

# Biomimetic mushroom-shaped fibrillar adhesive microstructure

S. Gorb<sup>1,\*</sup>, M. Varenberg<sup>1</sup>, A. Peressadko<sup>1</sup> and J. Tuma<sup>2</sup>

<sup>1</sup>*Department Arzt, Max Planck Institute for Metals Research, Heisenbergstrasse 3, Stuttgart 70569, Germany*

<sup>2</sup>*Gottlieb Binder GmbH, Bahnhofstrasse 19, Holzgerlingen 71088, Germany*

To improve the adhesive properties of artificial fibrillar contact structures, the attachment systems of beetles from the family Chrysomelidae were chosen to serve as a model. Biomimetic mushroom-shaped fibrillar adhesive microstructure inspired by these systems was characterized using a variety of measurement techniques and compared with a control flat surface made of the same material. Results revealed that pull-off force and peel strength of the structured specimens are more than twice those of the flat specimens. In contrast to the control system, the structured one is found to be very tolerant to contamination and able to recover its adhesive properties after being washed in a soap solution. Based on the combination of several geometrical principles found in biological attachment devices, the presented microstructure exhibits a considerable step towards the development of an industrial dry adhesive.

**Keywords:** biomimetics; microstructures; surface patterning; adhesion; attachment

## 1. INTRODUCTION

Fibrillar attachment systems have evolved several times independently in various animal groups (Gorb 2001; Gorb & Beutel 2001). Their geometrical features, such as contact splitting, high aspect ratio of single contact elements and their spatula-shaped heads, were found to be responsible for generating strong adhesion. The physical background of this phenomenon was intensively discussed in several recent publications (Arzt *et al.* 2003; Persson 2003; Persson & Gorb 2003; Chung & Chaudhury 2005; Gao *et al.* 2005). It was confirmed that adhesion-oriented geometry of biological fibrillar attachment systems may be employed to design artificial surfaces with enhanced adhesion.

There have been a few attempts to produce biomimetic fibrillar surfaces to amplify adhesion of a flat contact (Geim *et al.* 2003; Ghatak *et al.* 2004; Majidi *et al.* 2004; Peressadko & Gorb 2004; Northen & Turner 2005; Yurdumakan *et al.* 2005). However, the reported surfaces fabricated using various techniques ranging from laser technology to microlithography did not, in fact, mimic the natural adhesive structures, and they featured limited life cycle and overall gain in adhesion.

To improve the adhesive properties of artificial fibrillar contact structures, the attachment systems of beetles from the family Chrysomelidae were chosen to serve as a model. Based on the study of numerous species of these beetles, it has been previously shown that they are extremely specialized for adhering to smooth surfaces (Stork 1980, 1983; Pelletier &

Smilowitz 1987; Gorb 2001). Analysis of the functional morphology of adhesive hairs on their tarsi (figure 1) allows fabrication of an advanced adhesive microstructure.

The purpose of the present work is to report on biomimetic mushroom-shaped fibrillar adhesive microstructure that is inspired by these natural attachment systems and represents a considerable step towards the development of an industrial dry adhesive.

## 2. MATERIAL AND METHODS

### 2.1. Specimen preparation

Fibrillar specimens were produced by Gottlieb Binder GmbH (Holzgerlingen, Germany) at room temperature by pouring two-compound polymerizing polyvinylsiloxane (PVS; Coltène Whaledent AG, Altstätten, Switzerland) into the holed template lying on a smooth glass support. Prospective specimen height was defined by spacers between the support and a covering flat surface that was used to squeeze superfluous polymer out of the gap. After polymerization, the ready-to-use cast with Young's modulus of about 3 MPa (Peressadko & Gorb 2004) was removed from the template. The backside of the fibrillar casts was used for flat specimens.

### 2.2. Scanning electron microscopy

Two portions of a structured surface were used for scanning electron microscopy (SEM). The first part was employed to analyse the morphology of a fibrillar structure. To this end, the structure's backing was attached to a carbon double-sided tape fixed onto

\*Author for correspondence (s.gorb@mf.mpg.de).

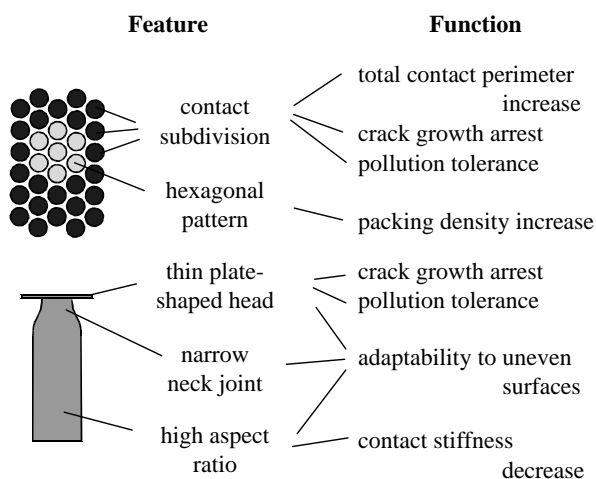


Figure 1. Schematic of the functional morphology of biological model attachment system.

a specimen holder. The second part was used to study the contact formed between artificial fibrils and flat smooth surface. For this purpose, its structured side was adhered to a small cover glass attached to the same specimen holder. The specimens were sputter coated with about 6 nm of gold–palladium for electrical conductivity and imaged in a Hitachi-4800 high-resolution SEM at accelerating voltage of 3 kV.

### 2.3. Pull-off test

The adhesion generated by structured and flat PVS surfaces in contact with a smooth flat glass substrate was tested on a home-made microtribometer. It consisted of a motorized precise translation stage (Physik Instrumente GmbH, Karlsruhe, Germany) and a fixed flexible cantilever (Tetra GmbH, Ilmenau, Germany), whose deflection, used to determine the contact forces, was measured with fibre-optic sensors (MTI Instruments, Inc., Albany, New York). The forces detected were calibrated with known precise weights immediately prior to experiments. To guarantee full intimate contact during measurements in a flat-on-flat contact scheme, a passive self-aligning system of specimen holders was used (Varenberg *et al.* 2006b). PVS specimens were discs of 2.9 mm diameter and 1.5 mm height mounted on the cantilever, and a glass cover slide of  $18 \times 7 \times 0.2 \text{ mm}^3$  in size fixed on the translation stage was used as a substrate. The roughness average ( $R_a$ ) of the glass and PVS surface was about 1 and 85 nm, respectively. Before the experiments, the specimens were washed with deionized water and liquid soap, and then dried in blowing nitrogen.

Each test started by preloading the specimens for 90 s. Under preloads between 50 and 130 mN, the real contact area was imaged with video zoom optics (Navitar Inc., Rochester, New York) to verify proper contact formation. Destructive interference of reflected white light in the glass–PVS interface resulted in a visualization of the real contact area as a much darker zone than the area that was out of contact. Subsequently, the pull-off force was measured while withdrawing the translation stage at a velocity of  $700 \mu\text{m s}^{-1}$ .

### 2.4. Peeling test

In these experiments, a PVS tape of 0.4 mm thickness and 25 mm width was adhered with its structured or flat side to a clean smooth glass substrate for a contact time of 30 s. Then, a constant load was applied to the tape at a variable angle from the substrate by fixing a dead weight to the tape edge and tilting the substrate. The angle between the tape and the substrate corresponding to a peeling crack speed of about  $100 \mu\text{m s}^{-1}$  was determined, and the process was repeated for different loads varied in the range of 70–300 mN. Before the experiments, the specimens were washed with deionized water and liquid soap, and then dried in blowing nitrogen.

A peeling test was also used to estimate the effect of contamination on the proper function of both structured and flat surfaces. In these experiments, the peeling angle was kept equal to  $12^\circ$ , and the peeling load corresponding to a crack speed of about  $900 \mu\text{m s}^{-1}$  was determined with clean and gradually contaminated specimens of both types. The degree of contamination was increased by placing the specimens on a dusty laboratory shelf between the evaluations of the peeling load. The shelf has not been touched over the last four years and dust particles ranging from a few micrometres to a few hundreds of micrometres in size have formed a uniform dust layer of a few hundreds of micrometres in height. Each time the specimens were placed on an untouched shelf surface, the increasing number of dust particles, which adhered to the specimens' surface in a naturally uncontrolled way, led to a subsequent decrease in their peeling strength. After finishing the contamination tests, the specimens were cleaned according to the above procedure and the peeling force was determined again to test their recovery ability. Portions of contaminated surfaces were also examined in SEM.

## 3. RESULTS AND DISCUSSION

### 3.1. Morphology

To reproduce the adhesive characteristics of the model biological system, its functional morphology was analysed and is represented in a simplified form in figure 1. Inspired by these design principles, a biomimetic mushroom-shaped fibrillar adhesive microstructure was fabricated. It is demonstrated in figure 2.

While principles of contact subdivision (Autumn *et al.* 2000; Scherge & Gorb 2001; Arzt *et al.* 2003) and hexagonal patterning (Ball 2001; Gorb 2001) were sufficiently discussed as mechanisms of adhesion enhancement in biological hairy attachment devices, the form of natural terminal contact elements has not yet received proper attention. The thin plate at the biological adhesive hair tip (figure 1) is very different from the flat-punch geometry used in most previously reported artificial patterned adhesives (Geim *et al.* 2003; Glassmaker *et al.* 2004; Peressadko & Gorb 2004; Crosby *et al.* 2005). Since the most effective biological attachment systems use spatula- or mushroom-shaped rather than the flat-punch geometry, this feature, which seems to be very important from the viewpoint of stress concentration and crack arrest, was implemented in the presented artificial adhesive system.

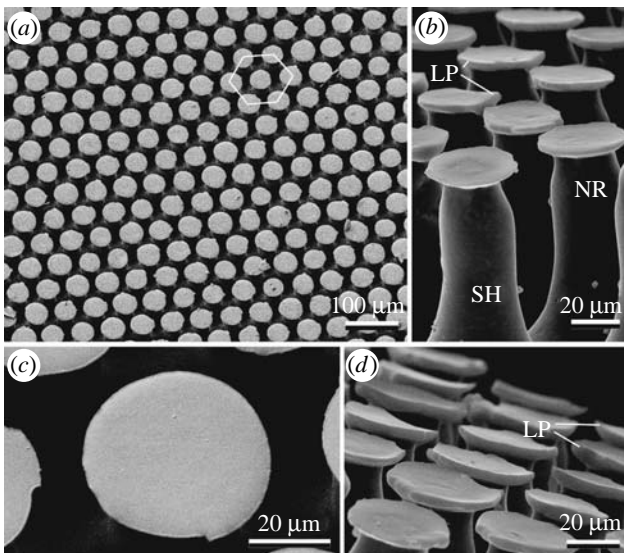


Figure 2. Biomimetic mushroom-shaped fibrillar adhesive microstructure made of PVS. (a, c) View from above; (b, d) side view. LP, contact plate lip; NR, narrow neck; SH, pillar shaft.

Structured in a hexagonal order (figure 2a), this system consisted of mushroom-shaped pillars of about 100  $\mu\text{m}$  in height, 60  $\mu\text{m}$  in base diameter, 35  $\mu\text{m}$  in middle diameter and 25  $\mu\text{m}$  in diameter at the narrowed region just below the terminal contact plates. These plates were of about 40  $\mu\text{m}$  in diameter and 2  $\mu\text{m}$  in thickness at the lip edges (figure 2b, d). The area density of the terminal contact plates was about 40%.

### 3.2. Performance

The contact formed by the studied PVS microstructure adhered to a smooth glass substrate is depicted in figure 3. It was found that the presence of terminal thin plates greatly improved the adaptability of the contact system. For instance, the terminal plates of the pillars, which were not ideally oriented perpendicular to the substrate, could still form a partial contact (figure 3b). These plates were even able to resist small surface unevenness or contaminations (figure 4d, e). Normally, only a part of the terminal plate was disabled in this case, while the rest of the plate remained in contact. This would be hardly possible having a flat-punch geometry.

Figure 4 presents the pull-off force measured between the PVS specimens and the glass substrate. Results revealed that the structured specimens featured a pull-off force more than twice that of the flat specimens, while in both cases it was independent of the preload. Two typical binarized images of structured and flat real contact area are also shown. Image analysis demonstrated that the flat surface had formed a real contact area about twice that of the structured surface, thus confirming that adhesion does not depend on the contact area (Varenberg et al. 2006a).

A peeling strength analysis was performed according to the following expression:

$$\left(\frac{F}{b}\right)^2 \frac{1}{2Ed} + \left(\frac{F}{b}\right)(1 - \cos \Theta) - R = 0, \quad (3.1)$$

where  $F$  is the peeling force;  $d$  is the thickness of the adhesive tape;  $b$  is the width of the tape;  $E$  is the

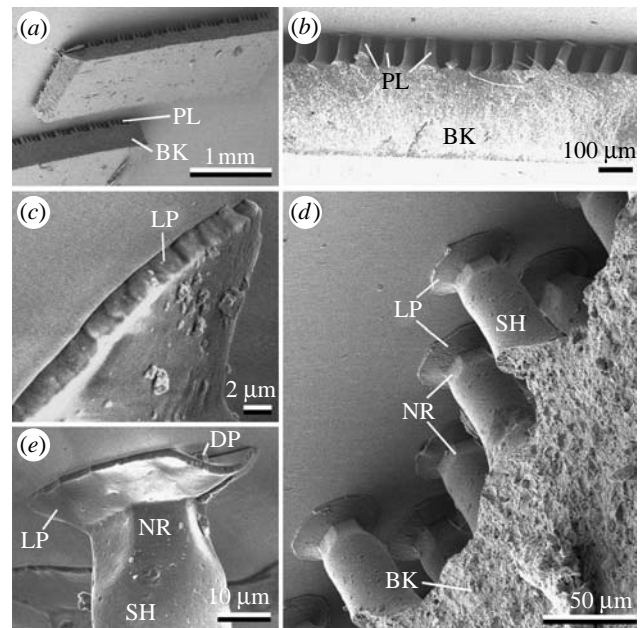


Figure 3. Structured PVS surface in contact with glass substrate. BK, backing; DP, dirt particle; LP, contact plate lip; NR, narrow neck; PL, pillar; SH, pillar shaft.

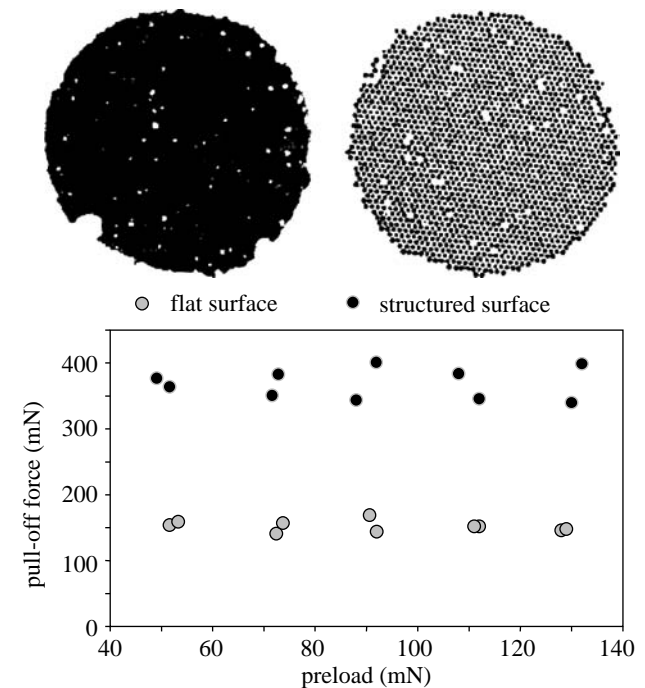


Figure 4. Pull-off force measured between PVS and glass specimens as a function of preload. Inserts are binarized images of real contact area for flat and structured surfaces.

Young's modulus of the tape material;  $\Theta$  is the peeling angle; and  $R$  is the energy required to fracture a unit area of interface (Kendall 1975). The fracture energy,  $R$ , of both structured and flat specimens was estimated using predefined values of peeling force and respective experimentally determined peeling angles. The mean fracture energy corresponding to a peeling crack speed of 100  $\mu\text{m s}^{-1}$  was found to be 1.38 and 0.51  $\text{J m}^{-2}$  for structured and flat tape, respectively. These values demonstrate again that the structured specimens featured adhesion more than twice that of the flat specimens. These results are also represented in figure 5,

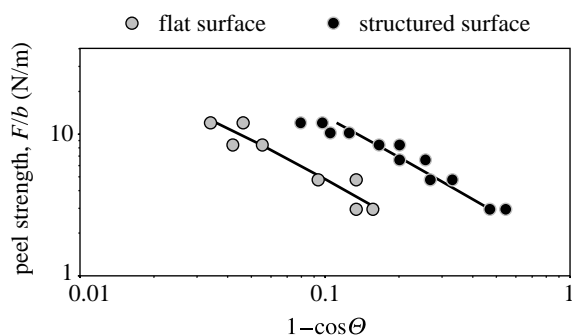


Figure 5. Dependence of peel strength on peel angle for a constant crack speed of  $100 \mu\text{m s}^{-1}$  obtained for flat and structured PVS surfaces. Markers correspond to experimental data and solid lines to theoretical predictions.

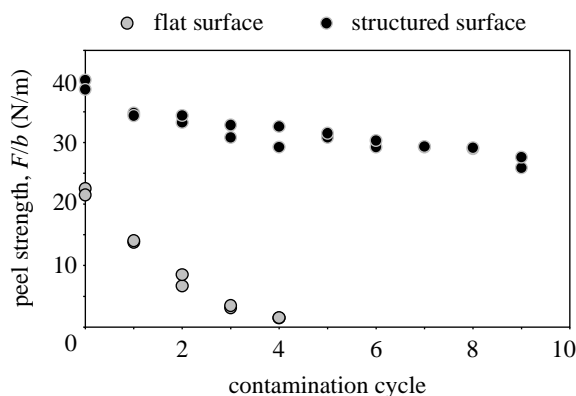


Figure 6. Effect of contamination on the peel strength of flat and structured PVS surfaces.

where the experimental data are plotted along with the theoretical predictions calculated from expression (3.1) using the mean values of the fracture energy.

Figure 6 demonstrates the effect of contamination on the peel strength of both flat and structured specimens. It is clearly seen that while the flat specimens experience dramatic decrease in peel strength and almost completely lose their ability to adhere during the very first contamination cycles, the structured specimens are much more tolerant to the degree of contamination and continue functioning steadily. Washed surfaces of both types were able to recover in full their peel strength, thus demonstrating that the observed adhesion reduction has resulted solely from contamination.

### 3.3. Analysis

The most recent empirical explanation of adhesion enhancement with contact subdivision is that adhesion is proportional to a total contact perimeter (Varenberg *et al.* 2006a). This results in a scaling effect with finer structures exhibiting stronger adhesion. However, attachment properties of any surface are defined not only by its ability to adhere, but also by the forces resisting contact deformation required to maximize contact zone and generate adhesion. Adhesion and the forces resisting contact deformation act in the opposite directions and the resulting pull-off force, or the external load needed to detach one surface from another, is the difference between the two. Thus, an ultimate

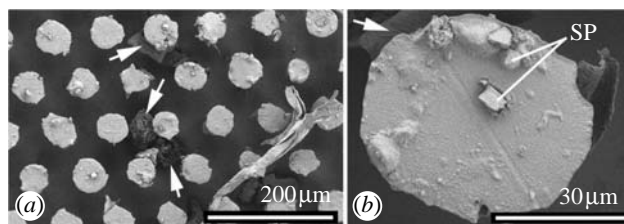


Figure 7. Contaminated structured PVS surface. Arrows point to dirt particles sunk into the space between the pillars. SP, small dirt particles adhered to flat contact plate.

requirement for an effective attachment system is to generate maximal adhesion with minimal elastic strain energy. For instance, the structured contact composed of pillars having a uniform height will perform a better pull-off force than the same structure having a stochastic distribution of pillar heights. This despite both contacts will presumably generate approximately the same adhesion in a loaded compressed state.

Low contact stiffness of a fibre array results in minimizing contact forces and providing a high adaptability of a structured surface to a substrate. The presence of structural hierarchical levels, which are widely spread in biological systems, is one of the features used to achieve this purpose. For instance, insects use two levels of outgrowths in their attachment organs, spiders evolved three levels (Gorb 2001) and geckos employ four levels of hierarchy (Hiller 1968; Autumn *et al.* 2000; Huber *et al.* 2005; Rizzo *et al.* 2006). In contrast to most previously reported artificial patterned adhesives (Geim *et al.* 2003; Glassmaker *et al.* 2004; Peressadko & Gorb 2004; Crosby *et al.* 2005), which have only one hierarchy level of structuring, the presented mushroom-shaped fibrillar adhesive involves two levels. This presumably makes the system more tolerant to uneven real surfaces. The first hierarchic level is represented by pillars, which provide some general adaptability, and the second level is represented by the flat terminal plates, which can adapt to local surface irregularities owing to their thin lips and narrowed flexible joints with pillars. These two hierarchical levels resemble, in a way, a recently reported sandwich-shaped structured adhesive, where pillars were enclosed between their backing and an upper covering thin film (Glassmaker *et al.* 2006). Another advantage of this second hierarchic level is that thin lips at terminal plates are more effective in arresting crack growth, which is a very important result of contact subdivision (Chung & Chaudhury 2005).

One more remarkable feature of contact splitting is related to its pollution resistance. Experimental evidence for the reduction of contamination in a gecko hairy adhesive system has been recently reported for the first time (Hansen & Autumn 2005). Self-cleaning in gecko setae was explained by an energetic disequilibrium between the adhesive forces attracting a dirt particle to the substrate and those attracting the same particle to one or more spatulae. SEM observations of contaminated artificial surfaces studied in the present work show that the advantage of the structured systems can be explained by several additional effects: (i) the thin contact plate terminating each pillar is flexible enough to form a reliable contact even in the

presence of small dirt particles (figure 3e); (ii) large particles can slip into the space between pillars (figure 7) and be accommodated there with a negligible effect on contact features; and (iii) disabled pillars do not alter the performance of their neighbours.

#### 4. CONCLUSION

Adhesive properties of a biomimetic mushroom-shaped fibrillar adhesive microstructure and a control flat surface made of the same material were characterized using a variety of measurement techniques. Despite rather coarse patterning, adhesive features of the structured surface were more than twice as effective as of the flat one. The higher tolerance of the patterned surface to contamination as well as its ability to recover by washing in a liquid soap solution were experimentally verified.

Based on the combination of several principles found in biological attachment devices, the presented microstructure exhibits a considerable step towards the development of an industrial dry adhesive. It can be currently produced in any size up to an A4 format and is suitable for use in a protective tape for sensitive glass surfaces or in reusable pads for the attachment of small objects to smooth surfaces. In fact, it was successfully used in the attachment feet of a 120 g wall-walking robot (Daltorio et al. 2005).

This work was supported by the Federal Ministry of Education, Science and Technology, Germany to S.G. (project BioFuture 0311851).

#### REFERENCES

- Arzt, E., Gorb, S. & Spolenak, R. 2003 From micro to nano contacts in biological attachment devices. *Proc. Natl Acad. Sci. USA* **100**, 10 603–10 606. (doi:10.1073/pnas.1534701100)
- Autumn, K., Liang, Y. A., Hsieh, S. T., Zesch, W., Chan, W. P., Kenny, T. W., Fearing, R. & Full, R. J. 2000 Adhesive force of a single gecko foot-hair. *Nature* **405**, 681–685. (doi:10.1038/35015073)
- Ball, P. 2001 *The self made tapestry: pattern formation in nature*. Oxford, UK: Oxford University Press.
- Chung, J. Y. & Chaudhury, M. K. 2005 Roles of discontinuities in bio-inspired adhesive pads. *J. R. Soc. Interface* **2**, 55–61. (doi:10.1098/rsif.2004.0020)
- Crosby, A. J., Hageman, M. & Duncan, A. 2005 Controlling polymer adhesion with “pancakes”. *Langmuir* **21**, 11 738–11 743. (doi:10.1021/la051721k)
- Daltorio, K. A., Gorb, S., Peressadko, A., Horchler, A. D., Ritzmann, R. E. & Quinn, R. D 2005 A robot that climbs walls using micro-structured polymer feet. In *Proc. Int. Conf. Climbing and Walking Robots, London, UK*, pp. 131–138.
- Gao, H. J., Wang, X., Yao, H. M., Gorb, S. & Arzt, E. 2005 Mechanics of hierarchical adhesion structures of geckos. *Mech. Mater.* **37**, 275–285. (doi:10.1016/j.mechmat.2004.03.008)
- Geim, A. K., Dubonos, S. V., Grigorieva, I. V., Novoselov, K. S. & Zhukov, A. A. 2003 Microfabricated adhesive mimicking gecko foot-hair. *Nat. Mater.* **2**, 461–463. (doi:10.1038/nmat917)
- Ghatak, A., Mahadevan, L., Chung, J. Y., Chaudhury, M. K. & Shenoy, V. 2004 Peeling from a biomimetically patterned elastic film. *Proc. R. Soc. A* **460**, 2725–2735. (doi:10.1098/rspa.2004.1313)
- Glassmaker, N. J., Jagota, A., Hui, C.-Y. & Kim, J. 2004 Design of biomimetic fibrillar interfaces: 1. Making contact. *J. R. Soc. Interface* **1**, 23–33. (doi:10.1098/rsif.2004.0004)
- Glassmaker, N. J., Jagota, A., Chaudhury, M. K. & Hui, C.-Y. 2006 Contact and adhesion mechanics of biomimetic fibrillar interfaces. In *Proc. Adhes. Soc.* 93–95.
- Gorb, S. N. 2001 *Attachment devices of insect cuticle*. New York, NY: Springer.
- Gorb, S. N. & Beutel, R. G. 2001 Evolution of locomotory attachment pads of hexapods. *Naturwissenschaften* **88**, 530–534. (doi:10.1007/s00114-001-0274-y)
- Hansen, W. R. & Autumn, K. 2005 Evidence for self-cleaning in gecko setae. *Proc. Natl Acad. Sci. USA* **102**, 385–389. (doi:10.1073/pnas.0408304102)
- Hiller, U. 1968 Untersuchungen zum Feinbau und zur Funktion der Haftborsten von Reptilien. *Z. Morphol. Tiere* **62**, 307–362. (doi:10.1007/BF00401561)
- Huber, G., Gorb, S. N., Spolenak, R. & Arzt, E. 2005 Resolving the nanoscale adhesion of individual gecko spatulae by atomic force microscopy. *Biol. Lett.* **1**, 2–4. (doi:10.1098/rsbl.2004.0254)
- Kendall, K. 1975 Thin-film peeling—the elastic term. *J. Phys. D: Appl. Phys.* **8**, 1449–1452. (doi:10.1088/0022-3727/8/13/005)
- Majidi, C., Groff, R. & Fearing, R. 2004 Clumping and packing of hair arrays manufactured by nanocasting. In *Proc. ASME IMECE*, 62142.
- Northen, M. T. & Turner, K. L. 2005 A batch fabricated biomimetic dry adhesive. *Nanotechnology* **16**, 1159–1166. (doi:10.1088/0957-4484/16/8/030)
- Pelletier, Y. & Smilowitz, Z. 1987 Specialized tarsal hairs on adult male Colorado potato beetles, *Leptinotarsa decemlineata* (Say), hamper its locomotion on smooth surfaces. *Can. Entomol.* **119**, 1139–1142.
- Peressadko, A. & Gorb, S. N. 2004 When less is more: experimental evidence for tenacity enhancement by division of contact area. *J. Adhes.* **80**, 1–15. (doi:10.1080/00218460490430199)
- Persson, B. N. J. 2003 On the mechanism of adhesion in biological systems. *J. Chem. Phys.* **118**, 7614–7621. (doi:10.1063/1.1562192)
- Persson, B. N. J. & Gorb, S. 2003 The effect of surface roughness on the adhesion of elastic plates with application to biological systems. *J. Chem. Phys.* **119**, 11 437–11 444. (doi:10.1063/1.1621854)
- Rizzo, N. W., Gardner, K. H., Walls, D. J., Keiper-Hrynko, N. M., Ganzke, T. S. & Hallahan, D. L. 2006 Characterization of the structure and composition of gecko adhesive setae. *J. R. Soc. Interface* **3**, 441–451. (doi:10.1098/rsif.2005.0097)
- Scherge, M. & Gorb, S. N. 2001 *Biological micro- and nanotribology: nature's solutions*. Berlin, Germany: Springer.
- Stork, N. E. 1980 Experimental analysis of adhesion of *Chrysolina polita* (Chrysomelidae, Coleoptera) on a variety of surfaces. *J. Exp. Biol.* **88**, 91–107.
- Stork, N. E. 1983 The adherence of beetle tarsal setae to glass. *J. Nat. Hist.* **17**, 583–597.
- Varenberg, M., Peressadko, A., Gorb, S. & Arzt, E. 2006a Effect of real contact geometry on adhesion. *Appl. Phys. Lett.* **89**, 121 905. (doi:10.1063/1.2356099)
- Varenberg, M., Peressadko, A., Gorb, S., Arzt, E. & Mroczek, S. 2006b Advanced testing of adhesion and friction with a microtribometer. *Rev. Sci. Instrum.* **77**, 066105. (doi:10.1063/1.2214692)
- Yurdumakan, B., Raravikar, N. R., Ajayan, P. M. & Dhinojwala, A. 2005 Synthetic gecko foot-hairs from multiwalled carbon nanotubes. *Chem. Commun.* **30**, 3799–3801. (doi:10.1039/b506047h)