

Docetaxel and gemcitabine activity in NSCLC cell lines and in primary cultures from human lung cancer

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Summary The activity of the following drugs was investigated in two established NSCLC cell lines: docetaxel, gemcitabine, vinorelbine, paclitaxel, doxorubicin (0.01, 0.1, 1 $\mu\text{g ml}^{-1}$), cisplatin, ifosfamide (1, 2, 3 $\mu\text{g ml}^{-1}$) and carboplatin (2, 4, 6 $\mu\text{g ml}^{-1}$). The cytotoxic activity was evaluated by the sulphorhodamine B assay. The two most active drugs, docetaxel and gemcitabine, used singly and in association, were investigated as a function of treatment schedule. The sequence docetaxel→gemcitabine produced only a weak synergistic interaction in RAL but a strong synergism in CAEP cells. The synergistic interaction increased in both cell lines after a 48-h washout between the drug administrations. Flow cytometric analysis showed that in docetaxel→gemcitabine sequence, docetaxel produced a block in G2/M phase and, after 48 h, provided gemcitabine with a large fraction of recovered synchronized cells in the G1/S boundary, which is the specific target phase for gemcitabine. Conversely, simultaneous treatment induced an antagonistic effect in both cell lines, and the sequential scheme gemcitabine→docetaxel produced a weak synergistic effect only in RAL cells. Moreover, the synergistic interaction disappeared when washout periods of 24 or 48 h between two drug administrations were adopted. The synergistic activity of docetaxel→ 48-h washout→gemcitabine was confirmed in 11 of 14 primary cultures, which represents an important means of validating experimental results before translating them into clinical practice. © 1999 Cancer Research Campaign

Keywords: combination regimens; docetaxel; gemcitabine; NSCLC cell lines; NSCLC primary cultures; preclinical therapy

Non-small-cell lung cancer (NSCLC) is considered one of the most chemoresistant tumours, and in a recent overview the pessimism about the absolute survival benefits from chemotherapy was underlined (Non-small Cell Lung Cancer Collaborative Group, 1995). In particular, the meta-analysis revealed a survival benefit of 10% at 1 year in a supportive care setting and an increased median survival of 1.5 months in patients treated with platinum-based chemotherapy regimens, thus emphasizing the need for new, effective drugs and drug combination regimens.

Phase I–II clinical studies have shown that new drugs such as gemcitabine and taxanes, used singly (Carino et al, 1997; Cortes-Funes et al, 1997; Belani et al, 1998; Boyer et al, 1998; Natale et al, 1998; Takada et al, 1998) or in combination (Georgoulas et al, 1997a, 1997b), are active in NSCLC. Clinical protocols for cancer chemotherapy tend to use two or more drugs rather than single agents. Polychemotherapeutic protocol design is very complex. It is mainly based on information derived from experimental *in vitro* and *in vivo* studies and has favoured combinations of drugs with complementary mechanisms of action. Conversely, drug delivery schedules are planned without experimental preclinical information.

Preclinical studies have shown different interaction patterns of cisplatin and gemcitabine (Peters et al, 1995; van Moorsel et al, 1998), and paclitaxel and gemcitabine (Kroep et al, 1998) activity as a function of treatment schedule in some human tumour cell lines.

Moreover, recent studies have shown the importance of recognizing specific perturbations induced by the different drugs on cell cycle when designing combination or sequential therapies in order to increase additive or synergistic effects and avoid antagonistic effects (Hahn et al, 1993; Theodossiou et al, 1998; Zoli et al, 1999).

We investigated the cytotoxic activity of docetaxel and gemcitabine, as well as their interaction as a function of treatment schedule and attributed their activity to induced cell cycle perturbations. With a view to translating preclinical information to clinical practice, the study was conducted on two cell lines derived from an epidermoid carcinoma and from an adenocarcinoma obtained and characterized in our laboratory, and on primary lung cancer cultures, considered the *in vitro* system which best reproduces the biology of clinical tumours.

MATERIALS AND METHODS

Established cell lines

The study was performed in two established NSCLC cell lines (obtained and characterized in our laboratory) representative of different lung cancer histotypes: the CAEP cell line derived from an epidermoid carcinoma and the RAL cell line derived from an adenocarcinoma (Gasperi-Campani et al, 1998). Cells were maintained as a monolayer at 37°C and subcultured weekly. Culture medium was composed of Dulbecco's modified Eagle's medium (DMEM)/Ham's F12 (1:1) supplemented with fetal calf serum (FCS; 10%), glutamine (2 mM), non-essential amino acids (1%) and insulin (10 $\mu\text{g ml}^{-1}$). Cells in the exponential growth phase were used for all the experiments.

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Primary cell cultures

Tumour material was obtained from 14 patients who underwent thoracotomy for primary lung cancer: six adenocarcinomas, four squamous cell carcinomas, three atypical and one typical carcinoids. The areas of gross necrosis were removed from samples, and tumour tissue was carefully minced in a small volume of culture medium (Ham's F12 supplemented with FCS (12%), L-glutamine (1%), insulin (1%), polymyxin B (50 U ml⁻¹), fungizone (5 µg ml⁻¹) and penicillin–streptomycin (50 U ml⁻¹) and reduced to fragments of about 1 mm. Samples were repeatedly passed through hypodermic needles of decreasing diameter. The resulting suspension was filtered through a nylon mesh (50 gauge), and a suspension consisting of single cells or small groups of cells was obtained. Cells were then washed two or three times in culture medium, collected by centrifugation for 5 min at 200 *g* and finally resuspended in fresh medium [DCCM supplemented with FCS (0.5%), 3,3'-triiodo-L-thyronine (1 × 10⁻⁸ M), epidermal growth factor (5 ng ml⁻¹), insulin (1%), glutamine (1%), hydrocortisone (0.1 µg ml⁻¹), 17β-estradiol (1 × 10⁻⁹ M), choleric toxin (1 ng ml⁻¹)] to perform the sulphorhodamine B (SRB) assay.

Drugs

Vinorelbine (Pierre Fabre Pharma, Milan, Italy), carboplatin (Bristol Meyers Squibb, Latina, Italy), and docetaxel (Rhône-Poulenc Rorer, Varese, Italy) were diluted with sterile physiological solution at a concentration of 10 mg ml⁻¹, paclitaxel (Bristol Meyers Squibb) at 6 mg ml⁻¹, gemcitabine (Lilly, Sesto Fiorentino [FI], Italy), doxorubicin (Pharmacia, Milan, Italy) and ifosfamide (Asta Medica, Milan, Italy) at 1 mg ml⁻¹, and cisplatin (Iketon, Milan, Italy) at a concentration of 0.5 mg ml⁻¹, divided into aliquots, and stored at -20°C. Drug stocks were freshly diluted in culture medium before any experiment.

In vitro chemosensitivity assay

The SRB assay according to the method of Skehan et al (1990) was used. Briefly, cells in the exponential phase of growth were collected by trypsinization, counted and plated in 96-well flat-bottomed microtitre plates (100 µl of cell suspension per well). Experiments were run in octoplet, and each experiment was repeated three times. Eighteen to 24 h after plating (a sufficient time for exponential growth recovery), 100 µl of culture medium containing or not the specific drugs were added to the wells. At the

end of drug exposure, cells were fixed with 50% trichloroacetic acid at 4°C (10 µl per well, final concentration 10%) for 1 h. After five washes with tap water, cells were stained with 0.4% SRB dissolved in 1% acetic acid (50 µl per well) for 30 min and subsequently washed four times with 1% acetic acid to remove unbound stain. Plates were air-dried, and bound protein stain was solubilized with 100 µl of 10 mM unbuffered Tris base [Tris (hydroxymethyl)aminomethane]. The optical density of treated cells was read, at a wavelength of 540 or 510 nm, by means of a fluorescence plate reader.

Single drug exposure

After preliminary experiments, drugs were used at scalar concentrations of 0.01, 0.1 and 1 µg ml⁻¹ for docetaxel and gemcitabine. Cells were exposed to the single drugs for 48 h.

Drug combinations

Docetaxel and gemcitabine were tested using different combination and sequence schedules. Exposure time to each of the two drugs was 24 h. Docetaxel and gemcitabine were tested at all three concentrations (0.01, 0.1 and 1 µg ml⁻¹) in combination schemes and when they were used as the first drug in the sequential schemes. The lowest concentration (0.01 µg ml⁻¹) was used when docetaxel or gemcitabine was administered as the second drug in the sequential schemes.

Primary cell cultures were treated with the drug combination, schedule and timing that proved most effective in the established cell lines.

Flow cytometric analysis

For the analysis of cell cycle perturbations, exponentially growing cells were trypsinized, rinsed and plated (3 × 10⁵ cells per dish) into 60-mm Petri dishes and incubated for 18–24 h at 37°C before drug exposure. Medium was aspirated from the plates, and 0.01 µg ml⁻¹ of docetaxel or gemcitabine was added to the exponentially growing cells. Control dishes were cultured using the same conditions, with comparable media changes. After a 24-h exposure to the drugs, cells were trypsinized, washed twice with phosphate-buffered saline and resuspended in 1 ml of 4,6-diamino-2-phenylindole (DAPI). For the determination of DNA content and S phase cell fraction in primary cultures, surgical samples were minced in 2 ml of DAPI for 3 min. Cells from cell lines and human tumours were then filtered through a disposable 40-µm filter assembly (RATCOM, Inc., Miami, FL, USA). Human lymphocytes were utilized as internal standard. For every sample, 30 000 cells were analysed by flow cytometry (RATCOM), and the data obtained were elaborated using Modfit (DNA Modeling System) software.

Statistical analysis

To quantify deviations from additive effects induced by the sequential administration of two drugs, a statistical Student's *t*-test was employed (Drewinko et al, 1976). For a given drug dose, we determined a surviving fraction (Sf) of cells: SfA for the first drug used in the sequential schemes and SfB for the second. Following combined administration, we determined SfAB. Additivity held, resulting in SfAB = SfA × SfB, so that our estimate of deviation from additivity was the quantity SfAB – (SfA × SfB). The ratio of

Table 1 Cytotoxic effect of drugs on CAEP and RAL cell lines

Drugs	Concentrations (µg ml ⁻¹)	Mean IC ₅₀ (µg ml ⁻¹) ^a	
		CAEP	RAL
Docetaxel	0.01, 0.1, 1	0.030 ± 0.01	0.085 ± 0.01
Gemcitabine	0.01, 0.1, 1	0.034 ± 0.009	0.530 ± 0.10
Navelbine	0.01, 0.1, 1	0.037 ± 0.01	Not reached
Paclitaxel	0.01, 0.1, 1	0.360 ± 0.12	0.833 ± 0.06
Doxorubicin	0.01, 0.1, 1	Not reached	Not reached
Cisplatin	1, 2, 3	Not reached	Not reached
Ifosfamide	1, 2, 3	Not reached	Not reached
Carboplatin	2, 4, 6	Not reached	Not reached

^aAfter a 48-h treatment.

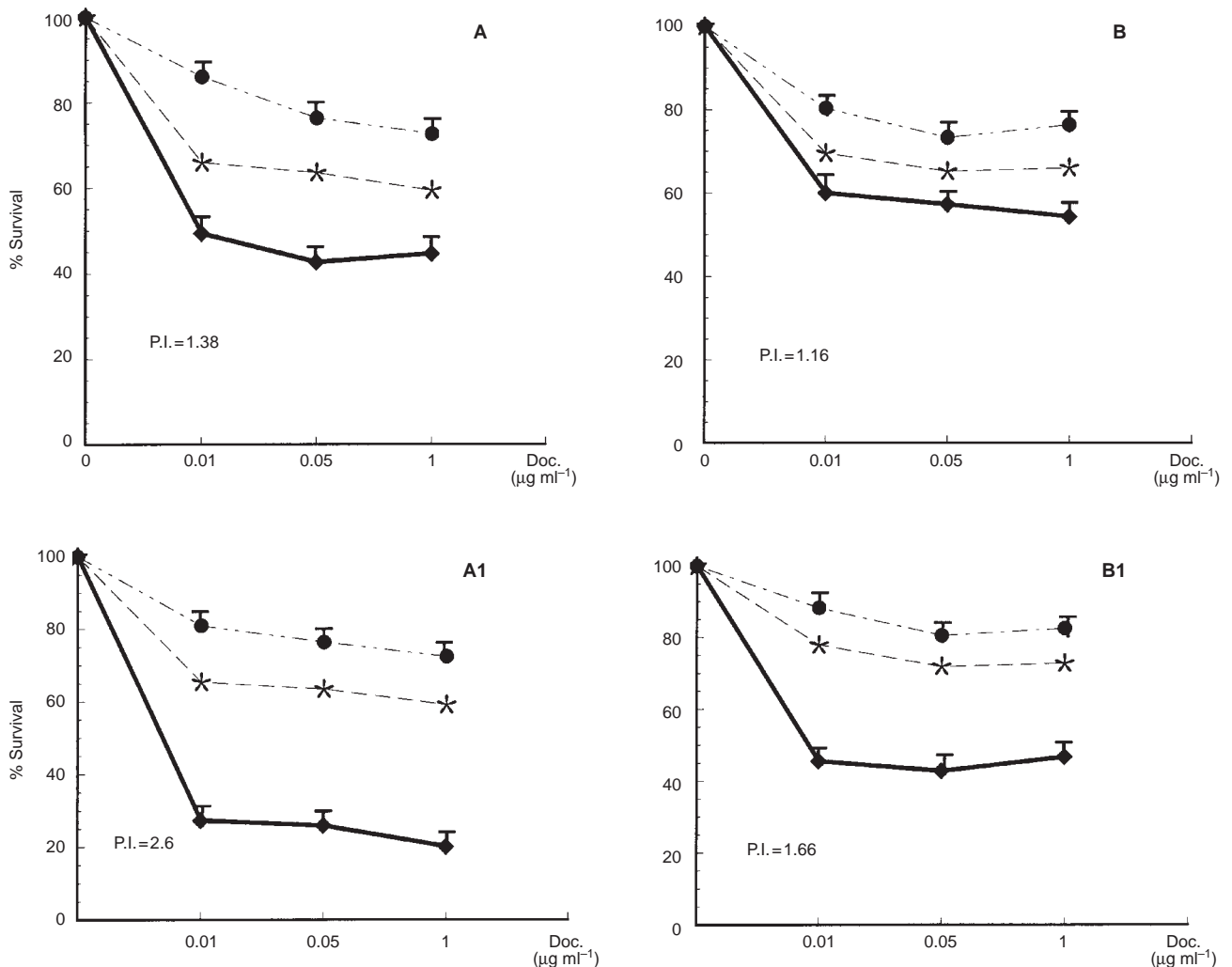


Figure 1 Dose-response survival curves of CAEP (A and A1) and RAL (B and B1) cells exposed to: (A) docetaxel for 24 hr followed by gemcitabine ($0.01 \mu\text{g ml}^{-1}$) for 24 hr; (A1) docetaxel for 24 hr followed by a 48-hr washout and then gemcitabine ($0.01 \mu\text{g ml}^{-1}$) for 24 hr; (B) docetaxel for 24 hr followed by gemcitabine (0.01mg ml^{-1}) for 24 hr; (B1) docetaxel for 24 hr followed by a 48-hr washout and then gemcitabine ($0.01 \mu\text{g ml}^{-1}$) for 24 hr. Docetaxel, observed survival —♦—; docetaxel→gemcitabine, —♦—; expected survival, —*—.

differences between observed versus expected survival and the square root of the relative variances for all drug combinations examined were, in fact, distributed normally, with the average equalling 0 and the variance equalling 1. The results obtained were defined according to the following criteria: $SfAB = SfA \times SfB$ indicated an additive effect, $SfAB < SfA \times SfB$, a synergistic effect, and $SfAB > SfA \times SfB$, an antagonistic effect. In drug combination studies, the performance index (PI) statistic model was used to evaluate type of interaction (Drewinko et al, 1976).

RESULTS

Established cell lines

The CAEP cell line was generally more sensitive to all the drugs than the RAL cell line. In particular, of the eight tested drugs, docetaxel and gemcitabine were the two most effective in both cell lines (Table 1).

The 24-h treatment with docetaxel followed by 24-h with gemcitabine (Figure 1) produced only a weak synergistic interaction in

the RAL cell line (PI = 1.16) but a strong synergism in the CAEP cell line (PI = 1.38). The synergistic interaction following the sequence was further increased by a 48-h washout in between the two-drug treatments in RAL cells (PI = 1.65) and even more so in the most sensitive cell line, CAEP (PI = 2.6) (Figure 1).

The sequential scheme of a 24-h treatment with gemcitabine immediately followed by docetaxel produced (Figure 2) a weak synergistic effect only in the RAL cell line (PI = 1.15). The synergistic interaction disappeared when washout periods of 24 or 48 h in between the two-drug treatments were adopted (Figure 2).

The simultaneous administration of docetaxel and gemcitabine induced (Figure 3) an antagonistic interaction in both cell lines at all docetaxel concentrations. The antagonistic effect was consistently observed when a single concentration of docetaxel and increasing concentrations of gemcitabine were tested (data not shown).

Cell cycle perturbations were analysed by flow cytometric analysis in an attempt to explain the mechanism underlying the synergistic interaction. In RAL cells, a 24-h treatment with gemcitabine caused an increase in G0/G1 phase cells and a dramatic decrease in the G2/M phase, which were still present at the 72-h

Table 2 Percentage of cells in different cell cycle phases at different times after a 24-h treatment with gemcitabine ($0.01 \mu\text{g ml}^{-1}$)

Cell cycle phase	Control samples	Times following treatment			
		0 h	24 h	48 h	72 h
RAL					
G ₀ -G ₁	50.2	70.1	70.2	69.7	67.4
S	35.6	28.0	28.0	29.1	32.3
G ₂ -M	14.2	1.9	1.8	1.2	0.3
Debris	9.6	23.7	5.3	5.7	8.6
CAEP					
G ₀ -G ₁	55.1	65.4	63.6	70.7	63.5
S	26.1	24.5	26.5	28.1	30.5
G ₂ -M	18.8	10.1	9.9	1.2	6.0
Debris	21.6	22.4	12.2	5.7	6.4

Table 3 Percentage of cells in different cell cycle phases at different times after a 24-h treatment with docetaxel ($0.01 \mu\text{g ml}^{-1}$)

Cell cycle phase	Control samples	Times following treatment			
		0 h	24 h	48 h	72 h
RAL					
G ₀ -G ₁	50.2	30.1	11.4	13.4	20.2
S	35.5	34.4	22.2	28.4	42.6
G ₂ -M	14.3	35.5	66.4	58.2	37.2
Debris	10.0	17.4	58.8	64.1	57.2
CAEP					
G ₀ -G ₁	55.1	20.5	11.2	13.4	34.1
S	26.0	21.5	10.2	27.4	27.0
G ₂ -M	18.9	58.0	78.6	59.2	48.9
Debris	21.6	50.7	75.6	54.2	36.0

In previous studies on human breast cancer cell lines and primary breast cancer cultures, we showed (Amadori et al, 1996) that treatment with doxorubicin followed by paclitaxel was more cytotoxic than simultaneous drug administration or the inverse sequence of

paclitaxel followed by doxorubicin. We also showed that the sequence defined at a preclinical level actually resulted in a high therapeutic efficacy in advanced breast cancer patients (Amadori et al, 1996; Frassinetti et al, 1997). In a further preclinical study on human breast cancer cell lines (Zoli et al, 1999), we observed a major therapeutic improvement using the sequence doxorubicin → paclitaxel → 48-h washout → gemcitabine. The schedule-dependent activity of multidrug regimens observed by us and other authors thus emphasizes the importance of preclinical studies (Peters et al, 1995; Kroep et al, 1998; van Moorsel et al, 1998).

In the present study, we used two NSCLC cell lines which reproduce a clinical situation since they proved to be highly sensitive to docetaxel and gemcitabine, in agreement with results from preclinical and phase I-II studies on NSCLC (Carino et al, 1997; Cortes-Funes et al, 1997; Boyer, 1998; Natale et al, 1998; Takada et al, 1998). The sequence gemcitabine → 48-h washout → docetaxel produced a low synergistic effect in both cell lines. This can, in part, be attributed to a block induced by gemcitabine in the G₀/G₁ phase that recovered after 72 h. Such a block may prevent exposure of the cells to the cytotoxic effect of docetaxel when the drug is administered immediately or 48 h after gemcitabine treatment. Conversely, the cytotoxic effect obtained with the opposite schedule, docetaxel → 48-h washout → gemcitabine, produced an evident synergistic effect in CAEP and a strong synergistic effect in RAL cell lines. Cell cycle perturbation analysis following this treatment schedule indicated that docetaxel produced an initial block in the G₂/M phase, thus providing a large fraction of recovered synchronized cells in the G₁/S boundary, which is the specific target phase for the antimetabolite (Hertel et al, 1990; Theodossiou et al, 1998), for the subsequent treatment with gemcitabine. It should be pointed out that the synergistic effect was already present at the lowest drug concentration.

Our findings are in agreement with those of Theodossiou et al (1998), who observed an antagonistic effect of gemcitabine and paclitaxel when administered simultaneously in the A549 lung cancer cell line. The same study showed a slightly less than additive cytotoxic effect when gemcitabine administration preceded that of paclitaxel, or after the inverse sequence. Conversely, using

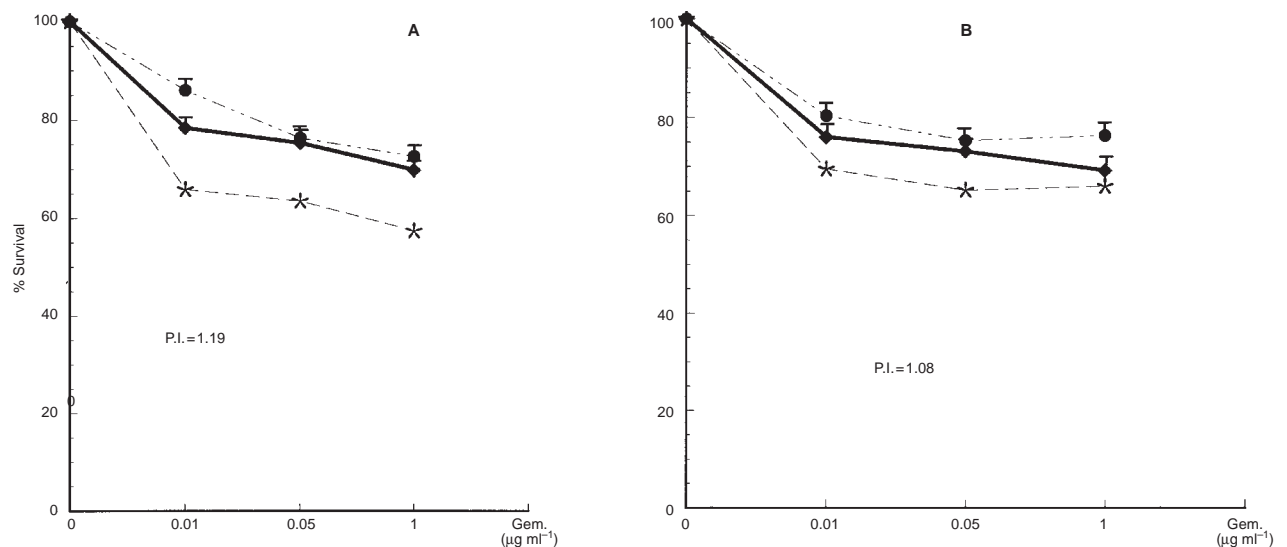
**Figure 3** Dose-response survival curves of CAEP (A) and RAL (B) cells exposed to: docetaxel plus gemcitabine ($0.01 \mu\text{g ml}^{-1}$) for 24 hr. Docetaxel, observed survival —●—●—●—●—; docetaxel→gemcitabine, —*—*—*—*— expected survival, —◆—◆—◆—◆— observed survival.

Table 4 Cytotoxic effects of sequential treatment, docetaxel (0.01, 0.1 and 1 µg ml⁻¹ for 24 h) followed by a 48-h washout and then gemcitabine (0.01 µg ml⁻¹ for 24 h), observed in 14 primary lung cancer cultures

Sample	Histotype	Cytotoxic effect	Performance index	DNA content	FCM-S (%)
1	Adenocarcinoma	Synergistic	2.40	Multiploid	NA ^a
2	Adenocarcinoma	Synergistic	1.55	Near-diploid	9.7
3	Adenocarcinoma	Synergistic	1.46	Hyperdiploid	16.8
4	Adenocarcinoma	Synergistic	1.37	Multiploid	3.8
5	Adenocarcinoma	Synergistic	1.31	Multiploid	NA
6	Adenocarcinoma	Antagonistic	1.29	Multiploid	NA
7	Squamous cell ca.	Synergistic	4.03	Near-diploid	11.5
8	Squamous cell ca.	Synergistic	3.53	Multiploid	NA
9	Squamous cell ca.	Synergistic	1.41	Near-diploid	11.1
10	Squamous cell ca.	Additive	–	Near-diploid	11.4
11	Atypical carcinoid	Synergistic	4.10	Multiploid	12.5
12	Atypical carcinoid	Synergistic	1.20	Hypodiploid	4.4
13	Typical carcinoid	Additive	–	Hypodiploid	1.0
14	Neuroendocrine with atypical carcinoid	Synergistic	1.56	Hyperdiploid	14.8

^aNot assessable for the partial overlapping of DNA histograms belonging to the different clones.

the other taxane, docetaxel, and following the sequential treatment docetaxel–gemcitabine, we observed a synergistic cytotoxic effect that increased significantly when the second drug was given after a 48-h washout. This finding was confirmed in most of the primary cultures from clinical human lung cancer tumour we used, which represents an important step in validating preclinical results before translating them into clinical practice (Villa et al, 1992).

The results from the present study reinforce the importance, previously evidenced for other drugs and tumour types (Citro et al, 1991; Savini et al, 1992; Dogliotti et al, 1996; De Lena et al, 1997; Silvestrini et al, 1997; Amadori et al, 1998; Fischel et al 1998; Van Moorsel et al 1998, 1999; Zoli et al, 1999) of preclinical research to define the best treatment scheduling. Considering the recently published results (Spiridonidis et al, 1998; Georgoulis et al, 1999), which show that gemcitabine and docetaxel can be safely used in combination, a protocol on advanced NSCLC will shortly be activated in Italy based on the findings from the present study.

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