

# PERSPECTIVES

## Molecular basis for the transmural distribution of the transient outward current

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Regional differences in electrical properties of cardiac cells contribute to the normal function of the heart as well as to the inscription of the J wave and T wave of the ECG. Amplification of these electrical heterogeneities can lead to the development of life-threatening cardiac arrhythmias and sudden death. A number of ionic distinctions have been shown to contribute to the different action potential morphologies of epicardial, M and endocardial ventricular cells as well as to the distinctive responses of these three cell types to pharmacological agents and pathophysiological states (for reviews see Antzelevitch *et al.* 1999; Antzelevitch & Dumaine, 2000).

Ventricular M and epicardial cells, but not endocardial cells, typically display action potentials with a prominent notch or phase 1, due to the presence of a large 4-aminopyridine (4-AP)-sensitive transient outward current ( $I_{to}$ ). The transmural distribution of  $I_{to}$  is among the most striking examples of electrical heterogeneity encountered in the ventricles of the canine and human heart. Regional differences in  $I_{to}$ , first suggested on the basis of action potential data, have now been demonstrated using voltage clamp techniques in canine, feline, rabbit, rat and human ventricular myocytes (Fig. 1; see Antzelevitch *et al.* 1999 for references). Transmural differences in the magnitude of the  $I_{to}$ -mediated action potential notch give rise to a transmural voltage gradient, which is responsible for the inscription of the electrocardiographic J wave. Accentuation of this gradient leads to the appearance of pathophysiological J waves, ST segment elevation

and the development of ventricular tachycardia and fibrillation in experimental models of the Brugada syndrome as well as under conditions of acute ischaemia.

Although the *Kv4.3* potassium channel gene has long been implicated in the expression of  $I_{to}$  channels in dog and man (Dixon & McKinnon, 1994; Kaab *et al.* 1998), investigators have been puzzled by the fact that *Kv4.3* message is not distributed across the ventricular wall in such a way as to account for the steep transmural gradient in the expression of the current. A report by Rosati and coworkers in this issue of *The Journal of Physiology* appears to have solved this mystery by providing evidence in support of the hypothesis that differential expression of *KChIP2*, a gene that strongly modulates the expression of the *Kv4.3* channel, is responsible for the transmural distribution of  $I_{to}$  in both canine and human hearts. *KChIP2* mRNA levels were found to be 25-fold higher in epicardium than in endocardium, whereas *Kv4.3* mRNA was expressed evenly across the ventricular wall. On the basis of this observation and the demonstration that *KChIP2*, when co-expressed with *Kv4.3* in *Xenopus* oocytes, dramatically increases  $I_{to}$ , the authors suggest that transmural differences in the expression of this ancillary ( $\beta$ ) subunit are responsible for the transmural gradient of  $I_{to}$  density. It is not as yet clear whether *KChIP2* merely increases sarcolemmal expression of *Kv4.3* protein ( $\alpha$  subunit of the  $I_{to}$  channel) or whether it influences channel function by increasing single channel conductance. Preliminary evidence points to increased surface membrane expression as the predominant mechanism by which *KChIP2* modulates  $I_{to}$ . The authors demonstrate a similar sensitivity of  $I_{to}$  in epicardial and endocardial cells to flecainide, suggesting that the current is produced principally by channels with *Kv4.3*, rather than *Kv1.4*,  $\alpha$  subunits in the two cell types.

Finally, the pioneering study by Rosati *et al.* (2001) highlights the divergence of molecular mechanisms regulating  $I_{to}$  in the hearts of small *vs.* large mammals. In small mammals, *Kv4.2*, sometimes in combination with *Kv4.3*,

is largely responsible for  $I_{to}$  (Brahmajothi *et al.* 1999; Dixon & McKinnon, 1994). In rat and other small mammals a gradient of *Kv4.2* mRNA across the ventricular wall appears to account for the transmural gradient in the density of  $I_{to}$ . The present study shows that *KChIP2* mRNA is expressed at uniform levels across the ventricular wall of the rat, although at levels much higher than in dog and human. Thus, in small mammals it is the distribution of the  $\alpha$  subunit (*Kv4.2*), whereas in large mammals it is the distribution of the  $\beta$  subunit (*KChIP2*), that underlies the transmural gradient of  $I_{to}$ . Whether similar transcriptional mechanisms are involved in the transmural expression of *Kv4.2* and *KChIP2* message remains to be established.

In addition to transmural distinctions, significant inter-ventricular and apico-basal differences in  $I_{to}$  have been described (Di Diego *et al.* 1996). Whether mechanisms similar to those described above are involved remains to be established and is of critical importance from the standpoint of our understanding and ability to deal with a number of life-threatening pathophysiological states. As one example, the Brugada syndrome, a sudden death syndrome affecting young adults, is a right ventricular disease owing to the fact that  $I_{to}$  is most prominent in the right ventricle of the heart.

ANTZELEVITCH, C. & DUMAINE, R. (2001). In *Handbook of Physiology, The Heart*, ed. PAGE, E., FOZZARD, H. A. & SOLARO, R. J. Oxford University Press, New York (in the Press).

ANTZELEVITCH, C., SHIMIZU, W., YAN, G. X., SICOURI, S., WEISSENBERGER, J., NESTERENKO, V. V., BURASHNIKOV, A., DI DIEGO, J. M., SAFFITZ, J. E. & THOMAS, G. P. (1999). *Journal of Cardiovascular Electrophysiology* **10**, 1124–1152.

BRAHMAJOTHI, M. V., CAMPBELL, D. L., RASMUSSEN, R. L., MORALES, M. J., TRIMMER, J. S., NERBONNE, J. M. & STRAUSS, H. C. (1999). *Journal of General Physiology* **113**, 581–600.

DI DIEGO, J. M., SUN, Z. Q. & ANTZELEVITCH, C. (1996). *American Journal of Physiology* **271**, H548–561.

DIXON, E. J. & MCKINNON, D. (1994). *Circulation Research* **75**, 252–260.

KAAB, S., DIXON, J., DUC, J., ASHEN, D., NABAUER, M., BEUCKELMANN, D. J., STEINBECK, G., MCKINNON, D. & TOMASELLI, G. F. (1998). *Circulation* **98**, 1383–1393.

ROSATI, B., PAN, Z., LYPEN, S., WANG, H.-S., COHEN, I., DIXON, J. E. & MCKINNON, D. (2001). *Journal of Physiology* **533**, 119–125.

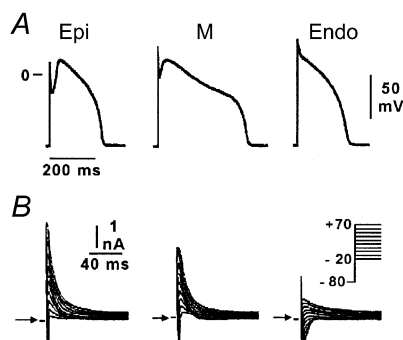


Figure 1

A, action potentials recorded from myocytes isolated from the epicardial (Epi), endocardial (Endo) and M regions of the canine left ventricle. B, transient outward current ( $I_{to}$ ) recorded from the three cell types during depolarizing steps from a holding potential of  $-80$  mV to test potentials ranging between  $-20$  and  $+70$  mV. Modified from Antzelevitch *et al.* (1999), with permission.