# HIV EVOLUTION AND ESCAPE

#### DOUGLAS D. RICHMAN<sup>1,2</sup>, and by invitation SUSAN J. LITTLE<sup>2</sup>, DAVEY M. SMITH<sup>2</sup>, TERRI WRIN<sup>3</sup>, CHRISTOS PETROPOULOS<sup>3</sup>, JOSEPH K. WONG $^{1,2}$

LA JOLLA, CALIFORNIA

#### ABSTRACT

Human immunodeficiency virus (HIV) exemplifies the principles of Darwinian evolution with a telescoped chronology. Because of its high mutation rate and remarkably high rates of replication, evolution can be appreciated over periods of days in contrast to the durations conceived of by Darwin. Certain selective pressures that drive the evolution of HIV include chemotherapy, anatomic compartmentalization and the immune response. Examples of these selective forces on HIV evolution are described.

## INTRODUCTION

"Natural selection is daily and hourly scrutinising, throughout the world, the slightest variations; rejecting those that are bad, preserving and adding up all that are good; silently and insensibly working, whenever and wherever opportunity offers.  $\dots$  We see nothing of these slow changes in progress, until the hand of time has marked the lapse of ages, and the so imperfect is our view into long-past geological ages, that we see only that the forms of life are now different from what they formerly were."  $-Darwin$ , "On the Origin of Species" (1)

Charles Darwin with astute observation of the natural world and perspicacious insight proposed his theory of evolution by means of natural selection. The principles of Darwinian evolution were succinctly summarized in a brilliant review of a new edition of On the Origin of Species by Robert Pollack in 1997 (2) (Figure 1). Darwin generated these principles almost 150 years ago with neither a glim-

<sup>&#</sup>x27;Veterans Affairs San Diego Healthcare System, San Diego, CA 92161

<sup>2</sup>University of California San Diego, La Jolla, CA 92093

<sup>&#</sup>x27;ViroLogic, Inc., South San Francisco, CA

Corresponding Author: Douglas D. Richman, M.D. Veterans Affairs San Diego Healthcare System and University of California San Diego Departments of Pathology and Medicine, 0679 9500 Gilman Drive La Jolla, CA 92093-0679 Tel: (858) 552-7439 Fax: (858) 552-7445 E-mail: drichman@ucsd.edu

## Principles of Darwinian Evolution

- \* All life shares a common ancestry
- **\* Variation is intrinsic to life**
- \* All differences among living things are the result of differential viability among the variants of previous forms
- \* Variation is intrinsic to life; therefore, a species can attain neither perfect form nor perfect stability

FIG. 1. Principles of Darwinian Evolution derived from reference 2.

mer of appreciation about the existence of nucleic acid nor its basis for genetic information. Nevertheless his insights have stood the test of time. The role (or opportunity) for those of us following his footsteps is to embellish his theory and to generate supportive documentation.

One application of his theory, which Darwin could not have foreseen, is the microbial world. Microbes outnumber us multicellular creatures both in numbers of species and population size. They also have more rapid replication rates. As a result they evolve more quickly. As Pollack observed, "The strategy our ancestors have followed since they first assembled into multicellular creatures bets on genetic stability and complexity to create a species made up of individuals, each with a reasonable chance of survival. The microbial strategy takes the opposite tack. Their genetic simplicity and malleability allow them to discard almost all progeny, always leaving a few genetic variants to survive any contingency." (2)

The introduction of human immunodeficiency virus (HIV) into the human species in the past century (3) has become a global scourge promising to be the primary infectious cause of mortality for years to come (4). Its success as a pathogen and in evading therapeutic and vaccination strategies is largely attributable to its ability to accelerate Darwinian evolution. HIV, like all microbes with single stranded RNA genomes, replicates with the high mutation rate of approximately one nucleotide change per genome per replication cycle (5,6). This is attributable to a reverse transcriptase enzyme with poor fidelity and the lack of proof reading mechanisms which are available to eukaryotic DNA. The rate of evolution of HIV is accelerated by a prodigious rate of replication with  $10^{10}-10^{11}$  virus particles generated daily in each infected individual (7), thus every possible mutation and many combinations of multiple mutations are generated on a daily basis. Moreover, this high rate of mutation and high rate of replication is occurring today in close to 50 million people (4).

Much translational research on HIV has proven to represent the need to deal with the capacity of the virus to adapt to selection pressures. My colleagues and <sup>I</sup> have been investigating the evolution of HIV in response to three selective pressures: chemotherapy, anatomic compartments and neutralizing antibody. Selected aspects of these investigations are summarized.

### HIV DRUG RESISTANCE

Following the clinical trials with zidovudine (AZT), the first drug shown to have antiretroviral activity (8,9), we examined serial isolates from patients receiving AZT monotherapy. Progressive incremental reductions in AZT susceptibility occurred in these isolates (Figure 2). These increases in drug resistance were later shown to be attributable to the cumulative acquisition of mutations in reverse transcriptase, the gene for the target enzyme (10,11). This observation is consistent with the adaptation of the virus during ongoing replication in the presence of the selective pressure of active drug. In 1996, we proposed that in the absence of selective drug pressure the probability of a resistant population emerging as predominant is negligible (Figure 3) (12). As the selective pressure of increasing drug activity is imposed, the probability of a resistant population emerging increases (13). At yet higher



FIG. 2. Zidovudine (AZT) susceptibility of sequential isolates of HIV-1 from a patient administered AZT (from reference 10).



FIG. 3. Proposed theoretical relationship between antiviral drug activity and the probability of the emergence of a drug resistant population. The opposing factors of selective drug pressure and rates of virus replication are indicated (from reference 12).

levels of selective antiviral pressure replication is sufficiently restricted to diminish the likelihood of outgrowth of a resistant population. Finally, when replication is completely suppressed, although selective drug pressure is high, resistant mutants cannot emerge. This proposal was not confirmed until potent combination chemotherapy could be designed, which permitted the dramatic suppression of plasma HIV RNA below the levels of detection of standard assays (Figure 4) (14,15).

Over the past several years testing for HIV drug resistance has become the standard of practice in the management of the chemotherapy of HIV patients and their drug regimens (16-18). As with much antimicrobial chemotherapy, the selection for widespread resistance has become a consequence of the success of these drugs. Similarly transmission of drug resistance to newly infected patients is becoming all too common (19).

# ANATOMIC COMPARTMENTS

The composition of cell types, the distribution of cellular and humoral immune responses, and drug disposition differ between the



weeks

FIG. 4. The suppression of HIV replication in patients receiving potent combination antiretroviral therapy. Patients were randomized to zidovudine plus lamivudine ZDV/ 3TC which was the standard of care in 1995, to indinavir (IDV) an investigational protease inhibitor, or all 3 drugs. The proportion of patients achieving the virological endpoint of plasma HIV RNA being below the limits of detection is depicted (from reference 14).

circulation and anatomic compartments. The central nervous system (CNS) is important both as a target for HIV pathology and as a drug, and possibly an immunologic, sanctuary. The genital tract is important as the primary source of new HIV transmissions.

Since the early 1990s, investigators have described distinctive envelope HIV sequences in the CNS compared to the circulation among patients with established disease (20-24). This contrasts with subjects studied during primary infection when partial sequence of the viral envelope from CSF and blood are indistinguishable (unpublished data) consistent with the transmission of a single genetic variant during primary infection and the subsequent independent evolution of virus populations during the course of infection. The selection for these variants could be distinctive host cell tropism or possibly differential immune selection. Doms and colleagues observed a trade-off between resistance of HIV variants to antibody neutralization and their ability to infect cells with a low density of the primary or secondary viral receptors (25,26). This observation may explain the intra-host evolution of HIV in the CNS where putative target cells, the microglia and to lesser extent astrocytes and microvascular endothelial cells, show low primary and secondary receptor expression while effective neutral-



FIG. 5. The anatomic compartmentalization of genetic variants examining the sequence of the reverse transcriptase of HIV is depicted. In this unrooted phylogenetic tree variants found in the lymphoid and central nervous systems have evolved independently (from reference 30).

izing antibody concentrations may differ from that in the systematic circulation (27,28).

Variable penetration of antiretroviral drugs into the CNS constitutes an additional and potentially clinically important selective pressure shaping viral evolution (29). Wong et al examined autopsy tissues, including brain, from 4 patients previously on failing antiretroviral therapy and found that genetic resistance patterns differed between virus populations in brain and those in lymph node and spleen (Figure 5) (30). Examples were found of the discordant absence of some resistance conferring mutations in brain when non-CNS-penetrating antiretrovirals were used and of the discordant persistence of some mutants in brain from past drug exposure, documenting the role of both the imposition and withdrawal of antiretroviral drug selection on the evolution of reverse transcriptase in the CNS (30). Recently, several groups have observed that the prevalence of discordant resistance between virus in cerebrospinal fluid and plasma approach 50% in selected patient populations (31,32). These studies provide some insight into the complexity of viral evolution in vivo but there is clearly a need for more systematic and longitudinal studies in the future to better understand the rates and determinants of viral genetic divergence in the CNS and their impact on disease progression and treatment response.

Similar to the central nervous system, the male genital tract is an anatomic compartment with a distinct hormonal, cytokine and immunologic milieu, which is specific for the development and maturation of sperm (33-35). This specialized environment also shapes the viral population that is harbored there. Sequence analysis has revealed that in some individuals, virus extracted from seminal plasma is similar to that found in blood plasma, while in others it differs, with few correlates to indicate why (35-39). It has been suggested that free virus may be exchanged between the blood and genital tract compartments while a subpopulation replicates in cells that are specific to the tissues of the genital tract (34,39-41). Reasons for a viral population that is specific for the genital tract could include adaptations that enhance interactions with cells important in the infection process such as dendritic cells. These cells have recently been shown to express a lectin, DC-SIGN (42) that interacts with HIV envelope and potentiates infection. Currently, we are using machine learning and phylogenetic techniques (43) in an attempt to identify conserved genetic regions of HIV derived from semen. Identification of such "sequence signatures" may have significant bearing on future vaccine development, since the vast majority of new HIV infections occur through exposure to HIV contained in genital secretions (40).

The male genital tract is not a homogenous compartment. It is comprised of many different tissues contributing fluids and cells to male genital secretions. Each of these tissues may represent subcompartments with their own environments for the development of HIV, including differential pharmacologic penetration (38,40,44). Since many antimicrobial medications do not penetrate well into the fibromuscular prostate, allowing bacterial and fungal pathogens to be harbored there, we investigated it as a reservoir for HIV when individuals are treated with antiretrovirals (39,45). Individuals who were treated with antiretroviral medications and had HIV RNA viral loads that were below the level of detection in the blood  $(<50 \text{ copies/ml})$  were also undetectable in their seminal plasma (<25 copies/ml) except when their prostate was stimulated by a digital massage before ejaculation (46). The recovery of cell free virus only after prostate stimulation implicated the prostate as a sanctuary for HIV during antiretroviral treatment, which could have significant transmission implications in both treated and untreated individuals (39).

# NEUTRALIZING ANTIBODY

The protective efficacy of most viral vaccines correlates with the elicitation of neutralizing antibody (47). The induction of neutralizing antibody to HIV by a candidate vaccine has been hampered by the inability to design immunogens that induce neutralizing antibody to most primary isolates of HIV (48). Even the role of neutralizing antibody in the natural history of HIV infection has been difficult to characterize because assays for neutralizing antibody have been labor intensive, slow and imprecise and the isolation of primary isolates from each patient to assay autologous antibody responses is slow, expensive and often difficult to achieve.

The development of an assay for neutralizing antibody, which is precise, applicable to virus from the plasma of patients, and amenable to high throughput, has permitted us to characterize the neutralizing antibody response of patients with HIV infection (49). A recombinant virus assay initially developed to measure antiretroviral drug resistance during a single round of virus replication was adapted to measure virus entry and its inhibition by neutralizing antibody (50). Briefly, full length envelope is amplified from plasma HIV RNA and co-transfected in an expression vector with an HIV-genomic vector deleted in envelope and expressing an indicator firefly luciferase gene. The pseudovirions expressing patient HIV envelope that are produced are used in an assay for neutralizing antibody using serial dilutions of patient plasma. The inhibition of infectivity is measured by reduction of luciferase activity.

We first investigated <sup>14</sup> study subjects who presented to the San Diego Acute and Early Infection Disease Research Program 30-65 days after their estimated date of HIV infection and who elected to defer or delay antiretroviral therapy. Plasma samples (3-13 per patient) were obtained at presentation to the clinic and at regular intervals for 6-39 months of follow-up. Autologous envelope-antibody pairs were assessed to characterize the development and evolution of autologous neutralizing antibody. Figure 6 demonstrates the ability of this assay to detect the emergence of autologous neutralization activity directed against the virus present at presentation of primary HIV infection (month 0) and in serial plasma samples (0, 6, and 12 months).

The neutralization activities of sequential plasma samples against sequential virus envelope proteins from the same patient (autologous responses) or against two reference viruses (heterologous responses), are displayed in Figure 7. This patient exemplified 12 of 14 patients who developed appreciable neutralizing antibody responses shortly after HIV infection. Of note each sequential virus tested escaped the



FIG. 6. Neutralization of autologous HIV. The neutralizing activity of plasmas obtained from an untreated patient 0, 6 and 12 months after presentation with primary infection is assayed against virus from months 0 and 12. The titer is defined as the reciprocal of the dilution of plasma that produces 50% inhibition of virus replication (dashed lines). The error at each dilution reflects the standard error of duplicate wells (from reference 49).

concurrent neutralizing antibody response in that patient. The neutralizing antibody responses to early viruses were continually boosted as an example of Original Antigenic Sin (51,52).

There was no relationship between the neutralizing antibody response and the disease course. The magnitude of the neutralizing antibody response correlated neither with steady state plasma HIV RNA level nor with CD4 cell count in the year after infection. Thus the neutralizing antibody response does not significantly restrict HIV replication. Rather it is an immunologic response to a constantly escaping antigen. In fact these rates exceed those observed with the emergence of HIV drug resistance mutations. Immunoglobin has a half life of three to four weeks while HIV virions in the plasma have a half life of hours (7). Studies in progress suggest that the rate of evolution in the envelope gene of HIV is determined by the magnitude of each patient's neutralizing antibody response. These observations further support the argument that the magnitude of the neutralizing antibody re-

#### DOUGLAS D. RICHMAN ET AL



FIG. 7. Neutralizing HIV antibody titers of sequential plasma specimens against autologous virus. Serial plasmas were obtained from an untreated patient presenting with primary HIV infection. The titer of each plasma against its concurrent virus specimen is shaded. Control viruses include an amphotropic murine leukemia virus  $(AMPHO)$ , a neutralization-sensitive  $X4$ —tropic virus  $(NL4-3)$ , and a relatively neutralization-resistant R5 - tropic virus (JR-CSF) (from reference 49).

sponse does not effectively impact disease course, but rather drives the rate of escape and evolution of the virus.

Using a second group of subjects with recent HIV infection, we investigated the impact of the administration of potent antiretroviral therapy on the neutralizing antibody response. To conduct these studies, a genomic HIV vector was constructed using a pol gene derived from a patient virus that was highly resistant to protease and reverse transcriptase inhibitors. This vector, in conjunction with patient virus envelope expression vectors, was used to measure neutralizing antibody accurately despite the presence of inhibitory drugs in plasma of treated patients that confound standard neutralization assays. Autologous antibody neutralization activities were measured in longitudinal plasma samples collected from five patients who were administered antiretroviral drugs shortly after presentation and who had sustained suppression of plasma HIV RNA below <sup>50</sup> copies/ml. In all five subjects, antibody titers plateaued at relatively low titers  $($  <math>1:500</math>) and their spectrum of activity evolved very little.

Of particular concern was the failure of patients to generate appreciable heterologous neutralizing antibody responses in the first year of infection despite generating high titers to their autologous virus. Cross-neutralization assays were performed with isolates shortly after infection from 13 patients. Compared with autologous viruses, neutralization of heterologous viruses was absent or at best negligible during

the first year of HIV infection. This observation is a particular concern regarding the ability of a candidate vaccine to generate broadly reactive neutralizing antibody.

# **DISCUSSION**

Because of its high rates of replication and mutation, HIV generates a remarkably wide array of genetic variants every day in each patient. Recombination readily occurs as well in vitro and in vivo which further drives the generation of variants (53). The imposition of selective pressures accelerates the evolution of HIV at rates never imagined by Darwin. The extensive use of antiretroviral therapy provides one of the most dramatic examples of the impact of human intervention on evolution in an ecological system (54). Evolution in different anatomic compartments like the central nervous system and genital tract impacts pathogenesis, drug resistance and transmission. Neutralizing antibody drives remarkable variability in the surface glycoprotein of HIV and imposes a major challenge to the development of an effective vaccine.

These observations provide a retrospective appreciation of Darwin's brilliant insights and a prospective need to address the challenges of designing effective treatments and vaccines. The lessons taught by HIV may help to address other microbial challenges like hepatitis C virus and emerging infections.

### ACKNOWLEDGMENTS

Supported by the VA San Diego Healthcare System Research Center for AIDS and HIV Infection, and VA Merit, the NIH Acute and Early Infectious Disease Research Program (AI 43683), the UCSD Center for AIDS Research (Al 36214), NIH Drug Resistance grant (Al 29164), Social & Scientific Systems (AI 38858), the NIH HIV Neurocognitive Disorders (MH 58076), the NIH Viral Dynamics grant (Al 47745), the NIH HIV Latency Study (AI 43752), the NIH K23 (Al 55276) and the Centers for Disease Control.

#### REFERENCES

- 1. Darwin C. On the Origin of Species by Means of Natural Selection, or the Preservation of Favoured Races in the Struggle for Life. The Easton Press, Norwalk, Conn, 1993, page 63
- 2. Pollack RE. [Book Review] On the origin of species by means of natural selection, or the preservation of favoured races in the struggle for life. N Engl <sup>J</sup> Med 337:137-138, 1997
- 3. Korber B, Muldoon M, Theiler J, Gao F, Gupta R, Lapedes A, Hahn BH, Wolinsky S,

Bhattacharya T. Timing the ancestor of the HIV-1 pandemic strains. Science 288:1789-1796, 2000

- 4. Piot P, Bartos M, Ghys PD, Walker N, Schwartlander B. The global impact of HIV/AIDS. Nature 410:968-973, 2001
- 5. Drake JW. Rates of spontaneous mutation among RNA viruses. Proc Natl Acad Sci USA 90:4171-4175, 1993
- 6. Mansky LM and Temin HM. Lower in vivo mutation rate of human immunodeficiency virus type 1 than that predicted from the fidelity of purified reverse transcriptase. J Virol 69:5087-5094, 1995
- 7. Perelson AS, Neumann AU, Markowitz M, Leonard JM, Ho DD. HIV-1 dynamics in vivo: virion clearance rate, infected cell lifetime, and viral generation time. Science 271:1582-1586, 1996
- 8. Fischl MA, Richman DD, Grieco MH, Gottlieb MS, Volberding PA, Laskin OL, Leedom JM, Groopman JE, Mildvan D, Schooley RT, Jackson GG, Durack DT, King D, AZT Collaborative Working Group. The efficacy of azidothymidine (AZT) in the treatment of patients with AIDS and AIDS-related complex. A double-blind, placebocontrolled trial. N Engl J Med 317:185-191, <sup>1987</sup>
- 9. Richman DD, Fischl MA, Grieco MH, Gottlieb MS, Volberding PA, Laskin OL, Leedom JM, Groopman JE, Mildvan D, Hirsch MS, Jackson GG, Durack DT, Nusinoff-Lehrman S, AZT Collaborative Working Group. The toxicity of azidothymidine (AZT) in the treatment of patients with AIDS and AIDS-related complex. A double-blind, placebo-controlled trial. N Engl J Med 317:192-197, <sup>1987</sup>
- 10. Larder BA, Darby G, Richman DD. HIV with reduced sensitivity to zidovudine (AZT) isolated during prolonged therapy. Science 243:1731-1734, 1989
- 11. Larder BA and Kemp SD. Multiple mutations in HIV-1 reverse transcriptase confer high-level resistance to zidovudine (AZT). Science 246:1155-1158, 1989
- 12. Richman DD. The implications of drug resistance for strategies of combination antiviral chemotherapy. Antiviral Res 29:31-33, 1996
- 13. Richman DD, Grimes JM, Lagakos SW. Effect of stage of disease and drug dose on zidovudine susceptibilities of isolates of human immunodeficiency virus. J AIDS 3:743-746, 1990
- 14. Gulick RM, Mellors JW, Havlir D, Eron JJ, Gonzalez C, McMahon D, Richman DD, Valentine FT, Jonas L, Deutsch P, Meibohm A, Holder D, Schleif WA, Condra JH, Emini EA, Chodakewitz JA. Treatment with indinavir, zidovudine, and lamivudine in adults with human immunodeficiency virus infection and prior antiretroviral therapy. N Engl <sup>J</sup> Med 337:734-739, <sup>1997</sup>
- 15. Gunthard HF, Wong JK, Ignacio CC, Guatelli JC, Riggs NL, Havlir DV, Richman DD. Human immunodeficiency virus replication and genotypic resistance in blood and lymph nodes after a year of potent antiretroviral therapy. J Virol 72:2422-2428, 1998
- 16. Hirsch MS, Conway B, D'Aquila RT, Johnson VA, Brun-Vezinet F, Clotet B, Demeter LM, Hammer SM, Jacobsen DM, Kuritzkes DR, Loveday C, Mellors JW, Vella S, Richman DD. Antiretroviral drug resistance testing in adults with HIV infection. JAMA 279:1984-1991, 1998
- 17. Hirsch MS, Brun-Vezinet F, D'Aquila RT, Hammer SM, Johnson VA, Kuritzkes DR, Loveday C, Mellors JW, Clotet B, Conway B, Demeter LM, Vella S, Jacobsen DM, Richman DD. Antiretroviral drug resistance testing in adult HIV-1 infection. JAMA 283:2417-2426, 2000
- 18. Hirsch MS, Brun-Vezinet F, Clotet B, Conway B, Kuritzkes DR, D'Aquila RT, Demeter LM, Hammer SM, Johnson VA, Loveday C, Mellors JW, Jacobsen DM, Richman DD. Antiretroviral drug resistance testing in adults infected with human immuno-

deficiency virus type 1: 2003 recommendations of an International AIDS Society-USA panel. Clin Infect Dis 37:113-128, 2003

- 19. Little SJ, Holte S, Routy JP, Daar ES, Markowitz M, Collier AC, Koup RA, Mellors JW, Connick E, Whitcomb JM, Hellmann NS, Richman DD. Antiretroviral-drug resistance among patients recently infected with HIV. N Engl J Med 347:385-394, 2002
- 20. Korber BTM, Kunstman KJ, Patterson BK, Furtado M, McEvilly MM, Levy R, Wolinsky SM. Genetic differences between blood- and brain-derived viral sequences from human immunodeficiency virus type 1-infected patients: evidence of conserved elements in the V3 region of the envelope protein of brain-derived sequences. J Virol 68:7467-7481, 1994
- 21. Haggerty S and Stevenson M. Predominance of distinct viral genotypes in brain and lymph node compartments of HIV-1-infected individuals. Viral Immunology 4:123- 131, 1991
- 22. Pang S, Vinters HV, Akashi T, O'Brien WA, Chen ISY. HIV-1 env sequence variation in brain tissue of patients with AIDS-related neurologic disease. JAIDS 4:1082- 1092, 1991
- 23. Epstein LG, Kuiken C, Blumberg BM, Hartman S, Sharer LR, Clement M, Goudsmit J. HIV-1 V3 domain variation in brain and spleen of children with AIDS: Tissuespecific evolution within host-determined quasispecies. Virology 180:583-590, 1991
- 24. Power C, McArthur JC, Johnson RT, Griffin DE, Glass JD, Perryman S, Chesebro B. Demented and nondemented patients with AIDS differ in brain-derived human immunodeficiency virus type 1 envelope sequences. J Virol 68:4643-4649, 1994
- 25. Edwards TG, Hoffman TL, Baribaud F, Wyss S, LaBranche CC, Romano J, Adkinson J, Sharron M, Hoxie JA, Doms RW. Relationships between CD4 independence, neutralization sensitivity, and exposure of a CD4-induced epitope in a human immunodeficiency virus type 1 envelope protein. J Virol 75:5230-5239, 2001
- 26. Puffer BA, Pohlmann S, Edinger AL, Carlin D, Sanchez MD, Reitter J, Watry DD, Fox HS, Desrosiers RC, Doms RW. CD4 independence of simian immunodeficiency virus Envs is associated with macrophage tropism, neutralization sensitivity, and attenuated pathogenicity. J Virol 76:2595-2605, 2002
- 27. Gorry PR, Taylor J, Holm GH, Mehle A, Morgan T, Cayabyab M, Farzan M, Wang H, Bell JE, Kunstman K, Moore JP, Wolinsky SM, Gabuzda D. Increased CCR5 affinity and reduced CCR5/CD4 dependence of a neurovirulent primary human immunodeficiency virus type 1 isolate. J Virol 76:6277-6292, 2002
- 28. Edinger AL, Mankowski JL, Doranz BJ, Margulies BJ, Lee B, Rucker J, Sharron M, Hoffman TL, Berson JF, Zink MC, Hirsch VM, Clements JE, Doms RW. CD4 independent, CCR5-dependent infection of brain capillary endothelial cells by a neurovirulent simian immunodeficiency virus strain. Proc Natl Acad Sci U <sup>S</sup> A 94:14742-14747, 1997
- 29. Reddy YS, Kashuba A, Gerber J, Miller V. Roundtable report: importance of antiretroviral drug concentrations in sanctuary sites and viral reservoirs. AIDS Res Hum Retroviruses 19:167-176, <sup>2003</sup>
- 30. Wong JK, Ignacio CC, Torriani F, Havlir D, Fitch NJS, Richman DD. In vivo compartmentalization of HIV: evidence from the examination of pol sequences from autopsy tissues. J Virol 70:2059-2071, 1997
- 31. Venturi G, Catucci M, Romano L, Corsi P, Leoncini F, Valensin PE, Zazzi M. Antiretroviral resistance mutations in human immunodeficiency virus type <sup>1</sup> reverse transcriptase and protease from paired cerebrospinal fluid and plasma samples. J Infect Dis 181:740-745, 2000
- 32. Cunningham PH, Smith DG, Satchell C, Cooper DA, Brew B. Evidence for indepen-

dent development of resistance to HIV-1 reverse transcriptase inhibitors in the cerebrospinal fluid. AIDS 14:1949-1954, 2000

- 33. Srinivasakumar N, Chazal N, Helga-Maria C, Prasad S, Hammarskjold M-L, Rekosh D. The effect of viral regulatory protein expression on gene delivery by human immunodeficiency virus type 1 vectors produced in stable packaging cell lines. J Virol 71(8):5841-5848, 1997
- 34. Dezzutti CS, Guenthner PC, Cummins JE, Jr., Cabrera T, Marshall JH, Dillberger A, Lal RB. Cervical and prostate primary epithelial cells are not productively infected but sequester human immunodeficiency virus type 1. J Infect Dis 183:1204- 1213, 2001
- 35. Eyre RC, Zheng G, Kiessling AA. Multiple drug resistance mutations in human immunodeficiency virus in semen but not blood of a man on antiretroviral therapy. Urology 55:5912000
- 36. Gupta P, Leroux C, Patterson BK, Kingsley L, Rinaldo C, Ding M, Chen Y, Kulka K, Buchanan W, McKeon B, Montelaro R. Human immunodeficiency virus type <sup>1</sup> shedding pattern in semen correlates with the compartmentalization of viral quasispecies between blood and semen. J Infect Dis 182:79-87, 2000
- 37. Kiessling AA, Fitzgerald LM, Zhang D, Chhay H, Brettler D, Eyre RC, Steinberg J, McGowan K, Byrn RA. Human immunodeficiency virus in semen arises from a genetically distinct virus reservoir. AIDS Res Hum Retroviruses <sup>14</sup> Suppl 1:S33- S411998
- 38. Krieger JN, Nirapathpongporn A, Chaiyaporn M, Peterson G, Nikolaeva I, Akridge R, Ross SO, Coombs RW. Vasectomy and human immunodeficiency virus type <sup>1</sup> in semen. J Urol 159:820-825, 1998
- 39. Xu C, Politch JA, Tucker L, Mayer KH, Seage GR, III, Anderson DJ. Factors associated with increased levels of human immunodeficiency virus type <sup>1</sup> DNA in semen. J Infect Dis 176:941-947, 1997
- 40. Coombs RW, Reichelderfer PS, Landay AL. Recent observations on HIV type-1 infection in the genital tract of men and women. AIDS 17:455-480, 2003
- 41. Poss M, Rodrigo AG, Gosink JJ, Learn GH, de Vange PD, Martin HL, Jr., Bwayo J, Kreiss JK, Overbaugh J. Evolution of envelope sequences from the genital tract and peripheral blood of women infected with clade A human immunodeficiency virus type 1. J Virol 72:8240-8251, 1998
- 42. Geijtenbeek TB, Kwon DS, Torensma R, van Vliet SJ, van Duijnhoven GC, Middel J, Cornelissen IL, Nottet HS, KewalRamani VN, Littman DR, Figdor CG, van Kooyk Y. DC-SIGN, a dendritic cell-specific HIV-1-binding protein that enhances transinfection of T cells. Cell 100:587-597, 2000
- 43. Pillai S, Good B, Richman D, Corbeil J. A new perspective on V3 phenotype prediction. AIDS Res Hum Retroviruses 19:145-149, <sup>2003</sup>
- 44. Paranjpe S, Craigo J, Patterson B, Ding M, Barroso P, Harrison L, Montelaro R, Gupta P. Subcompartmentalization of HIV-1 quasispecies between seminal cells and seminal plasma indicates their origin in distinct genital tissues. AIDS Res Hum Retroviruses 18:1271-1280, 2002
- 45. Larsen RA, Bozzette S, McCutchan JA, Chiu J, Leal MA, Richman DD, The California Collaborative Treatment Group. Persistent cryptococcus neoformans infection of the prostate after successful treatment of meningitis. Ann Intern Med 111:125- 128, 1989
- 46. Smith DM, Kingery JD, Ignacio CC, Havlir DV, Richman DD, Little SJ. The prostate as a reservoir for HIV-1, The 10th Conference on Retroviruses and Opportunistic Infections Boston, MA:February 10-14 (2003) (Abstract)
- 47. Parren PW and Burton DR. The antiviral activity of antibodies in vitro and in vivo. Adv Immunol 77:195-262, 2001
- 48. Burton DR. A vaccine for HIV type 1: The antibody perspective. Proc Natl Acad Sci U <sup>S</sup> A 94:10018-10023, <sup>1997</sup>
- 49. Richman DD, Wrin T, Little SJ, Petropoulos CJ. Rapid evolution of the neutralizing antibody response to HIV type <sup>1</sup> infection. Proc Natl Acad Sci U <sup>S</sup> A 100:4144- 4149, 2003
- 50. Petropoulos CJ, Parkin NT, Limoli KL, Lie YS, Wrin T, Huang W, Tian H, Smith D, Winslow GA, Capon DJ, Whitcomb JM. A novel phenotypic drug susceptibility assay for human immunodeficiency virus type 1. Antimicrob Agents Chemother 44:920- 928, 2000
- 51. Francis T, Jr. Influenza: the newe acquayantance. Ann Intern Med 39:203-221, 1953
- 52. Fazekas de St Groth S and Webster RG. Disquisitions of Original Antigenic Sin. I. Evidence in man. J Exp Med 124:331-345, 1966
- 53. Moutouh L, Corbeil J, Richman DD. Recombination leads to the rapid emergence of HIV-1 dually resistant mutants under selective drug pressure. Proc Natl Acad Sci USA 93:6106-6111, 1996
- 54. Palumbi SR. Humans as the world's greatest evolutionary force. Science 293:1786- 1790, 2001

### DISCUSSION

DuPont, Houston: Doug, it was a thorough and helpful presentation, with depressing results. <sup>I</sup> wonder if you would speculate down 10, 20, 30 years from now where you think we'll be with AIDS control. Will we be successful with either drug therapy or immunotherapy in this disease? AIDS looks more like cancer than an infectious disease to me the way it will be conquered.

Richman, LaJolla: <sup>I</sup> didn't have a chance, obviously, to describe all the aspects of treatment and pathogenesis of HIV disease. <sup>I</sup> think the accomplishments of chemotherapy have been one of the most dramatic accomplishments in American medicine. In just twenty years since the discovery of the virus the introduction of antiretroviral therapy in the developed world has resulted in a dramatic impact on morbidity and mortality. Medicine wards fifteen years ago were like a war zone, and now the only patients that are being admitted are either those with emotional or substance abuse problems who can't take their medicines, or frighteningly, a third of patients in this country who are infected and didn't know it until they first presented with their infection with AIDS related complications. We do need better ways to recognize our hundreds of thousands of infected, asymptomatic citizens who are unaware of or denying their condition. Access to therapy and the proper use of therapy can prevent disease and turn it into a manageable, chronic disease like hypertension or diabetes. The hope is that these drugs can be distributed and made available in the developing world, where  $97%$  of the infected patients now exist. <sup>I</sup> think we can significantly impact infected patients; moreover, if we can suppress virus replication, patients will no longer be likely to be transmitters. To have a major impact on the epidemic globally; however, the development of a vaccine is ultimately the only hope. This is going to be a long, long, hard struggle. Despite promising research, vaccine is not going to impact the epidemic <sup>I</sup> would think for at least a decade.