids) in 1 ml of phosphate buffer (10 mM $\text{Na}_2\text{HPO}_4/\text{NaH}_2\text{PO}_4$, 1 mM EDTA, and 0.1 mM NaN_3) at pH 7.5, vortexing and freeze-thawing six to eight times to create uniform vesicles of $\approx 200\text{-nm}$ diameter (40). M2TMP powder was codissolved with the detergent octyl- β -D-glucopyranoside (OG) in 2 ml of phosphate

buffer to reach an OG concentration of 30 mg/ml. The M2TMP/OG solution then was mixed with the DLPC vesicle solution, giving a final OG concentration of 15 mg/ml. The mixture was vortexed for 1 h, allowed to stand for 6–8 h at room temperature, and then dialyzed with a 3.5-kDa cutoff against 1 liter of phosphate buffer at 4°C for 3 days with buffer changes every 8–12 h to ensure complete removal of the detergent. The dialyzed M2TMP/DLPC solution was centrifuged at $150,000 \times g$ for 3 h at 10°C to give a wet pellet with $\approx 50\text{ wt } \%$ water. The final peptide/lipid (P/L) molar ratio is 1:15. UV-visible spectrum of the supernatant indicated $\approx 98\%$ binding of the peptide to the membrane. For amantadine-bound samples, 10 mM amantadine hydrochloride was added to the phosphate buffer throughout the lipid vesicle formation and peptide assembly process.

For orientation measurements, ^{15}N -labeled M2TMP was codissolved with DLPC lipids in trifluoroethanol at a P/L of 1:20, lyophilized, and then rehydrated to 50 wt % water with a pH 8.1 phosphate buffer. For amantadine-bound samples, 2 mmol of amantadine hydrochloride was added to the dry M2TMP/lipid mixture before dissolution in trifluoroethanol.

SSNMR Spectroscopy. Most NMR experiments were carried out on a Bruker (Karlsruhe, Germany) AVANCE-600 (14.1-T) spectrometer by using a 4-mm triple-resonance MAS probe. ^{13}C – ^{13}C and ^{15}N – ^{13}C 2D correlation and torsion angle experiments were conducted at 243 K to freeze peptide motion. All other parameters, including ^{15}N CSA, ^{15}N – ^1H dipolar coupling, and ^{13}C T_2 relaxation times, were measured at 303 K or 313 K where the peptide is uniaxially mobile in the LC phase of the DLPC bilayer. Typical radiofrequency pulse lengths were 5 μs for ^{13}C and 3.5–4.0 μs for ^1H . ^1H TPPM (41) or SPINAL (42) decoupling of 60–70 kHz were applied. ^{13}C chemical shifts were referenced to the α -Gly C' signal at 176.49 ppm on the TMS scale, and ^{15}N chemical shifts were referenced to the ^{15}N signal of *N*-acetyl-valine at 122 ppm on the liquid ammonia scale. For G34 torsion angles extraction from TALOS, the ^{13}C chemical shifts were converted to the 3-(trimethylsilyl)-propionate scale by adding 1.82 ppm to the measured shifts.

2D DQ-filtered ^{13}C – ^{13}C correlation spectra were measured by using a SPCS sequence (43) $<7\text{-kHz}$ MAS. DQ filtration removes lipid background ^{13}C signals, thus simplifying assignment of the protein signals. 2D ^{15}N – ^{13}C correlation spectra were measured by using a REDOR pulse train (44) of 0.7–2.1 ms for ^{13}C – ^{15}N coherence transfer (45).

ϕ angles were measured under 6.5-kHz MAS by using the HNCH technique, with doubling of the N–H dipolar coupling to enhance the angular resolution (22, 46). ^1H – ^1H homonuclear coupling was removed by an FSLG sequence (47). The HNCH data were simulated by using a doubled N–H coupling of 12.0 kHz and a C–H dipolar coupling of 12.5 kHz, both scaled by the FSLG scaling factor of 0.577. These values were directly measured by C–H and N–H DIPSHIFT correlation experiments on the protein at 243 K. ψ angle was measured with the NCCN experiment (23) correlating the N_i – C^α_i and C'_i – N_{i+1} bond orientations. Spinning speeds of 4 and 5 kHz were used to obtain multiple time points on the angle-dependent curve. χ_{H} torsion angles (H^α – C^α – C^β – H^β) were measured by correlating the C^α – H^α and C^β – H^β bond orientations by using a modified HCCH technique (25) under 9-kHz MAS. A HORROR sequence with a resonance condition of $\omega_1 = \omega/2$ (48) was used to selectively excite the C^α – C^β DQ coherence, followed by a dipolar-doubled C–H DIPSHIFT period. A doubled and FSLG-scaled C–H dipolar coupling of 26.0 kHz was used to simulate the angle-dependent curves. All these torsion angles have an inherent double degeneracy caused by the uniaxial nature of the dipolar coupling. The wrong angle is readily identified by the fact that it falls into either unpopulated regions of the Ramachandran diagram or the β -sheet region, which contradicts NMR chemical shifts.

^{15}N – ^1H dipolar couplings for orientation determination were obtained from a dipolar-doubled DIPSHIFT experiment (46, 49, 50) under 7-kHz MAS. An FSLG sequence with an effective field of 76.5 kHz was used for ^1H homonuclear decoupling.

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