

## Septic arthritis in Western and sub-Saharan African children - a review

Christopher B. D. Lavy

Received: 13 April 2006 / Revised: 18 April 2006 / Accepted: 18 April 2006 / Published online: 2 June 2006

© Springer-Verlag 2006

**Abstract** This article reviews what is known about the incidence, aetiology, presentation, bacteriology and management of septic arthritis in children. It compares where possible the different presentations and characteristics of this condition in the Western and sub-Saharan African regions.

**Résumé** Cet article est une revue de ce qui est connu sur l'incidence, l'étiologie, la présentation, la bactériologie et le traitement de l'arthrite septique chez l'enfant. Il compare les différentes caractéristiques de ces arthrites entre les pays de l'ouest et les pays sub-sahariens.

### Incidence

Septic arthritis in children is rare in the West and common in sub-Saharan Africa. There is little more detail on the incidence or prevalence than this in the published literature. In 1990 Shaw [106] in a review of septic arthritis in infancy and childhood reported that it is approximately twice as common as osteomyelitis in this age group, but that its relative incidence decreases, so that by adolescence the two conditions occur with a similar incidence. There are several reported studies of septic arthritis from the West and from

sub-Saharan Africa [56, 65, 66, 79, 86], which again suggests that it is much more common in Africa. These reports all discuss patients that presented at single institutions, and none of them has a denominator or source population so that no accurate estimate of incidence can be made.

### Aetiology and pathogenesis

A few cases of septic arthritis occur because of direct inoculation of bacteria into the joint, through injury or snake or animal bites in a rural community or by iatrogenic causes such as hip infection following femoral artery puncture [84], but the majority of cases are believed to occur by internal dissemination of bacteria [77, 95]. The bacteria may arrive in the joint via the blood stream as haematogenous spread or by direct spread from adjacent structures. In the hip and the shoulder, part of the metaphyseal shaft is intra-articular, and osteomyelitis of the femoral or humeral shaft may spread to the adjacent joint [85]. Trueta [119] in 1959 showed that neonates have small transphyseal blood vessels which allow direct spread from the bone to the epiphysis and thus to the joint. These vessels disappear at around 6 months. This may explain the different pattern of the clinical appearance of septic arthritis between neonates and older infants. With the former it is more common to have an associated osteomyelitis, indeed in some series 60–100% of cases of neonatal septic arthritis have adjacent osteomyelitis [106].

The presence of bacteria in a synovial joint either by direct or haematogenous spread does not necessarily cause septic arthritis. Many children have a severe persistent bacteraemia, but do not develop septic arthritis [40]. It is likely that there is a combination of other factors involved in addition to the presence of bacteria. It is also very likely

---

C. B. Lavy (✉)  
Department of Orthopaedic Surgery, College of Medicine,  
Private Bag 360,  
Blantyre, Malawi  
e-mail: chris.lavy@virgin.net

#### Present address:

C. B. Lavy  
Nuffield Department of Orthopaedic Surgery,  
Windmill Road,  
Oxford OX37LD, UK

that trauma is one of these factors. In many cases of septic arthritis there is evidence of preceding trauma, and it is a plausible theory that capillary stasis as a result of this trauma causes a nidus of infection that may develop into septic arthritis. Microtrauma at the capillary level may also reduce oxygen tension locally and decrease the efficiency of the natural humoral and cellular defence response [95]. The joints of the lower limb in the West are more commonly involved in trauma than the upper limb and have correspondingly more septic arthritis. Olney's work on rabbits [92] supports this theory, that microtrauma in the presence of a co-existing bacteraemia renders joints susceptible to infection.

When a blood-borne pathogenic bacteria arrives at a susceptible synovial joint a cascade of events is set in motion. The synovium is extremely vascular and contains no basement membrane, with the result that bacteria and white cells leak into the joint space [106]. Polymorphonuclear leucocytes are activated by the presence of bacteria and produce both collagenase and neutral and acid proteases [20, 21]. The white cells are not the only source of destructive enzymes as the synovial lining cells also produce enzymes [29], as do some bacteria, especially *Escherichia coli* and *Staphylococcus aureus* [112]. These proteolytic enzymes destroy the mucopolysaccharide ground substance of articular cartilage and allow collagen fibres in the cartilage to be further destroyed by friction as the joint moves. William Hunter had no knowledge of the existence of enzymes but gave an apt description 300 years ago when he noted in 1743 the destructive effects of sepsis on articular cartilage, stating:

When a cartilage is inflamed and soaked in a purulent material, the connecting fibres will be the soonest to give way and the cartilage will become soft and red [53].

Today we know a little more about the mechanism, but the basic pathological description remains valid. Phemister in 1924 was an early worker in the field of joint infection and found that incubation of cartilage with *Staphylococcus aureus* alone did not result in any cartilage breakdown, but the addition of staphylococcal pus to the ferment caused destruction of the cartilage [99].

Smith [108] showed that cartilage destruction starts to occur as early as 8 h after infection. Early administration of antibiotics helps to slow down the process, but even if intravenous antibiotic therapy is started within the first 24 h of infection, significant glycosaminoglycan destruction and collagen disruption occurs. Potent inhibitors of these proteolytic enzymes have been found in joint fluid, so it is likely that there is a complex interplay of enzymes within the joint [42].

In addition to enzymes from bacteria, white cells and synovium, the chondrocytes themselves may also play a

part in the destruction of cartilage. Ultrastructural analysis of chondrocytes in experimentally produced septic arthritis has shown an increase in lysosomal electron-dense bodies, suggestive of the production of proteolytic enzymes in both superficial and deep layers of articular cartilage. Chondrocytes, in common with polymorphonuclear leukocytes, have both neutral and acid proteases and may be stimulated to release these either by bacterial lipopolysaccharides or by interleukin 1 (IL-1) [38, 57, 78].

The source of IL-1 is generally thought to be the monocyte [46]. IL-1 acts as an inflammatory hormone rather than having any intrinsic enzymic or degrading activity itself. It can lead to increased amounts of prostaglandin E and collagenase from both the chondrocyte and synovial cells. In mature cartilage without sepsis, chondrocytes respond to IL-1 by breaking down the surrounding proteoglycan matrix.

Recent work on joint destruction in septic arthritis suggests that in addition to the acute inflammatory mechanism above, there is also a delayed immune response that does not require viable bacteria. Arthritis can be induced experimentally in animals by systemic injection of bacterial antigens, such as peptidoglycans [41, 60]. These antigens are preferentially deposited in the synovial tissue of remote joints and incite a sustained immune response resulting in arthritis. Laboratory work with mice shows that strains of *Staphylococcus* that produce exoproteins (i.e., enterotoxin) cause more severe arthritis in infected joints than do strains that do not produce exoproteins [3]. Following on from this, it was discovered that specific inhibition of T-lymphocyte proliferation decreases the severity of arthritis, while generalised inhibition of the immune system increases the severity of arthritis [1, 2]. This leads to the possibility that bacterial antigens and bacterial exotoxins stimulate T-lymphocyte proliferation and that this can occur even though the bacteria have been killed. T lymphocytes then degrade the ground substance and destroy articular cartilage, thus playing a similar role in infective arthritis to that which they play in non-infective chronic arthritis [12].

### Predisposing causes

Joint infection is perhaps surprisingly uncommon in HIV positive adults, but where it is found it is often associated with intravenous drug abuse, haemophilia and a CD4 count in the region of 250 [120]. Children who are HIV positive have an increased risk of septic arthritis [51], and anaemic, malnourished, underweight children in sub-Saharan Africa are also at high risk [65, 66, 79]. In adults, *Salmonella* septic arthritis is associated with systemic lupus erythema-

tosis (SLE), liver disease, schistosomiasis and avascular necrosis [17]. It has also been reported after iguana bites [91]. Salmonellosis, osteomyelitis and joint infections are also common in sickle cell disease [5]. The cause for this is probably the fact that intravascular sickling causes capillary occlusion, which devitalises and possibly infarcts the gut, permitting salmonella invasion. Reduced function of the liver and spleen together with interference of reticuloendothelial system function due to erythrophagocytosis also suppresses clearance of *Salmonella* from the blood stream. Abnormal opsonisation and complement function probably also play a role [5].

### Clinical features

A child with acute septic arthritis is typically unwell, with a fever. The joint is usually swollen, warm to the touch and acutely painful. The pain is exacerbated by movement, and the child holds the limb still. The position of most comfort varies with the joint, thus the septic hip is held in slight flexion, external rotation and abduction, the knee in slight flexion and the shoulder in internal rotation and abduction. These positions represent the position of maximum joint volume and therefore minimum pressure. If the condition is not treated then the infection may spread to cause local cellulitis and swelling of the whole limb. The child may also become toxic. Having outlined the typical case, however, the infant and neonate with a less well-developed immune system may present with much less severe symptoms and signs. The clinician must be alert to the possibility of septic arthritis in an infant with a swollen joint, minimal pain and no or mild fever [106]. In the author's Zambian series several of the children presented with so called 'pseudoparalysis' [66]. The affected limb was floppy and not actively used. If it was examined there did not appear to be significant pain. There are no comparative figures on the prevalence of pseudoparalysis in Western and sub-Saharan children with septic arthritis, but it is the observation of many clinicians that it is relatively more common in Africa. This may be due to the poor nutritional state of patients in sub-Saharan Africa and to the reduction in immune response [25].

### Joints involved

There is a major difference in terms of the site of infection between septic arthritis in children in the West and children from sub-Saharan Africa. The main reason for this is the large number of infections of the shoulder that are seen in sub-Saharan Africa. Jackson and Nelson

reviewed 514 infected joints in 471 Western children and found the knee to be the most commonly affected with 41%, followed by the hip with 23%, the ankle with 14%, the elbow 12% and the wrist and shoulder 4% each [56]. In Gillespie's series of 102 children the shoulder only represented 3% [33]. Molyneux's series from Malawi in 1982 reported the shoulder as being involved in 28% of cases, second only to the knee with 51% [79]. In the author's own series from Zambia the shoulder was involved in 19 out of the 34 prospectively studied cases, representing 56% of all infected joints [66]. Molyneux is the only author to have proposed a mechanism to explain why the shoulder is so commonly involved. She has observed that mothers in sub-Saharan Africa carry their children on their backs and swing them up by the arm. She has postulated the theory that microtrauma to the joint may make it susceptible to seeding of infection when there is a bacteraemia [79].

Boys are more commonly affected by septic arthritis than girls [33, 65, 87]. There is no obvious reason for this gender difference, but it may be that boys are more likely to be involved in activities that lead to repetitive minor joint trauma [109].

### Complications

Untreated septic arthritis can cause cartilage destruction by the mechanisms outlined above. Infection can then spread to the underlying growth plate, causing destruction of the physis with consequent loss of growth or tethering of the plate causing deformity [96]. Epiphyseal separation can also occur [7, 76]. Joint infection can also spread to the adjacent bone and cause osteomyelitis [6]. The presence of infection in the joint can cause a reactive capsular thickening which may reduce movement and result in fibrous ankylosis and even joint fusion. It may also lead to the opposite, namely capsular stretching and joint laxity or dislocation. Long-standing joint sepsis can discharge to the outside and cause chronic sinus formation and subsequent superinfection [6].

### Bacteriology

In the West, septic arthritis may be caused by a wide spectrum of bacteria, but there is a definite age relation to the common pattern [77]. In neonates less than 2 months old infected in the community the common organisms are group B *Streptococci*, followed by *Staphylococcus aureus* and gram-negative rods. If the infection was acquired in the hospital situation then *Staphylococcus* is more common and is reported as being the cause in up to 62% of cases [22]. In

the infant from 2 months to 4 years in the West *Haemophilus influenzae* has in the past been reported to be the most common cause of septic arthritis [106]; however, with increasing and effective vaccination campaigns the influence of this bacterium is diminishing [11, 50] and *Staphylococcus* and *Streptococcus* are again the common causes [77]. Some authors also report an increase in the incidence of joint infection by *Kingella kingae*, which mirrors the decline of *H. influenzae* [63, 64].

### High prevalence of *Salmonella* infections in sub-Saharan Africa

The picture is completely different in sub-Saharan Africa where *Salmonella* has a very high prevalence. It was cultured in 26 out of 34 (59%) of the author's cases in Zambia [66]. All of these children were under 3 years of age. *Salmonella* was grown in 40% of the culture-positive cases in Molyneux's Malawi series [79] and in 60% of the cultures in Ndauti's series in Kenya [86].

The reason for the high prevalence of *Salmonella* in septic arthritis is probably because it is the single most prevalent organism found in the blood of sub-Saharan children [69, 73, 89, 122]. Most cases of *Salmonella* bacteraemia are found in children between 6 months and 5 years of age, with the highest incidence between the ages of 10 and 14 months [39, 40]. It is also strongly associated with anaemia, poor nutritional status and malaria [39, 40, 69, 73, 89, 122]. In the author's Zambian series all the 26 children with *Salmonella* septic arthritis were anaemic and all were underweight [66]. It is thus likely that the high incidence of *Salmonella* septic arthritis is secondary to the high prevalence of *Salmonella* bacteraemia, which is in turn secondary to the poor nutritional status of the children in this part of Africa. The argument is strong if not compelling that *Salmonella* septic arthritis in sub-Saharan children is a disease of poverty.

### Diagnosis

The diagnosis of septic arthritis is essentially clinical. It has already been discussed above that in young children, and especially malnourished children, there can be a less acute clinical presentation and diagnosis is more difficult. In well-nourished children with a normal immune response there is typically an elevated ESR and white cell count, but, as with the clinical appearance, in the anaemic underweight child these parameters may be normal. The author's Zambian series had many cases where the ESR and neutrophil count were normal [66]. Where C reactive protein (CRP) can be

measured it has been reported as being more sensitive in both diagnosis and monitoring [121].

The definitive diagnostic test is bacteriological examination of the joint fluid collected by aseptic needle aspiration of the affected joint. This should be performed with a wide-bore needle (at least 20 gauge) to ensure adequate aspiration. The fluid should be gram stained, cultured, and white cells, glucose and lactate should be measured. The gram stain alone may confirm the diagnosis in up to 50% of cases [80]. It can also give guidance for early antibiotic selection before culture and sensitivity results are available. Some series report positive culture rates of as low as 60% [86]. However, other reports have positive culture rates of 80% [47]. Ike [54] suggests that increased efficiency of joint fluid culture can be obtained by the immediate transfer of the joint aspirate to blood culture bottles. The synovial fluid white cell count in septic arthritis is variable, ranging from 25,000 to 250,000 cells per millilitre; however, the differential consistently reveals around 90% polymorphonucleocytes [37, 48, 61, 84]. Synovial fluid glucose levels in septic arthritis decrease relative to serum glucose levels and are often below 40 mg per decilitre [37]. Comparisons between serum and joint glucose levels are often made harder because of time differences in sample taking and intravenous infusions in seriously ill patients, which may alter serum levels. Lactic acid levels in synovial fluid may be elevated, except in gonococcal infections [114]. Immunoelectrophoresis may be performed to look for antigens to *Haemophilus*, *Meningococcus*, *Strep. pneumoniae* and other bacteria [114]. PCR (polymerase chain reaction) assay, where it is available, may also be used to detect remnants of bacteria in the face of negative culture [54].

Imaging of infected joints is not easy. Plain X-rays often show no bony changes in the first 10 days of an infection. There may, however, be evidence of a widened joint space relative to the uninvolved side [106]. This increased space, and the fluid that is causing it, can also be demonstrated with ultrasound scans. Radioisotope scanning using technetium or gallium- or indium-labeled white cell scans may be performed, but are not generally very helpful in the diagnosis as they can be positive in both septic arthritis and adjacent osteomyelitis. [9, 47, 117]. Some authors, however, are more optimistic about their usefulness [6]. Computerised tomography (CT) and magnetic resonance imaging (MRI) scans are seldom used in the diagnosis of septic arthritis, but if available can show the presence of fluid in the joint and early changes in the adjacent bone. They may also show reactive changes or spread of infection to the surrounding tissues. MRI can also differentiate between septic arthritis and transient synovitis [68].

## Differential diagnosis

This includes chronic infections such as tuberculosis and fungal infections, trauma, juvenile chronic arthritis and other non-infective arthropathies, rheumatic fever, adjacent osteomyelitis, sickle-cell disease, haemophilia, neoplasia, and Henoch Schonlein Purpura [106]. Mechanical problems also need to be considered, including Perthe's disease and slipped femoral epiphysis in the hip, and cartilage problems or other causes of internal derangement in the knee.

## Treatment

The above review of the literature shows that septic arthritis involves an inflamed joint that contains bacteria and pus. On purely empirical grounds it has long been felt that the treatment should involve removal of the pus as rapidly and as completely as possible and that this should be combined with the administration of antibiotics. Few authors would disagree with that outline. However, the method of draining the pus remains a matter of considerable debate [49]. The general principle of removing pus from an infected joint is not questioned and is usually assumed to be self evident, although as with many established ideas in medicine, there is no prospective study evidence that removal of the pus gives a better outcome than leaving it in the joint. As with many long-established ideas in medicine, it is now difficult to question and it would be hard to get ethical permission to conduct a study that compared removing pus to leaving it in the joint.

There are a number of possible ways of removing the pus, ranging from invasive surgery where the joint is formally opened via minimally invasive surgery such as arthroscopy [8], so-called tidal irrigation [55], where the joint is aspirated then saline or other lavage fluid is washed in and out of the joint through wide bore needles, and simple aspiration. There has never been a prospective comparison of all the above methods of removing pus, or even a prospective comparison between any of them. Different authors tend to favour their chosen method. Goldenburg [35] and Lane [63] have both observed that the method of pus removal offered to a patient with septic arthritis depends largely on the specialty of the clinician under whom the patient is admitted. Patients being looked after by paediatricians and rheumatologists tend to have needle aspiration, while patients looked after by surgeons have a surgical method of pus removal. Advocates of their own methods make claims for their particular technique; for example, Parisien [94] claims that arthroscopy "is the most reasonable alternative to repeated aspirations or arthrotomy in the management of pyarthrosis in accessible joints". Chung [18] also advocates arthroscopy and gives examples of successful treatment, but offers no control group.

Other authors have written case reports of successful treatment by one or another of the methods of pus removal [31]. Goldenburg in 1975 tried to throw light on the controversy and wrote a retrospective review comparing needle aspiration to surgery as modes of initial drainage [35]. Broy in 1986 returned to the same question and reviewed the literature from 1959 to 1984 to find the answer [15]. Neither paper was conclusive. In 1993 Ho entitled his editorial "How best to drain an infected joint. Will we ever know for certain?" [49]. Three years previously Shaw in a major review of acute septic arthritis [106] described open surgical drainage as the "gold standard" for removal of pus, against which all other methods are measured. Bertone in a study on septic arthritis in horses showed that arthrotomy eradicated joint infection more completely than arthroscopy, but that secondary wound infection was a problem [8]. Nord, in a study on septic arthritis in goats, showed that giving antibiotics with arthrotomy, or arthroscopy, or needle aspiration or even just giving antibiotics on their own gave similar results [90]. These last two studies are interesting attempts to solve the problem, but they involve an animal model, and it is questionable as to whether their results hold for humans.

Some authors have concluded that aspiration is a satisfactory method for all joints except the hip, and others that the hip joint can be satisfactorily aspirated [123]. Many take the midline view that they will start with joint aspiration, and if it fails then move on to surgery [48]. It is hard to escape the observation that there is as yet no clear answer to this question.

How antibiotics are administered and for how long are two more questions for which there are no definitive studies. Most clinicians agree that intravenous antibiotics are advised in the early stages, with a change to oral when the patient is afebrile. Many clinicians also give antibiotics for 4 to 6 weeks, but again there is no scientific backing for this period. There is animal evidence that steroids administered with antibiotics may have a protective effect in reducing cartilage damage [115].

After the pus has been removed and antibiotics have been given, most clinicians will allow the patient to move the joint as the pain allows. One of the fathers of British orthopaedics, Robert Jones, had a dictum that rest after infection of a joint should be "prolonged, uninterrupted and enforced" [104]. This has gradually been discarded as Salter in a seminal study on rabbits in 1981 showed that continuous passive motion gave improved clinical and pathological results after joint infection and injury [104]. The study has not been repeated in humans, but early movement has become the accepted practice. It will probably never be tested because

of the difficulty of restraining people, especially children, who want to move, and because of the many other advantages of being active.

### Prognosis

Septic arthritis is a serious condition in any part of the world. The potential for major complications makes its diagnosis and early treatment an emergency. There is no doubt that the major factor in the improved management of septic arthritis in the last century was the discovery of antibiotics. In 1920 the mortality for a child with septic arthritis was around 10%, and this was reduced to less than 1% by 1970 [33]. There have been no prospective studies looking specifically for prognostic factors in septic arthritis; nevertheless, certain factors have regularly been associated with poor outcome. These are as follows:

- Age less than 1 year [49], which is probably due to immaturity of the immune system.
- Joint site, especially the hip and the shoulder, and polyarticular infections [24]. The shoulder and the hip both have an epiphysis that is completely contained within the joint and perhaps more vulnerable to infection and vascular embarrassment, and they both have intra-articular metaphyseal bone. Thus, the spread of infection from adjacent osteomyelitis is possible. The presence of infection in more than one joint may indicate a reduced host response to infection, which is itself likely to be linked to a worse prognosis.
- Underlying serious illness, e.g., renal failure, diabetes, cirrhosis and malignancy [49]. A poor outcome in this group is likely to be due to the combination of chronic disease and acute infection.
- Immunosuppressive drugs, e.g., corticosteroids and cytotoxic agents [24]. This is only to be expected as the drugs reduce the body's ability to resist infection.
- Long duration of symptoms and long delay in treatment [83, 108]. If the degree of damage to the joint by bacterial and other enzymes is related to the length of time these enzymes are in action, then delay in treatment is likely to be detrimental to the outcome.
- Virulent organisms, especially *Staphylococcus aureus* and gram-negative bacilli [34]. This may be due to associated cell-mediated immunity caused by bacterial exoprotein.

The above factors are taken from literature relating to patients from the West, but are likely to be of relevance also to the prognosis in septic arthritis in sub-Saharan Africa.

### References

1. Abdelnour A, Bremell T, Holmdahl R, Tarkowski A (1994) Clonal expansion of T lymphocytes causes arthritis and mortality in mice infected with toxic shocks syndrome toxin-producing staphylococci. *Eur J Immunol* 24:1116–1166
2. Abdelnour A, Bremell T, Holmdahl R, Tarkowski A (1994) Role of lymphocytes in experimental staphylococcus aureus arthritis. *Scand J Immunol* 39:403–408
3. Abdelnour A, Bremell T, Tarkowski A (1994) Toxic shock syndrome toxin 1 contributes to the arthritogenicity of *Staphylococcus aureus*. *J Infect Dis* 170:94–99
4. Adeyokunnu AA, Hendrickse RG (1980) Salmonella osteomyelitis in childhood. *Arch Dis Child* 55:175–184
5. Anand AJ, Glatt AE (1994) Salmonella osteomyelitis and arthritis in sickle cell disease. *Semin Arthritis Rheum* 24 (3):211–221
6. Aronson J, Garvin K, Seibert J, Glasier C, Tursky EA (1992) Efficiency of the bone scan for occult limping toddlers. *Z Kinderheilk* 12(1):38–44
7. Aroojis AJ, Johari AN (2000) Epiphyseal separations after neonatal osteomyelitis and septic arthritis. *J Pediatr Orthop* 20 (4):544–549
8. Belton AL, Davis DM, Cox HU et al (1992) Arthrotomy versus arthroscopy and partial synovectomy for treatment of experimentally induced infectious arthritis in horses. *Am J Vet Res* 53:585–591
9. Borman R, Johnson RA, Sherman FC (1986) Gallium scintigraphy for diagnosis of septic arthritis and osteomyelitis in children. *J Pediatr Orthop* 6:317
10. Bos CFA, Mol L JCD, Obermann WR, Tjin a Ton ER (1998) Late sequelae of neonatal septic arthritis of the shoulder. *J Bone and Joint Surg (Br)* 80:645–650
11. Bowerman SG, Green NE, Mencio GA (1997) Decline of bone and joint infections attributable to haemophilus influenzae type B. *Clin Orthop Relat Res* 341:128–133
12. Bremell T, Abdelnour A, Tarkowski A (1992) Histopathological and serological progression of experimental *Staphylococcus aureus* arthritis. *Infect Immun* 60:2976–2985
13. Bremell T, Holmdahl R, Tarkowski A (1994) Protective role of a sialoprotein (CD43) expressing cells in experimental staphylococcus aureus infection. *Infect Immun* 62:4637–4640
14. Brown R, Hussain M, McHugh K, Novelli V, Jones D (2001) Discitis in young children. *J Bone Jt Surg* 83-B:106–107
15. Broy SB, Schmid FR (1986) A comparison of medical drainage (needle aspiration) and surgical drainage (arthrotomy or arthroscopy) in the initial treatment of infected joints. *Clini Rheum Dis* 12:501–522
16. Callaghan M (2000) Prevalence of HIV infection in trauma patients in the paediatric wards of Queen Elizabeth Central Hospital Blantyre. Personal communication
17. Chen JY, Luo SF, Wu YJ, Wang CM, Ho HH (1998) Salmonella septic arthritis in systemic lupus erythematosus and other systemic diseases. *Clin Rheumatol* 17(4):282–287
18. Chung W, Slater GL, Bates EH (1993) Treatment of septic arthritis of the hip by arthroscopic lavage. *J Pediatr Orthop* 13 (4):444–446
19. Cohen JI, Bartlett JA, Corey GR (1987) Extra-intestinal manifestations of salmonella infections. *Medicine* 66 (4):349–381
20. Curtis PH Jr, Klein L (1965) Destruction of articular cartilage in septic arthritis: II in vitro studies. *J Bone Jt Surg* 47A:1596
21. Curtis PH Jr, Klein L (1963) Destruction of articular cartilage in septic arthritis: I. in vitro studies. *J Bone Jt Surg* 45A:797

22. Dan M (1984) Septic arthritis in young infants: clinical and microbiological correlations and therapeutic implications. *Rev Infect Dis* 6:147
23. Daniel D, Akeson W, Amiel D, Ryder M, Boyer J (1976) Lavage of septic joints in rabbits: Effects of Chondrolysis. *J Bone Jt Surg* 58(3):393–395
24. Dhar S (1996) Septic arthritis and osteomyelitis in children. *The Medicine Group (Journals) Ltd* 236–240
25. Doherty JF, Golden MH, Raynes JG, Griffen GE, McAdam KP (1993) Acute phase protein response is impaired in severely malnourished children. *Clin Sci* 83(2):169–175
26. Donatto KC (1998) Orthopaedic management of septic arthritis. *Rheum Dis Clin North Am* 24(2):275–287
27. Ebrahim GJ, Grech P (1966) Salmonella osteomyelitis in infants. *J Bone Jt Surg* 48(2):350–353
28. Epstein JH, Zimmerman B, Ho G Jr (1986) Polyarticular septic arthritis. *J Rheumatol* 13:1105–1107
29. Fell HB, Jubb RW (1977) The effect of synovial tissue on the breakdown of articular cartilage in organ culture. *Arthritis Rheum* 20:1359
30. Fisher LL, Douglas RG (1990) Salmonella arthritis. *N Y State J Med* 266–267
31. Forward DP, Hunter JB (2002) Arthroscopic washout of the shoulder for septic arthritis in infants - a new technique. *J Bone Jt Surg* 84B(8):1173–1175
32. Gelberman RH, Menon J, Austerlitz MS, Weisman MH (1980) Pyogenic arthritis of the shoulder in adults. *J Bone Jt Surg* 62A(4):550–553
33. Gillespie R (1973) Septic arthritis of childhood. *Clin Orthop Relat Res* 96:152–159
34. Goldenberg DL, Brandt KD, Cathcart ES, Cohen AS (1974) Acute arthritis caused by gram-negative bacilli. A clinical characterization. *Medicine* 53:197–208
35. Goldenberg DL, Brandt KD, Cohen AS, Cathcart ES (1975) Treatment of septic arthritis. *Arthritis Rheum* 18(1):83–90
36. Goldenberg DL (1998) Septic arthritis. *Lancet* 351:196–203
37. Goldenberg DL, Reed JI (1985) Bacterial arthritis. *N Eng J Med* 312:764
38. Gower M, Wood D, Inrie E (1984) Stimulation by human interleukin-1 of cartilage breakdown and production of collagenase and proteoglycanase by human chondrocytes but not osteoblasts. *Biochimica et Biophysica Acta* 797:186
39. Graham SM, Walsh AL, Molyneux EM, Phiri AJ, Molyneux ME (2000) Clinical presentation of non-typhoid salmonella bacteraemia in Malawian children. *Trans R Soc Trop Med Hyg* 94:310–314
40. Graham SM, Molyneux EM, Walsh AL et al (2000) Non-typhoidal salmonella infections of children in tropical Africa. *Pediatr Infect Dis J* 19:1189–1196
41. Gromartie WJ, Graddock JG, Schwab MH, Anderj SK, Yang CH (1977) Arthritis in rats after systemic injection of streptococcal cells or cell walls. *Exp Med* 146:1585–1602
42. Hadler NM, Johnson AM, Spitznagel J K, Quiinet RJ (1981) Protease inhibitors in inflammatory synovial effusions. *Ann Rheum Dis* 40:55
43. Hamed KA, Tam JY, Prober CG (1996) Pharmacokinetic optimisation of the treatment of septic arthritis. *Clin Pharmacokin* 2:156–163
44. Henderson RC, Rosenstein BD (1989) Salmonella septic and aseptic arthritis in sickle-cell disease. *Clin Orthop Relat Res* 248:261–264
45. Hendrickse RG, Collard P (1960) Salmonella osteitis in Nigerian children. *Lancet* 80–82
46. Herman JH, Greenblatt D, Khosla RC, Appel AM (1984) Cytokine modulation of chondrocyte proteinase release. *Arthritis Rheum* 27:79
47. Herndon WA, Alexieva BT, Schwindt ML, Scott KN, Shaffer WO (1985) Nuclear imaging for musculoskeletal infections in children. *J Pediatr Orthop* 5(3):343–347
48. Herndon WA, Knauer S, Sullivan JA, Gross RH (1986) Management of septic arthritis in children. *J Pediatr Orthop* 6(5):576–578
49. Ho George (1993) How best to drain an infected joint. Will we ever know for certain? *J Rheumatol* 20(12):2001–2005
50. Howard AW, Viskontas D, Sabbagh C (1999) Reduction in osteomyelitis and septic arthritis related to Haemophilus influenzae type B vaccination. *J Paediatr Orthop* 19(6):705–709
51. Hughes LO, Aronson J (1994) Skeletal infections in children. *Curr Opin Paediatr* 6(1):90–93
52. Hughes RA, Rowe IF, Shanson D, Keat ACS (1992) Septic bone, joint and muscle lesions associated with human immunodeficiency virus infection. *Br J Rheumatol* 31:381–388
53. Hunter W (1743) On the functions of the articulating cartilage. *Philos Trans R Soc Lond* 42:514
54. Ike RW (1998) Bacterial arthritis. *Curr Opin Rheumatol* 10(4):330–334
55. Ike RW (1993) Tidal irrigation in septic arthritis of the knee: a potential alternative to surgical drainage. *J Rheumatol* 20:2104–2111
56. Jackson MA, Nelson JD (1982) Etiology and medical management of acute suppurative bone and joint infections in paediatric patients. *J Pediatr Orthop* 2:313–323
57. Jasiu HE (1983) Bacterial lipopolysaccharides induce in vitro degradation of cartilage matrix through chondrocyte activation. *J Clin Invest* 72:2014
58. Kelly PJ, Martin WJ, Coventry MB (1970) Bacterial (suppurative) arthritis in the adult. *J Bone Jt Surg* 52(8):1595–1601
59. Klein DM, Barbera C, Gray ST, Spero CR, Perrier G, Teicher JL (1997) Sensitivity of objective parameters in the diagnosis of paediatric septic hips. *Clin Orthop Relat Res* 338:153–159
60. Kohashi O, Pearson, Y, Pearson CM, Watanabe Y, Kotani S (1977) Preparation of arthritogenic hydrosoluble peptidoglycans from both arthritogenic and non-arthritogenic bacteria cell walls. *Infect Immun* 16:861–866
61. Krey P, Bailen DA (1979) Synovial fluid leukocytosis - a study of extremes. *Am J Med* 67:436
62. Lacour M, Duarte M, Beutler A, Auckenthaler R, Suter S (1991) Osteoarticular infections due to *Kingella kingae* in children. *Eur J Pediatr* 150(9):612–618
63. Lane JG, Falahee MH, Wojtys EM, Hankin FM, Kaufer H (1990) Pyarthrosis of the knee: treatment considerations. *Clin Orthop Relat Res* 252:198–204
64. Laundry DW, Kehl DK (1998) Increasing prevalence of *Kingella kingae* in osteoarticular infections in young children. *J Pediatr Orthop* 18(2):262–267
65. Lavy CBD, Lavy VR, Anderson I (1995) Salmonella septic arthritis in Zambian children. *Trop Doct* 25:163–166
66. Lavy CBD, Lavy VR, Anderson I (1996) Salmonella septic arthritis of the shoulder in Zambian children. *J R Coll Surg Edinb* 41:196–199
67. Lavy CBD, Schmidt C, Kalua A, Phuka J (2001) The resistable rise of surgical Sepsis in Malawi. *Malawi Me J* 12(2):36
68. Lee SK, Suh KJ, Kim YW, Ryom HK, Kim YS, Lee JM, Chang Y, Kim YJ, Kang DS (1999) Septic arthritis versus transient synovitis at MR imaging: preliminary assessment with signal intensity alterations in bone marrow. *Radiology* 211(2):459–465
69. Lepage P, Bogaerts J, Van Goethem C et al (1987) Community acquired bacteraemia in African children. *Lancet* 1:1458–1461
70. Leslie BM, Harris JM, Driscoll D (1989) Septic arthritis of the shoulder in adults. *J Bone Jt Surg* 71(10):1516–1522
71. Lossos IS, Yossepowitch O, Kandel L, Yardeni D, Arber N (1998) Septic arthritis of the glenohumeral joint. *Medicine* 77:177–187

72. Lou NP, Perera CU (1991) Salmonella septic arthritis (letter). *J Infect* 23:101
73. Mabey DC, Brown A, Greenwood BM (1987) Plasmodium falciparum malaria and salmonella infections in Gambian children. *J Infect Dis* 155:1319–1321
74. Macnicol MF (2001) Patterns of musculoskeletal infection in childhood. *J Bone Jt Surg* 83B:1–2
75. Malnick SDH, Beergabel M, Lurie Y (1988) Septic arthritis (letter). *Lancet* 351:1060
76. Marx RG, Wright JG (1999) Slipped capital femoral epiphysis after septic arthritis of the hip in an adolescent: report of a case. *Can J Surg* 42(2):145–148
77. Matan AJ, Smith JT (1997) Pediatric septic arthritis. *Orthopaedics* 20(7):630–635
78. McGuire-Goldrins MB, Meats JE, Wood DD, Inrie EJ (1984) In vitro activation of human interleukin-1-like factor. *Arthritis Rheum* 27–654
79. Molyneux E, French G (1982) Salmonella joint infection in Malawian children. *J Infect* 4:131–138
80. Molyneux E, Lavy CBD, Lavy VR, Walsh AL (1988) Septic arthritis (letter). *Lancet* 351:1060–1061
81. Morgan DS, Fisher D, Merianos A, Currie BJ (1996) An 18 year clinical review of septic arthritis from tropical Australia. *Epidemiol Infect* 117:423–428
82. Morgan MG, Forbes KJ, Gillespie SG (1990) Salmonella septic arthritis. *J Infect* 21:195–203
83. Morrey BF, Bianco AJ, Rhodes KH (1976) Suppurative arthritis of the hip in children. *J Bone Jt Surg* 58(3):388–392
84. Morrissy R, Haynes D (1989) Acute hematogenous osteomyelitis: a model with trauma as an aetiology. *J Pediatr Orthop* 9:447
85. Nade S (1983) Acute septic arthritis in infancy and childhood. *J Bone Jt Surg* 65B:234
86. Nduati RW, Wamola IA (1991) Bacteriology of acute septic arthritis. *J Trop Pediatr* 37:172–175
87. Nelson JD (1972) The bacterial etiology and antibiotic management of septic arthritis in infants and children. *Pediatrics* 50:437
88. Nelson JD, Koontz WC (1978) Septic arthritis in infants and children. A review of 117 cases. *Pediatrics* 38:966
89. Nesbitt A, Mirza NB (1989) Salmonella septicaemia in Kenyan children. *J Trop Pediatr* 35:359
90. Nord KD, Dore DD, Deeney VF, Armstrong AL et al (1995) Evaluation of treatment modalities for septic arthritis with histological grading and analysis of levels of uronic acid, neutral protease and interleukin 1. *J Bone Jt Surg* 77A:258–265
91. Nowinski RJ, Albert MC (2000) Salmonella osteomyelitis secondary to iguana exposure. *Clin Orthop Relat Res* (372):250–253
92. Olney BW, Papasian CJ, Jacobs RR (1987) Risk of iatrogenic septic arthritis in the presence of bacteremia: a rabbit study. *J Pediatr Orthop* 7(5):524–526
93. Ostenson A, Geborek P (1991) Septic arthritis as a non-surgical complication in rheumatoid arthritis: relation to disease severity and therapy. *Br J Rheumatol* 30:35–38
94. Parisien JS, Shaffer B (1992) Arthroscopic management of pyarthrosis. *Clin Orthop Relat Res* 275:243–247
95. Perry CR (1999) Septic Arthritis. *Am J Orthop* 28(3):168–178
96. Peter W, Irving J, Letts M (1992) Long-term effects of neonatal bone and joint infection on adjacent growth plates. *J Pediatr Orthop* 12(6):806–810
97. Petty RE (1990) Septic arthritis and osteomyelitis in children. *Curr Opin Rheumatol* 2:616–621
98. Pfeiffenberger J, Meiss L (1996) Septic conditions of the shoulder. *Arch Orthop Trauma Surg* 115:325–331
99. Plemister DB (1924) The effect of pressure on articular surfaces in pyogenic and tuberculosis arthritides and its bearing on treatment. *Ann Surg* 4:481–500
100. Pritchett JW (1991) Growth plate activity in the arm. *Clin Orthop Relat Res* 268:235–242
101. Rasool MN (2001) Primary subacute haematogenous osteomyelitis in children. *J Bone Jt Surg* 83(1):93–98
102. Roy S, Bhawar J (1975) Ultrastructure of articular cartilage in pyogenic arthritis. *Arch Path* 99:44
103. Ryan MJ, Kavanagh R, Wall PG, Hazleman BL (1997) Bacterial joint infections in England and Wales: analysis of bacterial isolates over a four year period. *Br J Rheumatol* 36:370–373
104. Salter RB, Bell RS, Keeley FW (1981) The protective effect of continuous passive motion on living articular cartilage in acute septic arthritis. *Clin Orthop Relat Res* 159:223–247
105. Schmidt D, Mubarak S, Gelberman R (1981) Septic shoulders in children. *J Pediatr Orthop* 1:67–72
106. Shaw BA, Kasser JR (1990) Acute septic arthritis in infancy and childhood. *Clin Orthop Relat Res* 257:212–225
107. Shetty AK, Gedalia A (1998) Septic arthritis in children. *Infectious Arthritis* 24(2):286–305
108. Smith RL, Schurman DJ, Kajiyama G, Mell M, Gilkerson E (1987) The effect of antibiotics on the destruction of cartilage in experimental infectious arthritis. *J Bone Jt Surg* 69(7):1063
109. Smith SP, Thyoka M, Lavy CBD, Pitani AD (2002) Septic arthritis of the shoulder in children in Malawi. A randomized prospective study of aspiration versus arthrotomy and washout. *J Bone Jt Surg Br* 84(8):1167–1172
110. Smith SP, Thyoka M, Lavy CBD, Pitani AD (2002) The Blantyre septic joint score: a new scoring system for septic arthritis. *Trop Doct* 32:250–251
111. Smith RL, Schurman DJ (1983) Comparison of cartilage destruction between infectious and adjuvant arthritis. *J Orthop Res* 1:136
112. Smith RL, Merchant TC, Shurman DJ (1982) In vitro cartilage degradation by *E. coli* and *Staph. aureus*. *Arthritis Rheum* 25:441
113. Spencer J, Cattermole G, Andrade T, Dryden M, Fowler J (1999) Salmonella osteoarticular infection without predisposing factors. *J R Soc Med* 92:363–364
114. Steigbigel NH (1983) Diagnosis and management of septic arthritis. In: Remington JS, Schwartz MN (eds) Current clinical topics in infectious diseases, vol 4. McGraw-Hill, New York, p I
115. Stricker SJ, Lozman PR, Makwski AL, Gunja-Smith Z (1996) Chondroprotective effect of betamethasone in lapine pyogenic arthritis. *J Pediatr Orthop* 15(2):231–236
116. Studahl M, Bergman B, Kalebo P, Lindberg J (1994) Septic arthritis of the knee: a 10-year review and long-term follow-up using a new scoring system. *Scand J Infect Dis* 26:85–93
117. Sunburg SB, Savage JP, Foster BK, (1989) Technetium phosphate bone scan in the diagnosis of septic arthritis in childhood. *J Pediatr Orthop* (5):579–585
118. Tien YC, Chih HW, Lin GT, Hsien SH, Lin SY (1999) Clinical application of ultrasonography for detection of septic arthritis in children. *Kaohsiung J Med Sci* 15(9):542–549
119. Trueta J (1959) The three types of acute haematogenous osteomyelitis. *J Bone Jt Surg* 41(4):670–680
120. Vassilopoulos D, Chalasani P, Jurado RL, Workowski K, Agudelo CA (1977) Musculoskeletal infections in patients with human immunodeficiency virus infection. *Medicine* 76(4):284–294
121. Wall EJ (1998) Childhood osteomyelitis and septic arthritis. *Curr Opin Pediatr* 10(1):73–76
122. Walsh AL, Phiri AJ, Graham SM, Molyneux EM, Molyneux ME (2000) Bacteremia in febrile Malawian children: clinical and microbiological features. *Pediatr Infect Dis J* 19:312–318
123. Wilson NIL, Di Paola M (1986) Acute septic arthritis in infancy and childhood. The Royal Hospital for Sick Children, Glasgow. *J Bone Jt Surg* 64(4):584–587
124. Web site of Malawi National Statistical Office. <http://www.nso.malawi.net>