Properties of a Deoxyribonucleic Acid Ligase Mutant of *Escherichia coli:* X-Ray Sensitivity

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A deoxyribonucleic acid ligase-deficient mutant is X-ray sensitive relative to the parent strain, suggesting that deoxyribonucleic acid ligase functions in repair of X-ray-induced, single-strand scissions.

One of the primary consequences of X-irradiation to bacterial cells is the breakage of phosphodiester bonds in the deoxyribonucleic acid (DNA) backbone, resulting in single-strand scissions (3, 7). There is evidence that these singlestrand scissions can be repaired (1, 5, 7). An enzyme, DNA ligase, has been described which catalyzes the formation of covalent bonds between polynucleotides (2, 8); possibly this enzyme functions in the repair of X-ray-induced, singlestrand scissions. Should this be true, one would expect a ligase-deficient mutant to be X-ray sensitive. In our laboratory, we isolated a temperature-sensitive, ultraviolet (UV) radiationsensitive mutant of Escherichia coli TAU-bar, ts-7 (4), which is deficient in DNA ligase (9, 10). Accordingly, we have investigated ts-7 for X-ray sensitivity.

Cultures were grown aerobically at 25 C in a glucose-salts medium with appropriate supplements (4, 6) to early log phase (approximately 2×10^8 cells/ml), harvested by centrifugation, and suspended in the salts medium without glucose or supplements. The cells were placed in a covered, plastic petri dish and were continuously stirred during irradiation. X rays were produced at a rate of 1 kr/min by a Westinghouse Quadrocondex X-ray tube, operating at 250 kVp and 15 ma at a target distance of 22 cm. Dosimetry was made with a Victoreen dosimeter. At appropriate intervals samples were taken, and viable cell counts were determined by the agar overlay method by using the glucose-salts medium with added supplements and 15 g of agar per liter.

Data for survival of TAU-bar and ts-7 are presented in Fig. 1. From these data, ts-7 appears to be X-ray sensitive. At a dose of 20 kr, survival of TAU-bar is 0.07%, whereas that of ts-7 is 0.001%. The TAU-bar survival curve has a small shoulder at low doses; this shoulder is absent for

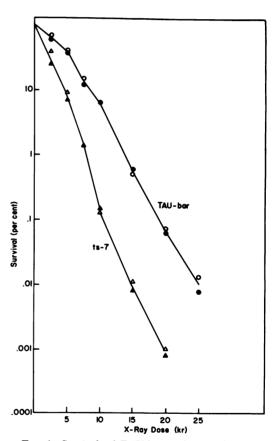


FIG. 1. Survival of TAU-bar and ts-7 after X-irradiation. Cells growing exponentially at 25 C were irradiated for the doses indicated, and viable cell counts determined. The open symbols represent plates incubated at 25 C; the closed symbols, plates incubated for 2 hr at 40 C immediately after plating and then transferred to 25 C. The circles represent TAU-bar; the triangles, ts-7.

Vol. 102, 1970

ts-7, and the slope of the curve is steeper. With UV irradiation, TAU-bar shows a slight temperature effect; survival of this strain when it is incubated for 2 hr at 40 C before incubation at 25 C is greater than survival when incubated only at 25 C (9, 10). This temperature effect is not expressed with X-irradiation. With UV irradiation, survival of ts-7 is markedly reduced when incubated for 2 hr at 40 C followed by incubation at 25 C, compared to survival when incubated only at 25 C (9, 10). In contrast to this result, ts-7 does not appear to be temperature-sensitive with respect to X-ray sensitivity. This behavior may be due to the observation that in vitro DNA ligase activity of ts-7 is not temperature-sensitive but is reduced at either temperature in comparison to TAU-bar activity (9).

We have shown that ts-7 is X-ray sensitive in comparison to the parent strain TAU-bar. Three independently isolated revertants of ts-7 are identical to TAU-bar with respect to temperature sensitivity, UV sensitivity, kinetics of accumulation of single-stranded fragments of newly replicated DNA (C. Pauling and L. Hamm, Proc. Nat. Acad. Sci. U.S.A., 64: 1195-1202), and X-ray sensitivity; we conclude that the phenotype of ts-7 is the consequence of a single revertible mutation. These results suggest that a common DNA ligase functions in the repair of X-rayinduced, single-strand scissions and the dark repair of UV radiation-induced lesions, and confirm our prediction that a DNA ligase-deficient mutant would be X-ray sensitive.

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LITERATURE CITED

- Boyce, R. P., and M. Tepper. 1968. X-ray induced single strand breaks and joining of broken strands in superinfecting lambda DNA in *E. coll* lysogenic for lambda. Virology 34:344-351.
- Gellert, M. 1967. Formation of covalent circles of lambda DNA by *E. coli* extracts. Proc. Nat. Acad. Sci. U.S.A. 57: 148-155.
- Ginoza, W. 1967. The effects of ionizing radiation on nucleic acids of bacteriophages and bacterial cells. Annu. Rev. Microbiol. 21:325-362.
- Hanawalt, P. C. 1963. Involvement of synthesis of RNA in thymineless death. Nature 198:286.
- Kaplan, H. S. 1966. DNA strand scission and loss of viability after X-irradiation of normal and sensitized bacterial cells. Proc. Nat. Acad. Sci. U.S.A. 55:1442-1446.
- Maaløe, O., and P. C. Hanawalt. 1961. Thymine deficiency and the normal DNA replication cycle. J. Mol. Biol. 3:144– 155.
- McGrath, R. A., and R. W. Williams. 1966. Reconstruction in vivo of irradiated Escherichia coli deoxyribonucleic acid; the rejoining of broken pieces. Nature 212:534-535.
- Olivera, B. M., and I. R. Lehman. 1967. Linkage of polynucleotides through phosphodiester bonds by an enzyme from *Escherichia coli*. Proc. Nat. Acad. Sci. U.S.A. 57:1426–1433.
- Pauling, C. 1968. The role of polynucleotide ligase in normal DNA replication. Cold Spring Harbor Symp. Quant. Biol. 33:148-150.
- Pauling, C., and L. Hamm. 1968. Properties of a temperature sensitive radiation-sensitive mutant of *Escherichia coli*. Proc. Nat. Acad. Sci. U.S.A. 60:1495-1502.