

RESEARCH PAPER

Chronic heart rate reduction by ivabradine prevents endothelial dysfunction in dyslipidaemic mice

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Background and purpose: High resting heart rate is a predictor for total and cardiovascular mortality independent of other risk factors in patients with coronary artery disease. We tested the hypothesis that a reduction of resting heart rate with the cardiac pacemaker I_f current inhibitor ivabradine prevents the endothelial dysfunction associated with dyslipidaemia.

Experimental approach: Three-month-old dyslipidaemic (DL) male mice expressing the human ApoB-100 were assigned or not (DL, $n=16$), to treatment for 3 months with ivabradine ($10 \text{ mg kg}^{-1} \text{ d}^{-1}$, $n=17$). Wild-type C57Bl/6 mice (WT, $n=15$) were used as controls. Heart rate was measured at 3, 4.5 and 6 months. Dilatation to acetylcholine (ACh) of isolated cerebral and renal arteries was investigated at 6 months.

Key results: Heart rate remained stable in anaesthetized WT mice, increased (25%, $P<0.05$) with age in DL mice but was limited (11%, $P<0.05$) by ivabradine. At 6 months, left ventricular maximal pressure was similar in all groups. The minimal and end-diastolic left ventricular pressures were increased ($P<0.05$) in DL (10.2 ± 1.0 and 18.7 ± 1.4 mm Hg) compared to WT (-0.4 ± 0.7 and 6.3 ± 1.0 mm Hg) and reduced ($P<0.05$) by ivabradine (4.2 ± 1.3 and 11.5 ± 1.5 mm Hg). ACh-induced maximal dilatation was impaired ($P<0.05$) in renal and cerebral arteries isolated from DL compared to WT (56 ± 7 versus $83 \pm 3\%$ in renal arteries; 22 ± 2 versus $42 \pm 2\%$ in cerebral arteries). Ivabradine completely prevented ($P<0.05$) this dysfunction in renal and cerebral arteries.

Conclusions and implications: Selective heart rate reduction with ivabradine limits cardiac dysfunction and prevents the renovascular and cerebrovascular endothelial dysfunction associated with dyslipidaemia.

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Abbreviations: DL, dyslipidaemic mice; DL + IVA, dyslipidaemic mice treated with ivabradine; INDO, indomethacin; L-NNA, N^onitro-L-arginine; NAC, N-acetyl-L-cysteine; WT, wild-type mice

Introduction

In the 1980's, an epidemiological report established that increased resting heart rate was a prognostic factor for coronary artery disease and mortality (Dyer *et al.*, 1984). This was confirmed by data from the Framingham study (Kannel, 1987) and more recently by data from our group (Diaz *et al.*, 2005). Although it was shown, experimentally, that a stress-related β -adrenoceptor-mediated increase in heart rate could promote atherosclerosis in monkeys (Kaplan *et al.*, 1987; Strawn *et al.*, 1991), the link between cardiovascular diseases and heart rate remains largely unknown. Stress-related tachycardia has been used to investigate the mechanisms of enhanced atherogenesis at high heart rate. The proportion

of dysfunctional coronary artery endothelial cells in monkeys exposed to behavioral stress was much higher in untreated, tachycardic animals than in those treated with a β -blocker (Strawn *et al.*, 1991). This endothelial dysfunction associated with high heart rate may represent an important mechanism of increased atherogenesis. Whether the beneficial effects of β -blockade in this setting are specifically due to the decrease in heart rate or rather to the reduction of other deleterious impacts of the hyperadrenergic state remains unknown.

Du *et al.* (2004) demonstrated that ivabradine, an inhibitor of the pacemaker I_f current in the sino-atrial node (Tardif *et al.*, 2005), reduces heart rate in conscious and anaesthetized mice independently of sympathetic activation. Specific inhibition of I_f current also does not affect blood pressure or myocardial contractility, intracardiac conduction or ventricular repolarization (Vilaine, 2006). Ivabradine is therefore an ideal agent to test the hypothesis that selective reduction of heart rate prevents endothelial dysfunction

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independently of the level of activation of the sympathetic nervous system. For this study, we used mice expressing the human apoprotein-B, because we have shown that these mice develop mild dyslipidaemia associated with changes in the endothelial pathways leading to arterial dilatation (Krummen *et al.*, 2005; Gendron *et al.*, 2007), and an accelerated endothelial dysfunction thereafter (Krummen *et al.*, 2006).

Methods

The procedures and protocols were performed in accordance with our institutional guidelines and the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH publication nos. 85-23, revised 1996). Three-month-old male C57Bl/6 dyslipidaemic mice (DL) ($n=55$) expressing human apoprotein B-100 (Sanan *et al.*, 1998; Krummen *et al.*, 2005) and 3-month-old wild-type (WT) C57Bl/6 control male mice ($n=20$) were used. The plasma concentration of cholesterol was 2.7 ± 0.1 mM in WT and 3.7 ± 0.3 mM in DL mice ($P<0.05$). Triglyceride levels were increased ($P<0.05$) from 1.3 ± 0.2 mM in WT to 3.1 ± 0.5 mM in DL mice. Hypercholesterolaemia was maintained in 6-month-old DL mice and unaffected by a 3-month treatment with ivabradine.

Dyslipidaemic mice were assigned to receive 3 months of treatment (from 3 to 6 months) with ivabradine ($10 \text{ mg kg}^{-1} \text{ day}^{-1}$ in drinking water; Du *et al.*, 2004), metoprolol ($80 \text{ mg kg}^{-1} \text{ day}^{-1}$ in drinking water), with the doses adjusted to body weight (DL + IVA, $n=17$; DL + METO, $n=13$) or no treatment (DL, $n=16$). At the age of 3 and 4.5 months, all mice were anaesthetized with isoflurane (2.5%) in O_2 (0.51 min^{-1}) to monitor heart rate by echocardiography (7 MHz). At the age of 6 months, mice were killed by exsanguination under anaesthesia (isoflurane) after heart rate and cardiac function had been measured using a Millar catheter inserted in the left ventricle via carotid artery. During the course of the study, three DL mice and one ivabradine-treated mouse were killed at the age of 4.5 months, and three DL and two ivabradine-treated mice were killed at the age of 6 months.

The right and left renal arteries, as well as the middle and posterior communicating cerebral arteries, were isolated and placed in ice-cold physiological salt solution of the following composition (mM): NaCl 130, KCl 4.7, KH_2PO_4 1.18, MgSO_4 1.17, NaHCO_3 14.9, CaCl_2 1.6, EDTA 0.023 and glucose 10, aerated with 12% O_2 /5% CO_2 /83% N_2 (37 °C, pH 7.4). All experiments were conducted on segments of 2–3 mm in length and pressurized in a pressure myograph (Living System, Burlington, VT, USA) at 100 (renal) or 60 mm Hg (cerebral) (Drouin *et al.*, 2007; Gendron *et al.*, 2007). An equilibration time of 45 min was allowed before every experiment.

Telemetry

In a separate experiment, eleven 3-month-old DL mice and five age-matched WT mice were simultaneously instrumented, under isoflurane anaesthesia, with OpenHeart radio-frequency transmitters (Data Sciences International, Arden

Hills, MN, USA) as described previously (Brouillette *et al.*, 2007). Electrocardiogram lead placement represents conventional lead II position. The 16 signals corresponding to the 16 mice were acquired simultaneously. Recordings were analysed with ECG Auto (version 1.7; EMKA Technologies, Paris, France) and heart rates calculated. Six of the DL mice were treated with ivabradine ($10 \text{ mg kg}^{-1} \text{ day}^{-1}$) *per os* as described above.

Experimental protocols

Dilator responses to acetylcholine (ACh, 1 nM to $30 \mu\text{M}$) were measured in vessels pre-contracted with phenylephrine ($10 \mu\text{M}$). The level of pre-contraction was similar in all groups. Only one concentration–response curve was obtained by arterial segment. In renal artery segments, ACh-induced dilator responses were measured after inhibition of nitric oxide synthase (NOS) using N^{ω} -nitro-L-arginine (L-NNA, $10 \mu\text{M}$) and in the presence of the antioxidant *N*-acetyl-L-cysteine (NAC, $1 \mu\text{M}$), because free radicals impair renal endothelial function of DL mice (Gendron *et al.*, 2007). In cerebral artery segments, ACh-induced dilator responses were measured after inhibition of the cyclooxygenases (COX) with indomethacin (INDO, $10 \mu\text{M}$) and in the presence of catalase ($100 \mu\text{ml}^{-1}$; extract from bovine liver, EC no. 232-577-1 with 2940 U mg^{-1} protein), an inactivator of H_2O_2 , which is an endothelium-derived relaxing factor in these arteries (Drouin *et al.*, 2007). The inability of ivabradine to induce direct vascular effects was tested separately: cerebral and renal arteries isolated from WT mice were exposed to ivabradine ($0.1 \mu\text{M}$, compared with a plasmatic concentration of $\approx 0.025 \mu\text{M}$ at a treatment dose of $10 \text{ mg kg}^{-1} \text{ day}^{-1}$; Du *et al.*, 2004) directly in the organ bath. This did not limit either myogenic tone, the level of precontraction induced by phenylephrine or the dilatation induced by ACh ($1 \mu\text{M}$) (data not shown). Only the control dilator responses induced by ACh were measured in mice treated with metoprolol. All inhibitors were added to the bath 30 min before the start of the protocol.

Statistical analysis

In every case, n refers to the number of animals used in each protocol. Continuous variables are expressed as mean \pm s.e.mean. The distribution of each parameter was investigated and observed to be normal. Half-maximum effective concentrations of ACh were measured from each individual concentration–response curve using a logistic curve-fitting programme (Allfit; Dr André Deléan, Université de Montréal). Vascular sensitivity (pD_2) values, the negative log of the half-maximum effective concentrations, were obtained. For each protocol, the basal diameter in the no-flow condition was determined at the end of the 45-min equilibration period. Myogenic tone, which is a reduction in diameter induced by an increase in luminal pressure, was measured and is expressed as a percentage of the maximal diameter, obtained at the end of the experiment using a Ca^{2+} -free physiological salt solution containing $10 \mu\text{M}$ of

sodium nitroprusside. ACh-induced dilatation is expressed as a percentage of the maximal diameter.

Haemodynamic parameters and myogenic tone were studied using one-way analysis of variance. When *P*-value for group effect was statistically significant, *post hoc* tests were performed to compare the three groups of mice. Two-way repeated-measure analyses of variance were performed to study the change over time of heart rate and body weight of the three groups of mice. When the interaction between time and group was significant, *post hoc* tests were used to investigate the effects of group and time. The effects of L-NNA, NAC, COX and INDO on ACh-induced dilatations, in each group, were compared using one-way analysis of variance followed, in case of significance, by *post hoc* tests. To assess the effects of L-NNA, NAC, COX and INDO on ACh-induced dilatations between groups, the change in each measurement from pretreatment (control) to post treatment (in presence of L-NNA, NAC, COX and INDO) was calculated for each of the three groups. Comparisons between groups (WT versus DL, DL versus DL + IVA and DL + IVA versus WT) were evaluated by *t*-test. The results were considered to be statistically significant when the *P*-value was <0.05.

Materials

Metoprolol and all antagonists and inhibitors were purchased from Sigma-Aldrich Canada Ltd. (Oakville, Ontario, Canada). Ivabradine was provided by Institut de Recherches Internationales Servier (Courbevoie, France).

Results

Heart rate and cardiac function

The heart rate of the WT mice remained unchanged over the 3-month period (Table 1). This contrasts with the age-dependent rise in heart rate in DL mice at 4.5 and 6 months ($P < 0.05$ versus WT mice). In DL mice treated with ivabradine, the rise in heart rate observed in the untreated DL animals was prevented at 4.5 months and reduced at 6 months ($P < 0.05$ versus DL at both time points). The minimal (minimal left-ventricular pressure (P_{\min})) and end-diastolic (P_{ed}) left-ventricular pressures were increased in DL mice ($P < 0.05$ versus WT), without change in maximal (P_{max}) left-ventricular systolic pressure and contractility ($\pm dP/dt$) and the relaxation constant τ (Table 2). The isovolumic

contraction time and relaxation time were also increased in 6-month-old DL mice ($P < 0.05$ versus WT). Treatment with ivabradine prevented the changes in isovolumic contraction time ($P < 0.05$ versus DL) and isovolumic relaxation time, and limited the rise in P_{\min} and P_{ed} ($P < 0.05$ versus DL for all parameters).

Using telemetric measures in conscious mice, the heart rate-lowering effect of ivabradine was confirmed over a 6-week period (Figure 1). Heart rate (beats per minute) before initiation of treatment was similar in all groups (WT: 600 ± 11 , DL: 586 ± 14 and DL + IVA: 607 ± 9) and did not change in untreated mice (WT: 608 ± 3 and DL: 613 ± 39). However, ivabradine significantly ($P < 0.05$) reduced heart rate by 17% in DL mice from the first (515 ± 3) to the sixth week (507 ± 4 beats min^{-1}) of treatment (Figure 1).

Mouse body weight

The body weight of the animals rose significantly and regularly from the age of 3–4.5 months ($P < 0.05$) and from 4.5–6 months ($P < 0.05$) in WT ($n = 15$) mice (27 ± 0 , 33 ± 1 and 38 ± 1 g, respectively). In DL mice ($n = 16$ – 10), evolution of body weight was similar (28 ± 1 , 33 ± 2 and 37 ± 2 g). In the presence of ivabradine ($n = 17$ – 15), body weight gain was slower from 29 ± 1 g at 3 months to 31 ± 1 g at 4.5 months ($P < 0.05$ versus 3 months) and 33 ± 2 g at 6 months ($P < 0.05$ versus WT at 6 months).

Renal artery reactivity

Pressure induced a myogenic response of renal arteries in the three groups, although myogenic tone was reduced in DL

Table 1 Heart rate (beats min^{-1}) of WT, DL and DL + IVA mice measured under isoflurane anaesthesia at 3, 4.5 and 6 months of age

Age (months)	WT	n	DL	n	DL + IVA	n
3	385 ± 8	15	373 ± 9	16	385 ± 10	17
4.5	405 ± 11	15	$431 \pm 14^*$	13	$375 \pm 13^{\S}$	16
6	381 ± 15	15	$465 \pm 10^{*,\ddagger}$	10	$422 \pm 11^{*,\ddagger,\S}$	14

Abbreviations: DL, dyslipidaemic mice; DL + IVA, dyslipidaemic mice treated with ivabradine; WT, wild-type mice.

DL + IVA mice were treated with ivabradine (IVA, $10 \text{ mg kg}^{-1} \text{ day}^{-1}$) from the age of 3 to 6 months. Data are mean \pm s.e.mean. $^*P < 0.05$ compared with 3 months; $^{\ddagger}P < 0.05$ compared with 4.5 months; $^{\S}P < 0.05$ compared with the WT; $^{\S}P < 0.05$ compared with DL. *n* refers to the number of animals used in each group.

Table 2 Cardiac function and left-ventricular pressures of WT ($n = 15$), DL ($n = 10$) and DL + IVA ($n = 14$) mice measured under isoflurane anaesthesia at 6 months of age with a Millar catheter

Groups	P_{max} (mm Hg)	P_{\min} (mm Hg)	P_{ed} (mm Hg)	$+dP/dt$ (mm Hg s^{-1})	$-dP/dt$ (mm Hg s^{-1})	IVCT (ms)	IVRT (ms)	τ (ms)
WT	100 ± 1	-0.4 ± 0.7	6.3 ± 1.0	5364 ± 114	-5041 ± 134	12.9 ± 0.1	34.4 ± 0.7	6.6 ± 0.5
DL	108 ± 3	$10.2 \pm 1.0^*$	$18.7 \pm 1.4^*$	5253 ± 110	-4849 ± 129	$14.7 \pm 0.6^*$	$39.9 \pm 0.4^*$	6.6 ± 0.2
DL + IVA	102 ± 2	$4.2 \pm 1.3^{*,\ddagger}$	$11.5 \pm 1.5^{*,\ddagger}$	5277 ± 107	-4958 ± 92	$13.1 \pm 0.3^{\ddagger}$	36.9 ± 1.6	7.0 ± 0.2

Abbreviations: τ , relaxation time constant; $\pm dP/dt$, contractility index; DL, dyslipidaemic mice; DL + IVA, dyslipidaemic mice treated with ivabradine; IVCT, isovolumic contraction time; IVRT, isovolumic relaxation time; P_{ed} , left ventricular end-diastolic pressure; P_{max} , maximal left ventricular systolic pressure; P_{\min} , minimal left ventricular pressure; WT, wild-type mice.

DL + IVA mice were treated with ivabradine (IVA, $10 \text{ mg kg}^{-1} \text{ day}^{-1}$) from the age of 3–6 months. Data are mean \pm s.e.mean. $^*P < 0.05$ compared with WT; $^{\ddagger}P < 0.05$ compared with DL.

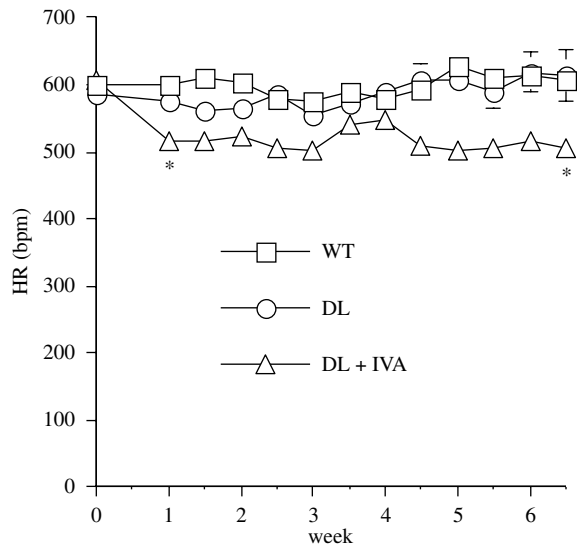


Figure 1 The reduction of heart rate (HR) by ivabradine (IVA, $10 \text{ mg kg}^{-1} \text{ day}^{-1}$) measured by telemetry over a 6-week treatment period in conscious instrumented DL mice ($n=6$) compared to untreated DL ($n=5$) and WT ($n=5$) mice. All signals were recorded in parallel using a 16-channel EMKA telemetry acquisition set up. Data are mean \pm s.e.mean. * $P < 0.05$ compared with WT and DL (error bars are encompassed in the thickness of the symbols). DL, dyslipidaemic; WT, wild type.

Table 3 Basal vascular parameters in control conditions of pressurized (100 mm Hg) renal arteries isolated from WT ($n=14$), DL ($n=8-14$) and DL + IVA ($n=9-16$) mice

Groups	D_{max} (μm)	Myogenic tone (% of D_{max})	Contraction to PE $10 \mu\text{M}$ (% of D_{max})	Contraction to high K^+ (% of D_{max})
WT	385 ± 22	33 ± 3	64 ± 2	66 ± 3
DL	399 ± 20	$16 \pm 3^*$	64 ± 3	61 ± 4
DL + IVA	403 ± 16	$9 \pm 2^*$	64 ± 3	68 ± 3

Abbreviations: DL, dyslipidaemic mice; DL + IVA, dyslipidaemic mice treated with ivabradine; D_{max} , maximal diameter; high K^+ , 40 mM KCl physiological solution; PE, phenylephrine; WT, wild-type mice.

DL + IVA mice were treated with ivabradine (IVA, $10 \text{ mg kg}^{-1} \text{ day}^{-1}$) from the age of 3–6 months. Data are mean \pm s.e.mean. * $P < 0.05$ compared with the WT.

mice and ivabradine-treated mice (Table 3). There was no difference in the amplitude of the precontracting tone induced by phenylephrine or high potassium level, as well as in the maximal vessel diameters measured at the end of the experiment in calcium-free solution containing sodium nitroprusside. These data suggest that the smooth muscle of these vessels is not altered by dyslipidaemia and/or ivabradine.

The endothelium-dependent dilatation induced by ACh was significantly impaired at 6 months in arteries isolated from DL mice compared with WT mice ($P < 0.05$; Figure 2). However, 3 months of treatment with ivabradine completely prevented this abnormality: this was evidenced not only by the maintenance of the maximal dilatation induced by ACh, but also by an increased vascular sensitivity to ACh (Table 4).

We previously demonstrated that acute exposure to NAC, an antioxidant, restored maximal dilatation to ACh in DL

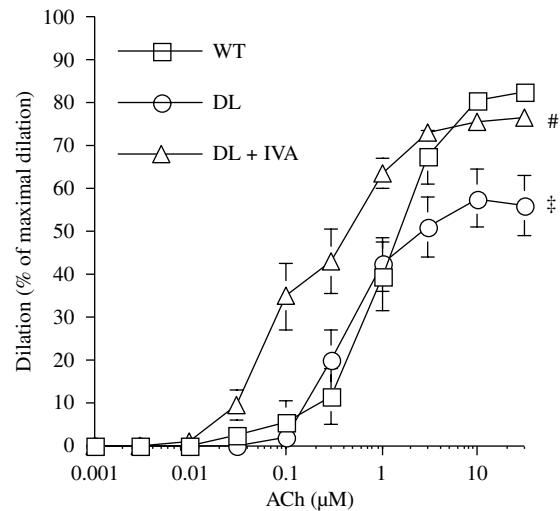


Figure 2 Endothelium-dependent dilatation in response to ACh of pressurized (100 mm Hg), precontracted with phenylephrine, renal arteries isolated from 6-month-old WT ($n=13$), DL ($n=13$) and DL + IVA ($n=15$) mice. DL + IVA mice were treated with ivabradine (IVA, $10 \text{ mg kg}^{-1} \text{ day}^{-1}$) from the age of 3–6 months. Data are mean \pm s.e.mean. ‡ $P < 0.05$ compared with WT; # $P < 0.05$ compared with DL. ACh, acetylcholine; DL, dyslipidaemic; DL + IVA, dyslipidaemic mice treated with ivabradine; WT, wild type.

Table 4 Maximal dilatation and vascular sensitivity to ACh of pressurized renal (100 mm Hg) arteries isolated from WT ($n=8-14$), DL ($n=7-13$) and DL + IVA ($n=11-15$) mice

	WT		DL		DL + IVA	
	pD_2	E_{max}	pD_2	E_{max}	pD_2	E_{max}
Control	6.0 ± 0.1	83 ± 3	6.2 ± 0.2	$56 \pm 7^\ddagger$	$6.8 \pm 0.1^{\ddagger, \S}$	$77 \pm 3^\S$
L-NNA	5.8 ± 0.1	$59 \pm 9^*$	5.6 ± 0.2	40 ± 7	$6.3 \pm 0.1^{*, \ddagger, \S}$	$56 \pm 6^*$
NAC	$6.2 \pm 0.1^\ddagger$	81 ± 7	6.3 ± 0.2	$83 \pm 2^*$	$6.2 \pm 0.2^*$	78 ± 5

Abbreviations: DL, dyslipidaemic mice; DL + IVA, dyslipidaemic mice treated with ivabradine; E_{max} , % of maximal dilatation; L-NNA, N-nitro-L-arginine; NAC, N-acetyl-L-cysteine; pD_2 , vascular sensitivity; WT, wild-type mice.

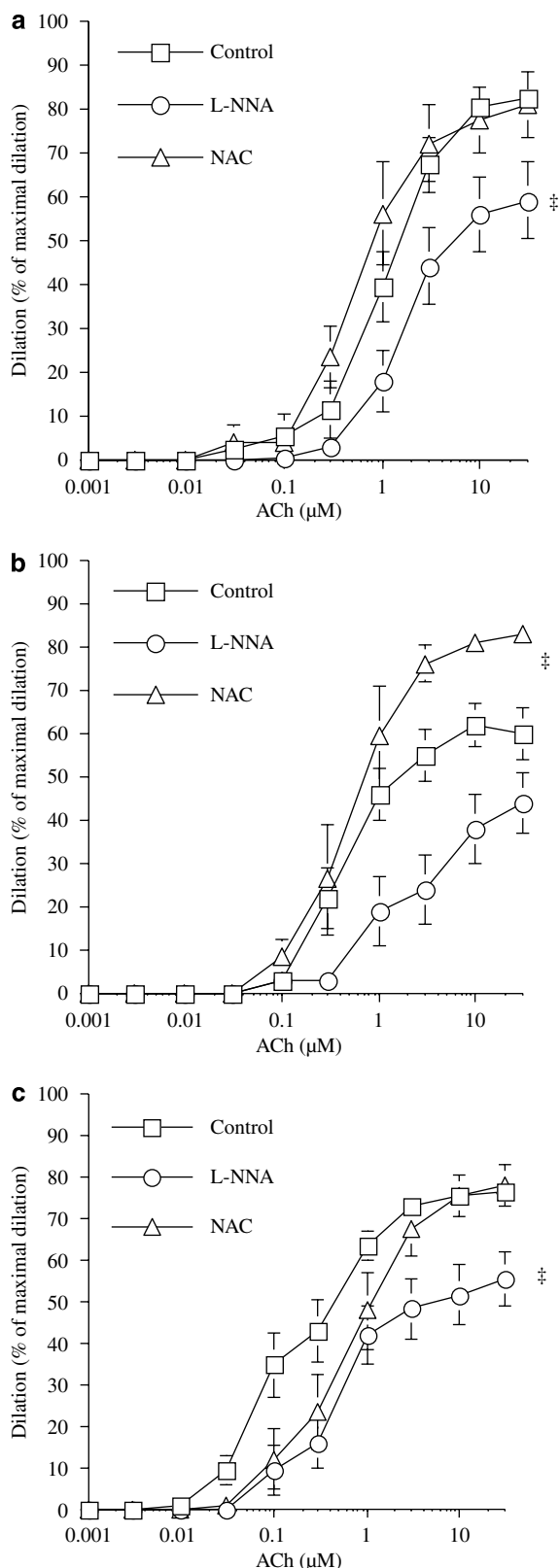
DL + IVA mice were treated with ivabradine (IVA, $10 \text{ mg kg}^{-1} \text{ day}^{-1}$) from the age of 3–6 months. Responses were obtained in control conditions (control) either in the presence of eNOS inhibition (L-NNA, $10 \mu\text{M}$), or in the presence of the antioxidant NAC ($1 \mu\text{M}$). Data are mean \pm s.e.mean. * $P < 0.05$ compared with control; † $P < 0.05$ compared with L-NNA; ‡ $P < 0.05$ compared with the WT; § $P < 0.05$ compared with DL.

mice (Krummen *et al.*, 2006; Gendron *et al.*, 2007). This response was confirmed, as shown in Figure 2, whereas NAC had no effect on vessels isolated from WT mice (Figure 3a). In ivabradine-treated mice, endothelial function was maintained and NAC had no effect (Figure 3c), as was the case in WT mice.

We also tested the effect of endothelial nitric oxide synthase (eNOS) inhibition, using L-NNA, on the dilatation induced by ACh. In renal arteries isolated from WT mice, L-NNA significantly reduced dilatation induced by ACh (Figure 3a), as observed previously (Gendron *et al.*, 2007). In DL mice, the reduced dilatation induced by ACh was not further impaired by NOS inhibition (Figure 3b). However, ivabradine revealed that dilatation induced by ACh was sensitive to L-NNA, indicating significant involvement of NO in these treated DL mice.

Cerebral artery reactivity

In the three groups, the myogenic tone of the cerebral arteries induced by an internal pressure of 60 mm Hg was similar (Table 5). In addition, there was no difference in the



amplitude of the precontracting tone induced by phenylephrine or high external potassium, or in the maximal vessel diameters measured at the end of the experiment in calcium-free solution containing sodium nitroprusside. These data indicate that vascular smooth-muscle contractile and dilator mechanisms are not affected by dyslipidaemia and/or ivabradine.

The endothelium-dependent dilatation induced by ACh was significantly impaired at 6 months in arteries isolated from DL mice compared with WT mice ($P < 0.05$; Figure 4). Ivabradine prevented this endothelial dysfunction, as revealed by maintenance of maximal dilatation induced by ACh in the treated group ($P < 0.05$ for ivabradine-treated versus DL mice; Table 6).

We recently demonstrated (Drouin *et al.*, 2007) that eNOS-derived hydrogen peroxide production is an important endothelium-derived relaxing factor in mouse cerebral arteries (Figure 5a). From this we hypothesized that, in pathological conditions associated with oxidative stress, this pathway would be altered. This hypothesis is confirmed by the lack of effect of catalase in vessels isolated from DL mice (Figure 5b). In ivabradine-treated mice, the inhibitory effect of catalase was present ($P < 0.05$ versus control; Figure 5c) and similar to that of WT mice (Figure 5a). COX inhibition is also known to limit ACh-induced dilatation of mouse cerebral arteries. However, COX inhibition did not further limit the dilator response to this muscarinic agonist in DL mice (Figure 5b). The functional effect of indomethacin on ACh-induced dilatation was normal in cerebral arteries isolated from ivabradine-treated DL mice ($P < 0.05$ versus control; Figure 5c), further demonstrating that ivabradine protected these arteries from the endothelial damage associated with dyslipidaemia.

Table 5 Basal vascular parameters in control conditions of pressurized (60 mm Hg) cerebral arteries isolated from WT ($n = 12$), DL ($n = 8-9$) and DL + IVA ($n = 6-8$) mice

Groups	D_{max} (μm)	Myogenic tone (% of D_{max})	Contraction to PE $10 \mu\text{M}$ (% of D_{max})	Contraction to high K^+ (% of D_{max})
WT	190 ± 5	18 ± 2	48 ± 3	60 ± 4
DL	216 ± 6	15 ± 2	50 ± 3	55 ± 2
DL + IVA	212 ± 5	16 ± 4	55 ± 4	61 ± 5

Abbreviations: DL, dyslipidaemic mice; DL + IVA, dyslipidaemic mice treated with ivabradine; D_{max} , maximal diameter; high K^+ , 40 mM KCl physiological solution; PE, phenylephrine; WT, wild-type mice.

DL + IVA mice were treated with ivabradine (IVA, $10 \text{ mg kg}^{-1} \text{ day}^{-1}$) from the age of 3-6 months. Data are mean \pm s.e.mean.

Figure 3 Endothelium-dependent dilatation in response to ACh (control) of pressurized (100 mm Hg) renal arteries isolated from 6-month-old WT ($n = 8-14$) (a), DL ($n = 7-13$) (b) and DL + IVA ($n = 11-15$) (c) mice. The effects of eNOS inhibition by L-NNA ($10 \mu\text{M}$) and the antioxidant NAC ($1 \mu\text{M}$) were tested in separate segments of renal arteries isolated from the same animal. DL + IVA mice were treated with ivabradine (IVA, $10 \text{ mg kg}^{-1} \text{ day}^{-1}$) from the age of 3-6 months. Data are mean \pm s.e.mean. $^{\ddagger}P < 0.05$ compared with the control. ACh, acetylcholine; DL, dyslipidaemic; DL + IVA, dyslipidaemic mice treated with ivabradine; L-NNA, N^{G} -nitro-L-arginine; NAC, N -acetyl-L-cysteine; WT, wild type.

Metoprolol-induced reduction in heart rate and endothelial function

The β -adrenoceptor antagonist metoprolol prevented the rise in heart rate associated with ageing in anaesthetized DL mice

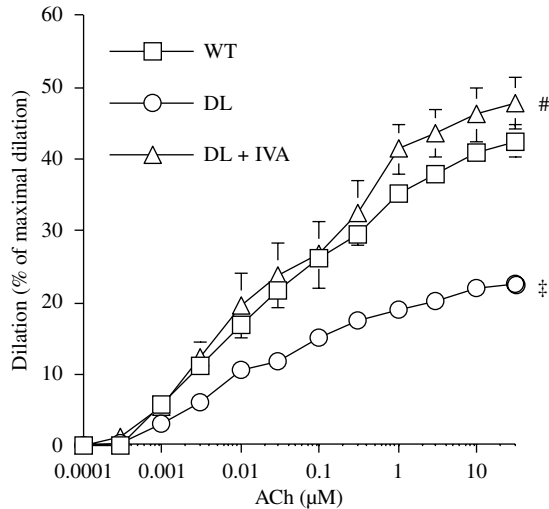


Figure 4 Endothelium-dependent dilatation in response to ACh of pressurized (60 mm Hg) cerebral arteries, preconstricted with phenylephrine, isolated from 6-month-old WT ($n=12$), DL ($n=8$) and DL + IVA ($n=8$) mice. DL + IVA mice were treated with ivabradine (IVA, 10 mg kg⁻¹ day⁻¹) from the age of 3–6 months. Data are mean \pm s.e.mean. $\ddagger P < 0.05$ compared with WT; $\# P < 0.05$ compared with DL. ACh, acetylcholine; DL, dyslipidaemic; DL + IVA, dyslipidaemic mice treated with ivabradine; WT, wild type.

Table 6 Maximal dilatation and vascular sensitivity to ACh of pressurized cerebral (60 mm Hg) arteries isolated from WT ($n=11$ –12), DL ($n=8$) and DL + IVA ($n=7$ –8) mice

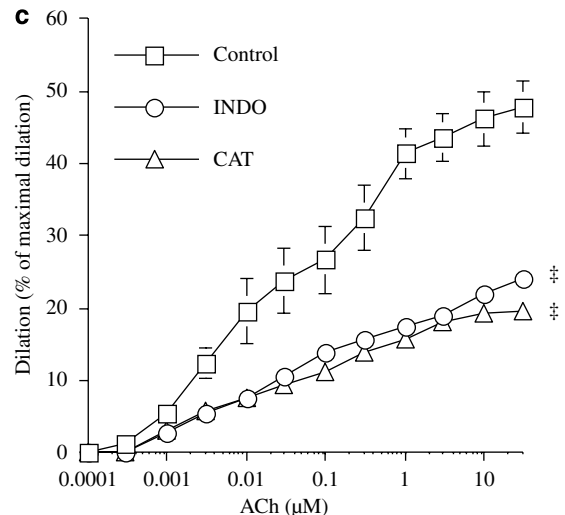
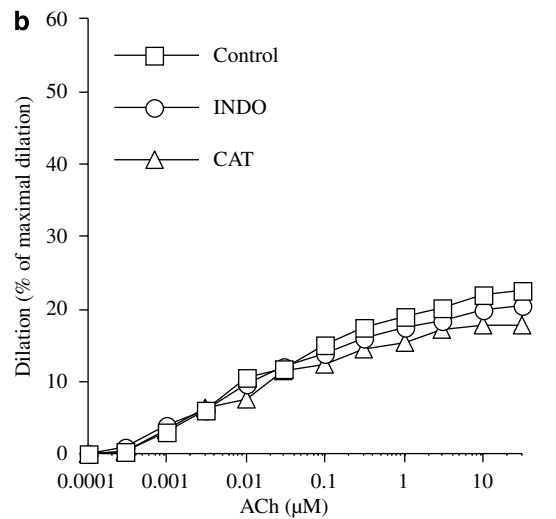
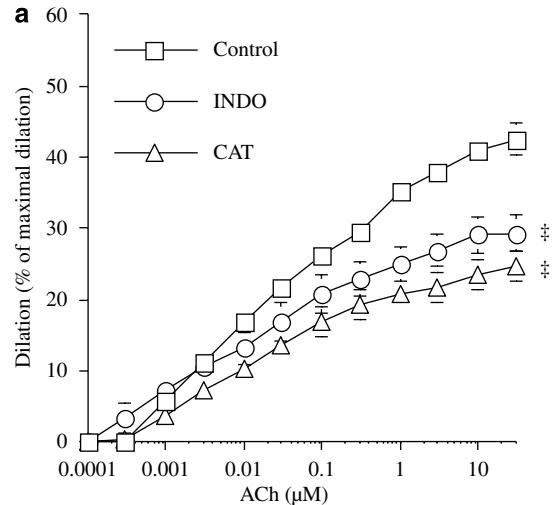
	WT		DL		DL + IVA	
	pD_2	E_{max}	pD_2	E_{max}	pD_2	E_{max}
Control	7.5 \pm 0.1	42 \pm 2	7.6 \pm 0.1	22 \pm 2 [†]	7.5 \pm 0.2	48 \pm 4 [‡]
CAT	7.7 \pm 0.1	25 \pm 2*	7.9 \pm 0.1	18 \pm 1 [†]	7.4 \pm 0.2 [‡]	20 \pm 1* [‡]
INDO	7.8 \pm 0.2	27 \pm 3*	7.8 \pm 0.2	21 \pm 2 [†]	7.0 \pm 0.3 ^{‡,†}	24 \pm 1* [‡]

Abbreviations: CAT, catalase; DL, dyslipidaemic mice; DL + IVA, dyslipidaemic mice treated with ivabradine; E_{max} , % of maximal dilatation; INDO, indomethacin; pD_2 , vascular sensitivity; WT, wild-type mice.

DL + IVA mice were treated with ivabradine (IVA, 10 mg kg⁻¹ day⁻¹) from the age of 3–6 months. Responses were obtained in control conditions (control), either in the presence of H₂O₂ inactivation by catalase (100 U ml⁻¹) or in the presence of the COX inhibitor INDO (10 μ M). Data are mean \pm s.e.mean. * $P < 0.05$ compared with control; [†] $P < 0.05$ compared with the WT; [‡] $P < 0.05$ compared with DL.

Figure 5 Endothelium-dependent dilatation in response to ACh (control) of pressurized (60 mm Hg) cerebral arteries isolated from 6-month-old WT ($n=11$ –12) (a), DL ($n=8$) (b) and DL + IVA ($n=7$ –8) (c) mice. The effects of H₂O₂ inactivation by catalase (100 U ml⁻¹) and COX inhibition by INDO (10 μ M) were tested in separate segments of cerebral arteries isolated from the same animal. DL + IVA mice were treated with ivabradine (IVA, 10 mg kg⁻¹ day⁻¹) from the age of 3–6 months. Data are mean \pm s.e.mean. $\ddagger P < 0.05$ compared with the control. ACh, acetylcholine; COX, cyclooxygenase; DL, dyslipidaemic; DL + IVA, dyslipidaemic mice treated with ivabradine; INDO, indomethacin; WT, wild type.

as efficiently as ivabradine (Figure 6a). However, the protective effect of the heart rate reduction observed under ivabradine therapy was not fully reproduced by metoprolol. Renal artery sensitivity to ACh was not improved (6.1 \pm 0.1, $n=13$) compared with vessels isolated from DL mice treated



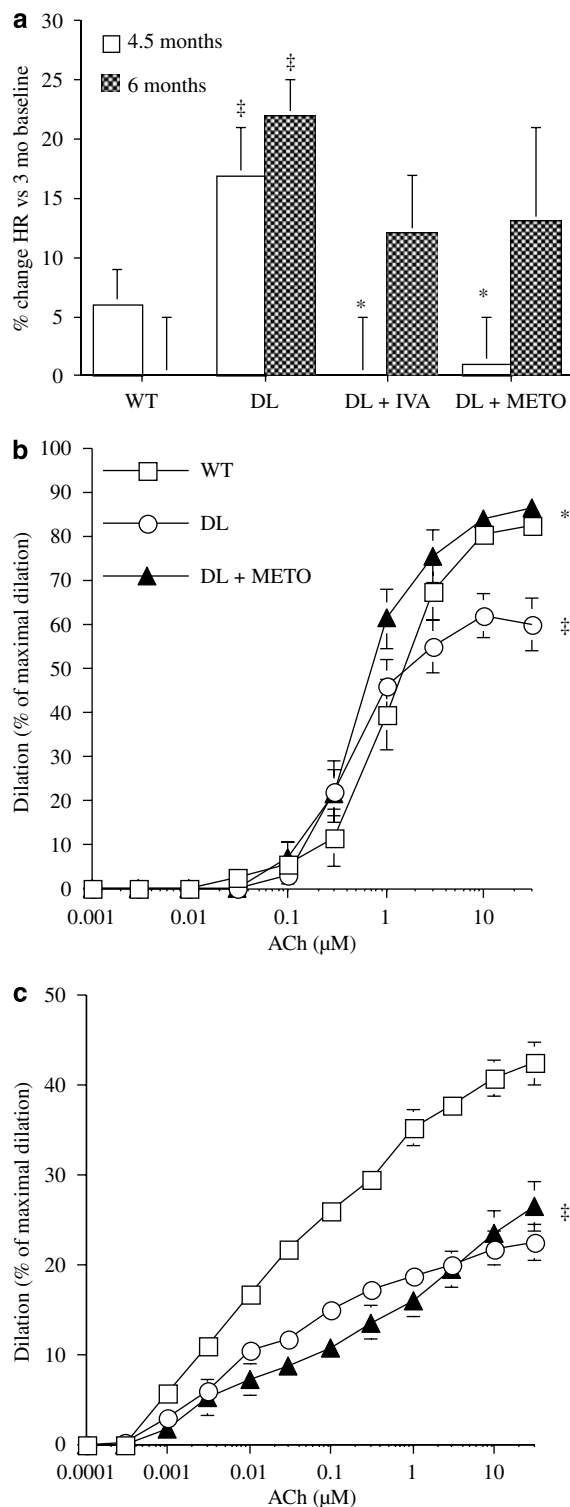


Figure 6 The effects of chronic treatment, from the age of 3-months, with ivabradine (IVA) and metoprolol (METO) on (a) the increase (%) in heart rate (HR) measured under anaesthesia in DL mice observed at 4.5 and 6 months of age. (b and c) Endothelium-dependent dilator responses to ACh of pressurized renal (b) and cerebral (c) arteries, precontracted with phenylephrine, isolated from 6-month-old WT ($n=13$ and 12), DL ($n=13$ and 8) and DL + METO ($n=13$ and 13, respectively) mice. DL + METO mice were treated with metoprolol ($80 \text{ mg kg}^{-1} \text{ day}^{-1}$) from the age of 3–6 months. Data are mean \pm s.e.mean. $^{\ddagger}P < 0.05$ compared with WT; $*P < 0.05$ compared with DL. ACh, acetylcholine; DL, dyslipidaemic; WT, wild type.

with ivabradine (Table 4, $P < 0.05$). The maximal dilatation induced by ACh was significantly improved ($P < 0.05$) in vessels isolated from metoprolol-treated mice compared with responses in vessels from untreated DL mice and not significantly different ($P > 0.05$) from those obtained in vessels from WT and from DL mice treated with ivabradine (Figure 6b). However, in cerebral arteries metoprolol did not prevent the decrease in endothelial function associated with DL (Figure 6c).

Discussion

The main finding of this study is that mild heart rate reduction by ivabradine, but not metoprolol, prevents deterioration of the endothelial dilator function of renal and cerebral arteries associated with dyslipidaemia in mice.

Several large-scale clinical reports with extended follow-up have demonstrated the relationship between heart rate and future cardiovascular events (Dyer *et al.*, 1984; Kannel, 1987; Diaz *et al.*, 2005). Smaller studies have also linked heart rate with the rate of atherosclerosis progression (Perski *et al.*, 1992). Indeed, stress-induced tachycardia has been shown experimentally to be associated with an enhancement of atherosclerosis and endothelial coronary artery cells dysfunction (Strawn *et al.*, 1991). Slower heart rate (either spontaneously or by sino-atrial node ablation) in cynomolgus monkeys was associated with a reduction of plaque burden in coronary (Beere *et al.*, 1984) and carotid (Beere *et al.*, 1992) arteries. More recently, Korshunov and Berk (2004) demonstrated that heart rate strongly predicted vascular wall remodelling in low-shear stress conditions. Even though high heart rate has been shown to be associated with endothelial dysfunction, the effect of a pure reduction in heart rate on endothelial dysfunction remained largely unknown. Cardiac I_f is a mixed sodium/potassium current responsible for the pacemaker activity in the sinus node. Ivabradine is a selective I_f current inhibitor reducing heart rate to a similar extent in conscious and anaesthetized mice (Du *et al.*, 2004), allowing us to test for the effect of pure heart rate reduction on endothelial function.

Dyslipidaemia induces an endothelial dysfunction in which oxidative stress-dependent damage plays a significant role (Krummen *et al.*, 2005, 2006; Gendron *et al.*, 2007). Our data clearly demonstrate that chronic reduction in heart rate with ivabradine preserves the function of the endothelium in mice exposed to dyslipidaemia. In ivabradine-treated DL mice, the endothelial function was normal in both vascular beds investigated. In contrast, a similar heart rate reduction using metoprolol, a β -adrenoceptor antagonist, was ineffective in preserving cerebrovascular endothelial function. In renal arteries, the vascular sensitivity to ACh was significantly enhanced in ivabradine-treated mice compared with WT mice (but identical to that measured at 3 months; see Gendron *et al.*, 2007) and the NO pathway was preserved, confirming the important protection effected by ivabradine. Metoprolol preserved the maximal dilator effect of ACh, but without maintaining vascular sensitivity. The mitigated effects of metoprolol may be related to the coupling between endothelial β -adrenoceptors and eNOS (Ciccarelli *et al.*,

2007; Kou and Michel, 2007): inhibition of this pathway could counterbalance the beneficial effects of a pure heart rate reduction on the arterial tree.

Ivabradine normalized the response of the endothelium to pharmacological interventions such as the antioxidant *N*-acetylcysteine in renal arteries as well as the H₂O₂ inactivator catalase and the COX inhibitor indomethacin in cerebral arteries. The absence of an increase in ACh-induced vasodilatation with *N*-acetylcysteine in WT mice and ivabradine-treated DL mice suggests that these animals were not subjected to increased oxidative stress, unlike untreated dyslipidaemic mice (Gendron *et al.*, 2007). Furthermore, the marked effect of catalase, which inactivates H₂O₂, on ACh-induced vasodilatation in cerebral arteries demonstrates that the eNOS-dependent pathway in this arterial bed (Drouin *et al.*, 2007) is intact in WT mice and ivabradine-treated mice, in contrast to untreated DL mice.

Since ivabradine has no direct antioxidant properties, endothelial protection must be effected by another mechanism. These protective pathways may include an improvement of the shear stress-dependent stimulation of the endothelium, favouring eNOS expression and/or preventing NO or H₂O₂ degradation. Another possibility is that heart rate reduction may result in less strain being applied onto the arterial wall, so reducing its mechanical fatigue.

Of note, all the endothelial function changes occurred without alterations in smooth-muscle contractility or maximal dilator potential. Vascular smooth-muscle contractility to phenylephrine and high potassium were not affected by either DL or ivabradine. In addition, the maximal diameter was also similar in all groups, suggesting that the relaxing mechanisms were also intact. Hence, ivabradine had no functional interactions with calcium channels and the intracellular mechanisms regulating smooth-muscle reactivity.

The chronic reduction in heart rate also limited the cardiac dysfunction that occurred in dyslipidaemic animals. The significant increase in heart rate observed between the ages of 3–6 months in DL mice was accompanied by diastolic dysfunction revealed by increases in left ventricular minimal and end-diastolic pressures and isovolumic relaxation time. Ivabradine significantly improved these diastolic function parameters in DL mice. Mulder *et al.* (2004) had previously shown that treatment with ivabradine improves diastolic dysfunction in rats subjected to coronary artery ligation. Our results are the first to demonstrate that diastolic dysfunction associated with dyslipidaemia is also improved by ivabradine.

In conclusion, this experimental study supports the concept that a chronically elevated heart rate is deleterious to the cardiovascular system. Conversely, the pharmacological reduction of heart rate with the selective I_f current blocker ivabradine is beneficial in limiting the progression of both ventricular and vascular dysfunctions.

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Conflict of interest

Dr Florence Mahlberg-Gaudin is an employee of Laboratoires Servier. Dr Tardif has received honoraria from Laboratoires Servier.

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