## Nucleotide Sequence of VP4 and VP7 Genes of Human Rotaviruses with Subgroup I Specificity and Long RNA Pattern: Implication for New G Serotype Specificity

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We sequenced the genes coding for the two neutralization proteins, VP4 and VP7, of human rotavirus strains L26 and L27 with subgroup I specificity but the long RNA pattern. The deduced VP7 amino acid sequence of strains L26 and L27 showed a low homology (73.6 to 81.9%) to those of rotavirus strains of the established serotypes. This finding, together with the previous serological characterizations, suggests that the VP7 (G) serotype of the L26 and L27 strains is distinct from those of strains of the previously established serotypes. In contrast, the VP4 sequences of the L26 and L27 strains were quite similar to those of virulent serotype 2 strains (DS-1, S2, and RV-5).

Group A rotaviruses have two neutralization proteins, VP4 and VP7, in the outer layer of the double-shelled particle (15, 27, 31). By cross-neutralization tests with hyperimmune sera, 11 distinct serotypes have been identified in group A rotaviruses so far (2, 5, 16, 24, 28, 35, 36). The serotype specificity of rotaviruses is ascribed mainly to the antigenic specificity of glycoprotein VP7 (5, 20). According to the recent proposal for the binary terminology of rotavirus serotypes, the antigenic specificity of VP7 of the 11 established serotypes can be designated G1 to G11 by using the prefix G for glycoprotein. VP4, a target protein for trypsin activation of infectivity, also has an independent antigenic specificity (15, 27). The proposed designation for the type specificity carried by VP4 is to use the prefix P for protease (P1, P2, and so on).

Recent comparative studies on the nucleotide sequences of VP4 genes from representative human rotavirus strains suggested the presence of at least four P types of VP4 (10, 32). In this article we tentatively designate them as follows: P1, found in virulent strains with G1, G3, or G4 specificity; P2, in virulent strains with G2 specificity; P3, in asymptomatic strains; and P4, in virulent strain K8 with G1 specificity.

Five additional types of VP4 in bovine, simian, and porcine rotaviruses have been suggested by sequence analyses (19, 23, 25, 26). However, systematic antigenic typing of VP4 from human and animal rotavirus strains has not yet been performed. Furthermore, it has been found that several cross-reactive neutralization epitopes exist at least in P1 and P2 VP4s and in P1, P2, and P3 VP4s (31), suggesting ambiguity in the serotyping of VP4 compared with that of VP7.

We recently characterized 20 unusual human rotavirus strains derived from Philippine infants with diarrhea which had subgroup I specificity and the long RNA pattern typical of subgroup II rotaviruses (21). By cross-neutralization tests with the four Philippine strains isolated in cell culture and other established strains, these unusual strains were found In this study, we sequenced the genes encoding the two neutralizing proteins VP4 and VP7 of the Philippine strains L26 and L27. A comparison of their sequence with those of established serotypes supported the idea that strains L26 and L27 have a new G serotype on VP7, while their P type on VP4 is similar to that of P2 found in strains with the G2 serotype.

Strains L26 and L27 were grown in MA-104 cells in the presence of trypsin (1  $\mu$ g/ml), and single-shelled particles were purified by differential centrifugation, fluorocarbon treatment, and CsCl gradient centrifugation after EDTA treatment. Viral mRNA was synthesized in vitro by the endogenous transcriptase present in single-shelled particles. Synthetic oligonucleotide primers were used to sequence mRNA by reverse transcriptase in the presence of dideoxynucleotides as described previously (11). The terminal 100 nucleotides of the RNA were determined by using denatured double-stranded virion RNA.

Figure 1 shows the complete nucleotide sequence and the deduced amino acid sequence of the VP7 gene of strain L26. The fundamental structure of the VP7 gene from strain L26 was similar to those of other rotavirus strains (5, 20). The entire VP7 gene of the L26 strain consisted of 1,062 nucleotides, with two potential open reading frames beginning at an ATG at positions 49 to 51 or 136 to 138 and ending at a TAG at positions 1027 to 1029. The open reading frames had the capacity to code for a VP7 of 297 or 326 amino acids. Two potential glycosylation sites were found at amino acid positions 69 to 71 and 238 to 240. Eight cysteine residues were conserved, as has been found for all strains examined to date. The VP7 sequence of strain L27 was identical to that of strain L26 VP7.

Except for serotype 7 strains, sequence data for which are not yet available, the nucleotide and amino acid sequences of strains representing the established serotypes, including non-serotype 6 bovine strain 61A (possible serotype 10)

not to be antigenically related to any other established serotypes except for serotypes 7, 10, and 11, antisera for which were not available (S. Urasawa, T. Urasawa, F. Wakasugi, N. Kobayashi, K. Taniguchi, I. C. Lintag, M. C. Saniel, and H. Goto, Arch. Virol., in press).

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GGC TTT AAA AGA GAG AAT TTC CGT TTG GCT AGC GGT TAG CTC CTT TTA ATG TAT GGT ATT 60 м Y G 4 Т 120 GAA TAT ACC ACA ATT CTA ACC ATT TTG ATA TCA ATC GTT CTA CTA AAT TAT ATA TTA AAA E Y т Т Ι L т I L I S Ι v L L N Y I L ĸ 24 TCG ATA ACT AGT ATG ATG GAC TTT ATT ATA TAT AGA TTC TTA CTA GTT TTT GTT ATC GTA 180 т S М M D F F L 44 Ι Ι Y R F L L 240 CTG CCA TTT ATT ANA GCT CAN ANC TAT GGA ATA ANT CTT CCA ATC ACA GGT TCT ATG GAT L Ρ F I K A Q N Y G I N L Р Ι т G S м D 64 300 ACT GCA TAT GTA AAC TCT ACG CAA CAA GAA AGT TTT ATG ACT TCC ACT TTA TGC TTG TAT Y v N N S Т 0 0 Е S F Y S т L С L 84 A TAT CCG AAT TCA GTT ACG ACT GAA ATA ACT GAC CCC GAT TGG ACG CAT ACA CTA TCA CAA 360 Р S Ý Е 104 Y N Т Т Ι Т D Р D W т н т L S 0 CTA TTT CTG ACT AAA GGA TGG CCA ACA AAT TCT GTT TAC TTC AAG AGT TAT GCT GAC ATA 420 L F L т K G W Ρ т N S v Y F K S Y A D I 124 GCG TCC TTC TCT GTA AAT CCA CAG TTA TAC TGT GAT TAC AAT ATC GTG TTA GTA CAA TAT 480 S A s F N P 0 L Y C D v 144 Y N Ι L 0 Y CAA AAT TCA TTA GCG TTA GAT GTT TCG GAA CTC GCT GAT TTA ATT TTA AAT GAA TGG TTA 540 0 N S L A L D v S E L D W 164 A L Ι L N Е г TGT AAT CCG ATG GAC GTA ACG TTA TAT TAT TAT CAA CAG ACT GAC GAA GCC AAT AAA TGG 600 С N Р М D v т L Y Y Y 0 ο т D Е A N K 184 ATA TCA ATG GGA GAT TCA TGT ACA GTT AAA GTA TGT CCT TTA AAT ATG CAA ACG TTA GGA 660 S M G D S С v 204 Ι т v K С Р L N М 0 т L G ATT GGA TGT ACA ACA ACC GAC GTC GCA ACA TTT GAA GAA GTA GCA AAC CCG GAA AAG TTA 720 I G С т т т D v A т F Е Е v A N A Е K Г 224 GTA ATT ACT GAT GTT GTA GAC GGA GTC AAT CAT AAG ATC AAT ATT ACA TTG AAT ACA TGC 780 v Ι т D v v D G v N H K т 244 Ι N Ι Т L N С ACT ATA CAA AAT TGT AAA AAA TTG GGA CCT AGA GAA AAC GTA GCA ATT ATA CAA GTA GGT 840 т I Q N С K K L G Р R Е N V A Ι I v 264 0 G GGT TCT GAC ATC ATA GAT ATA ACA GCA GAT CCA ACA ACA ATT CCA CAA ACT GAA AGA ATA 900 S D G Ι Ι D I Т A D P т т P Е R 284 I 0 Т I ATG CGA ATA AAT TGG AAA AAA TGG TGG CAA GTG TTT TAT ACC GTA GTA GAT TAC ATA AAT 960 M R N W Ι K K W W ο v F Y т v D 304 v Y Ι N CAA ATA GTT CAG GTA ATG TCT AAA CGA TCT AGA TCA CTA AAT TCA GCT GCA TTT TAT TAC 1020 Ι v v M S Q Q ĸ R S R S L N S Α A F Y Y 324 AGA ATT TAG ATA TAG CTT AGG TTA GAG TTG GTC GAT GTG ACC 1062 326

FIG. 1. Complete nucleotide sequence and deduced amino acid sequence of the VP7 gene of strain L26. Two potential glycosylation sites are shown in boxes.

recently isolated in Thailand (32a), were compared with those of strain L26. The nucleotide and amino acid sequences determined were only 72.9 to 77.4% and 73.6 to 81.9% homologous, respectively (Table 1). These low homology values contrast strongly with the 91 to 100% homology found among the strains of a given serotype (14).

VP7 has six serotype-specific regions, designated A to F (amino acids 39 to 50, 87 to 101, 120 to 130, 143 to 152, 208 to 221, and 233 to 242, respectively) (14). Amino acids in these regions are well conserved among strains of the same serotype but differ considerably among strains belonging to different serotypes. Three regions in particular, B, D, and E, are considered the major antigenic sites, since mutants resistant to anti-VP7 serotype-specific neutralizing monoclonal antibodies had amino acid changes only in these three regions (3, 5, 22, 29). A comparison of amino acid sequences in these regions between strain L26 and strains with serotype

1, 2, 3, 4, 5, 6, 8, 9, 11, or possibly 10 specificity revealed a great difference (Fig. 2). The amino acid sequence homologies in the three regions were only 34 to 51%. In our previous studies (21; Urasawa et al., in press), neutralizing monoclonal antibodies specific for serotype 1, 2, 3, or 4 could not recognize strains L26 and L27 either in an enzyme-linked immunoassay or a neutralization test. Furthermore, two-way cross-neutralization tests with hyperimmune sera showed no serological relationship of the two strains to other serotypes except for serotypes 7, 10, and 11, for which antisera were not available. Thus, the present sequence data strongly support the notion that these strains represent a new G (VP7) serotype.

The complete nucleotide sequences of VP4 genes from strains L26 and L27 were also determined. The VP4 nucleotide sequence of the two strains was 2,359 bases long and contained a single long open reading frame beginning with

Strain	G Serotype	B region	D region	E region
		87 101	143 152	208 221
L26	12	NSVTTEITDPDWTHT	QYQNSLALDV	TTTDVATFEEVANA
KU	1	TEAS-Q-A-GKD-	КQВМ	QN-DSMI-EN
Wa	1	TEAS-Q-A-GKD-	КQЕМ	QN-DSMI-EN
S2	2	AEAKNS-DE-EN-	R-D-TSB	KIDISS
DS-1	2	ABAKNS-DE-EN-	R-D-TSEA	KNISS
Р	3	TEAAN-NS-KD-	K-DAT-QM	LTNT-
SA11	3	TBAAN-NS-KD-	K-DAT-QM	L
ST-3	4	SEAP-Q-S-TE-KD-	RFVSGEEI	QNTTDS
VA70	4	SEAP-Q-S-NE-KD-	KFASGEEI	QNMDS
OSU	5	-ЕЛЛЛ-ТК-КЕ-	K-DGN-QM	SINST
NCDV	6	VEASNA-TE-KD-	K-DSTQEM	LI-NPDT-TM
UK	6	VEASNA-TE-KD-	K-DSTQEM	LI-NPDTTT
69M	8	VBABA-SS-KDH	K-NANSBM	LTTT-
B37	8	VEABA-SS-KDH	K-NANSEM	LTTT-
WI61	9	ABAS-Q-G-TE-KD-	K-DST-EM	NTAS
F45	9	AEAS-Q-G-TE-KD-	K-DST-EM	NTAS
61 <b>a</b>	10?	TBARN-NB	R-NSEM	QNTRT-
YM	11	HEAA-Q-A-DK-KD-	K-DGNSQM	LPTS-

FIG. 2. Comparison of the VP7 amino acid sequence in three antigenic regions (B, D, and E) of strain L26 with those of human rotavirus strains with different G serotypes. The entire VP7 amino acid sequences of strains other than L26 have been reported previously (4, 5, 8, 9, 12–14, 17, 28, 32a). NCDV, Nebraska calf diarrhea virus.

ATG at positions 10 to 12 and terminating with TAG at positions 2335 to 2337. The entire deduced amino acid sequences of strains L26 and L27 are shown in Fig. 3. Four nucleotide sequences, three of which caused amino acid changes, were different between the two strains: AGA (codon 51), AAC (codon 324), GAA (codon 392), and TTT (codon 405) in strain L26 were GGA, AAT, AAA, and TGT, respectively, in strain L27.

The nucleotide and amino acid sequences of L26 and L27 were compared with those of human rotavirus strains recovered from symptomatic and asymptomatic patients. The VP4 nucleotide sequence of both strains showed higher homology (95%) to those of strains (DS-1 and RV-5) with P2 specificity than to those of strains with P1, P3, and P4 specificities (87, 74, and 67%, respectively).

Figure 4 shows the comparison of the amino acid sequences in a selected region (amino acids 361 to 430) where P2-specific amino acid sequences are found. In our previous study (30), we showed that amino acid residue 392 is crucial to neutralization epitope specific to P1 and P2 by using KU

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TABLE 1. Nucleotide and amino acid sequence homology of
strain L26 VP7 with VP7 from other rotavirus strains with
different G serotype specificities

	% Homology with VP7 of strain L26		
Strain (G serotype)	Nucleotide sequence	Amino acid sequence	
KU (1)	74.7	76.9	
DS-1 (2)	73.3	76.1	
SA11 (3)	77.4	81.3	
Rhesus rotavirus (3)	76.1	81.9	
VA70 (4)	73.1	74.2	
Gottfried (4)	72.9	73.6	
OSU (5)	75.5	80.7	
Nebraska calf diarrhea virus (6)	76.0	78.2	
69M (8)	74.2	79.2	
WI61 (9)	76.3	80.9	
61A (10?)	73.8	77.5	
YM (11)	74.3	80.8	

(P1) and DS-1 (P2) antigenic mutants resistant to each of the anti-VP4 neutralizing monoclonal antibodies specific to P1 or P2. L27 VP4 had a different amino acid from L26 at position 392, Glu in L26 and Lys in L27. Interestingly, the DS-1 mutant resistant to the P2-specific neutralizing monoclonal antibody (S2-2F2) had an amino acid substitution at this residue (Lys-392 to Glu). Indeed, S2-2F2 antibody, which was reactive with strain L27, did not recognize strain L26 in a neutralization test (21). Strain L26 may be a naturally occurring variant which acquired resistance to the antibody directed to the epitope involving amino acid residue 392 during an infection process in an individual who had preexisting antibodies directed to that epitope.

In general, there was a cosegregation between VP4 and VP7 genes of human rotaviruses detected in nature; strains with G1, G3, or G4 have P1 specificity, while strains with G2 have P2 specificity (10, 30, 31). An exception is found in strains recovered from asymptomatic infections in neonates, which have P3 specificity and either G1, G2, G3, or G4 specificity (6, 10, 11). In our recent study (32), we reported another exception, strain K8 with G1 and unique VP4 (P4). However, strains like L26 and L27 having the P2 type on VP4 and a G serotype other than G2 on VP7 have not been described. The L26 and L27 strains have subgroup I specificity on VP6, which is generally found in serotype 2 strains

MASLIYRQLLTNSYSVDLHDEIEQIGSEKTQNVTINPGPFAQTRYAPVNWRHGEINDSTTVEPVLDGPYQPTTFKPPNDY G	80
WLLISSNTDGVVYESTNNSDFWTAVIAVEPRVSQTNRQYILFGENKQFNIENNSDKWKFFEMFKGSSQSNFSNRRTLTSN	160
NRLVGMLKYGGRVWTFHGETPRATTDSSNTADLNNISIVIHSEFYIIPRSQESKCNEYINNGLPPIQNTRNVVPLSLSSR	240
SIQYRRAQVNEDITISKTSLWKEMQYNRDIIIRFKFGNSVIKLGGLGYKWSEISYKAANYQYSYSRDGEQVTAHTTCSVN	320
G VNN F SYNGGSLPTDFSISRYEVIKEN SYVYIDYWDDSKAFRNM VYVRSLAANLN SVKCAGGSYN FRLPVGEWPIMNGGA K	400
VSLHFAGVTLSTQFTNFVSLNSLRFRFSLTVDEPSFSIIRTRTVNLYGLPAANPNNGNEYYEMSGRFSLISLVPTNDDYQ C	480
TPIMNSVTVRQDLERQLSDLREEFNSLSQEIAMSQLIDLALLPLDMFSMFSGIKSTIDLTKSMATSVMKKFRKSKLATSI	560
SEMTNSLSDAASSASRSASVRSNLSVISNWTDASKSTSNITDLVNDVSTQTSTISKKLRLKEMITQTEGMSFDDISAAVL	640
KTKIDMSTQIGKNTLPDIVTEASEKFIPKRSYRVLKDNEVMEINTEGKFFAYKVDTLNEIPFDINKFAELVTDSPVISAI	720
IDFKTLKNLNDNYGITRMEALNLIKSNPNVLRNFINQNNPIIRNRIEQLILQCKL 775	

FIG. 3. Complete amino acid sequences of VP4 from strains L26 and L27. Only the amino acids of strain L27 which are different from those of strain L26 are shown below the L26 amino acid sequence.

Strain	P type			
		361 (362)	392 (393)	430(431)
KU	P1	FRNMVYVRSLAANLNSV	KCTGGSYDFSIPVGAWPVMNGGAVSLHFA	GVTQFTDFVSL
P	P1			
RV-5	P2		RLGI	
DS-1	P2		N-RLKI	
V-S2-2F2	2 (DS-1) 1	2	N-RLEI	
L26	(P2)		N-RLEI	
L27	(P2)		N-RLKI	
M37	P3		SN-N-QLSS	
1076	Р3		SN-N-QMS	
K8	P4			LS-QYTDY

FIG. 4. Comparison of amino acid sequences of a selected region (amino acids 361 to 430) on VP4 of strains L26 and L27 with those of VP4 from human rotaviruses of different P types. The numbers above the sequence refer to amino acid positions. The numbers in parentheses show the amino acid positions in strain K8, which has an insertion of one amino acid after residue 135 (32). The entire VP4 amino acid sequences of strains other than L26 and L27 have been reported previously (10, 18, 23, 29, 32).

having P2 on VP4 and G2 on VP7. Therefore, these strains, which still reserve the cosegregation between P2 specificity and subgroup I specificity, might be naturally occurring reassortants between a serotype 2 human rotavirus and a strain with novel serotype specificity (G12) on VP7. Despite the frequent occurrence of human rotavirus reassortment in vitro (7, 34), the appearance of such reassortants in nature seems to be rare. There may be some constraints, which have not been elucidated, on the occurrence of human rotavirus reassortants in nature.

Although the L26 and L27 strains have P2 specificity on their VP4, they showed little relationship with reference strains having VP4 of P2 specificity in cross-neutralization tests (Urasawa et al., in press), indicating low immunogenicity of VP4 of these strains. This finding is in accord with the general evidence that antibody reactivity in hyperimmune sera is directed largely against VP7 in most wild strains. In contrast, the reassortants prepared in vitro by coinfection with two different rotavirus strains exhibited high immunogenicity of VP4 (16, 27, 34). The surface configuration formed by different combinations of VP4 and VP7 might affect their relative contribution to immunogenicity, as suggested from reassortment studies in which the recipient genetic background affected phenotypic properties such as plaque size (1). Characterization of more naturally occurring unusual strains, most of which may be reassortants, would facilitate understanding of this issue. The prevalence in human and animals of strains with the G12 serotype, like the L26 and L27 strains, is being examined by the use of G12-specific neutralizing monoclonal antibodies and by **RNA-RNA** hybridization.

Nucleotide sequence accession numbers. The L26 VP7 and L27 VP4 gene sequences have been given GenBank accession numbers M36396 and M36397, respectively.

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## LITERATURE CITED

- Chen, D., J. W. Burns, M. K. Estes, and R. F. Ramig. 1989. The phenotypes of rotavirus reassortants depend upon the recipient genetic background. Proc. Natl. Acad. Sci. USA 86:3743–3747.
- Clark, H. F., Y. Hoshino, L. M. Bell, J. Groff, G. Hess, P. Bachman, and P. A. Offit. 1987. Rotavirus isolate WI61 representing a presumptive new human serotype. J. Clin. Microbiol. 25:1757-1762.
- 3. Dyall-Smith, M. L., I. Lazdins, G. W. Tregear, and I. H. Holmes. 1986. Location of the major antigenic sites involved in rotavirus serotype-specific neutralization. Proc. Natl. Acad.

Sci. USA 83:3465-3468.

- 4. Elleman, T. C., P. A. Hoyne, M. L. Dyall-Smith, I. H. Holmes, and A. A. Azad. 1983. Nucleotide sequence of the gene encoding the serotype-specific glycoprotein of UK bovine rotavirus. Nucleic Acids Res. 11:4689–4701.
- 5. Estes, M., and J. Cohen. 1989. Rotavirus gene structure and function. Microbiol. Rev. 53:410-449.
- Flores, J., K. Midthun, Y. Hoshino, K. Green, M. Gorziglia, A. Z. Kapikian, and R. M. Chanock. 1986. Conservation of the fourth gene among rotaviruses recovered from asymptomatic newborn infants and its possible role in attenuation. J. Virol. 60:972-979.
- Garbarg-Chenon, A., F. Bricout, and J. C. Nicolas. 1986. Serological characterization of human reassortant rotaviruses. J. Virol. 59:510–513.
- Glass, R. I., J. Keith, O. Nakagomi, T. Nakagomi, J. Askaa, A. Z. Kapikian, R. M. Chanock, and J. Flores. 1985. Nucleotide sequence of the structural glycoprotein VP7 gene of Nebraska calf diarrhea virus rotavirus: comparison with homologous genes from four strains of human and animal rotaviruses. Virology 141:292–298.
- Gorziglia, M., Y. Aguirre, Y. Hoshino, J. Esparza, I. Blumentals, J. Askaa, M. Thompson, R. I. Glass, A. Z. Kapikian, and R. M. Chanock. 1986. VP7 serotype-specific glycoprotein of OSU porcine rotavirus: coding assignment and gene sequence. J. Gen. Virol. 67:2445-2454.
- Gorziglia, M., K. Green, K. Nishikawa, K. Taniguchi, R. Jones, A. Z. Kapikian, and R. M. Chanock. 1988. Sequence of the fourth gene of human rotaviruses recovered from asymptomatic or symptomatic infections. J. Virol. 62:2978–2984.
- Gorziglia, M., Y. Hoshino, A. Buckler-White, I. Blumentals, R. Glass, J. Flores, A. Z. Kapikian, and R. M. Chanock. 1986. Conservation of amino acid sequence of VP8 and cleavage region of 84-kDa outer capsid protein among rotaviruses recovered from asymptomatic neonatal infection. Proc. Natl. Acad. Sci. USA 83:7039-7043.
- Gorziglia, M., K. Nishikawa, K. Green, and K. Taniguchi. 1988. Gene sequence of the VP7 serotype specific glycoprotein of Gottfried porcine rotavirus. Nucleic Acids Res. 16:775.
- Green, K. Y., Y. Hoshino, and N. Ikegami. 1989. Sequence analysis of the gene encoding the serotype-specific glycoprotein (VP7) of two new human rotavirus serotypes. Virology 168:429– 433.
- Green, K. Y., K. Midthun, M. Gorziglia, Y. Hoshino, A. Z. Kapikian, R. M. Chanock, and J. Flores. 1987. Comparison of the amino acid sequences of the major neutralization protein of four human rotavirus serotypes. Virology 168:429–433.
- Hoshino, Y., M. M. Sereno, K. Midthun, J. Flores, A. Z. Kapikian, and R. M. Chanock. 1985. Independent segregation of two antigenic specificities (VP3 and VP7) involved in neutralization of rotavirus infectivity. Proc. Natl. Acad. Sci. USA 82:8701-8704.
- Hoshino, Y., R. G. Wyatt, H. B. Greenberg, J. Flores, and A. Z. Kapikian. 1984. Serotypic similarity and diversity of rotaviruses of mammalian and avian origin as studied by plaque-reduction

neutralization. J. Infect. Dis. 149:694-702.

- 17. Hum, C. P., M. L. Dyall-Smith, and I. H. Holmes. 1989. The VP7 gene of a new G serotype of human rotavirus (B37) is similar to G3 proteins in the antigenic C region. Virology 170:55-61.
- Kantharidis, P., M. L. Dyall-Smith, and I. H. Holmes. 1987. Marked sequence variation between segment 4 genes of human RV-5 and simian SA11 rotaviruses. Arch. Virol. 93:111–121.
- Kantharidis, P., M. L. Dyall-Smith, G. W. Tregear, and I. H. Holmes. 1988. Nucleotide sequence of UK bovine rotavirus segment 4: possible host restriction of VP3 genes. Virology 166:308-315.
- Kapikian, A. Z., and R. M. Chanock. 1990. Rotaviruses, p. 1353–1404. *In* B. N. Fields, D. N. Knipe, R. M. Chanock, J. L. Melnick, B. Roizman, and R. E. Shope (ed.), Virology. Raven Press, New York.
- Kobayashi, N., I. C. Lintag, T. Urasawa, K. Taniguchi, M. C. Saniel, and S. Urasawa. 1989. Unusual human rotavirus strains having subgroup I specificity and "long" RNA electropherotype. Arch. Virol. 109:11–23.
- Mackow, E. R., R. D. Shaw, S. M. Matsui, P. T. Vo, D. A. Benfield, and H. B. Greenberg. 1988. Characterization of homotypic and heterotypic VP7 neutralization sites of rhesus rotavirus. Virology 165:511-517.
- 23. Mackow, E. R., R. D. Shaw, S. M. Matsui, P. T. Vo, M. N. Dang, and H. B. Greenberg. 1988. Characterization of the rhesus rotavirus VP3 gene: location of amino acids involved in homologous and heterologous rotavirus neutralization and identification of a putative fusion region. Proc. Natl. Acad. Sci. USA 85:645-649.
- Matsuno, S., A. Hasegawa, A. Mukoyama, and S. Inouye. 1985. A candidate for a new serotype of human rotavirus. J. Virol. 54:623-624.
- 25. Nishikawa, K., and M. Gorziglia. 1988. The nucleotide sequence of the VP3 gene of porcine rotavirus OSU. Nucleic Acids Res. 16:847.
- Nishikawa, K., K. Taniguchi, A. Torres, Y. Hoshino, K. Green, A. Z. Kapikian, R. M. Chanock, and M. Gorziglia. 1988. Comparative analysis of the VP3 gene of divergent strains of rotaviruses simian SA11 and bovine Nebraska calf diarrhea virus. J. Virol. 62:4022–4026.
- Offit, P. A., and G. Blavat. 1986. Identification of the two rotavirus genes determining neutralization specificities. J. Virol. 57:46-49.
- 28. Ruitz, A. M., I. V. Lopez, S. Lopez, R. T. Espejo, and C. F.

Arias. 1988. Molecular and antigenic characterization of porcine rotavirus YM, a possible new rotavirus serotype. J. Virol.

- 62:4331-4336.
  29. Taniguchi, K., Y. Hoshino, K. Nishikawa, K. Y. Green, W. L. Maloy, Y. Morita, S. Urasawa, A. Z. Kapikian, R. M. Chanock, and M. Gorziglia. 1988. Cross-reactive and serotype-specific neutralization epitopes on VP7 of human rotavirus: nucleotide sequence analysis of antigenic mutants selected with monoclonal antibodies. J. Virol. 62:1870-1874.
- Taniguchi, K., W. L. Maloy, K. Nishikawa, K. Y. Green, Y. Hoshino, S. Urasawa, A. Z. Kapikian, R. M. Chanock, and M. Gorziglia. 1988. Identification of cross-reactive and serotype 2-specific neutralization epitopes on VP3 of human rotavirus. J. Virol. 62:2421-2426.
- Taniguchi, K., Y. Morita, T. Urasawa, and S. Urasawa. 1987. Cross-reactive neutralization epitopes on VP3 of human rotavirus: analysis with monoclonal antibodies and antigenic variants. J. Virol. 61:1726–1730.
- 32. Taniguchi, K., K. Nishikawa, T. Urasawa, S. Urasawa, K. Midthun, A. Z. Kapikian, and M. Gorziglia. 1989. Complete nucleotide sequence of the gene encoding VP4 of a human rotavirus (strain K8) which has unique VP4 neutralization epitopes. J. Virol. 63:4101–4106.
- 32a. Taniguchi, K., Y. Pongsuwanna, M. Choonthanom, and S. Urasawa. 1990. Nucleotide sequence of the VP7 gene of a bovine rotavirus (strain 61A) with different serotype specificity from serotype 6. Nucleic Acids. Res. 18:4613.
- Taniguchi, K., S. Urasawa, and T. Urasawa. 1985. Preparation and characterization of neutralizing monoclonal antibodies with different reactivity patterns to human rotaviruses. J. Gen. Virol. 66:1045-1053.
- Urasawa, S., T. Urasawa, and K. Taniguchi. 1986. Genetic reassortment between two human rotaviruses having different serotype and subgroup specificities. J. Gen. Virol. 67:1551– 1559.
- 35. Urasawa, S., T. Urasawa, K. Taniguchi, and S. Chiba. 1984. Serotype determination of human rotavirus isolates and antibody prevalence in pediatric population in Hokkaido, Japan. Arch. Virol. 81:1–12.
- Woode, G. N., S. Zheng, B. I. Rosen, N. Knight, N. E. K. Gourley, and R. F. Ramig. 1987. Protection between different serotypes of bovine rotavirus in gnotobiotic calves: specificity of serum antibody and coproantibody responses. J. Clin. Microbiol. 25:1052–1058.