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NEOCEPTORS: REENGINEERING GPCRS TO RECOGNIZE TAILORED LIGANDS

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Abstract

Efforts to model and reengineer the putative binding sites of G protein-coupled receptors (GPCRs) have led to an approach to combining small molecule “classical” medicinal chemistry and gene therapy. By this approach, complementary structural changes, for example, based on novel ionic or H bonds, are made in the receptor and ligand for selective enhancement of affinity. Thus, a modified receptor (neoceptor) is designed for activation by tailor-made agonists that do not interact with the native receptor. The neoceptor is no longer activated by the native agonist, but rather acts as scaffold for docking of novel small molecules (neoligands). In theory, the approach could verify the accuracy of GPCR molecular modeling, dissection of signaling, design of small molecules to rescue disease-related mutations, and small-molecule-directed gene therapy. The neoceptor-neoligand pairing may offer spacial specificity by delivering the neoceptor to a target site and temporal specificity by administering neoligand when needed.

Introduction

The use of GPCR agonists for therapy has inherent limitations [1] from desensitization and widespread receptor distribution leading to undesired side effects. We are developing an alternate approach to benefit from GPCR activation in a more spatially and temporally selective manner than the systemic administration of agonists to the native GPCR. This approach of neoceptors [2–4] combines small molecule “classical” medicinal chemistry and gene or cell therapy. By this rational design approach, complementary structural changes are made in the receptor and ligand for selective enhancement of affinity (Figure 1). The activation of a neoceptor in a spatially-selective manner would be achieved by cell- or organ-target delivery of the gene, given the development of an appropriate delivery method.

Molecular modeling based on homology to the best studied GPCR, rhodopsin [5,6], has been used widely to arrive at hypotheses for ligand docking, which are ideally validated using site-directed mutagenesis [7,8]. With this knowledge and the ability to tailor-make new analogues of a native agonist, one may design a matched neoceptor and neoligand, i.e. the binding site of a given GPCR may be engineered to recognize synthetic agonist ligands that do not activate the native receptor. As opposed to *de novo* receptor design [9], this approach uses the native receptor as a scaffold for docking of novel molecules. This reengineered GPCR (neoceptor) ideally retains its capacity to activate a particular second messenger pathway causing beneficial effects identical to those induced by the native receptor. The uniquely-matched ligands

(neoligands) are synthesized based on molecular complementarity with the neoceptor. The structure activity relationship (SAR) profile of such modified receptors need not correspond to that of the parent.

It is envisioned that the neoceptor DNA would be delivered by an appropriate organ-targeted gene or cell therapy. In the absence of the neoligand, the neoceptor would be “silent”, not subject to activation by the native agonist. The side effects normally associated with agonist therapy would not be expected, since the native receptor would not be activated by the tailored ligand. Thus, by design, the interacting pairs of receptor and ligand would have to be orthogonal with respect to the native pair.

The reengineering of enzymes, nuclear receptors, and other proteins is practiced in a variety of contexts [10–16]. Various kinases were reengineered to recognize modified ATP analogues, for example, by the creation of a “bump and hole” [10]. Microscopic complementarity of the β -adrenoceptor and its chemically modified ligands was studied [12]. GPCRs have been engineered for regulation by metal ions leading to insights into the activation mechanism [17]. Reengineered receptors have been proposed for rescue from genetic diseases [4,15].

Although not intended for therapeutic application, Conklin and colleagues have introduced RASSLs (Receptors Activated Solely by Synthetic Ligands) for mechanistic probing through conditional expression in transgenic mice [13]. RASSLs begin with a GPCR for which a synthetic high affinity agonist probe is known and then mutation reduces the affinity of the endogenous ligands with retention of affinity for the synthetic agonist. The neoceptor approach, however, is based on the rational reengineering of both the putative binding site (genetically) and the ligand. Although the term RASSL would seem to include the neoceptor approach as well, there is an important distinction. In the first reported class of RASSLs, i.e. chimeric κ -opiate receptors containing the second extracellular loop of the δ -opiate receptor, the recognition was not truly orthogonal – the synthetic ligand also activated the native receptor, indicating a reengineered receptor, but not an engineered ligand. A mutant histamine receptor that combines a 200-fold lower potency for the endogenous ligand with improved affinity of the 2-phenylhistamine class of H_1 receptor agonists was identified [18]. Random mutagenesis of the A_{2B} AR led to the identification of gain-of-function mutations [19].

This article describes the efforts using the adenosine receptors (ARs) as proof of the neoceptor concept by rationally applying insights from molecular modeling and the envisioned applications as research tools and possibly in a futuristic therapeutic modality.

Neoceptors – development

The neoceptor approach was validated for ARs, which respond to stress-elevated levels of extracellular adenosine and have diverse protective roles against ischemia and tissue damage [20,21]. Much experience has been gained to tailor ligands for the native ARs, and several selective agonists are in advanced clinical trials for inflammation, cancer, arthritis, and cardiac arrhythmias and imaging [20,22–24]. Because of the widespread distribution of native ARs, agonist therapy has been impeded by side effects. The A_{2A} and A_3 ARs have been developed as test cases of the neoceptor approach [2–4] to address the issue of inherent nonselectivity of agonist-based therapies.

In general, hypotheses for receptor docking of nucleosides and the proposed conformational changes of GPCRs that are associated with activation have guided the design of neoligands. Modeling of the putative ligand-binding site of the A_3 AR receptor [8] led to the identification of a conserved site for mutagenesis, i.e. a His residue (7.43) that has been implicated in agonist recognition [8]. Modeling of A_1 , A_{2A} and A_3 ARs places His(7.43) within a hydrophilic ribose-binding region of the putative agonist binding site. This residue corresponds to the Lys of

rhodopsin which forms the Schiff base with retinal, and in the AR has been proposed to be H-bonded to the 3'-hydroxyl group of adenosine [2]. Mutagenesis of A_{2A} and A₃ ARs indicates His(7.43) is associated with agonist binding and less important for antagonist binding.

Initially the A₃AR was converted into a neoreceptor that can recognize uniquely modified nucleosides that are inactive at the native ARs [2]. This concept is also dependent on the retention of the ability of the neoreceptor to activate signaling pathways known to be beneficial in cardiac myocyte cultures for antiischemic protection, such as A₃AR-activated phospholipase D [4].

His7.43 of the A₃AR receptor was mutated to Glu. A complementary functional group was incorporated in a synthetic neoligand, i.e. 3'-amino-3'-deoxyadenosine (MRS1960, Figure 2). A novel electrostatic pair forming between the neoreceptor and neoligand was intended for selective recognition of MRS1960 by the carboxylate-modified receptor [2]. The consequences of H272E mutation are: 1) Adenosine (100 μM) no longer binds to the receptor. However, the affinity of a standard agonist radioligand (¹²⁵I-I-AB-MECA) is fortuitously only 2-fold decreased. Thus, an important synthetic agonist tool may still be used to characterize the mutant receptor. MRS1960 and the mutant H272E A₃ AR form a suitable association to achieve a 6-fold enhancement of binding affinity. A novel electrostatic pair would account for the enhanced recognition and subsequent activation of second messenger systems by the tailored ligand.

Since the ratio of selective enhancement of MRS1960 was only modest, we improved the interaction based on prediction from molecular modeling [4,25]. A 3'-aminomethyl neoligand (MRS3176 [26]) achieved a 20-fold enhancement of affinity at the H272E mutant A₃ AR. The impetus for this structural change was the prediction from molecular modeling that the 3'-amino group may be at an excessive distance from the His imidazole group to form a direct H bond. This modified nucleoside also contained a substituted N⁶-benzyl group, which tends to enhance affinity at the wild type and mutant A₃ ARs.

Thus, ligands may be tailored with strategically-positioned amino groups for recognition by carboxylate mutants of the native receptor. However, a more dramatic demonstration of selective affinity enhancement and orthogonality was desired. The substituent at the 3'-position of ribose was varied in charge, size, and H bonding ability, and each analogue was examined for binding affinity at several neoreceptor variations. The optimal affinity enhancement and orthogonality followed the incorporation of a urea group in place of the 3'-hydroxyl group of adenosine analogues, as in MRS3481 (Figure 2). The urea group is capable of forming multiple H bonds with the neoreceptor and appears to preclude binding at the native ARs for steric reasons [27]. A model of this analogue docked in the H272E receptor showed a bidentate coordination of between the carboxylate group and the urea moiety (Figure 3).

It was necessary to probe the coupling pattern of the neoreceptor [4]. Although the coupling specificity in GPCRs is principally a function of the second and third intracellular loops [28], and ligand specificity is governed by functionality within the upper third of the TM regions, the preservation of the typical A₃AR second messengers could not be assumed. Coupling to one known effector pathway of the A₃AR, i.e. stimulation of phospholipase C through the Gβ,γ-subunits, is preserved upon activation of the H272E mutant receptor by MRS3481. Thus, at least part of the downstream signaling of this cytoprotective receptor is maintained. The stimulation of phospholipase C by MRS3481 occurred with an EC₅₀ value of ~100 nM at the neoreceptor, while it was inactive at 100 μM at the WT receptor similarly expressed in COS-7 cells. For comparison, the EC₅₀ for adenosine at the WT receptor was ~1 μM and >100,000 at the neoreceptor. The effects of MRS3481 acting through the H272E neoreceptor on other signaling systems (e.g., cyclic AMP, ion channels, and arrestin) remain to be determined.

In a chick cardiac myocyte culture, which is an established model for cardioprotection [29], the neoligand MRS3481 induced a potent antiischemic protection in cells expressing the H272E neoceptor [4]. Also, this protection correlated with the activation of PLD, as occurs with the native A₃ AR transfected in the same cell system. Thus, a neoceptor-neoligand pair has been demonstrated to be beneficial in inducing stages of a response to stress in a tissue known to respond similarly when the native parent receptor is present.

The anti-inflammatory A_{2A} AR was also converted into a neoceptor [3]. The same approach of mutation of the conserved His278 in TM7 could not be used, because mutation of the corresponding His residue to Asp or Glu did not lower the potency of native adenosine. However, a hydrophilic residue, i.e. T88 in TM3, on the other side of the putative subdomain for ribose binding to the ARs was selected for mutation. This residue in the A_{2A} AR is exclusively associated with agonist binding [3]. The T88D mutant receptor was unaffected in the ability to bind the nonselective AR antagonist CGS15943, however it failed to bind the nonselective AR agonist NECA, even at a concentration of 100 μM. The T88D mutant A_{2A} AR recognized a strategically 5'-modified amino derivative (MRS3366, Figure 2B). Moreover, the precise position and spacer length of the amino group was critical to achieving a selective affinity enhancement at the neoceptor. Other mutant A_{2A} ARs displayed even greater degrees of enhancement for neoligands. For example, MRS3417 was functionally enhanced at the N181D mutant receptor by 110-fold, however, this combination was not truly orthogonal since this modified receptor was still capable of being activated by known AR agonists.

In addition to mutating the ligand binding pocket, it is also possible to mutate sites involved in phosphorylation and desensitization to retard these processes in neoceptors. This is particularly important with regard to A₃ receptors since these are known to undergo exceptionally rapid desensitization. It should also be possible to mutate promoter regions of the neoceptor transcript. This could be used to enable induction of the neoceptor mRNA - thus adding an additional layer of control in the response to the neoligand.

Neoceptors – potential applications

Ligand docking in rhodopsin-based molecular models of GPCRs has been controversial. Neoceptors provide a means of verifying the accuracy of predictions based on molecular modeling of GPCRs, which have been subject to discrepancies, especially for agonist docking [6]. A gain-of-function mutation and a complementary ligand provide powerful evidence that the predicted binding pocket is correct. Moreover, neoceptors can be used for mechanistic probing of the role of a specific GPCR in cells or tissues.

Therapeutic applications are also envisioned for gene therapy, dependent on site specific gene delivery, e.g. in the cardiovascular system [30] (Table 1). Novel proposed applications include donor stem cells, which hold great promise in repairing or regenerating diseased tissues such as heart or other organs. Methods to enhance the survival of exogenous stem cells when they are implanted in the recipient subjects may greatly increase their ability to achieve repair. Although the native A₃ AR has a potent cytoprotective effect, its ubiquitous presence will cause significant side effects. The use of a tailor-made neoligand is proposed for selectively activating, as needed, a cytoprotective neoceptor to be expressed in the donor stem cells. Another potential therapeutic application of the orthogonal neoceptor-neoligand pair is in skeletal muscle, based on cytoprotection by adenosine acting at the A₁AR [32].

Additionally, increasing evidence suggests that mutations in genes encoding GPCRs are an important cause of human disease. The neoceptor approach and binding site modeling could be used to design small molecules to specifically rescue disease-related mutations.

Conclusions

Neoeptors represent a rational design approach for new pharmacological tools and possible therapies. By iterative steps of modeling and ligand design, one may identify and refine a neoeptor/neoligand pair. The success of the neoeptor strategy for the ARs validates the use of GPCR homology modeling, and provides a means for dissection of signaling, design of small molecules to rescue disease-related mutations, and small-molecule-directed gene therapy. The neoeptor-neoligand pairing may offer spacial specificity by delivering the neoeptor to a target site and temporal specificity by administering neoligand when needed. The success of this approach depends on orthogonality of the interaction, which is not required by the RASSL approach, to avoid undesirable, nonselective activation of the native receptor. This process may now be applied to other GPCRs, even in the absence of X-ray crystallographic structures.

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Glossary

SAR

structure activity relationships

Ligand docking

process of computational identification of an energetically favorable binding mode of a small molecule in its receptor site

Orthogonal

multiple systems in which individual elements interact only within a system and do not cross-react, from the Greek “ortho”, meaning “right” and “gonia”, meaning “angle”

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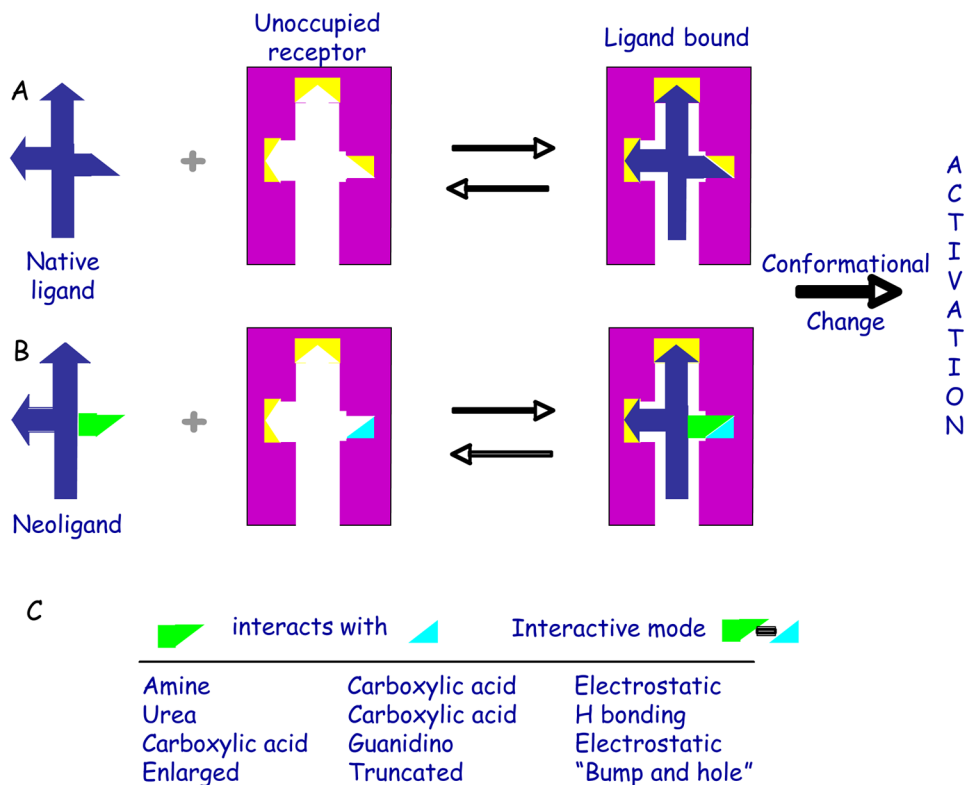
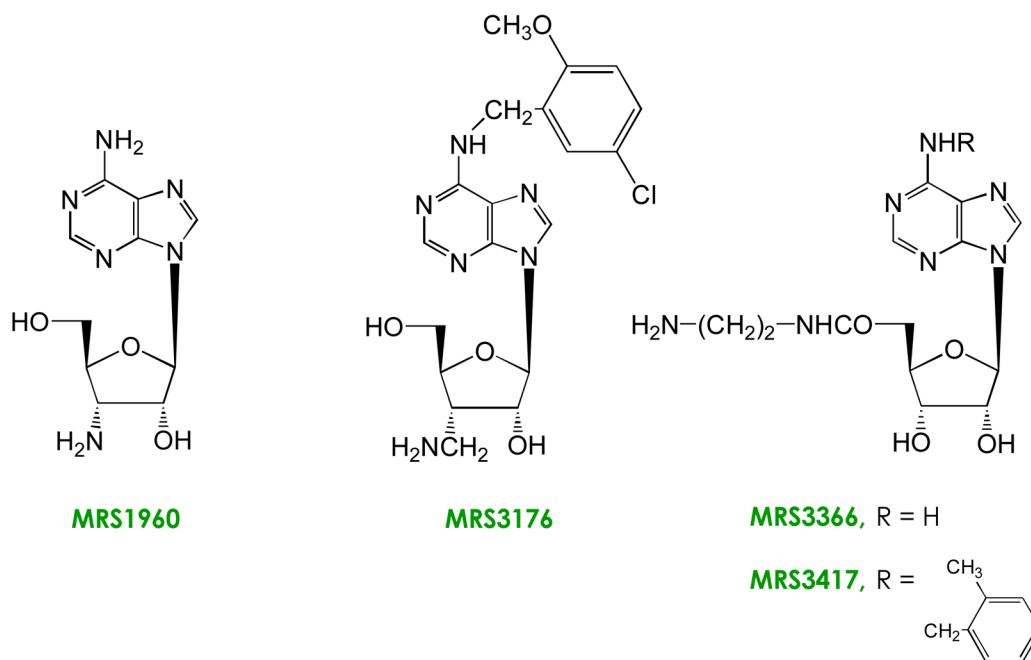
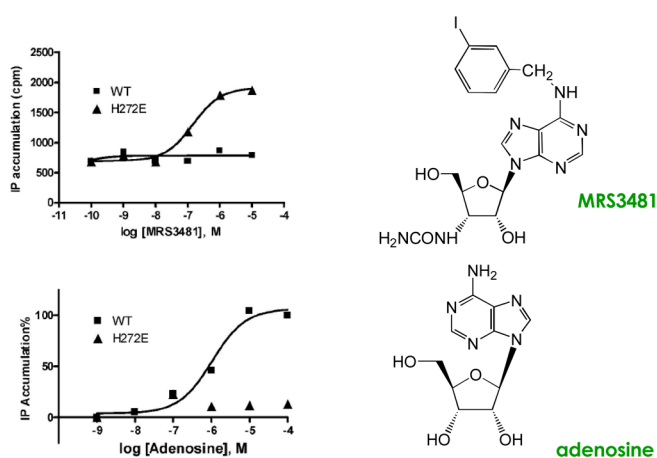


Figure 1. Schematic of reengineering of GPCR by mutation of a small region of the putative ligand binding site of the native receptor (A) to recognize a tailored ligand, to achieve the desired orthogonality of activation. Yellow polyhedra symbolize functional groups on the native receptor required for ligand recognition, and at least one of which is modified in the neoceptor. The pair of neoceptor and neoligand (B) is intended for therapeutics via organ-targeted delivery of a reengineered GPCR gene. (C) shows means by which selective affinity enhancement may be achieved.



A



B

Figure 2. Modifications of the structure of adenosine leading to neoligands for A_{2A} and A_3 ARs. Known SAR of adenosine derivatives have identified sites of modification for achieving receptor subtype selectivity and in some cases for tuning the efficacy. Cumulative efforts to characterize agonist SARs at ARs have focused on the substituent groups at 2, N^6 , 5' positions of the nucleoside and the conformation of the ribose moiety [25]. (A) Amine derivatives, which showed enhancements in binding of 6- (MRS1960) and 20-fold (MRS3176) at the H272E mutant A_3 AR; and 10-fold (MRS3366) and >300-fold (MRS3417) at mutant A_{2A} ARs. The large N^6 -substituents of MRS3176 and MRS3417 serve to increase the affinity at both wild type and mutant ARs, and the extra methylene group at the ribose 3' position was included to span a predicted gap in the A_3 AR docking model. (B) Comparison of the functional effects of

adenosine and the 3'-ureido neoligand MRS3481 at wild type and H272E mutant A₃ARs. The enhancement of binding was >200-fold.

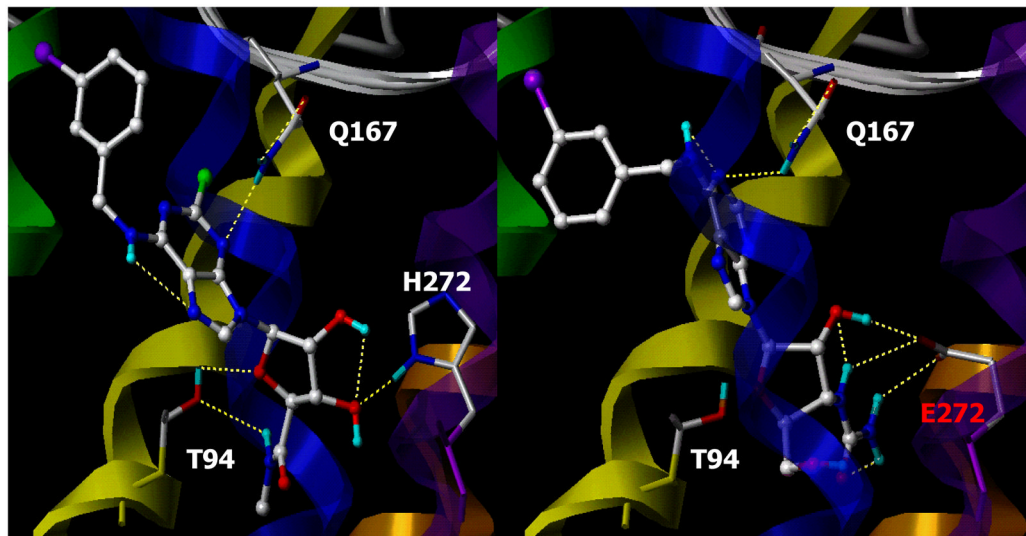


Figure 3. Docking of agonist Cl-IB-MECA in the native human A₃AR (left panel) and a neoligand, MRS3481, in the H272E neoreceptor (right panel). The homology models were derived from human A₃AR model based on rhodopsin and resembling the meta I state [6].

Table 1
 Future Therapeutic Applications of AR-Derived Neoreceptors.

Receptor	Target tissue	Effect	Reference ^a
A ₁	A-V node	Antiarrhythmic	31
A ₁	Skeletal myocytes	Antiischemic protection	32
A ₁	Kidneys	Reduced hyperfiltration	33
A ₁ , A ₃	Cardiac myocytes	Antiischemic protection	4
A ₁ , A ₃	Hematopoietic stem cells	Myeloprotection, enhanced survival	34
A _{2A}	Neutrophils, T cells	Antiinflammatory	35
A _{2A}	Platelets	Antithrombotic	36
A _{2A}	Liver, bowel	Cytoprotection	37,38
A _{2A} , A _{2B}	Endothelial cells/Vascular smooth muscle cells	Vasodilatation, angiogenesis	39,40
A ₃	Cancer cells/tumors	Cytostatic, anticancer effect	22
A ₃	Synoviocytes	Antiarthritic	23
A ₃	Lungs	Antiischemic protection	24

^aThe basis for these projected applications in the physiological effects of the native ARs is described in reference 25 and as indicated.