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Pro-Inflammatory Functions of Astrocytes Correlate with Viral Clearance and Strain-Dependent Protection from TMEV-Induced Demyelinating Disease

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Abstract

Intracerebral infection of susceptible strains of mice, *e.g.* SJL/J, with Theiler's murine encephalomyelitis virus (TMEV) leads to a persistent CNS infection accompanied by development of a chronic-progressive inflammatory CNS autoimmune demyelinating disease which is clinically and pathologically similar to human multiple sclerosis. In contrast, resistant strains of mice, *e.g.* C57BL/6 (B6), effectively clear TMEV from the CNS and do not develop demyelinating disease. Although CD8⁺ T cells are crucial for viral clearance in B6 mice, SJL mice also mount potent CD8⁺ T cell responses against virus, thus the reason for the viral persistence in the CNS in these mice is unclear. Here, we examined innate anti-viral responses of CNS-resident astrocytes as a potential determinant of viral persistence and disease susceptibility. We demonstrate that B6 astrocytes produce significantly higher levels of cytokines, chemokines and adhesion molecules in response to TMEV infection, or stimulation with IFN- γ and TNF- α or poly I:C than SJL mice. In addition, TMEV more effectively induces MHC I molecules on B6 astrocytes than SJL, corresponding with an increased ability to activate TMEV-specific CD8⁺ T cells directly *ex vivo*. These results suggest that enhanced anti-viral responses of B6 astrocytes contribute to the ability of these mice to clear TMEV from the CNS and therefore to their resistance to the development of autoimmune demyelinating disease.

Keywords

multiple sclerosis; Theiler's murine encephalomyelitis virus; astrocytes; cytotoxic T lymphocytes; disease susceptibility

Introduction

Intracerebral infection of susceptible strains of mice with the BeAn strain of Theiler's murine encephalomyelitis virus (TMEV) leads to a persistent central nervous system (CNS) infection accompanied by development of a chronic-progressive inflammatory demyelinating disease

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beginning approximately 30 days post-infection (dpi) (Dal Canto and Lipton, 1975; Lipton, 1975). TMEV-induced demyelinating disease (TMEV-IDD) is a relevant model for human multiple sclerosis (MS) because many of the clinical symptoms and pathologic correlates are similar, including CD4⁺ T cell and mononuclear phagocyte infiltration into the CNS, primary demyelination and spastic hindlimb paralysis (Dal Canto and Lipton, 1975; Hafler, 2004; Lipton, 1975). In contrast, resistant mice are able to efficiently clear TMEV from the CNS and do not progress to demyelinating disease (Chamorro et al., 1986).

Resistance to persistent CNS infection by TMEV is controlled by multiple genetic loci, with the strongest linkage to the class I H-2D MHC region (Brahic et al., 2005; Lipton and Melvold, 1984). Mice which carry an H-2^{s, f, p, r, v} or ^q haplotype are susceptible to TMEV-IDD, while those bearing H-2^{b, k, or d} haplotypes are resistant (Rodriguez and David, 1985). The particular importance of the H-2D region was demonstrated using hybrid strains which express H-2K^s and H-2D^b, which are resistant to TMEV-IDD, and those expressing H-2K^b and H-2D^s, which are susceptible (Clatch et al., 1985). Additionally, H-2^b mice which are genetically deficient in H-2D are susceptible to TMEV-IDD and the insertion of an H-2D^b gene into a normally susceptible H-2^q strain renders them resistant, further supporting a critical role of H-2D in determining genetic susceptibility to TMEV-IDD (Azoulay et al., 1994; Azoulay-Cayla et al., 2000; Lipton et al., 1995a).

The strong association of H-2D haplotype with development of demyelinating disease indicates a role of CD8⁺ T cells in disease which could be pathologic or protective. Most evidence points towards a protective role of CD8⁺ T cells via the clearance of TMEV from the CNS, because mice on a resistant background which are CD8-depleted via antibody or genetically deficient in CD8⁺ T cells fail to clear the virus and develop CNS demyelination and inflammation, although they develop only mild clinical symptoms (Borrow et al., 1992; Fiette et al., 1993; Ure and Rodriguez, 2002). Additionally, TMEV-infected CD8-deficient mice on a susceptible background exhibit decreased viral clearance, increased CNS inflammation and a more rapid and severe clinical disease course (Begolka et al., 2001).

Despite evidence demonstrating the importance of the CD8⁺ T cell response in protection from TMEV-IDD, susceptible strains of mice mount robust CD8⁺ T cell responses against TMEV, similar in frequency to those in resistant strains (Kang et al., 2002; Lindsley et al., 1991; Lyman et al., 2004), which raises the question of why susceptible strains of mice are unable to completely clear the virus from the CNS. It has recently been demonstrated using bone marrow chimeras that some genetic control of susceptibility depends on cells which are not of hematopoietic origin, suggesting these genes might control responses in the target organ (Aubagnac et al., 2002). Astrocytes are neuroectodermally derived cells which are critical for normal functions of the CNS including synaptic transmission, maintenance of the blood-brain barrier, metabolic support of axons, and ion and neurotransmitter buffering. We and other groups have demonstrated that activated astrocytes can also contribute to immune responses in the CNS through the production of cytokines, chemokines, anti-microbial molecules and the ability to activate CD4⁺ T cells in the presence of IFN- γ (Bailey et al., 2006; Carpentier et al., 2005; Carpentier et al., 2007). Although astrocytes generally express low to undetectable levels of MHC I molecules under resting conditions (Schnitzer and Schachner, 1981; Traugott, 1987), treatment with cytokines or infection with virus can upregulate MHC I and increase the ability of astrocytes to activate or be lysed by CD8⁺ T cells (Lavi et al., 1988; Liu et al., 1989; Skias et al., 1987; Suzumura et al., 1986; Wong et al., 1984). The upregulation of MHC I is highly variable between strains of mice and is controlled by genes within the MHC locus (Massa et al., 1989), suggesting that the expression of MHC I on astrocytes could account for some of the MHC-dependent genetic control of TMEV-IDD susceptibility. Astrocytes in susceptible mice are activated in response to TMEV *in vivo* and harbor viral antigen (Zheng et al., 2001). It has been suggested that astrocytes are the major source of replicating virus *in*

vivo, although other groups contend that microglia/macrophages contribute more to viral load (Lipton et al., 1995b; Zheng et al., 2001).

Here, we demonstrate that astrocytes derived from TMEV-IDD resistant B6 mice are less efficiently infected by TMEV *in vitro* than those from susceptible SJL mice. Additionally, B6 astrocytes express increased levels of cytokines, chemokines and adhesion molecules in response to various other pro-inflammatory stimuli compared to SJL astrocytes. Finally, we demonstrate that B6 astrocytes infected with TMEV more readily upregulate MHC I and more efficiently activate CD8⁺ T cells than SJL astrocytes. We hypothesize that the augmented antiviral responses of B6 astrocytes contribute to the enhanced ability of these mice to clear TMEV from the CNS and therefore contribute to their resistance to the development of chronic demyelinating disease.

Results

TMEV infection of SJL and B6 Astrocytes

We have previously observed that astrocytes from TMEV-IDD susceptible SJL mice can be directly infected with TMEV, expressing both viral RNA and protein antigens (Carpentier et al., 2007). To ascertain if there are strain differences in the astrocytic response to TMEV infection between resistant B6 and susceptible SJL mice, we first determined if TMEV infection of B6 astrocytes differs from SJL astrocytes. Fewer B6 astrocytes express TMEV antigens at 48 h post infection, indicating that they are less efficiently infected than SJL astrocytes (Figure 1A,B; $p=0.05$). Consistent with our previous observations, there was a small but statistically significant increase in cell death after 72 h of infection with TMEV in both SJL and B6 cultures. The amount of cytotoxicity in TMEV-infected cultures is quite small in comparison to a freeze-thaw positive control. There was no difference in the amount of cytotoxicity observed between the mouse strains, indicating that TMEV does not produce a strong lytic infection in astrocytes from either strain (Figure 1C).

Innate immune functions of SJL and B6 Astrocytes

We next examined the innate immune functions of astrocytes induced by TMEV infection. We have previously completed time course experiments with astrocytes infected with TMEV and observed that production of type I IFNs, cytokines and chemokines increases through 72 h, and we have therefore chosen this time to examine these innate immune functions (Carpentier et al., 2007). There is a trend towards increased production of type I IFNs in B6 astrocytes after TMEV infection (Figure 2), but it did not reach statistical significance ($p=0.19$ for IFN- α and $p=0.09$ for IFN- β). It therefore appears that type I IFN production is likely not the mechanism by which B6 astrocytes better control TMEV infection. B6 astrocytes, however, induce higher levels of a variety of cytokines and chemokines after TMEV infection and poly I:C treatment (Figure 3). In response to treatment with poly I:C, both SJL and B6 astrocytes express similar levels of IL-6, CCL2, CCL5, CXCL1 and CXCL10 (Figure 3A, C, F-H). However, B6 astrocytes treated with poly I:C express significantly higher levels of TNF- α , CCL3, and CCL4 than SJL astrocytes (Figure 3B, D, E; $p=0.012$ CCL3; $p=0.038$ CCL4; $p=0.032$ TNF- α). In response to TMEV infection, as we have previously observed (11), SJL astrocytes express IL-6, CCL2, CCL5 and CXCL10 (Figure 3A, C, F, H). B6 astrocytes infected with TMEV produce significantly increased levels IL-6 and CXCL10 (Figure 3A, H; $p=0.026$ IL-6; $p=0.013$ CXCL10) and also show a trend towards increased production of TNF- α , CCL2 and CCL5 (Figure 3B, C, F). B6 astrocytes infected with TMEV also induce CCL3 and CCL4, which is significantly increased compared to SJL (Figure 3D, E; $p=0.043$ CCL3; $p=0.038$ CCL4). Combined exposure to IFN- γ and TNF- α induced significant expression of CCL2, CCL5 and CXCL10 in both strains, and CXCL10 production under this condition was slightly enhanced in B6 astrocytes (Figure 3C, F, H; $p=0.046$). CCL3 and CCL4 were also slightly enhanced in

B6 IFN- γ and TNF- α treated astrocytes, but the levels remained quite low compared to the other treatments (Figure 3D, E; $p=0.0058$ CCL3; $p=0.028$ CCL4). Importantly, many of these chemokines produced by astrocytes have been demonstrated to be critical for recruitment CD8⁺ T cells to sites of viral infection (Thomsen et al., 2003).

We have also previously demonstrated that stimulation of astrocytes with TLR ligands or cytokines upregulated their expression of the adhesion molecules ICAM-1 and VCAM-1 (Carpentier et al., 2005). These adhesion molecules are crucial for the infiltration of leukocytes into the CNS (Ransohoff et al., 2003), and it has been proposed that their expression on astrocytes is particularly crucial for the penetration of leukocytes deep into the CNS parenchyma (Gimenez et al., 2004). There is no significant difference in the levels of adhesion molecules expressed in mock infected SJL or B6 astrocytes (Figure 4A,F and data not shown). IFN- γ and TNF- α treatment upregulate ICAM-1 and VCAM-1 similarly in SJL and B6 mice (Figure 4 B,G,E,J). Poly I:C more strongly upregulates ICAM-1 in B6 than SJL (Figure 4 C,E; $p=0.04$), and there is a trend towards stronger upregulation of VCAM-1 although it does not reach significance (Figure 4 H,J; $p=0.07$). The increased upregulation of adhesion molecules on B6 astrocytes is most marked and consistent with TMEV infection (Figure 4 D,E,I,J; $p=0.03$ ICAM-1, $p=0.04$ VCAM-1), which may contribute towards the increased early infiltration of CD8⁺ T cells into the CNS of B6 mice. This increased expression of adhesion molecules is not restricted to cells which are actually infected with TMEV, which is a minority of cells in B6 cultures, because there is a whole population shift in ICAM-1 and VCAM-1 expression.

CD8⁺ T cell activation by astrocytes

We next examined the ability of astrocytes to directly activate CD8⁺ T cells, which are crucial to clearance of TMEV from the CNS (Begolka et al., 2001; Borrow et al., 1992; Fiette et al., 1993). In order to activate CD8⁺ T cells, a cell must express the appropriate peptide in the context of MHC class I, which astrocytes can express in response to virus infection or cytokine treatment (Fontana et al., 1986; Lavi et al., 1988; Liu et al., 1989; Suzumura et al., 1986; Wong et al., 1984). We thus compared the constitutive expression and regulation of MHC I molecules H-2K and H-2D in SJL and B6 astrocytes, respectively. We chose these molecules because all known TMEV epitopes are restricted to H-2K in SJL and H-2D in B6 mice (Kang et al., 2002; Lyman et al., 2002). SJL astrocytes express little or no H-2K^s in the unstimulated state (Figure 5A), but it is upregulated by treatment with IFN- γ and TNF- α or poly I:C (Figure 5B, C). In contrast, infection with TMEV is a relatively poor inducer of H-2K^s expression by SJL astrocytes (Figure 5D). B6 astrocytes also express low constitutive levels of H-2D^b (Figure 5E), but upregulate it in response to IFN- γ and TNF- α , poly I:C, or TMEV infection (Figure 5F-H). Compared to SJL, B6 astrocytes upregulate H-2 expression significantly more in response to IFN- γ and TNF- α ($p=0.029$) and TMEV ($p=0.049$), and there is a trend towards higher upregulation in response to poly I:C as well ($p=0.09$).

These differences suggested that SJL astrocytes may be less able to activate the antiviral functions of CD8⁺ T cells due to their relative lack of MHC I expression. However, it is difficult to directly compare the levels of MHC I on B6 and SJL astrocytes because they are distinct molecules and analyzed with different antibodies. Therefore, to further test this hypothesis, we examined the ability of astrocytes to induce IFN- γ production by TMEV-specific CD8⁺ T cells. CD8⁺ T cells were isolated from the spleens of naïve or TMEV-infected mice at 8 dpi using magnetic cell sorting with anti-CD8 α microbeads. Purified samples averaged approximately 90% CD8 α ⁺ cells (data not shown). The majority of CD8 α ⁺ cells are also positive for CD90, indicating that they are T cells, although a small subset of CD8 α ⁺ cells co-express CD11c (data not shown). This is potentially a concern, because CD8 α ⁺ dendritic cells are known for their potent ability to activate CD8⁺ T cells (Ardavin, 2003). However, we did not observe any IFN- γ production above background in CD8⁺ T cell cultures from TMEV-infected mice which did

not contain astrocytes, even with specific peptide added (Figure 6B). Importantly, virtually none of the sorted cells were CD90⁺CD8 α ⁻, which represents CD4⁺ T cells and another potential contaminating source of IFN- γ (data not shown). We co-cultured CD8 α ⁺ T cells from TMEV-infected SJL and B6 mice with astrocytes that were mock infected, treated with IFN- γ and TNF- α , or infected with TMEV 48 h prior with or without the immunodominant CD8-restricted viral peptides, VP3₁₅₉₋₁₆₆ for SJL and VP2₁₂₁₋₁₃₀ for B6 (Kang et al., 2002; Lyman et al., 2002). Culture supernatants were harvested after 96 h and analyzed for IFN- γ levels. Neither SJL nor B6 astrocytes in the absence of CD8⁺ T cells produce significant levels of IFN- γ under any condition (not shown). CD8⁺ T cells isolated from naïve mice also failed to produce significant amounts of IFN- γ under any conditions (Figure 6A). SJL and B6 astrocytes were able to induce of IFN- γ from TMEV-specific CD8⁺ T cells upon the addition of specific peptide (Figure 6B). B6 astrocytes were more efficient at inducing peptide-specific CD8⁺ T cell activation when TMEV-infected cells were employed ($p=0.011$ effect of strain by two-way ANOVA; $p<0.05$, Bonferonni post-test SJL TMEV to B6 TMEV). Mock infected and especially IFN- γ and TNF- α treated astrocytes showed a similar ability to activate CD8⁺ T cells from TMEV-infected mice, indicating that the difference between these strains is limited to the astrocytic response to TMEV infection. This observation which correlates well with the increased levels of MHC I expression on TMEV-infected astrocytes we have previously noted (Figure 5). Importantly, B6 astrocytes infected with TMEV were also able to induce IFN- γ production from TMEV-specific CD8⁺ T cells without the addition of exogenous peptide, indicating that they are able to efficiently process and present endogenous viral antigen, whereas SJL astrocytes were not (Figure 6C; $p=0.037$ effect of strain by two-way ANOVA, $p<0.01$ by Bonferonni post-test, B6 TMEV to SJL TMEV). This difference is most likely the result of increased MHC I expression in TMEV-infected B6 astrocytes (Figure 5D,H) rather than a difference in viral antigen levels, since we observe somewhat decreased levels of viral antigen in TMEV-infected B6 cultures after 48h (Figure 1A,B). In order to confirm that there are no differences between the frequency or IFN- γ producing ability of SJL and B6 TMEV-specific CD8⁺ T cells which could potentially account for these results, we performed ELISPOTs from both strains of mice with the immunodominant and subdominant TMEV peptides at 7dpi (Figure 6D,E). Our results demonstrate that similar numbers of CD8⁺ T cells capable of producing IFN- γ in response to TMEV-specific peptides exist in the spleens of B6 and SJL mice acutely after infection, and further indicate that only the antigen processing and/or presentation ability of astrocytes differ between these two strains. Since astrocytes are infected *in vivo* with TMEV (Zheng et al., 2001), their ability to process and present viral antigen to CD8⁺ T cells could be crucial for CNS virus clearance.

Discussion

These findings demonstrate key functional differences between astrocytes from TMEV-IDD resistant B6 and susceptible SJL mice in their ability to mount efficient anti-viral responses. We first demonstrate that B6 astrocytes are more resistant to TMEV infection than SJL astrocytes. Astrocytes are infected in SJL mice *in vivo*, although whether astrocytes or microglia/macrophages are the major source of replicating virus has been controversial (Lipton et al., 1995b; Zheng et al., 2001). Our data indicates that astrocytes are less efficiently infected in resistant B6 mice, which could account for the somewhat decreased viral loads in B6 compared to SJL brains even early after infection (Lyman et al., 2004), and could significantly contribute to the inability to establish a persistent viral infection and result in demyelinating disease. Our data also demonstrates decreased innate immune functions and decreased ability to process and present antigen to CD8⁺ T cells in TMEV-infected SJL astrocytes compared to B6. Together, these data suggest the possibility that astrocytes, non-hematopoietic cells resident in the CNS, may significantly contribute to the ability of the immune system to efficiently clear virus in this target organ. We are working to confirm these results in an *in vivo* system using GFAP-GFP transgenic mice to allow astrocytes to be sorted from TMEV-infected brains and

compared for expression of pro-inflammatory molecules and ability to present antigen to CD8⁺ and CD4⁺ T cells. We are also pursuing studies in mice with genetic defects in various astrocyte-related inflammatory functions to determine which functions are important for susceptibility to TMEV-IDD *in vivo*.

The ability to clear TMEV from the CNS is critically dependent on CD8⁺ T cells (Borrow et al., 1992; Fiette et al., 1993; Ure and Rodriguez, 2002). Additionally, the strongest genetic linkage to TMEV susceptibility lies within the H-2D locus in the MHC region of the genome, further suggesting the critical importance of CD8⁺ T cells in viral clearance (Brahic et al., 2005). For this reason, most studies to date examining strain-dependent susceptibility to TMEV-IDD have focused on CD8⁺ T cell functions. Surprisingly, however, susceptible mice mount very strong CD8⁺ T cell responses to TMEV infection, and the activation, cytokine production and cytolytic functions of SJL CD8⁺ T cells are similar to those seen in resistant B6 mice (Kang et al., 2002; Lindsley et al., 1991; Lyman et al., 2004).

There is, however, a reported delay in the infiltration of CD8⁺ T cells into the CNS of TMEV-infected SJL mice compared to B6 (Lyman et al., 2004). Our results correlate well with this study, since we observe here that SJL astrocytes express lower levels of cytokines, chemokines and adhesion molecules, particularly in response to TMEV infection, which may cause less efficient recruitment and infiltration of anti-viral CD8⁺ T cells into the CNS. B6 astrocytes express higher levels of the cytokines IL-6 and TNF- α after TMEV infection or poly I:C treatment, which may damage the blood-brain barrier (BBB) and contribute to the ability of leukocytes to infiltrate the CNS (de Vries et al., 1996). B6 astrocytes also express significantly higher levels of CXCL10, CCL3 and CCL4 than SJL astrocytes and trend towards higher production of CCL2 and CCL5. These chemokines are of particular interest because they are critical for the recruitment of virus-specific CD8⁺ T cells to sites of infection throughout the body (Thomsen et al., 2003) and their induction has been described in the CNS in response to many viral infections, including TMEV (Glass et al., 2005; Hoffman et al., 1999; Lane et al., 2000; Nansen et al., 2000; Nuovo and Alfieri, 1996). Genetic deficiency or blockade of CCL3, CCL5, CXCL10 or CCR2 (the obligate receptor for CCL2) during acute CNS infection generally inhibits the infiltration of T cells and monocytes/macrophages into the CNS and is associated with increased viral loads after infection with MHV or TMEV (Chen et al., 2001; Lane et al., 2000; Liu et al., 2000; Trifilo et al., 2003; Ure et al., 2005). CXCR3, the receptor for CXCL9-11, and CCR5, the receptor for CCL3, CCL4, and CCL5, are also expressed on many CNS-infiltrating CD4⁺ and CD8⁺ T cells and likely important for the ability to gain access to the CNS (Giunti et al., 2003; Kivisakk et al., 2002; Nansen et al., 2000; Shacklett et al., 2004; Sorensen et al., 1999). Adhesion molecules expressed at higher levels by B6 astrocytes may also promote more efficient leukocyte entry, and particularly promote their ability to invade deep into the CNS parenchyma (Gimenez et al., 2004).

In addition to being recruited to the CNS, CD8⁺ T cell cytotoxic functions must be activated in order to clear TMEV from the CNS, which requires the recognition of virus peptides in the context of MHC I. Astrocytes are able to express MHC I in response to cytokines and virus (Fontana et al., 1986; Lavi et al., 1988; Liu et al., 1989; Suzumura et al., 1986; Wong et al., 1984), but a critical role for astrocytes during CD8⁺ T cell activation *in vivo* during CNS infection has yet to be demonstrated. We show here that astrocytes from resistant B6 mice are able to upregulate the MHC I molecule H-2D^b in response to TMEV infection and to induce the activation of TMEV-specific CD8⁺ T cells as indicated by the production of IFN- γ . In contrast, upregulation of H-2K^s in TMEV-infected SJL astrocytes is minimal, despite their high level of infection, and SJL astrocytes are inefficient at processing and presenting viral antigen to CD8⁺ T cells. Importantly, the frequency of splenic TMEV-specific CD8⁺ T cells able to produce IFN- γ in response to the respective immunodominant TMEV CD8 epitope in SJL and B6 mice is similar. Thus, the decrease in the ability of SJL CNS-resident cells to

efficiently activate CD8⁺ T cells may significantly contribute to the inability of SJL CD8⁺ T cells to completely clear the virus from the CNS, despite their ability to efficiently carry out anti-viral responses *in vitro*.

CD8⁺ T cells are largely protective in TMEV-IDD because of their importance in viral clearance, but there are scenarios in which CD8⁺ T cells can be pathologic as well. Resistant mice deficient in CD8⁺ T cells or the CD8⁺ T cell cytotoxic mediator perforin infected with the DA strain of TMEV develop demyelination, inflammation and persistent virus in the CNS, but display minimal clinical disease (Murray et al., 1998; Ure and Rodriguez, 2002). This has led to the hypothesis that CD8⁺ T cells may mediate axonal damage in TMEV-IDD and in MS, which is a major pathologic correlate of permanent neurologic disability (McDole et al., 2006; Trapp et al., 1998). Astrocytes reportedly express MHC I in MS lesions, and therefore could contribute to CD8⁺ T cell-mediated axonal damage by either direct or bystander mechanisms (Hoftberger et al., 2004; Traugott, 1987). Interestingly, SJL astrocytes exposed to the pro-inflammatory cytokines IFN- γ and TNF- α , as would be the case in T cell and macrophage rich MS lesions, are able to upregulate H-2K^S, and in this context MHC I expression by astrocytes may be pathologic. In conclusion, the contribution of the innate and adaptive immune functions of astrocytes therefore appears to be crucial to the balance of protective immunity vs. the development of autoimmune disease after CNS infection.

Materials and methods

Mice

Pregnant (15–17 days) SJL/J (SJL) mice and 5-to 7-week-old SJL mice were purchased from Harlan Labs (Bethesda, MD). Pregnant (11–15 days) and 5-to 7-week old C57BL/6 (B6) mice were purchased from Jackson Laboratories (Bar Harbor, ME). Mice were housed in the Northwestern University animal facility in accordance with university and National Institutes of Health animal care guidelines and afforded access to food and water *ad libitum*. Neonatal mice 1 to 3 days old were used for the isolation of astrocytes.

Reagents

Poly I:C was purchased from Sigma (St. Louis, MO) and used at 100 μ g/ml. Recombinant murine IFN- γ was purchased from Pharmingen (San Diego, CA) and used at 100 U/ml. Recombinant murine TNF- α was purchased from R&D Systems (Minneapolis, MN) and used at 500 U/ml. These doses selected were experimentally determined to elicit optimal responses (Carpentier et al., 2005). VP_{3159–166} (FNFTAPFI) and VP_{2121–130} (FHAGSLLVGM) were purchased from Genemed Synthesis Inc (South San Francisco, CA). The amino acid composition was verified by mass spectrometry and purity (>95%) was assessed by high-performance liquid chromatography.

Media

Mixed glial and astrocyte cultures were maintained in Dulbecco's modified Eagle medium (DMEM)-F12 (Sigma) supplemented with 10% fetal calf serum (FCS; Sigma), 6 g/l glucose (Sigma), 2.4 g/l sodium bicarbonate (Sigma), 0.37 g/l L-glutamine (Life Technologies, Gaithersburg, MD), 100 U/ml penicillin (Life Technologies) and 100 μ g/ml of streptomycin (Life Technologies; complete DMEM-F12 media). T cell assays were performed in DMEM with 5% FCS, 0.37 g/l L-glutamine, 100 U/ml penicillin, 100 μ g/ml streptomycin, 55 μ M 2-Mercaptoethanol (Gibco, Grand Island, NY), and 0.1 mM nonessential amino acids (Sigma; complete D5 media).

Astrocyte Isolation

T75 tissue culture flasks were coated from 3 h to overnight with 10 µg/ml poly-D-lysine (PDL; Sigma). Intact brains were removed from 1-to 3-day-old neonatal mice, hindbrains dissected away, and meninges removed. The cerebral hemispheres were transferred to a nylon mesh bag and gently dissociated. Cells in suspension were passed through #60 and #100 stainless steel screens (Sigma) to remove large pieces of debris and tissue. Cells were seeded at 2–3 brains per flask, and maintained in complete DMEM-F12 media at 37°C and 7.5% CO₂. At confluency (generally 14–21 days), microglia and oligodendrocytes were removed from the astroglial bed layer by orbital shaking for 24 h at 300 rpm. Astrocytes that adhered to the flask were removed with 1x trypsin-ethylenediamine-tetracetic acid (EDTA; 0.05 g/l trypsin, 0.02 g/l EDTA; Sigma) and re-plated until experimental use. Cultures prepared in this manner are ≥ 98% pure by staining for glial fibrillary acidic protein (GFAP), an astrocyte specific intermediate filament (Carpentier et al., 2005). We also routinely screen cultures for microglial contamination by flow cytometry for CD45, a marker of immune lineage cells, and typically find ≤ 1% of cells are CD45⁺ (data not shown).

Treatment/Infection of Primary Astrocytes

At time of infection, all media was removed from astrocyte cultures and replaced with a minimal volume of DMEM. Astrocytes were treated with media alone or stimulated with IFN-γ and TNF-α or poly I:C. TMEV was added at a multiplicity of infection of 10 (MOI; 10 plaque forming units/cell). All cultures were left at room temperature for 1 h to allow the virus to infect. After 1 h, complete F12 media was added to each culture and poly I:C or cytokine was accordingly added such that the total concentration remained constant.

Flow Cytometry

Cells were incubated with Accutase (BD Biosciences, San Jose, CA), removed from flasks, and washed in phosphate buffered saline (PBS) containing 0.5% bovine serum albumin (BSA; Sigma) and 2mM EDTA (Sigma). Intracellular staining with anti-TMEV was performed on cells fixed with Cytofix/Cytoperm solution containing formaldehyde and saponin for 20 minutes at 4°C (Pharmlingen). Cells were washed twice in Perm/Wash Buffer containing saponin (Pharmlingen) and nonspecific staining was blocked with anti-CD16/32 (eBioscience, San Diego, CA) and 10% normal goat serum (Sigma) for at least 20 minutes at 4°C. Cells were washed once and incubated with anti-TMEV serum generously donated by Dr. Howard Lipton (Northwestern University, Evanston, IL) for 30 minutes at 4°C. Cells were washed twice more and incubated with goat anti-rabbit IgG-FITC (Pharmlingen) for 20 minutes at 4°C. Cells were washed at least twice with Perm/Wash buffer and twice with PBS/BSA/EDTA before flow cytometric analysis. For staining of cell surface antigens, cells were blocked with 10% mouse and 10% rat serum (Sigma) and 1:100 purified anti-mouse CD16/32 (eBioscience) in PBS/BSA/EDTA for at least 20 minutes at 4°C. Cells were then incubated with primary antibodies or appropriate isotype controls (eBioscience or Pharmlingen) for at least 30 minutes at 4°C. After primary antibody incubation, astrocytes were washed three times with PBS/BSA/EDTA and incubated with secondary antibodies or streptavidin-APC (Pharmlingen) as necessary for 20 minutes at 4°C. Cells were washed three more times with PBS/BSA/EDTA prior to analysis. Flow cytometry was performed on a BD Biosciences LSR flow cytometer using FACs Diva acquisition software. Data analysis was performed using Tree Star FlowJo software. Plots shown are gated on forward/side scatter and absence of CD45 staining, to avoid analyzing possible contaminating microglia.

Analysis of Cytotoxicity

Astrocytes were seeded at 7.5×10^4 cells/ml in PDL coated 48-well plates in complete DMEM-F12 media and allowed to adhere overnight. Cultures were mock infected or infected with

TMEV (MOI=10). Supernatants were withdrawn at 24, 48 and 72 h and analyzed for lactate dehydrogenase (LDH) release as a measure of cell death using the Nonradioactive Cytotoxicity Assay kit (Promega) as per manufacturer's instructions.

Enzyme-Linked Immunoassays (ELISAs)

Astrocytes were seeded at 7.5×10^4 cells/ml in PDL coated 48-well plates in complete DMEM-F12 media and allowed to adhere overnight. Cultures were mock infected or infected with TMEV (MOI=10) and supernatants were withdrawn at 72 h. IFN- α and IFN- γ ELISAs were performed as per kit instructions (R&D Systems, Minneapolis, MN).

Analysis of Astrocyte Cytokine and Chemokine Protein Expression

Astrocytes were seeded at 7.5×10^4 cells/ml in PDL coated 48-well plates in complete DMEM-F12 media and allowed to adhere overnight. Cultures were mock infected, treated with IFN- γ (100 U/ml) and TNF- α (500 U/ml), treated with poly I:C (100 μ g/ml) or infected with TMEV (MOI=10) and supernatants were withdrawn at 72 h. Supernatants were analyzed for cytokine and chemokine expression by Invitrogen Biosource (Carlsbad, CA) Mouse Individual Flex Kit for IL-6, TNF- α , CCL2, CCL3, CCL4, CCL5, CXCL1, and CXCL10. Analysis was performed on a Luminex IS.100 system (Qiagen, Valencia, CA).

Infection of mice with TMEV

Mice were anesthetized with an inhaled mixture of isoflurane and oxygen and infected intracerebrally with 3×10^6 plaque forming units (PFU) of the BeAn strain of TMEV. Mice infected for 8 days were used for CD8⁺ T cell isolation.

CD8⁺ T Cell Activation Assay

SJL and B6 mice were infected with TMEV at day 0. At day 5, SJL or B6 astrocytes were plated at 1×10^5 /well in a PDL coated 96 well plate. At day 6, astrocyte cultures were mock infected, treated with IFN- γ and TNF- α (100 and 500 U/ml) or infected with TMEV (MOI=10). At day 8, spleens were collected from naïve or TMEV-infected mice and homogenized over a #100 wire mesh screen. Red blood cells were lysed with Tris-ammonium chloride for 5 minutes at room temperature and washed in serum-containing media. Cells were blocked with 10% mouse serum and 10% rat serum in PBS/BSA/EDTA for at least 30 minutes at 4°C and then incubated with anti-CD8 α microbeads (Miltenyi Biotec, Auburn, CA) for 15 minutes at 4°C. Positive cells were collected using the positive select double sort (posseld) program on an Automacs system (Miltenyi) and were approximately 90% pure on average as determined by flow cytometry (data not shown). Astrocyte cultures were washed twice with complete D5 media and incubated with 3×10^5 CD8⁺ T cells plus 0, 10 or 100 μ M of VP3₁₅₉₋₁₆₆ (SJL) or VP2₁₂₁₋₁₃₀ (B6) in complete D5 media. At day 12, culture supernatants were harvested and analyzed for cytokine expression using the Upstate Mouse Multi-Cytokine 5plex Kit containing beads specific for IFN- γ , TNF- α , IL-2, IL-4 and IL-10 as per manufacturer's instructions.

Enzyme-linked Immuno-SPOT (ELISPOT) assays

ELISPOT assays were carried out as previously described (Getts et al., 2007). Briefly, 5×10^5 bulk splenocytes were isolated at 7dpi and plated at 5×10^5 cells per well with 0 or 50 μ g of the indicated peptide. The number of spots, representing IFN- γ producing cells, was quantified for each strain and peptide. All ELISPOT data are presented as mean number of spots per 5×10^5 splenocytes \pm SEM.

Statistics

A two-way ANOVA with repeated measures and Bonferroni post tests was used to compare cytotoxicity over a 72 h time course and CD8⁺ T cell activation. Freeze-thaw positive controls were compared to uninfected cultures at 72 h using a one-tailed student's T test. A paired one-tailed Student's t test was used to compare means of percentage of infected cells, cytokine and chemokine production and adhesion molecule expression between SJL and B6 astrocytes. In all cases, $p < 0.05$ is considered significant.

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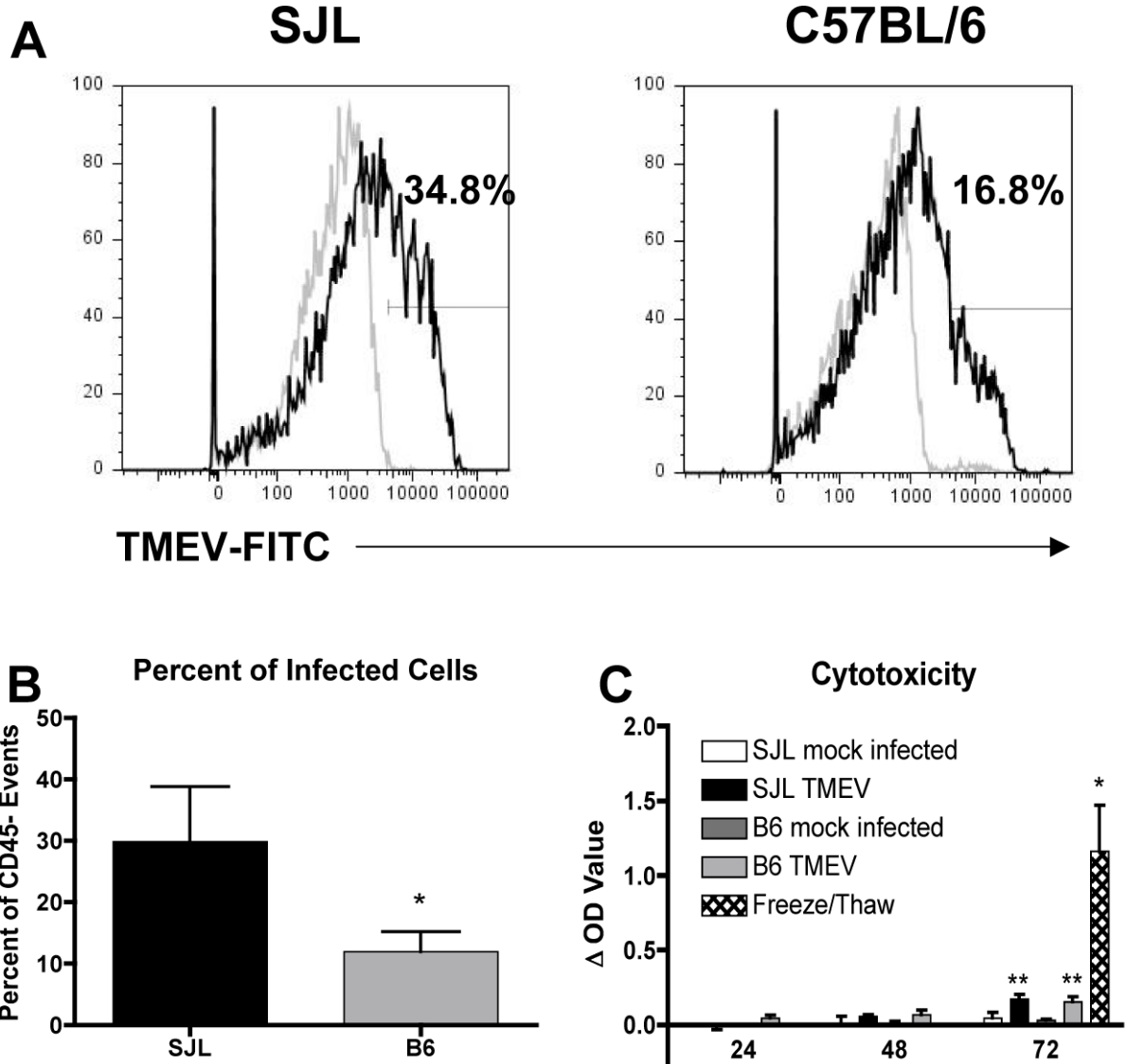


Figure 1. SJL astrocytes are infected with TMEV more efficiently than B6 astrocytes
 SJL or B6 astrocytes were mock infected or infected with TMEV (MOI=10). **(A)** Mock infected (grey line) and TMEV infected (black line) cultures were harvested at 48 h and stained with anti-TMEV antibody. The percentage of cells falling within the drawn gate in the TMEV-infected cultures is shown above the gate. Data are representative of 3 separate experiments. **(B)** The percent of TMEV-infected cells in SJL and B6 cultures was quantified. Data shown are the mean ± SEM of 3 separate experiments. *p=0.05, one-tailed Student's t test **(C)** Supernatants were collected at 24, 48 and 72 h post infection and analyzed for lactate dehydrogenase release as a measure of cell death. Data are expressed as the difference between the optical density (OD) of the mock infected samples at 24 h and the experimental samples and are the average ± SEM of 3 separate experiments. Freeze/thawed samples were used as a positive control. A two way ANOVA with repeated measures found significant effects of time (p=0.012) and treatment group (p=0.0014). **OD value significantly increased compared to control, p<0.01 by Bonferroni post test. *OD value significantly increased compared to mock infected cells at 72 h by one-tailed Student's t test, p<0.05

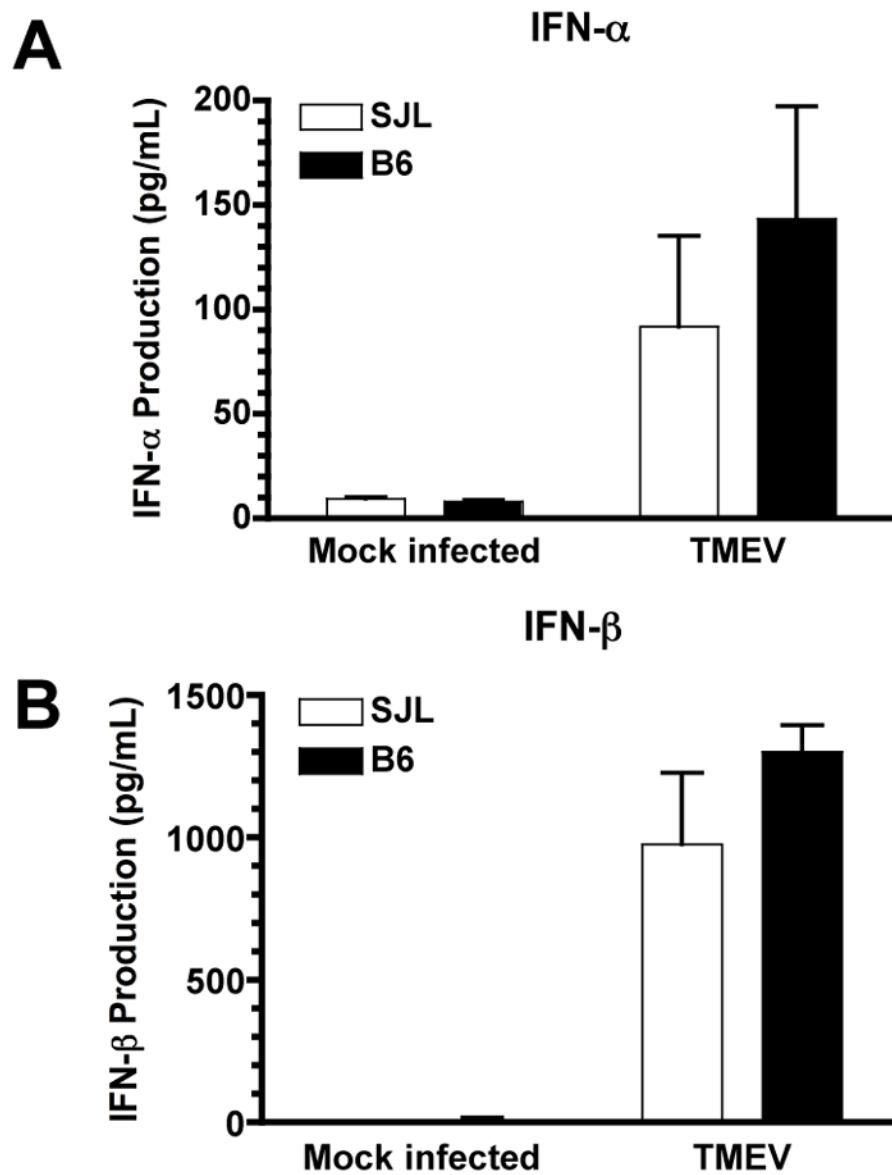


Figure 2. TMEV infection induces similar levels of type I IFNs in SJL and B6 astrocytes
SJL or B6 astrocytes were mock infected or infected with TMEV (MOI=10). Supernatants were harvested at 72 h post-infection and analyzed for the production of IFN- α and IFN- β by ELISA. Data shown are the average \pm SEM of 3–4 separate experiments.

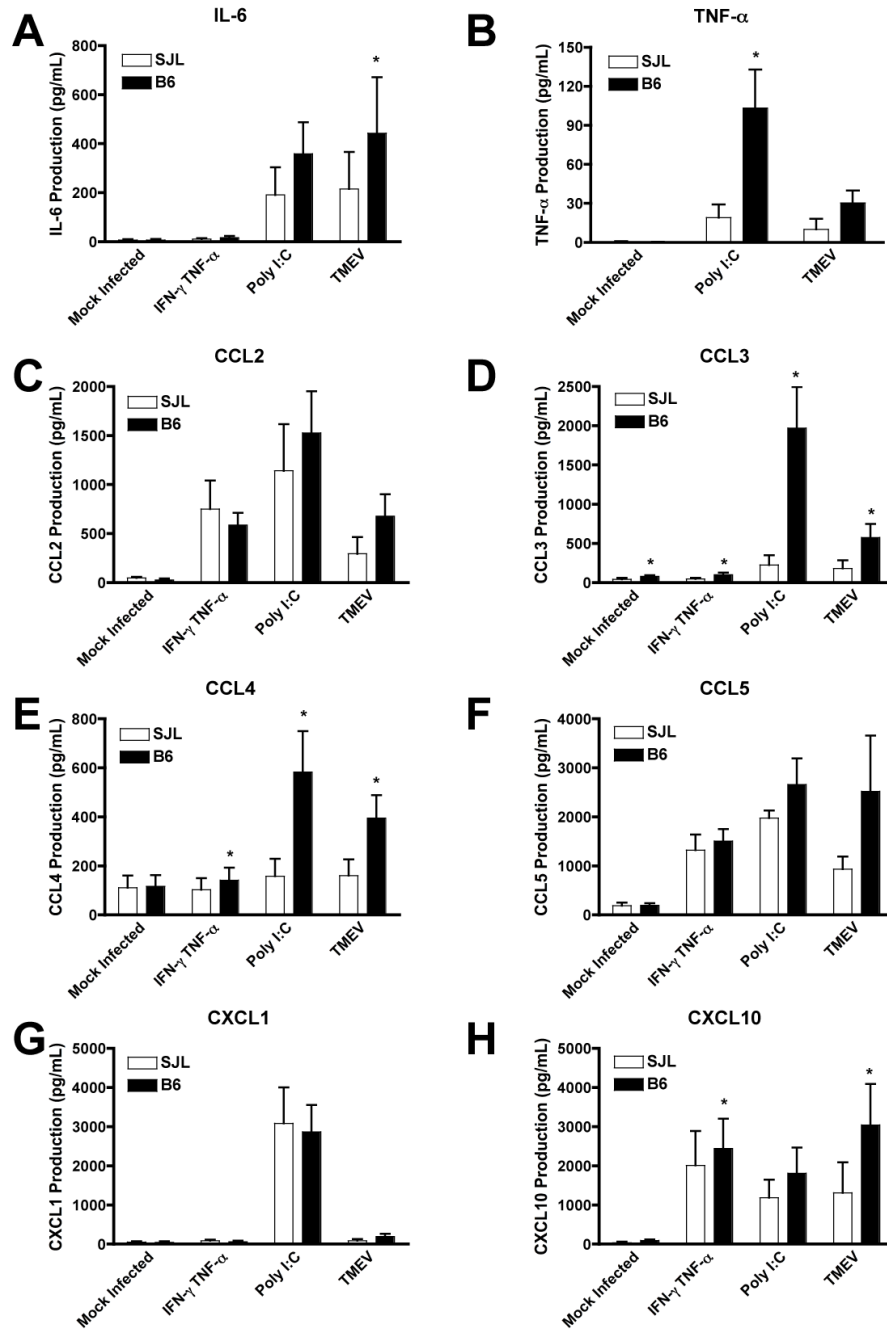


Figure 3. B6 astrocytes produce greater amounts of cytokines and chemokines than SJL astrocytes SJL or B6 astrocytes were mock infected, treated with IFN- γ and TNF- α (100 and 500 U/ml), poly I:C (100 mg/ml), or infected with TMEV (MOI=10). Supernatants were harvested at 72 h and analyzed for the production of cytokines and chemokines by Luminex multiplex bead assay. Data shown are the average \pm SEM of 5 separate experiments. *Levels significantly increased compared to SJL astrocytes by paired one-tailed Student's t test, p<0.05

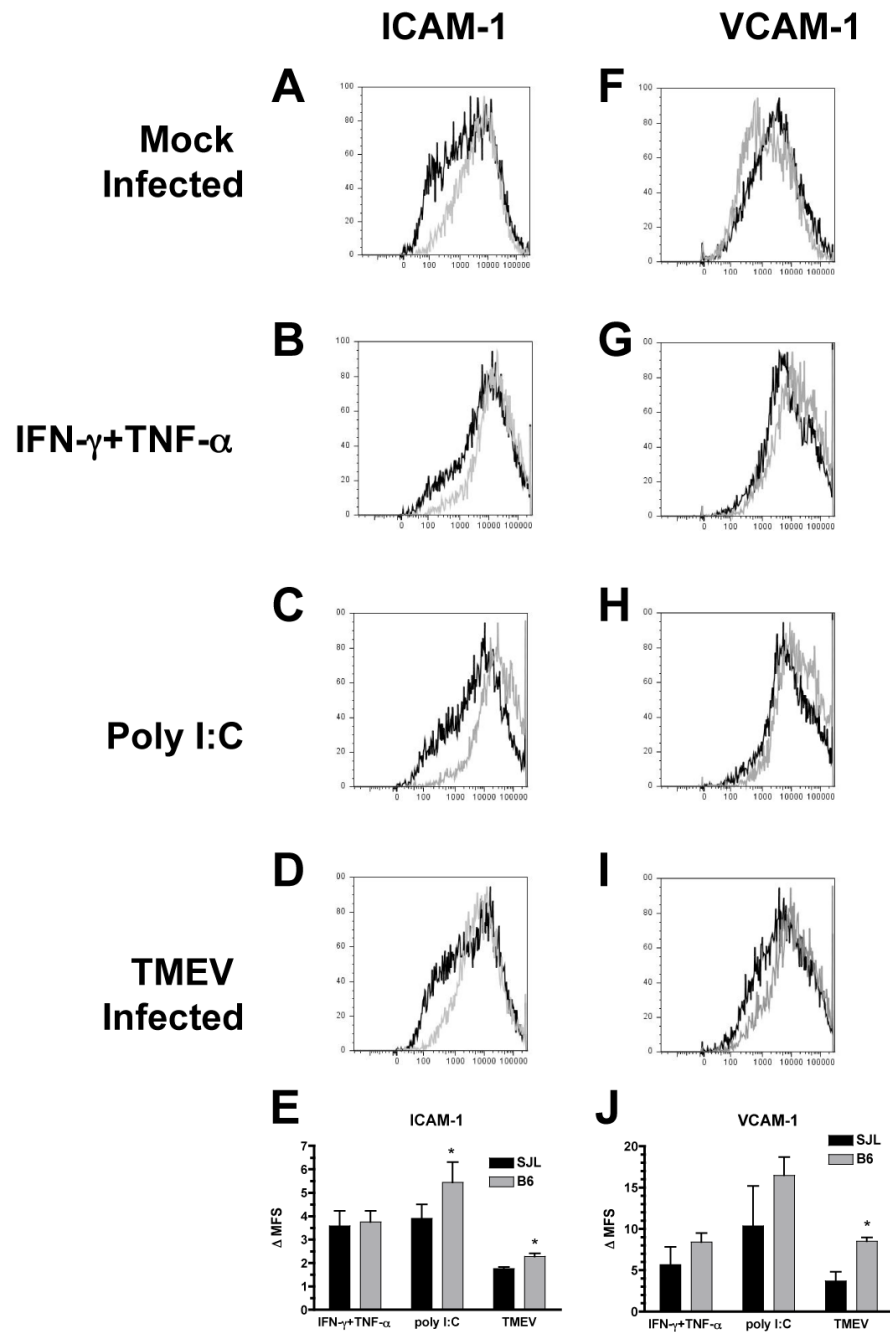


Figure 4. B6 astrocytes express higher levels of adhesion molecules than SJL astrocytes
Astrocyte cultures from B6 (gray lines) or SJL (black lines) mice were mock infected, treated with IFN- γ and TNF- α (100 and 500 U/ml) or poly I:C (100 μ g/ml), or infected with TMEV (MOI=10). After 48 h, cells were harvested and stained for ICAM-1 (A–D) and VCAM-1 (F–I) and analyzed by flow cytometry. Plots shown are representative of 3–4 separate experiments. (E,J) The fold increase in the mean fluorescence shift (Δ MFS) in the treated samples compared to the control was determined. Data shown are the average \pm SEM of 3–4 separate experiments. *Levels significantly increased compared to SJL astrocytes by paired one-tailed student's T test, $p < 0.05$

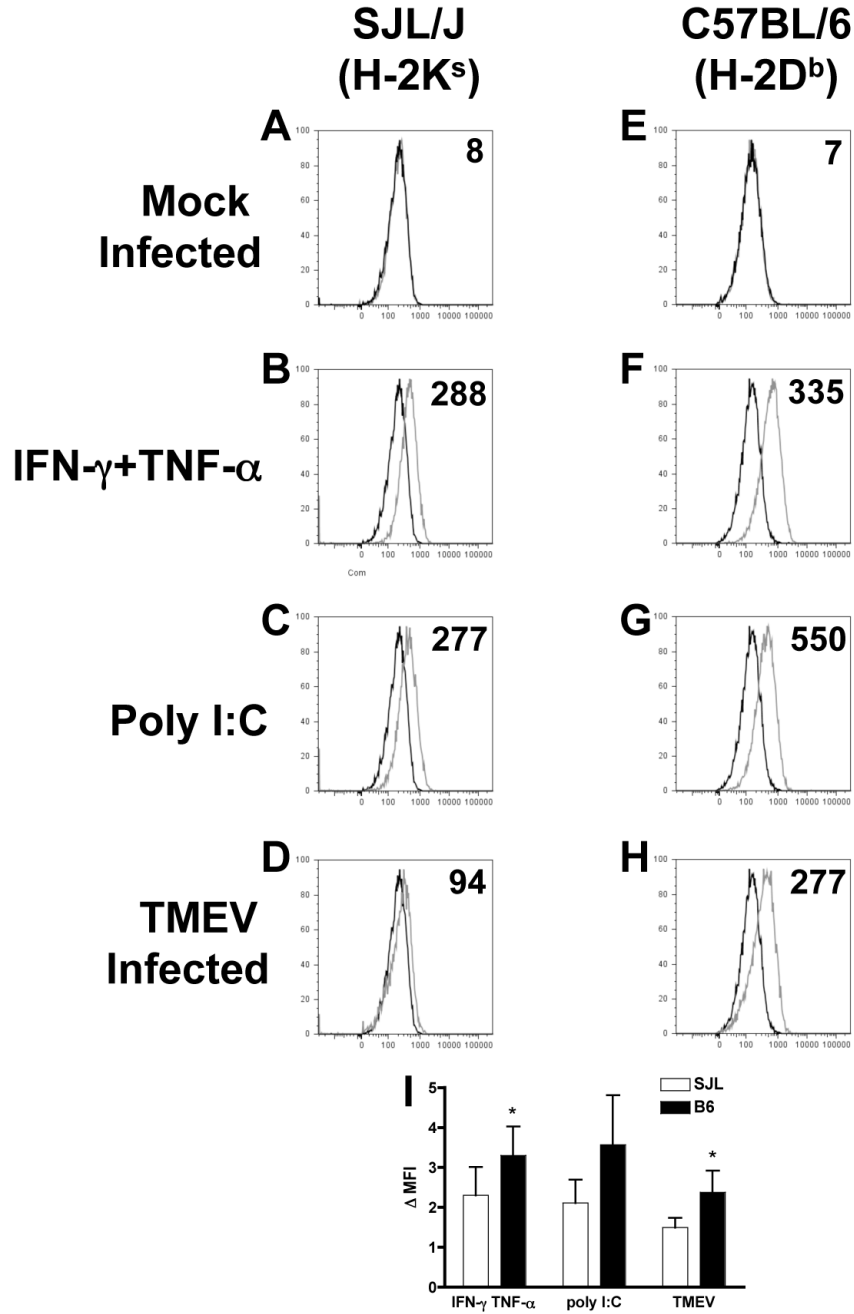


Figure 5. TMEV-infected B6 astrocytes upregulate MHC I to higher levels than SJL astrocytes
 Astrocyte cultures from SJL or B6 mice were mock infected (black line, all graphs), treated with IFN- γ and TNF- α (100 and 500 U/ml) or poly I:C (100 μ g/ml) or infected with TMEV (MOI=10; gray lines). After 48 h, cells were harvested and stained for H-2K^s (SJL) or H-2D^b (B6) MHC class I molecules and analyzed by flow cytometry. Isotype controls are indicated with gray lines in A, E. Plots shown are representative of 3–4 separate experiments (A–H). Numbers in each graph are the mean fluorescence shift (MFS) of the indicated treatment and strain. (I) The fold increase in the mean fluorescence intensity (Δ MFI) in the treated samples compared to the control was determined. Data shown are the average \pm SEM of 3

separate experiments. *Levels significantly increased compared to SJL astrocytes by paired one-tailed student's T test, $p < 0.05$.

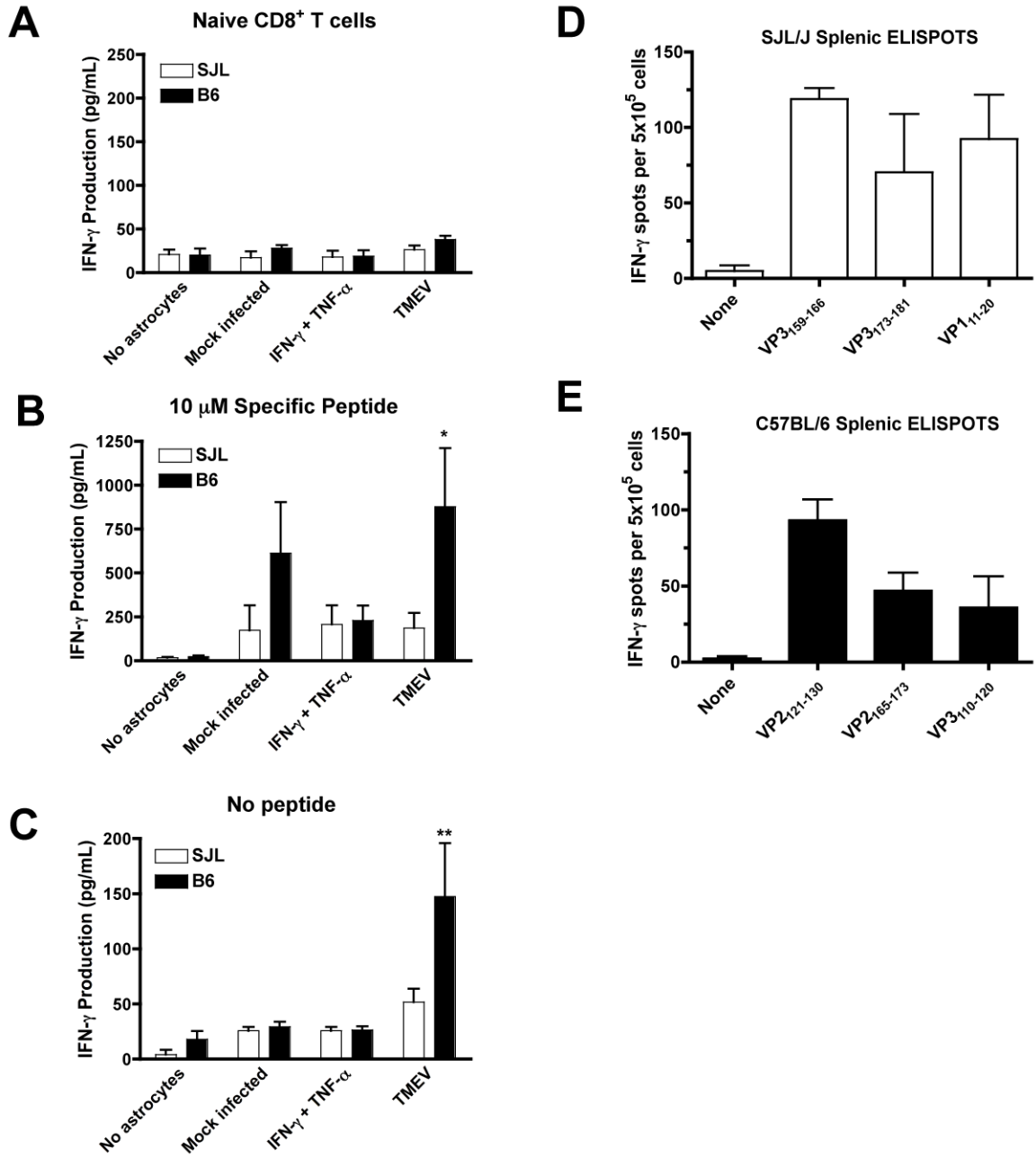


Figure 6. B6 astrocytes activate TMEV-specific CD8⁺ T cells more efficiently than SJL astrocytes
 Astrocytes were plated at 10⁵ cells/well and mock infected, treated with IFN- γ and TNF- α (100 and 500 U/ml) or infected with TMEV (MOI=10). At 48 h, CD8⁺ T cells were isolated by AutoMacs from naive (A) or TMEV-infected mice at day 8 pi (B,C). Astrocytes were washed and incubated with (A,B) or without (C) 10 μ M specific peptide (VP3₁₅₉₋₁₆₆ for SJL and VP2₁₂₁₋₁₃₀ for B6). Culture supernatants were collected at 96 h and analyzed for IFN- γ production by Luminex multiplex bead assay. Data shown is the average \pm SEM of 4 separate experiments. With specific peptide, the effect of strain was significant by two-way ANOVA ($p=0.0108$). Without specific peptide, both strain ($p=0.0371$) and treatment ($p=0.0022$) had significant effects. Levels significantly increased in B6 compared to SJL by Bonferroni post test, * $p<0.05$; ** $p<0.01$. The frequency of splenic T cells producing IFN- γ in response to the

dominant and subdominant viral epitopes from both SJL (**D**) and B6 (**E**) mice were analyzed at day 7 PI. Of note the frequency of IFN- γ -producing splenic CD8 T cells specific for the respective immunodominant CTL epitopes did not differ between the two mouse strains. Results are representative of two separate experiments.