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Helium-3 Diffusion MR Imaging of the Human Lung over Multiple Time Scales

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Abstract

Rationale and Objectives—Diffusion MRI with hyperpolarized ³He gas is a powerful technique for probing the characteristics of the lung microstructure. A key parameter for this technique is the diffusion time, which is the period during which the atoms are allowed to diffuse within the lung for measurement of the signal attenuation. The relationship between diffusion time and the length scales that can be explored is discussed, and representative, preliminary results are presented from ongoing studies of the human lung for diffusion times ranging from milliseconds to several seconds.

Materials and Methods—³He diffusion MR imaging of the human lung was performed on a 1.5T Siemens Sonata scanner. Using gradient-echo-based and stimulated-echo-based techniques for short and medium-to-long diffusion times, respectively, measurements were performed for times ranging from 2 ms to 6.5 s in two healthy subjects, a subject with sub-clinical chronic obstructive pulmonary disease and a subject with bronchopulmonary dysplasia.

Results—In healthy subjects, the apparent diffusion coefficient decreased by about 10-fold, from approximately 0.2 to 0.02 cm²/s, as the diffusion time increased from approximately 1 ms to 1 s. Results in subjects with disease suggest that measurements made at diffusion times substantially longer than 1 ms may provide improved sensitivity for detecting certain pathological changes in the lung microstructure.

Conclusion—With appropriately designed pulse sequences it is possible to explore the diffusion of hyperpolarized 3 He in the human lung over more than a 1000-fold variation of the diffusion time. Such measurements provide a new opportunity for exploring and characterizing the microstructure of the healthy and diseased lung.

Keywords

MRI of lung; diffusion MRI; hyperpolarized helium

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INTRODUCTION

The lung parenchyma and airspaces are poorly visualized with conventional, proton-based magnetic resonance imaging (MRI) due to the low concentration of water and the inhomogeneous magnetic environment, both of which diminish the strength of the MR signal compared to that from other organs in the body. In contrast, inhalation of *hyperpolarized* helium-3 (3 He) gas provides a strong signal from the lung airspaces that permits high-spatialresolution MR imaging (1,2). Prior to performing an MRI examination with 3 He, the gas is polarized outside of the scanner using a dedicated, laser-based device within which, over a period of minutes to hours (depending on the quantity of gas and details of system design), the nuclear polarization of the 3 He atoms builds up to tens of percent. This polarization is several orders of magnitude larger than that achieved for protons when the human body is placed in an MRI scanner, and offsets the low density of the gas so that high-quality MR images of 3 He in the airspaces of the lung can be obtained.

A number of imaging strategies have been used for hyperpolarized 3 He MRI of the lung to investigate diseases such as asthma and emphysema (3–5). Of particular interest is the application of diffusion MRI techniques to probe the characteristics of the lung microstructure. These methods provide a measurement that reflects the random Brownian motion of the 3 He gas atoms within the airspaces of the lung (as opposed to the trans-membrane diffusion of the gas) and the degree to which this random motion is restricted by the structure of the lung tissue. Diffusion results in attenuation of the MR signal, from which an *apparent* diffusion coefficient (ADC) can be calculated. The measured ADC varies inversely with the degree to which the diffusion of the gas is restricted. For example, the ADC for 3 He gas in healthy lung parenchyma is substantially smaller than the corresponding diffusion coefficient in an unrestricted space, and the ADC measured in a lung with severe emphysema is larger than that for a healthy lung, reflecting the enlargement of airspaces caused by the tissue destruction that occurs with emphysema. Thus, regional changes of the microstructure that occur in pulmonary diseases such as emphysema can be characterized by measuring the ADC of 3 He gas in the lung (5,6). Further, appropriately designed measurement protocols permit representative structural dimensions to be determined (7,8), and recent evidence suggests that ADC measurements may permit detection of subtle, sub-clinical structural changes before they become apparent on highresolution computed tomography (9).

The ADC derived from a hyperpolarized ³He diffusion measurement depends on the details of the imaging procedure. One of the key parameters is the diffusion time, which is the period during which the atoms are allowed to diffuse within the lung for the measurement of signal attenuation. The distance that the atoms diffuse is determined by the diffusion time, increasing as the diffusion time is increased, and by the degree of restriction imposed by the lung microstructure. The vast majority of ADC measurements have been performed by using a diffusion time of a few milliseconds, which corresponds to a diffusion distance of a few hundred microns in healthy lung. (The diameter of an alveolus in a healthy adult lung is approximately 250 µm.) Although preliminary results from this technique appear very promising, such *shorttime-scale* measurements interrogate a region the size of only a few healthy alveoli, and therefore provide little information about the connectivity among the airspaces. Considering the underlying complex, interconnected structure of the lung, it is reasonable to expect that the ADC will remain dependent on the diffusion time for periods up to at least several seconds (10). Measurements with diffusion times of tens of milliseconds (*medium time scale*) to several seconds (*long time scale*) may thus offer improved sensitivity for the detection of pathological changes that affect airspace connectivity (11,12) and permit characterization of structural changes that are not accessible with short-time-scale measurements.

The purpose of this work is to discuss the relationship between the diffusion time and the length scales that can be explored within the lung, and to present preliminary, representative results from ongoing studies at our institution of ADC measurements in the human lung over time scales ranging from milliseconds to several seconds. To provide the reader who is unfamiliar with the details hyperpolarized-gas diffusion methods with the requisite background, we begin with a brief overview of the most common method for measuring the ADC of 3 He gas in the lung.

ADC MEASUREMENT AND DIFFUSION TIME

Several approaches have been evaluated for measuring the ADC of 3 He gas in the lung (5, 11,13). Of these, the most commonly-used approach involves applying a matched pair of magnetic-field gradient pulses to the transverse magnetization in a manner analogous to that described for proton diffusion MRI many years ago by Stejskal and Tanner (14). The basic framework for making an ADC measurement based on a gradient-echo pulse sequence is illustrated in Fig 1a. After transverse magnetization is generated by an excitation radiofrequency (RF) pulse, a gradient pulse is applied along a selected direction and imparts a position dependent *phase tag* to the transverse magnetization. Subsequently, the atoms are allowed to diffuse during a waiting period (the diffusion time) after which a second gradient pulse is applied to *unwind* the phase tag imparted by the first gradient pulse. In the absence of diffusion (and ignoring effects of relaxation), the signal measured following the second gradient pulse will be the same as that which would have been measured just after the excitation RF pulse. However, in the presence of diffusion, the positions of the atoms will change between the two gradient pulses. Thus, the second gradient pulse cannot completely unwind the phase tag imparted by the first gradient pulse and the signal measured following the second gradient pulse will be attenuated compared to that measured in the absence of diffusion.

The effect of the tagging gradient pulse on the transverse magnetization is illustrated in Fig 1b. During the gradient pulse, the precession frequency of the transverse magnetization varies linearly with distance. As a result, the gradient pulse twists the magnetization vectors into a helical pattern that is characterized by the tag wavelength as shown. The tag wavelength is inversely proportional to the strength and duration of the tagging gradient pulse.

Together, the "tag" and "un-tag" gradient pulses that are shown in Fig 1a form what is commonly referred to as a bipolar gradient. The degree to which these gradient pulses and the intervening time delay result in attenuation of the MR signal due to diffusion is characterized by the *b* value; a higher *b* value corresponds to greater attenuation due to diffusion. For the case when the ramp times of the gradient pulses can be neglected, the *b* value for a bipolar gradient is proportional to the diffusion time and the square of the area under the tagging gradient pulse.

For the gradient-echo pulse-sequence configuration illustrated in Fig 1a, the maximum achievable diffusion time is limited by $T2^*$ decay of the ³He transverse magnetization in the lung. At 1.5 Tesla, which is the field strength that has been used for the majority of $3H$ e imaging studies in humans, $T2^*$ is on the order of 20 ms (15). To overcome this limitation on the diffusion time, a stimulated-echo pulse-sequence configuration can be used (10,16). Recall that a stimulated echo is formed by three RF pulses, wherein the phase state of the transverse magnetization is stored along the longitudinal axis between the second and third RF pulses, and thus the signal decay between these pulses is governed by T1 relaxation. Since the T1 for ³He gas in the lung is approximately 20 s $(\sim 1,000)$ times larger than T2*), much longer diffusion times can be accessed by using a stimulated-echo-based measurement.

The basic framework for making an ADC measurement based on a stimulated-echo pulse sequence is outlined in Fig 2. Compared to the gradient-echo-based method shown in Fig 1a,

an RF pulse is applied following the "tag" gradient pulse to store the phase-tagged magnetization along the longitudinal axis. After the waiting period, the third RF pulse returns the phase-tagged magnetization to the transverse plane for measurement. Additional details on the implementation of this method for ADC measurements with hyperpolarized ³He gas can be found in reference (10).

As noted above, the diffusion time determines the distance that the atoms are allowed to probe during the measurement. Given the free diffusion coefficient (*D*) of the gas mixture of interest, the mean distance (x_{RMS}) that the atoms diffuse in an unrestricted environment can be calculated from the well-known relationship $x_{\text{RMS}} = \sqrt{6Dt}$, where *t* is the time that the atoms are allowed to diffuse within a three-dimensional environment. (Actually, we calculate the square root of mean squared distance, termed the RMS distance, since the mean distance is zero.) The corresponding diffusion coefficient for dilute 3 He in air is approximately 0.85 cm²/s.

Figure 3 shows the RMS diffusion distances for 3 He in air for diffusion times ranging from 0.1 ms to 10 s. For the diffusion time corresponding to typical short-time-scale ADC measurements (\sim 1 ms), the corresponding RMS distance for freely diffusing 3 He in air is equivalent to only about three alveolar diameters, whereas the length of an acinus is several millimeters. Within the healthy lung parenchyma, the motion of 3 He atoms is significantly restricted and thus the diffusion distances are less than those for the unrestricted case. For example, the ADC measured in the lungs of healthy human adults for a diffusion time of approximately 1 ms is roughly $0.2 \text{ cm}^2/\text{s}$ (5,6). Using this value in the equation above, the corresponding RMS distance is $350 \mu m$. Therefore, diffusion times much greater than a few milliseconds are required to permit the ³He atoms to explore the connectivity among the smallest generations of airways (e.g., acini) as well as any collateral ventilation pathways that may exist (17,18).

MATERIALS AND METHODS

³He diffusion MR imaging was performed using a 1.5-T commercial scanner (Magnetom Sonata, Siemens Medical Solutions, Malvern, PA) that included the multi-nuclear imaging package and a vest-shaped RF coil (Clinical MR Solutions, Brookfield, WI) tuned to the ³He resonant frequency of 48.5 MHz. All studies were performed under a physician's Investigational New Drug application (IND 57,866) for imaging with hyperpolarized 3 He using a protocol approved by our institutional review board. All subjects gave written informed consent prior to participation in the study. The subject's heart rate and oxygen saturation level were monitored (Omni-Track Vital Signs Monitoring System, model 3100; Invivo Research Inc., Orlando FL) throughout the imaging session. All studies were supervised by a physician.

Representative ADC results obtained over a range of diffusion times are presented for 4 subjects, including 2 healthy volunteers (1 male, 1 female; ages 55, 66 yrs), 1 subject with subclinical chronic obstructive pulmonary disease (COPD) (male, age 70 yrs, $FEV₁$ 106 %) predicted, GOLD Stage 0), and 1 subject with bronchopulmonary dysplasia (BPD) (female, age 6 yrs, $FEV₁$ 37 % predicted). (The subject with "sub-clinical" COPD has imaging findings that are indicative of early emphysema, but has normal spirometry and no clinical symptoms of lung disease.) The forced vital capacity (FVC) and forced expiratory volume in one second $(FEV₁)$ were measured in each subject on the day of imaging using a model PB100 spirometer (Puritan Bennett; Lenexa, KS) and the Knudson 1983 reference tables for predicted normal limits (19).

 3 He gas was polarized to a level of approximately 30% by collisional spin exchange with an optically pumped rubidium vapor using a commercial system (Model 9600 Helium Polarizer; Magnetic Imaging Technologies Inc., Durham, NC). The desired volume of

hyperpolarized 3He gas (approximately 50 ml for global measurements, 200 ml for projection images and 400 ml for multi-slice measurements) was dispensed into a Tedlar plastic bag (Jensen Inert Products, Coral Springs, FL) and diluted with N_2 to a total volume of approximately 1 liter, transported to the scanner room and inhaled by the subject. The total volume of gas prepared for each subject was directly proportional to the subject's FVC, such that an inhalation volume of 1 liter corresponded to an FVC of 3.5 liters.

Short-time-scale ADC measurements were performed by using a gradient-echo-based diffusion technique as outlined in Fig 1a and described in reference (5). Medium- and longtime-scale ADC measurements were performed by using a stimulated-echo-based diffusion technique as outlined in Fig 2. This pulse sequence also included (not shown in Fig 2) the measurement of calibration data just prior to the first RF pulse of the stimulated-echo preparation. The calibration data permitted the measured signal decay to be corrected for the effects of relaxation and the RF pulses, and thus the corresponding ADC could be calculated from a single application of the pulse sequence, which is critical for practical use in human subjects. Additional technical details of the stimulated-echo-based pulse sequences can be found in references (10) and (20). Pulse-sequence parameter values for each of the ADC measurements are provided in the respective figure captions.

RESULTS AND DISCUSSION

Global (i.e., not spatially encoded and therefore integrated over the whole lung) ADC values from the lung of a healthy human volunteer for diffusion times ranging from 20 ms to 6.5 s are shown in Fig 4. ADC values for diffusion times from 20 ms to 1.5 s were measured during a breath-hold using a tag wavelength of 5 mm (solid line in Fig 4), and ADC values for diffusion times from 200 ms to 6.5 s were measured during a second breath-hold using a tag wavelength of 10 mm (dashed line in Fig 4). Both measurements were performed using a stimulated-echobased pulse sequence. To obtain sufficient signal attenuation for an accurate measurement at short diffusion times, a relatively short tag wavelength of 5 mm was used. However, the signal attenuation with this tag wavelength for diffusion times on the order of 1 s was so large that accurate measurements could not be obtained for relatively long diffusion times. Therefore, a longer tag wavelength of 10 mm was used (during a separate breath-hold period) to obtain ADC values for diffusion times up to 6.5 s.

We observe that the ADC decreased monotonically and substantially as the diffusion time increased. For a tag wavelength of 5 mm, the ADC decreased by approximately 9-fold as the diffusion time increased from 20 ms to 1.5 s, and for a tag wavelength of 10 mm the ADC decreased by approximately 5-fold as the diffusion time increased from 200 ms to 6.5 s. The consistency of this general behavior – a smooth and substantial decrease in ADC with increasing diffusion time – has been verified in a group of ten healthy subjects by Wang et al (10).

Comparing the ADC versus diffusion time curves for the two tag wavelengths, we see that for a given diffusion time the ADC for the 10-mm tag wavelength is always greater than that for the 5-mm tag wavelength. A previous study (10) explored this behavior for tag wavelengths ranging from 6 to 15 mm and also found that for a given diffusion time the ADC increased as the tag wavelength increased. The *b* value is inversely proportional to the tag wavelength, and thus the observed behavior can be equivalently stated as: the ADC from a two-point measurement (i.e., reference signal and signal corresponding to a given *b* value) increases as the *b* value decreases. This same behavior has been observed for short-time-scale measurements by several investigators (7,21,22). The dependence of (two-point) ADC values on tag wavelength (or, equivalently, *b* value) highlights the fact that there is need to agree on standardized diffusion-sensitization schemes to permit quantitative comparison of ADC

measurements among research groups as illustrated, for example, by the data in reference (23). On the other hand, the dependence of the ADC on *b* value provides an opportunity to derive additional useful information about the microstructure. This has already been demonstrated for short-time-scale measurements by Yablonskiy et al. (7); additional research is needed to determine what can be learned from similar measurements at medium or long time scales.

Coronal projection ADC maps from the lung of a healthy human volunteer for diffusion times ranging from 2 ms to 1.5 s are shown in Fig 5. The short-time-scale ADC map was acquired using the *b*-value pair 0 and 1.6 s/cm², and the medium- and long-time-scale ADC maps were acquired using a tag wavelength of 5 mm (diffusion time: 50 or 200 ms) or 10 mm (diffusion time: 1.5 s). Each ADC map was acquired during a separate breath-hold period. Consistent with the behavior observed in Fig. 4 for the global ADC values, the ADC values in the maps of Fig. 5 decreased monotonically and substantially as the diffusion time increased; the mean ADC decreased by approximately 12-fold as the diffusion time increased from 2 ms to 1.5 s. The mean ADC values in the medium- and long-time-scale ADC maps of Fig. 5 were similar to the global ADC values in Fig. 4 at the corresponding diffusion times. (The results shown in Fig 4 and Fig 5 are from different subjects.)

Figure 6 illustrates the appearance of short-time-scale (Fig 6a) and long-time-scale (Fig 6b) ADC maps in a subject with sub-clinical COPD. Overall, the relative increase in the ADC values in the long-time-scale map compared to those for a healthy subject (mean ADC 0.039 cm²/s for Fig 6b versus 0.013 cm²/s for Fig 5, right; 200% increase) was substantially larger than that for the ADC values in the short-time-scale ADC map (mean ADC $0.31 \text{ cm}^2/\text{s}$ for Fig 6a versus $0.16 \text{ cm}^2/\text{s}$ for Fig 5, left; 94% increase). In addition, as seen in Fig 6, the regional elevation of the ADC values in the lung apices was more conspicuous on the long-time-scale ADC map than on the short-time-scale ADC map. The ADC values along the dashed lines through the mid-section of the lung were 0.036 cm²/s and 0.34 cm²/s, respectively, whereas those along the dotted lines through the apices were $0.050 \text{ cm}^2/\text{s}$ (39% increase) and 0.41 cm² /s (21% increase), respectively (10). This result suggests that long-time-scale ADC measurements may be more sensitive than short-time-scale ADC measurements for detecting the microstructural alterations that occur in emphysema, and is consistent with measurements made in canine and explanted human lungs by Woods et al (11,12) using a somewhat different technique for measuring the long-time-scale ADC; recent studies in additional human subjects continue to support this hypothesis (24).

Our final example is a 6-year-old child with a clinical history of BPD. The ADC maps shown in Fig 7 are from a series of axial slices that were obtained during a single breath-hold with a pulse sequence that acquired both short-time-scale and long-time-scale measurements by using a gradient-echo-based technique immediately followed by a stimulated-echo-based technique (25). The ADC values within the lung parenchyma are fairly uniform in the short-time-scale ADC map; the elevated values in the central region correspond to 3 He gas in large airways. In contrast, local elevations in ADC values are seen in both lungs in the long-time-scale ADC map, and include a region of particularly elevated values in the lateral aspect of the left lung. Analogous to the results observed for early emphysema, it appears that long-time-scale ADC measurements may be more sensitive than short-time-scale ADC measurements for detecting certain microstructural alterations that occur in BPD.

CONCLUSIONS

By using a combination of gradient-echo-based and stimulated-echo-based pulse sequences, it is possible to explore the diffusion of hyperpolarized 3 He in the human lung over more than a 1000-fold variation of the diffusion time. Diffusion measurements made over such a range

of time scales provide a new opportunity to explore the microstructure of the healthy and diseased lung. In healthy human subjects, the ADC decreases by more than 10-fold as the diffusion time increases from approximately 1 ms to 1 s; the ADC remains dependent on diffusion time for times up to at least approximately 10 seconds. Preliminary experimental results suggest that measurements made at diffusion times substantially longer than 1 ms may be more sensitive for detecting certain pathological changes in the microstructure of the lung, such as those that occur in emphysema or bronchopulmonary dysplasia. Additional studies with these techniques are needed in both healthy and diseased lungs to better understand the relationship between microstructural changes and variations in the ADC with diffusion time, and to determine appropriate parameter combinations for optimum detection and characterization of pathological changes.

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Figure 1.

a: Basic framework for making an ADC measurement based on a gradient-echo pulse sequence. Spatial encoding gradients, which would be applied following the "un-tag" gradient pulse, are omitted for simplicity. **b:** The effect of the tagging gradient pulse on the transverse magnetization. The left side of the diagram shows a series of representative transverse magnetization vectors before application of the tagging pulse. The tips of these magnetization vectors fall along a straight line. The gradient twists the magnetization vectors into a helical pattern as shown on the right side of the diagram.

Figure 2.

Basic framework for making an ADC measurement based on a stimulated-echo pulse sequence. Spatial encoding gradients are omitted for simplicity. The "un-tag" gradient pulse is positive (instead of negative) because the second RF pulse phase conjugates the magnetization in the process of storing it.

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Figure 3.

Relationship between the diffusion time and the RMS distance that ³He atoms in air diffuse in an unrestricted environment. The approximate dimensions of several structures in the human lung are indicated for comparison.

Figure 4.

Global ADC values from the lung of a healthy human volunteer for diffusion times ranging from 20 ms to 6.5 s. ADC values for diffusion times from 20 ms to 1.5 s were measured during a breath-hold using a tag wavelength of 5 mm. ADC values for diffusion times from 200 ms to 6.5 s were measured during a second breath-hold using a tag wavelength of 10 mm. For a given tag wavelength, the ADC decreased monotonically with increasing diffusion time. Each of the two measurements was performed using a single application of a stimulated-echo-based pulse sequence with the following parameters: TR, 62 ms; TE for stimulated echoes, 6.0 ms; TE for calibration data, 0.5 ms; flip angle, 5°; number of ADC values per measurement, 24 (5mm tag wavelength) or 102 (10-mm tag wavelength).

Figure 5.

Coronal projection ADC maps from the lung of a healthy human volunteer for diffusion times ranging from 2 ms to 1.5 s. Each ADC map was acquired during a separate breath-hold period. The short-time-scale ADC map (diffusion time: 2 ms) was acquired using a gradient-echobased pulse sequence, and the medium- and long-time-scale ADC maps (diffusion times: 50, 200 and 1500 ms) were acquired using a stimulated-echo-based pulse sequence. The ADC values decreased monotonically with increasing diffusion time, consistent with the behavior for global ADC values illustrated in Fig 4. The artifactual dark regions near the base of the lung were caused by the large susceptibility interface at the diaphragmatic surface. Parameters for the gradient-echo acquisition included: TR, 6.3 ms; TE, 4.5 ms; flip angle, 10°; *b* values, 0 and 1.6 s/cm²; diffusion-sensitization direction, anterior-posterior. Parameters for the stimulated-echo acquisition included: TR, 8.0 ms; TE for stimulated echo, 7.0 ms; TE for diffusion-weighted image, 2.3 ms; TE for calibration data, 3.6 ms; flip angle, 5°; tag wavelength, 5 mm (diffusion time 50 or 200 ms) or 10 mm (diffusion time 1500 ms); diffusionsensitization direction, anterior-posterior. Parameters common to both acquisitions included: in-plane resolution, 5.9×5.9 mm²; slice thickness, projection. Adapted from Fig 2 in reference (20).

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Figure 6.

Coronal short-time-scale (**a**) and long-time-scale (**b**) ADC maps from a subject with subclinical COPD. The diffusion times were 2 ms and 1.5 s for the short-time-scale and long-timescale measurements, respectively. The long-time-scale ADC map exhibits markedly elevated ADC values in the lung apices; the values in the mid-section and base of the lung are also elevated compared to those for a healthy subject (e.g., Fig 5, right-most ADC map). In contrast, the short-time-scale ADC values in the lung apices are only mildly elevated compared to those in the rest of the lung. Parameters for the short-time-scale, gradient-echo acquisition included: TR, 6.3 ms; TE, 4.5 ms; flip angle, 10°; in-plane resolution, 5.0×10.0 mm²; slice thickness, projection; *b* values, 0 and 1.6 s/cm²; diffusion-sensitization direction, anterior-posterior. Parameters for the long-time-scale, stimulated-echo acquisition included: TR, 6.4 ms; TE for stimulated echoes, 7.0 ms; TE for calibration data, 1.3 ms; flip angle, 5°; in-plane resolution, 6.3×7.3 mm²; slice thickness, projection; tag wavelength, 10 mm; diffusion-sensitization direction, anterior-posterior. Adapted from Fig 9 in reference (10).

Figure 7.

Axial short-time-scale (**a**) and long-time-scale (**b**) ADC maps from a subject with bronchopulmonary dysplasia. The diffusion times were 2 ms and 1.0 s for the short-time-scale and long-time-scale measurements, respectively. The ADC values are quite uniform within the lung parenchyma in the short-time-scale ADC map (the elevated values are gas within large airways), while local elevations of the ADC are seen in both lungs in the long-time-scale ADC map. Parameters for the short-time-scale, gradient-echo acquisition included: TR, 11.0 ms; TE, 6.7 ms; flip angle, 3° ; *b* values, 0 and 1.6 s/cm²; diffusion-sensitization direction, head-foot. Parameters for the long-time-scale, stimulated-echo acquisition included: TR, 6.4 ms; TE for stimulated echoes, 7.0 ms; TE for calibration data, 1.3 ms; flip angle, 5°; tag wavelength, 10 mm; diffusion-sensitization direction, head-foot. Parameters common to both acquisitions included: in-plane resolution, 5.9×5.9 mm²; slice thickness, 40 mm.