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## Syntheses and optimization of new GS39783 analogues as positive allosteric modulators of GABA<sub>B</sub> receptors

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### Abstract

The optimization of GS39783 into potent, selective and safe positive allosteric modulators of GABA<sub>B</sub> receptors is presented.

The receptors for the major inhibitory neurotransmitter in the central nervous system, GABA, are subdivided into ionotropic GABA<sub>A</sub> and GABA<sub>C</sub> receptors and metabotropic GABA<sub>B</sub> receptors. Whereas GABA<sub>A</sub> and GABA<sub>C</sub> receptors form chloride-permeable ion channels, GABA<sub>B</sub> receptors are G-protein coupled receptors (GPCRs). These receptors were discovered in 1980 by Norman G. Bowery<sup>1</sup> and act post and pre-synaptically to inhibit neuronal excitability and neurotransmitter release, respectively. A possible role of GABA<sub>B</sub> receptors in a large number of CNS disorders such as cognition deficits, anxiety, depression, epilepsy, pain and drug addiction has been discussed.<sup>2</sup> Some of these diseases like anxiety, pain and drug addiction could potentially be treated by activation of GABA<sub>B</sub> receptors, which can be achieved by administration of either agonists or positive allosteric modulators. Whereas benzodiazepines are well known positive allosteric modulators of GABA<sub>A</sub> receptors, the first examples of allosteric enhancers for GABA<sub>B</sub> receptors have been described only recently.<sup>3</sup> One of the most interesting compound found was GS39783 (Figure 1).<sup>3b</sup> However, despite an interesting *in vitro* and *in vivo* profile,<sup>3b,4</sup> GS39783 was found to be genotoxic probably because of its aromatic nitro group (Figure 1)<sup>5</sup>.

This communication describes our efforts towards the identification of a novel, drug-like class of compounds acting as positive allosteric modulators for GABA<sub>B</sub> receptors.

In order to introduce molecular diversity in position 5 of the pyrimidine ring, a 4 steps procedure depicted in Scheme 1 was optimized in order to obtain compounds with a chlorine or with a hydrogen in position 6. 4,6-dichloro-2-methylpyrimidine was first substituted by cyclopentylamine and then iodinated to lead to compound **6**. This scaffold was then used in a Suzuki cross coupling<sup>6</sup> to give very efficiently a small focused library of substituted 4-amino-6-chloro-5-phenylpyrimidines (Cpds **7a–15a**) which were then hydrogenated under standard conditions to give the desired 4-amino-5-phenylpyrimidines (Cpds **7b–15b**). As a

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means to introduce molecular diversity at the last step in position 4 of the pyrimidine ring, a versatile way of synthesis was designed (Scheme 2). Starting from the commercially available 5-bromo-2,4-dichloropyrimidine, a regioselective nucleophilic substitution was performed in position 4 exclusively to give the 4-methylthio derivative **17**<sup>7</sup> which was then involved in a halogen exchange<sup>8</sup> to give 2-iododerivative **18**. The 2-methyl group and the 5-[(4-trifluoromethyl)phenyl] substituents were introduced by a Negishi cross coupling<sup>9</sup> and a Suzuki cross coupling,<sup>10</sup> respectively to afford **20**. Our first attempt was to oxidize the methylsulfide moiety of **20** to the corresponding methylsulfonyl (**21**) by treatment with mCPBA<sup>11</sup> and then to use **21** as a template. Unfortunately, only few amines reacted cleanly with **21** probably because of the steric hindrance near the leaving group. Our second attempt was to hydrolyze the methylsulfide moiety of **20** with hydrochloric acid<sup>12</sup> to give **23** (Scheme 3). This compound was then reacted with POCl<sub>3</sub> to give the corresponding 4-chloropyrimidine **24**. This useful intermediate was submitted to nucleophilic substitutions with a collection of amines to give compounds **25–34**. Finally the third library with molecular diversity in position 2 was synthesized starting from compound **16**, which was substituted first with exo-2-aminonorbormane<sup>13</sup> and then with MeSNa to give **36** (Scheme 4). The phenyl moiety was introduced by a Suzuki cross coupling to afford **37**. On one hand, a desulfurization by treatment with Ni-Raney in EtOH<sup>14</sup> leads to **39** and on the other hand **37** can also be oxidized into the corresponding 2-methylsulfonylpyrimidine **38** which was then reacted with various nucleophiles.

Thanks to a preliminary work with nitro-mimetics (Figure 2), we found that to replace the nitro group, we should have a lipophilic substituent (see Cpd **3**) with an electron-withdrawing effect (see Cpd **1** and **2**). One way to combine both of these effects is to substitute the position 5 of the pyrimidine ring by a substituted phenyl. Moreover, substantial efficacy in the biological assay<sup>15</sup> was observed for compounds with only one cyclopentylamine substituent in position 4 of the pyrimidine ring (compare **1** with **2** and **3** with **4**). Furthermore, it was also shown earlier that the replacement of the 2-methylthio substituent by a 2-methyl group is not detrimental for the activity.<sup>3b</sup> After screening for GABA<sub>B</sub> receptor positive modulatory activity of the first library of compounds (Table 1), we found that the 6-chloro substituent was detrimental to the efficacy at the receptor (compare for example **8a** with **8b** at 25 μM of compound, **12a** with **12b** and **15a**) with **15b**. Moreover, the introduction of a second cyclopentylamino substituent in position 6 led to only weakly active or inactive compounds (data not shown) confirming our hypothesis that the space in the receptor is very limited and that a proton is the best substituent for the position 6 of the pyrimidine ring. On the other hand, as postulated before, electron-withdrawing groups on the phenyl ring such as a 4-trifluoromethyl or a 4-trifluoromethoxy showed the best results (compare **9b** with **12b** and **10b** with **15b**). Indeed, the replacement of a 4-methylphenyl (**9b**) or 4-methoxyphenyl (**10b**) by a 4-trifluoromethylphenyl (**12b**) or a 4-trifluoromethoxyphenyl (**15b**) led to more efficacious positive modulators (Table 1). Other electron-withdrawing substituents were used on the phenyl ring but none of them had increased activity at the receptor compared to the trifluoromethyl or trifluoromethoxy groups (data not shown). The introduction of a substituent in position 3 of the phenyl ring led to a decrease of activity (compare **14b** with **15b**). To conclude, the best nitro-mimetic group identified in this series was a 4-(trifluoromethyl)phenyl substituent. Then we focused our attention on position 4 of the pyrimidine ring (Scheme 2 and Scheme 3).

This collection of compounds shows interesting structure activity relationships (Table 2). For this series it became obvious that an increased size of the cycloalkyl led to increased efficacy. This trend was first observed with compound **29** in which the cyclopentyl substituent was replaced by a cycloheptyl (compare **12bb** and **22** with **29** at 2.5 μM). Interestingly, the introduction of an additional bulky substituent on the cyclohexyl ring (**30**) led to a less active product (compare **30** with **22**). Moreover, the introduction of an aromatic ring onto the amino group was not tolerated by the receptor (compare **31**, **32**, **34** with **22**) therefore we thought that

a hydrophobic interaction with a spatially extended substituent was necessary at this position. Surprisingly, despite its 4-tert-butylamino substituent, **33** was found to be a weak positive modulator. The needs of bulky, cyclic aliphatic side chain was then confirmed when cycloalkyl chains were used such as a norbornyl (**27**), an adamantyl (**25**), (**26**) or a cycloheptyl (**29**). A substantial increase in activity was observed when comparing **29** to **12b** and to **22**. Despite its good efficacy, **29** was not investigated any further because of significant binding activities to other GPCRs (data not shown). We were then interested in compounds **25** and **26** bearing an adamantyl substituent on the amino group. Both compounds showed were potent positive allosteric modulators (Table 2) which led to the hypothesis that the substituents in position 4 of the pyrimidine ring bind into a large lipophilic pocket in the receptor. However, again these products were not considered for further evaluation despite an increased selectivity profile because of their high logPs ( $\log P > 5.9$ ) and low water solubilities ( $< 10 \text{ mg.L}^{-1}$ ). Finally, we focused our attention on **27** which bears a norbornyl group at its amino function. Both the *exo* isomer (**27**) and the *endo* isomer (**28**) were evaluated (Table 2). Compound **27** was more efficacious, and was of similar potency and efficacy compared to GS39783. Genotoxicity<sup>16</sup> and mutagenicity<sup>17</sup> assays were performed and **27** was found to be safe both in the micronucleus test and in the Ames test. For that reason **27** was considered for further *in vitro* and *in vivo* evaluations and the *exo*-2-norbornyl substituent was kept for the optimization of the position 2.

Finally the screening of the collection of compounds with molecular diversity in position 2 gave surprising results (Table 3). The replacement of the methyl moiety of **27** by an hydrogen led to a less active compound which indicated that a substitution at this position is mandatory. The introduction of a 2-SMe group (**37**), a methylsulfonyl (**38**) or a methylamino (**42**) were detrimental for activity (compare **37**, **38** and **42** with **27**). In contrast, the replacement of a 2-methyl group by a cyano (**40**), a methoxy (**41**) or a dimethylamino (**43**) gave compounds with a similar potency than GS39783. Surprisingly, the introduction of a N-methylpiperazin-1-yl (**44**) reduced the biological activity suggesting that the space in the receptor is limited and that only small substituents are tolerated. Compounds **40**, **41** and **43** are currently under evaluation for their physicochemical and pharmacokinetics properties as well as their full pharmacological characterization.

Positive allosteric modulators of GABA<sub>B</sub> receptors represent an interesting class of therapeutic agents for the treatment of anxiety and drug addiction. Despite its good *in vitro* and *in vivo* potency, GS39783 was only useful as a pharmacological tool because of its genotoxicity. We report herein a new class of positive allosteric modulators of GABA<sub>B</sub> receptors derived from GS39783 with an increased drug-likeness, a decreased toxicity and an excellent selectivity profile. Some *in vivo* investigations are currently on going in anxiety and drug addiction models and the results of these studies will be reported in due course.

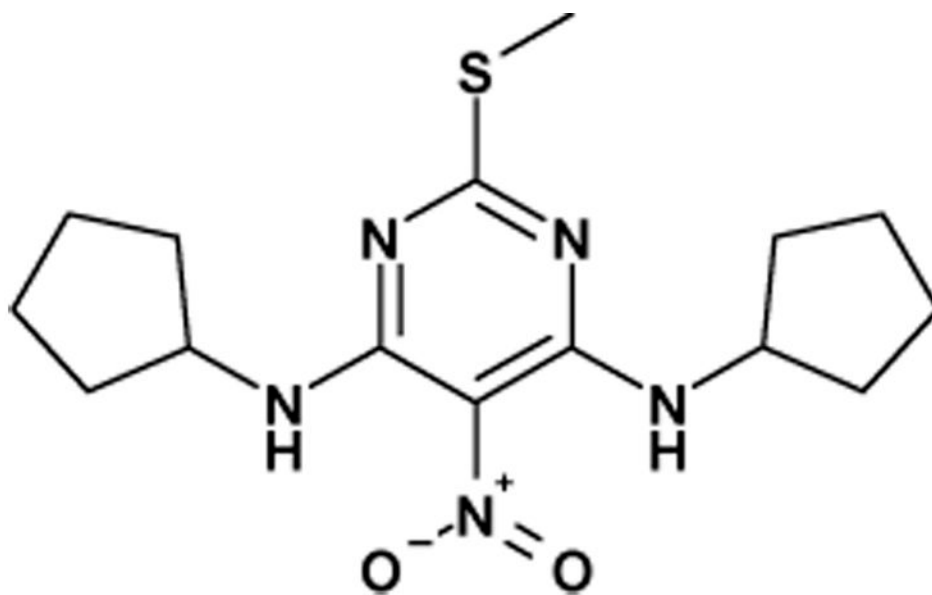
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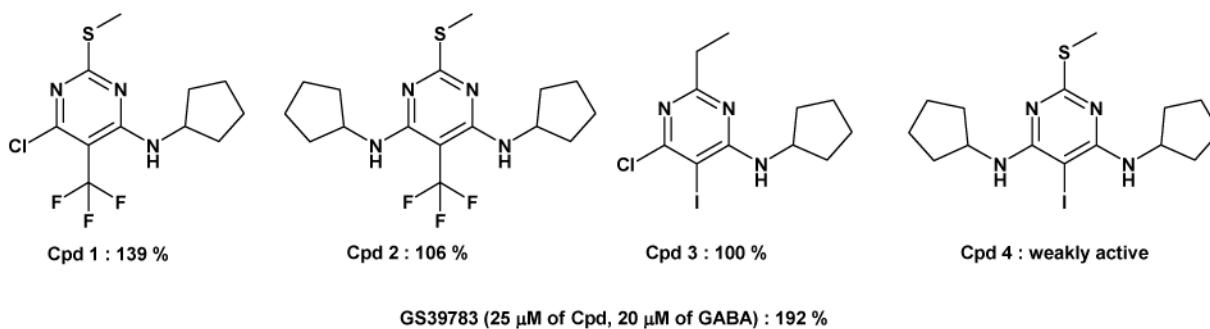
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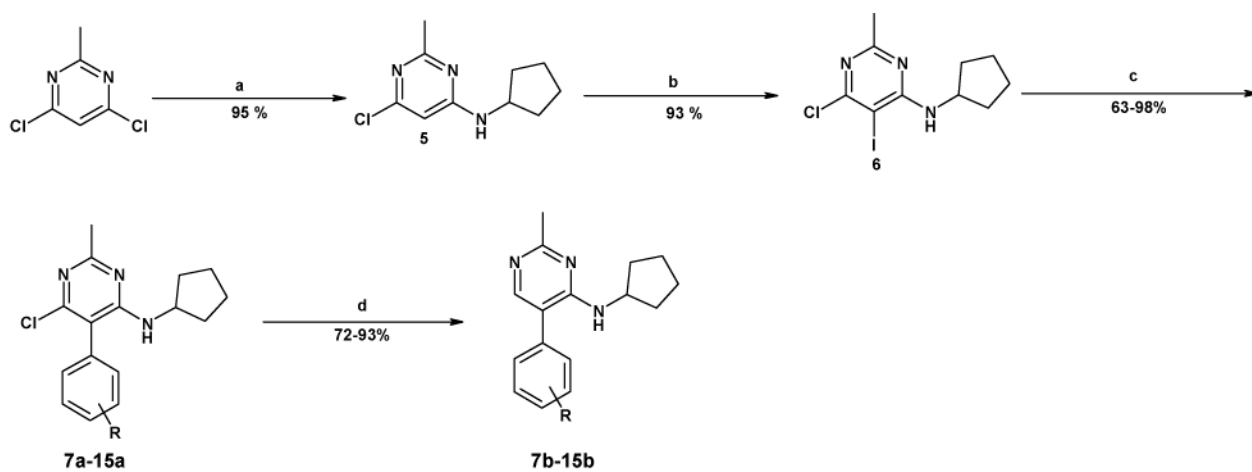
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  15. GTP( $\gamma$ )<sup>35</sup>S binding was used as functional assay for GABA<sub>B</sub> receptor activity (see reference 3b). To assay for positive modulatory activity the compounds were co-applied with GABA. 1 $\mu$ M GABA stimulates GABA<sub>B</sub> receptors to approximately EC<sub>20</sub> values, 20 $\mu$ M GABA stimulates to approximately EC<sub>80</sub> values. If co-application of the test compounds with GABA significantly increased the signal above EC<sub>20</sub> (at 1 $\mu$ M GABA) or EC<sub>80</sub> (at 20 $\mu$ M GABA) we concluded that the compound positively modulated the GABA response. In control experiments the test compounds were assayed in the absence of GABA. For selected compounds, concentration-response curves were generated and the EC<sub>50</sub> values and maximal stimulations at a GABA concentration of 1 $\mu$ M determined.
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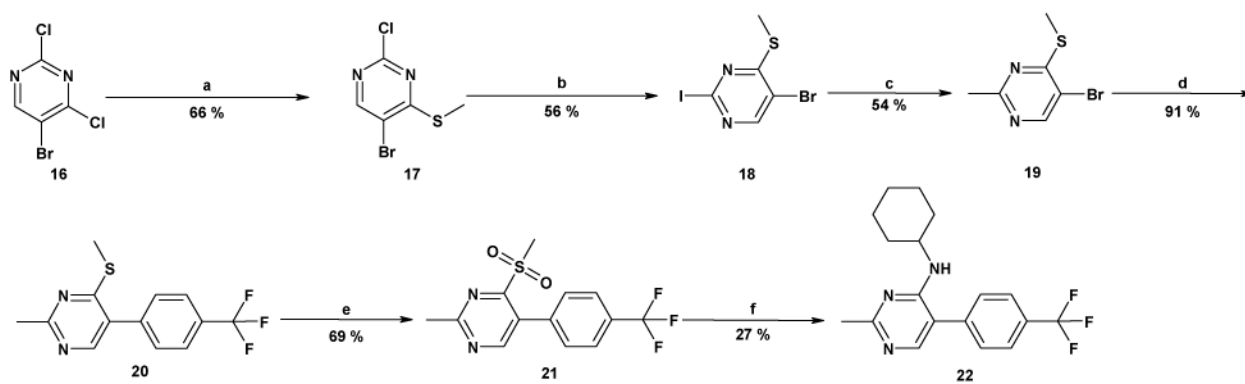
**Figure 1.**  
Structure of GS39783

**Figure 2.**

Biological activity of some GS39783 derivatives bearing nitro-mimetics in position 5. The potentiation of the GABA-induced stimulation of  $\text{GTP}(\gamma)^{35}\text{S}$  binding was measured using membranes from  $\text{GABAB}_{\text{B}(1\text{b}/2)}$  expressing CHO-K1 cells as described.<sup>3a</sup> The activities of compounds (25  $\mu\text{M}$ ) were determined in the presence of 20  $\mu\text{M}$  GABA. The data are normalized to the maximal effect (100%) obtained with a saturating concentration of GABA (1mM). 20  $\mu\text{M}$  GABA, when applied alone, stimulates to approximately  $\text{EC}_{80}$  levels (80%).

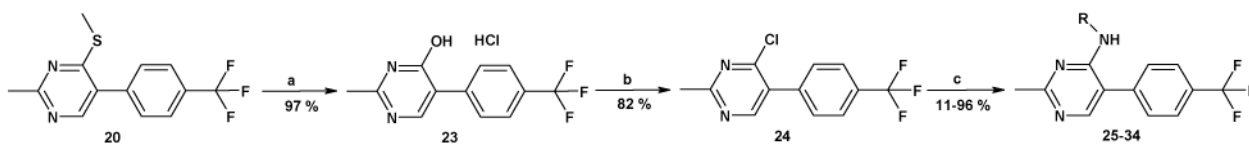
**Scheme 1.**

Reagents and conditions: (a): cyclopentylamine (4.0 eq.), dioxane, 0°C to RT, 24h; (b): NIS, DMF, 80°C, 24h ; (c): arylboronic acid (1.05 eq.), Pd(OAc)<sub>2</sub> (0.02 eq.), dppf (0.03 eq.), K<sub>3</sub>PO<sub>4</sub> (2.0 eq.), DME, water, 85°C ; (d): Pd/C 10%, AcONa (1.1 eq.), EtOH, H<sub>2</sub>, RT.

**Scheme 2.**

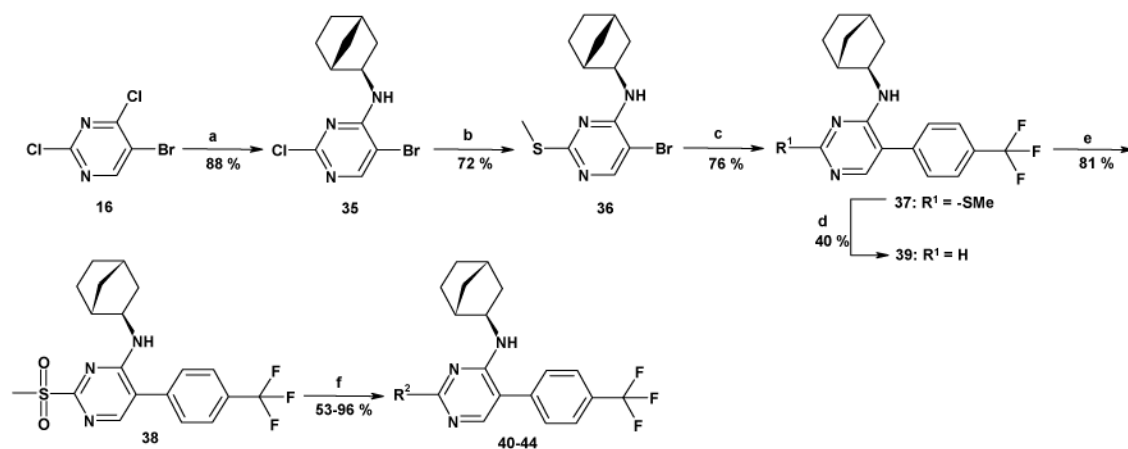
Reagents and conditions: (a): MeSNa, THF, RT; (b): HI 57%, RT; (c): MeZnCl, Pd(PPh<sub>3</sub>)<sub>4</sub>, THF, RT to 60°C; (d): 4-trifluoromethylbenzeneboronic acid, Na<sub>2</sub>CO<sub>3</sub>, Pd(PPh<sub>3</sub>)<sub>4</sub>, EtOH, Toluene, water, 110°C; (e): mCPBA, DCM, RT; (f): cyclohexylamine, DMF, 150°C, microwaves, 20 min.



**Scheme 3.**

Reagents and conditions: (a): HCl 37%, MeOH, reflux; (b): POCl<sub>3</sub>, one drop of DMF, 80°C;

(c): R-NH<sub>2</sub>, solvent, Base, 80°C.

**Scheme 4.**

Reagents and conditions: (a): *exo*-2-aminonorbornane, THF, 0°C to RT; (b): MeSNa, THF, 0°C to RT; (c): 4-trifluoromethylbenzeneboronic acid, Na<sub>2</sub>CO<sub>3</sub>, Pd(PPh<sub>3</sub>)<sub>4</sub>, toluene, EtOH, water, 110°C; (d): Ni-Raney, EtOH, 60°C; (e): mCPBA, DCM, RT, 1.5h; (f): Cpd **40**: KCN, DMSO, 80°C. Cpd **41**: Sodium methoxide, THF, RT. Cpd **42**: methylamine, EtOH, RT. Cpd **43**: dimethylamine, THF, 80°C. Cpd **44**: N-methylpiperazine, THF, 80°C.

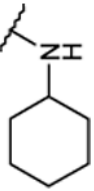
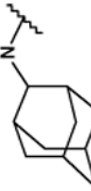
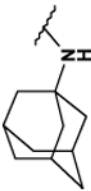


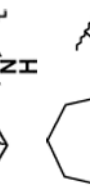
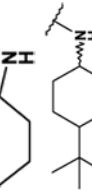
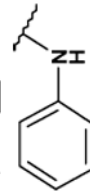
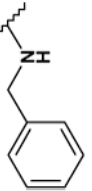
Table 1

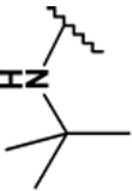
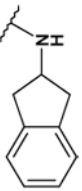
Biological activities obtained for compounds **7a–15b**. The compounds were assayed as described.<sup>3,15</sup> When applied alone, 1  $\mu$ M GABA stimulates to approximately EC<sub>20</sub> levels (20% stimulation). The effects (%) are normalized to the maximal effect of a saturating concentration of GABA (1 mM). Values were calculated from triplicate measurements. pEC<sub>50</sub> and E<sub>max</sub> values were calculated from concentration-response curves (8 concentrations) in the presence of 1  $\mu$ M of GABA.

Cpd	R	2.5 $\mu$ M of Cpd, 1 $\mu$ M of GABA Effect (%) $\pm$ sem	25 $\mu$ M of Cpd, 1 $\mu$ M of GABA Effect (%) $\pm$ sem	pEC <sub>50</sub> $\pm$ sem	E <sub>max</sub> (%) $\pm$ sem
GS39783	-	132 $\pm$ 4	153 $\pm$ 2	6.13 $\pm$ 0.06	146 $\pm$ 5
7a	4- <i>n</i> Butyl	27 $\pm$ 1	39 $\pm$ 1	-	-
7b	4-Ethyl	25 $\pm$ 1	18 $\pm$ 3	-	-
8a	4-Ethyl	23 $\pm$ 2	39 $\pm$ 2	-	-
8b	4-Ethyl	29 $\pm$ 1	61 $\pm$ 1	-	-
9a	4-Ethyl	23 $\pm$ 2	34 $\pm$ 1	-	-
9b	4-Ethyl	23 $\pm$ 2	41 $\pm$ 2	-	-
10a	4-Methoxy	21 $\pm$ 1	29 $\pm$ 1	-	-
10b	4-Methoxy	21 $\pm$ 1	38 $\pm$ 2	-	-
11a	3-Methyl	25 $\pm$ 1	36 $\pm$ 1	-	-
11b	3-Methyl	21 $\pm$ 0	28 $\pm$ 2	-	-
12a	4-CF <sub>3</sub>	17 $\pm$ 1	26 $\pm$ 1	-	-
12b	4-CF <sub>3</sub>	62 $\pm$ 3	114 $\pm$ 2	5.30 $\pm$ 0.03	127 $\pm$ 3
13a	4-COOEt	14 $\pm$ 1	5 $\pm$ 3	-	-
13b	4-COOEt	26 $\pm$ 0	31 $\pm$ 0	-	-
14a	3-OCF <sub>3</sub>	20 $\pm$ 1	33 $\pm$ 1	-	-
14b	3-OCF <sub>3</sub>	23 $\pm$ 1	25 $\pm$ 1	-	-
15a	4-OCF <sub>3</sub>	19 $\pm$ 1	35 $\pm$ 1	-	-
15b	4-OCF <sub>3</sub>	48 $\pm$ 3	100 $\pm$ 3	5.34 $\pm$ 0.07	117 $\pm$ 7

Table 2

Biological activities obtained for compounds **22**, **25–34**. The compounds were assayed as described under Table 1. Compounds **25–27**, **29** and **34** have a relatively low water solubility due to their high lipophilicity. This is reflected in this assay by a similar or reduced activity at 25  $\mu\text{M}$  versus 2.5  $\mu\text{M}$ . Concentration-dependent increase in activity however was observed in full concentration response curves, with maximal effects at 10  $\mu\text{M}$ .

Cpd	R	2.5 $\mu\text{M}$ of Cpd, 1 $\mu\text{M}$ of GABA Effect (%) $\pm$ sem	25 $\mu\text{M}$ of Cpd, 1 $\mu\text{M}$ of GABA Effect (%) $\pm$ sem	1 $\mu\text{M}$ of GABA pEC <sub>50</sub> $\pm$ sem	E <sub>max</sub> (%) $\pm$ sem
22		80 $\pm$ 1	93 $\pm$ 5	6.06 $\pm$ 0.05	83 $\pm$ 4
25		90 $\pm$ 1	80 $\pm$ 2	5.56 $\pm$ 0.13	132 $\pm$ 14
26		125 $\pm$ 4	141 $\pm$ 6	5.46 $\pm$ 0.10	185 $\pm$ 15
27		122 $\pm$ 3	110 $\pm$ 0	5.78 $\pm$ .03	183 $\pm$ 4
28		82 $\pm$ 5	52 $\pm$ 4	-	-
29		128 $\pm$ 4	70 $\pm$ 4	5.78 $\pm$ 0.03	137 $\pm$ 3
30		28 $\pm$ 1	32 $\pm$ 2	-	-
31		36 $\pm$ 2	69 $\pm$ 6	-	-
32		29 $\pm$ 1	39 $\pm$ 2	-	-

Cpd	R	2.5 $\mu$ M of Cpd, 1 $\mu$ M of GABA Effect (%) $\pm$ sem	25 $\mu$ M of Cpd, 1 $\mu$ M of GABA Effect (%) $\pm$ sem	pEC <sub>50</sub> $\pm$ sem	1 $\mu$ M of GABA E <sub>max</sub> (%) $\pm$ sem
33		40 $\pm$ 3	21 $\pm$ 7	-	-
34		58 $\pm$ 11	59 $\pm$ 4	-	-

**Table 3**  
 Biological activities obtained for compounds **37-44**. The compounds were assayed as described under Table 1.

Cpd	R	2.5 $\mu$ M of Cpd, 1 $\mu$ M of GABA Effect (%) $\pm$ sem	25 $\mu$ M of Cpd, 1 $\mu$ M of GABA Effect (%) $\pm$ sem	1 $\mu$ M of GABA pEC <sub>50</sub> ( $\mu$ M) $\pm$ sem	E <sub>max</sub> (%) $\pm$ sem
37	-SMe	82 $\pm$ 7	91 $\pm$ 4	-	-
38	Me-SO <sub>2</sub> -	24 $\pm$ 7	30 $\pm$ 12	-	-
39	H	48 $\pm$ 3	63 $\pm$ 4	-	-
40	-CN	117 $\pm$ 3	127 $\pm$ 1	5.92 $\pm$ 0.11	198 $\pm$ 21
41	-OMe	115 $\pm$ 2	131 $\pm$ 6	5.69 $\pm$ 0.13	191 $\pm$ 21
42	-NHMe	20 $\pm$ 3	25 $\pm$ 8	-	-
43	-NMe <sub>2</sub>	94 $\pm$ 4	126 $\pm$ 7	5.78 $\pm$ 0.08	167 $\pm$ 16
44	N-methylpiperazine	26 $\pm$ 8	112 $\pm$ 7	-	-