

MINIREVIEW

Vitamin D Signaling, Infectious Diseases, and Regulation of Innate Immunity[▽]

John H. White*

Departments of Physiology and Medicine, McGill University, Montreal, Quebec, Canada

Vitamin D was first identified as a cure for nutritional rickets, a disease of bone growth caused by an inadequate uptake of dietary calcium. It is now known that vitamin D can be obtained through two independent pathways: limited dietary sources and the photochemical action of solar UV light in the skin. Cod liver oil was discovered to be an excellent source of antirachitic activity in 1827, although it was not until several decades later that the active ingredient was identified as vitamin D₃. Even earlier, in 1822, a Polish physician studying children reached the remarkable conclusion that sunlight cured rickets after noting that rickets was relatively rare in the clearer air of rural areas. Almost 100 years later, in 1919, it was shown that artificial UV light cured rickets (49; www.beyonddiscovery.org/Includes/DBLink.asp?ID=1176). Indeed, secosteroidal vitamin D₃ is produced in the skin via photochemical and thermal conversion of 7-dehydrocholesterol in the presence of UVB light (~295 to 320 nm). While it seems that vitamin D is readily accessible via dietary or solar routes, vitamin D insufficiency or deficiency is, in fact, quite widespread. Solar UVB irradiation is absorbed by atmospheric ozone; consequently, the surface intensity of UVB varies markedly with latitude and time of year. Moreover, as vitamin D intake is generally inadequate in most diets (38–40), the rate of vitamin D insufficiency or deficiency increases with increasing latitude.

The term vitamin D refers collectively to vitamin D₃ and vitamin D₂, which is derived from irradiation of the steroid ergosterol in yeast. Biologically active vitamin D is generated via largely hepatic 25-hydroxylation catalyzed by CYP2R1, CYP27A1, and possibly other enzymes to produce 25-hydroxyvitamin D (25D) (21, 40, 43, 67, 81), which has a long half-life and is the major circulating vitamin D metabolite. 25D is modified by 1 α -hydroxylation catalyzed by CYP27B1, which produces hormonal 1,25-dihydroxyvitamin D (1,25D) (40, 43, 67). Vitamin D compounds are catabolized via 24-hydroxylation by CYP24, whose expression is strongly inducible by 1,25D, which constitutes a negative feedback loop (40, 43, 67).

While the kidneys are a major site of 1 α -hydroxylation of 25D, it has recently become clear that generation of hormonal 1,25D in peripheral tissues is critical to the full scope of the physiological actions of this compound. Renal 1 α -hydroxylation

is tightly controlled by calcium homeostatic signals, particularly circulating parathyroid hormone (PTH). Although initially characterized as a calcium homeostatic agent, vitamin D is now known to have pleiotropic actions, including a key role in immune system regulation (49). Importantly in this regard, recent research detailed here has uncovered critical, cell-specific differences in both the regulation of 1 α -hydroxylation of 25D and 24-hydroxylation that are relevant to the role of 1,25D as an immune system regulator.

VITAMIN D INSUFFICIENCY/DEFICIENCY AND DISEASE

While there is no strict definition, vitamin D deficiency is widely defined as circulating 25D levels of less than 20 ng/ml (50 nM) (11, 39, 40, 53, 87), whereas an individual is generally considered to be vitamin D sufficient if the circulating 25D concentration is greater than 30 to 32 ng/ml (75 to 80 nM) (19, 37, 87). 25D levels are inversely associated with circulating PTH levels until the 25D concentration is greater than 30 to 40 ng/ml, at which point PTH levels bottom out. While vitamin D intoxication can occur, it is not observed until 25D levels reach 150 ng/ml (375 nM) or more (40), and it is associated with hypercalcemia, which, if chronic, can result in urinary calculi (renal or bladder stones) and renal failure.

While cases of vitamin D toxicity do occur, vitamin D insufficiency/deficiency is far more common. In temperate regions, surface solar UVB irradiation is insufficient to induce cutaneous vitamin D₃ synthesis for periods around the winter solstice that are up to 6 months long or longer at higher latitudes (38), a period that is known as vitamin D winter. For obvious reasons, cutaneous vitamin D synthesis is also strongly influenced by skin color (55). Lack of cutaneous vitamin D synthesis, coupled with vitamin D-poor diets, has contributed to high levels of vitamin D insufficiency or deficiency in European and North American populations (38–40, 60). For example, a survey of healthy females across northern Europe found that there was widespread vitamin D deficiency (6), and a recent study found that 42% of African-American women in the United States were seriously 25D deficient (<15 ng/ml) (19).

Epidemiological studies have linked vitamin D deficiency to increased rates of cancer, as well as autoimmune and infectious diseases (80). In the United States the rates of bladder, breast, colon, ovary, and rectal cancer increase twofold from south to north (34). North-south gradients of autoimmune conditions, such as multiple sclerosis, Crohn's disease, and type 1 diabetes,

* Mailing address: Departments of Physiology and Medicine, McGill University, Montreal, Quebec, Canada. Phone: (514) 398-8555. Fax: (514) 398-7452. E-mail: john.white@mcgill.ca.

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have also been documented (2, 15, 47, 59). Connections between vitamin D insufficiency and infectious diseases go back over 100 years to the recognition in the 19th century that solar radiation was beneficial for patients suffering from tuberculosis (TB). Associations between vitamin D deficiency and TB susceptibility were described over 20 years ago (22, 33). A more recent study of a genetically homogeneous immigrant population of Gujarati Asians in the London area with high rates of TB found that there was an association between active disease and 25D deficiency and that there was an even stronger association of disease with undetectable serum levels of 25D (101). In addition, we have known for over 20 years that 1,25D inhibits the growth of *Mycobacterium tuberculosis* in cultured human macrophages (73). Interest in the connection between vitamin D supplementation and treatment of TB has been rekindled lately by the findings of several studies (50–52, 54, 75), not least of which was the recent observation of Martineau and colleagues in a double-blind randomized controlled trial that a single dose of 100,000 U (2.5 mg) of vitamin D₃ enhanced antimycobacterial immunity in healthy tuberculin skin test-positive donors (54). In addition, critical links described below have recently been made between molecular events controlling vitamin D signaling pathways and innate immune responses against mycobacterial infection.

While the potential protective effects against TB infection have attracted the most attention, data are accumulating from several sources that vitamin D may also be beneficial in combating a range of other bacterial or viral infectious agents. One small but intriguing study worthy of follow up found that elderly women undergoing long-term treatment with vitamin D as an antiosteoporosis agent had a significantly lower rate of *Helicobacter pylori* infections than women in an untreated control group (44). There have also been a number of studies examining the potential role of vitamin D in protection against upper and lower respiratory tract infections, which can be caused by a variety of etiological agents, many of which are viral (14, 57, 102). Subclinical vitamin D deficiency was associated with severe lower respiratory tract infection in an Indian study (97), and clinical vitamin D deficiency was associated with a 13-fold-increased risk of pneumonia in Ethiopian children (58). A Finnish study found that there was an association between serum 25D concentrations of less than 40 nM (16 ng/ml) and a range of acute respiratory infections (sinusitis, tonsillitis, otitis, bronchitis, pneumonia, pharyngitis, and laryngitis) in young army recruits (46). In addition, Cannell and several colleagues have persuasively argued, based on a range of epidemiological data, that cutaneous vitamin D production provides the “seasonal stimulus” associated with solar radiation that underlies the seasonality of epidemic influenza (16, 17). Finally, clinical and genetic evidence is accumulating that vitamin D may play a role in modulating human immunodeficiency virus (HIV) infection, although more work needs to be done to clarify the relationship between vitamin D physiology and HIV infection. A positive correlation was established between vitamin D supplementation and CD4-positive T-cell counts in seropositive individuals (94). A correlation between mortality from HIV infection and vitamin D deficiency has not been clearly established. However, interpretation of the vitamin D status of HIV-positive individuals is complicated by the potential confounding effects of antiretroviral therapy on vita-

min D metabolism (94). This is an area that merits further clarification, because, as detailed below, a potential role for 1,25D signaling in modulating HIV infection is supported by genetic studies on vitamin D receptor (VDR) gene polymorphisms.

VDR GENE POLYMORPHISMS AND INFECTIOUS DISEASES

The association between vitamin D physiology and infectious disease is also supported by genetic studies implicating polymorphisms in the gene encoding the VDR in disease susceptibility (91). There are numerous VDR polymorphisms, including a common *FokI* restriction fragment length polymorphism (RFLP) that shifts translational initiation to an ATG three codons downstream and *TaqI* and *BsmI* RFLPs in the 3′ untranslated region. Genetic studies have linked VDR polymorphisms with a number of infectious diseases, including susceptibility to *M. tuberculosis* infection and treatment outcome. A Peruvian study found an association between specific genotypes of both the *TaqI* and *FokI* RFLPs and time to microbiologic resolution of pulmonary TB (75). In a case-control study of 2,015 African subjects, homozygotes for *TaqI* polymorphism (genotype *tt*) were significantly underrepresented in TB patients (8). In the study on Gujarati Asians mentioned above, the *ff* genotype of the *FokI* RFLP was associated with the extent of pulmonary TB in 25D-deficient patients (101).

VDR polymorphisms have also been linked to leprosy, which is caused by a distinct mycobacterial agent, *Mycobacterium leprae* (29). Tuberculoid leprosy presents with few bacilli in macrophages and a strong cell-mediated response, whereas the more severe lepromatous leprosy is characterized by numerous bacilli and a weak cellular response. The *tt* VDR polymorphism was associated with tuberculoid leprosy in Bengali patients, whereas the *TT* genotype was associated with lepromatous leprosy (77). Although the *tt* genotype was associated with susceptibility to leprosy in a case-control study of patients in the Karonga district of Malawi (30), the expected frequency of *tt* homozygotes was low (5%), and apparent differences between patient and control populations could have been due to chance.

A recent analysis of young children found that the *ff* genotype was associated with adjusted relative odds of acute lower respiratory tract infection (predominantly viral bronchiolitis) that were seven times those of the *FF* genotype (76). VDR polymorphisms have been linked to HIV infection, although clear conclusions regarding the role of vitamin D signaling in controlling HIV infection have been difficult to draw. No associations were found between *BsmI* polymorphisms and HIV infection, whereas an association was established between the *BB* genotype and disease progression based on several criteria (7). However, it is difficult to ascribe variations in the *BsmI* genotype to changes in VDR function. Another recent study found no association between a specific polymorphism and protection against HIV infection in a population of injection drug users, but it did find a correlation between specific VDR haplotypes (blocks of polymorphisms) (23). The authors concluded that protective VDR polymorphisms were associated with reduced VDR function, consistent with vitamin D signal-

ing promoting HIV infection, and noted based on an in vitro study that the 1,25D-bound VDR could activate the HIV type 1 long terminal repeat (61).

MOLECULAR MECHANISMS OF ACTION OF VITAMIN D

Much of the action of 1,25D can be explained by its binding to and activation of the VDR. The VDR is a nuclear receptor and ligand-activated transcription factor (20, 49) composed of a highly conserved DNA binding domain and an α -helical ligand binding domain (72). The ligand-bound VDR activates transcription by heterodimerization with retinoid X receptors (RXRs), which is essential for high-affinity DNA binding to cognate vitamin D response elements (VDREs) located in the regulatory regions of 1,25D target genes. VDREs are composed of direct repeats of PuG(G/T)TCA motifs separated by 3 bp (DR3) or everted repeats with 6-bp spacing (ER6) (20, 26, 49, 89, 90). (Note that everted repeats are palindromic but have symmetry [toes pointing out] opposite that of the so-called inverted repeats [toes pointing in] originally identified as response elements for steroid receptors.) ER8 motifs can also function as response elements for the VDR and related retinoic acid receptors (86), thus partially integrating 1,25D and retinoid signaling. DNA-bound VDR/RXR heterodimers recruit numerous so-called coregulatory proteins, which control histone modifications, chromatin remodeling, and RNA polymerase II binding and transcriptional initiation (24, 31, 56, 70, 74). The ligand-bound VDR can also repress transcription. For example, in the presence of 1,25D, VDR/RXR heterodimers can displace DNA-bound nuclear factor of activated T cells, thus repressing cytokine gene expression (5, 85). While numerous VDREs have been identified in relatively promoter-proximal locations, recent work has provided evidence that the DNA-bound VDR can function at distances as great as 75 kb to regulate adjacent target gene transcription (45).

VITAMIN D SIGNALING AND METABOLISM IN THE IMMUNE SYSTEM

Evidence for a role of vitamin D signaling in the immune system in general and in innate immune responses in particular has been accumulating from a variety of sources. The VDR is present in most cells of the immune system, including T lymphocytes, neutrophils, and antigen-presenting cells, such as macrophages and dendritic cells (3, 10, 12, 62, 69). 1,25D is an inhibitor of maturation of dendritic cells, the most potent of the antigen-presenting cells, and acts directly on T lymphocytes to inhibit T-cell proliferation (92). 1,25D signaling represses the transcription of genes encoding key T helper 1 (Th1) cytokines, such as gamma interferon and interleukin-2 (5, 92). 1,25D is thus a suppressor of antigen presentation to and activation and recruitment of Th1 cells. The net effect of 1,25D action is to polarize T-helper responses toward a more regulatory Th2 phenotype, which is considered a key component of its capacity to suppress Th1-driven autoimmune responses (92).

In the last few years, researchers in the vitamin D field, and particularly researchers interested in the immunomodulatory functions of vitamin D, have come to appreciate the important

contributions of extrarenal 1 α -hydroxylase (CYP27B1) to vitamin D physiology. Activated macrophages and dendritic cells express CYP27B1 (1, 64, 65, 84), which, unlike the renal enzyme, is not regulated by Ca²⁺ homeostatic signals but is regulated primarily by immune inputs, mainly gamma interferon and agonists of the Toll-like receptor (TLR) pattern recognition receptors. Critically, this renders the immune system responsive to circulating levels of 25D. Liu and colleagues found in microarray studies that signaling through human macrophage TLR1/2 heterodimers stimulated with bacterial lipopeptides induced expression of both CYP27B1 and the VDR (50) (Fig. 1). Most importantly, they showed that in TLR2/1-stimulated human macrophages cultured in the presence of human serum, downstream VDR-driven responses were strongly dependent on serum 25D concentrations. VDR-driven responses were strongly attenuated or absent in serum from vitamin D-deficient individuals, a defect that could be overcome by 25D supplementation. Moreover, consistent with previous findings (60, 82), the 25D levels in serum from African-Americans used in the study were markedly lower than those of Caucasian Americans (50). This study thus provided a clear demonstration of the dependence of immune responses on circulating 25D levels. Similarly, stimulation of the TLR4/CD14 receptor complexes by lipopolysaccharide induces CYP27B1 expression (84; unpublished results), consistent with correlations that other workers have found between TLR4 and CYP27B1 expression (27, 28).

Remarkably, while expression of CYP24, the mitochondrial enzyme that initiates 1,25D catabolism, is exquisitely sensitive to the presence of 1,25D, the negative feedback loop appears to be defective in macrophages (Fig. 1). Ren and colleagues have recently shown that while expression of CYP24 transcripts is induced by 1,25D in macrophages as in other cells, the corresponding enzymatic activity is virtually undetectable (71). 1,25D induces the expression in macrophages of a splice variant form (CYP24-SV) that encodes a truncated enzyme lacking the critical amino-terminal mitochondrial targeting sequence (71). Although the substrate binding pocket of CYP24-SV is apparently functional, the enzyme, trapped in the cytosol, appears to be catalytically inactive. This suggests that, in macrophages, robust 1,25D signaling is maintained over an extended period of time, which would be advantageous for combating intracellular pathogens such as *M. tuberculosis*. It also provides at least part of the molecular basis for the excessive production of 1,25D by macrophages in granulomatous diseases such as sarcoidosis (41).

1,25D IS A DIRECT INDUCER OF ANTIMICROBIAL INNATE IMMUNITY

We have had molecular evidence that 1,25D is a regulator of innate immune responses for several years. It has been known since the early 1990s that expression of the coreceptor of TLR4, CD14, is strongly induced by 1,25D in human cells (63). This regulation is conserved in the mouse; for example, recent work showed that induction of CD14 expression by 25D was abrogated in mice lacking CYP27B1 (79). This study also showed that vitamin D signaling enhanced the expression of TLR2 approximately twofold in human keratinocytes. Given that signaling through either TLR2 or TLR4 enhances vitamin

homologues reduced the infectivity of lentiviral vectors (83), suggesting that vitamin D signaling may indeed induce antiretroviral activity.

SPECIES-SPECIFIC MECHANISMS OF AMP EXPRESSION

Although classes of AMPs are conserved, there is considerable interspecies variation in both gene sequence and number and in tissue distribution and regulation of expression. α -, β -, and θ -defensins contain six disulfide bond-forming cysteines (66), and subclasses are distinguished by different spacings of Cys residues. While β -defensins are widespread in vertebrates, α -defensins are mammalian, and θ -defensins are primate specific. The five human α -defensins are expressed in myeloid or enteric tissues, whereas the 19 murine genes (cryptidins) are only enteric. Cathelicidins are cationic and defined for their N-terminal cathelin domain, which is cleaved during maturation. Mice and humans have single cathelicidin genes, whereas there are multiple cathelicidin peptides in bovine species (103).

Apart from variations in gene number and the tissue distribution of expression, there are also differences in gene regulation between species. Notably, neither of the VDREs in the *CAMP* and *DEFB2* genes is conserved in mice, and Gombart et al. (32) noted that the *CAMP* VDRE is imbedded in an *Alu* repeat, which is a human- or primate-specific transposable element. This lack of conservation is noteworthy in light of differences that have emerged in regulation of AMP expression in humans and mice. It was established by 2001 that stimulation of TLR2 on either human or mouse macrophages led to induction of antimicrobial activity against TB infection (13, 88). Induction of antimicrobial activity in murine macrophages is dependent on inducible nitric oxide synthase activity. Remarkably, however, whereas inducible nitric oxide synthase inhibitors blocked induction of AMP activity in mouse macrophages, they had no such effect in human cells.

The mechanism of induction in human macrophages was unclear until the discovery of the TLR2/1-stimulated expression of both *CYP27B1* and *VDR* in human cells, leading to the induction of *CAMP* under conditions of 25D sufficiency (50) (Fig. 1). Moreover, in 25D-treated human cells, *CAMP* protein was shown to colocalize with mycobacteria in phagolysosomal structures. Subsequently, knockdown of *CAMP* expression in TB-infected human THP-1 macrophage-like cells confirmed that its induction is essential for 1,25D-stimulated antimycobacterial activity (52). Whether vitamin D signaling is induced in murine macrophages remains unclear. However, even if induction of *CYP27B1* and *VDR* does occur, it would be unlikely to lead to *CAMP* expression because of the lack of a VDRE in the murine *CAMP* gene.

CAMP expression is also strongly induced in human keratinocytes under epithelial wound healing conditions (18, 25, 36). In *CYP27B*^{-/-} mice, however, under conditions where *CYP27B1* ablation completely eliminated the strong injury-induced expression of CD14, induction of *CAMP* expression was mildly attenuated, but the effect did not achieve statistical significance (79). It has been argued that regulation of AMP expression in mice and humans has diverged because mice use nitric oxide as an intermediate in innate immune signaling and

are nocturnal, whereas humans acquire vitamin D from exposed skin during the daytime (50).

Taken together, the interplay between 1,25D and TLR signaling and the direct induction by 1,25D of AMP gene expression provide a strong molecular basis for epidemiological evidence documenting the protective effects of a vitamin D-replete state against infectious diseases. These factors also underline a growing consensus among researchers (11, 38, 40, 93) that the widespread vitamin D insufficiency/deficiency observed in North American and European populations strongly supports revising upward the recommendations for adequate daily intake of vitamin D (currently 200 IU for children and 400 to 600 IU for adults in these populations) and possibly extending vitamin D supplementation beyond dairy products, as is now practiced in the United States.

REFERENCES

- Adams, J. S., O. P. Sharma, M. A. Gacad, and F. R. Singer. 1983. Metabolism of 25-hydroxyvitamin D₃ by cultured pulmonary alveolar macrophages in sarcoidosis. *J. Clin. Invest.* **72**:1856–1860.
- Adorini, L. 2003. Tolerogenic dendritic cells induced by vitamin D receptor ligands enhance regulatory T cells inhibiting autoimmune diabetes. *Ann. N. Y. Acad. Sci.* **987**:258–261.
- Adorini, L. 2005. Intervention in autoimmunity: The potential of vitamin D receptor agonists. *Cell. Immunol.* **233**:115–124.
- Akutsu, N., R. Lin, Y. Bastien, A. Bestawros, D. J. Enepekides, M. J. Black, and J. H. White. 2001. Regulation of gene expression by 1 α ,25-dihydroxyvitamin D₃ and its analog EB1089 under growth inhibitory conditions in squamous carcinoma cells. *Mol. Endocrinol.* **15**:1127–1139.
- Alroy, I., T. Towers, and L. P. Freedman. 1995. Transcriptional repression of the interleukin-2 gene by vitamin D₃: direct inhibition NFATp/AP-1 complex formation by a nuclear hormone receptor. *Mol. Cell. Biol.* **15**:5789–5799.
- Andersen, R., C. Molgaard, L. T. Skovgaard, C. Brot, K. D. Cashman, E. Chabros, J. Charzewska, A. Flynn, J. Jakobsen, M. Karkkainen, M. Kiely, C. Lamberger-Allardt, O. Moreiras, A. M. Natri, M. O'Brien, M. Rogalska-Niedzwiedz, and L. Ovesen. 2005. Teenage girls and elderly women living in northern Europe have low winter vitamin D status. *Eur. J. Clin. Nutr.* **59**:533–541.
- Barber, Y., C. Rubio, E. Fernandez, M. Rubio, and J. Fibla. 2001. Host genetic background at CCR5 chemokine receptor and vitamin D receptor loci and human immunodeficiency virus (HIV) type 1 disease progression among HIV-seropositive injection drug users. *J. Infect. Dis.* **184**:1279–1288.
- Bellamy, R., C. Ruwende, T. Corrah, K. P. W. J. McAdam, M. Thursz, H. C. Whittle, and A. V. S. Hill. 1999. Tuberculosis and chronic hepatitis B virus infection in Africans and variation in the vitamin D receptor gene. *J. Infect. Dis.* **179**:721–724.
- Bergman, P., L. Walter-Jallow, K. Broliden, B. Agerberth, and J. Soderlund. 2007. The antimicrobial peptide LL-37 inhibits HIV-1 replication. *Curr. HIV Res.* **5**:410–415.
- Bhalla, A. K., E. P. Amento, B. Serog, and L. H. Glimcher. 1984. 1,25-Dihydroxyvitamin D₃ inhibits antigen-induced T cell activation. *J. Immunol.* **133**:1748–1754.
- Bischoff-Ferrari, H. A., E. Giovannucci, W. C. Willett, T. Dietrich, and B. Dawson-Hughes. 2006. Estimation of optimal serum concentrations of 25-hydroxyvitamin D for multiple health outcomes. *Am. J. Clin. Nutr.* **84**:18–28.
- Brennan, A., D. R. Katz, J. D. Nunn, S. Barker, M. Hewison, L. J. Fraher, and J. L. O'Riordan. 1987. Dendritic cells from human tissues express receptors for the immunoregulatory vitamin D₃ metabolite, dihydroxycholecalciferol. *Immunology* **61**:457–461.
- Brightbill, H., D. H. Libraty, S. R. Krutzik, R. B. Yang, J. T. Belisle, J. R. Bleharski, M. Maitland, M. V. Norgard, S. E. Plevy, S. T. Smale, P. J. Brennan, B. R. Bloom, P. J. Godowski, and R. L. Modlin. 1999. Host defense mechanisms triggered by microbial lipoproteins through Toll-like receptors. *Science* **285**:732–736.
- Brook, I. 2005. Microbiology and antimicrobial management of sinusitis. *J. Laryngol. Otol.* **119**:251–258.
- Brown, S. J. 2006. The role of vitamin D in multiple sclerosis. *Ann. Pharmacother.* **40**:1158–1161.
- Cannell, J. J., R. Vieth, J. C. Umhau, M. F. Holick, W. B. Grant, S. Madronich, C. F. Garland, and E. Giovannucci. 2006. Epidemic influenza and vitamin D. *Epidemiol. Infect.* **134**:1129–1140.
- Cannell, J. J., M. Zaslloff, C. F. Garland, R. Scragg, and E. Giovannucci. 2008. On the epidemiology of influenza. *Virology* **5**:Art 29.
- Carretero, M., M. J. Escamez, M. Garcia, B. Duarte, A. Holguin, L. Reta-

- mosa, J. L. Jorcano, M. del Rio, and F. Larcher. 2008. In vitro and in vivo wound healing-promoting activities of human cathelicidin LL-37. *J. Invest. Dermatol.* **128**:223–236.
19. Chapuy, M. C., P. Preziosi, M. Maamer, S. Arnaud, P. Galan, S. Hercberg, and P. J. Meunier. 1997. Prevalence of vitamin D insufficiency in an adult normal population. *Osteoporos. Int.* **7**:439–443.
 20. Chawla, A., J. Repa, R. M. Evans, and D. J. Mangelsdorf. 2001. Nuclear receptors and lipid physiology: opening the X-files. *Science* **294**:1866–1870.
 21. Cheng, J. B., M. A. Levine, N. H. Bell, D. J. Mangelsdorf, and D. W. Russell. 2004. Genetic evidence that the human CYP2R1 enzyme is a key vitamin D 25-hydroxylase. *Proc. Natl. Acad. Sci. USA* **101**:7711–7715.
 22. Davies, P. D. O., R. C. Brown, and J. S. Woodhead. 1985. Serum concentrations of vitamin D metabolites in untreated tuberculosis. *Thorax* **40**:187–190.
 23. De la Torre, M. S., C. Torres, G. Nieto, S. Vergara, A. S. Carrero, J. Macia, J. A. Pineda, A. Caruz, and J. Fibla. 2008. Vitamin D receptor gene haplotypes and susceptibility to HIV-1 infection in injection drug users. *J. Infect. Dis.* **197**:405–410.
 24. Dilworth, F. J., and P. Chambon. 2001. Nuclear receptors coordinate the activities of chromatin remodeling complexes and coactivators to facilitate initiation of transcription. *Oncogene* **20**:3047–3054.
 25. Dorschner, R. A., V. K. Pestonjamas, S. Tamakuwala, T. Ohtake, J. Rudisill, V. Nizet, B. Agerberth, G. H. Gudmundsson, and R. L. Gallo. 2001. Cutaneous injury induces the release of cathelicidin anti-microbial peptides active against group A streptococcus. *J. Invest. Dermatol.* **117**:91–97.
 26. Drocourt, L., J. C. Ourlin, J. M. Pascucci, P. Maurel, and M. J. Vilarem. 2002. Expression of CYP3A4, CYP2B6, and CYP2C9 is regulated by the vitamin D receptor pathway in primary human hepatocytes. *J. Biol. Chem.* **277**:25125–25132.
 27. Evans, K. N., H. Taylor, D. Zehnder, M. D. Kilby, J. N. Bulmer, F. Shah, J. S. Adams, and M. Hewison. 2004. Increased expression of 25-hydroxyvitamin D-1 alpha-hydroxylase in dysgerminomas—a novel form of humoral hypercalcemia of malignancy. *Am. J. Pathol.* **165**:807–813.
 28. Evans, K. N., L. Nguyen, J. Chan, B. A. Innes, J. N. Bulmer, M. D. Kilby, and M. Hewison. 2006. Effects of 25-hydroxyvitamin D-3 and 1,25-dihydroxyvitamin D-3 on cytokine production by human decidual cells. *Biol. Reprod.* **75**:816–822.
 29. Fitness, J., K. Tosh, and A. V. S. Hill. 2002. Genetics of susceptibility to leprosy. *Genes Immun.* **3**:441–453.
 30. Fitness, J., S. Floyd, D. K. Warndorff, L. Sichali, L. Mwaungulu, A. C. Crampin, P. E. Fine, and A. V. S. Hill. 2004. Large-scale candidate gene study of leprosy susceptibility in the Karonga district of northern Malawi. *Am. J. Trop. Med. Hyg.* **71**:330–340.
 31. Glass, C. K., and M. G. Rosenfeld. 2000. The coregulator exchange in transcriptional functions of nuclear receptors. *Genes Dev.* **14**:121–141.
 32. Gombart, A. F., N. Borregaard, and H. P. Koefler. 2005. Human cathelicidin antimicrobial peptide (CAMP) gene is a direct target of the vitamin D receptor and is strongly up-regulated in myeloid cells by 1,25-dihydroxyvitamin D-3. *FASEB J.* **19**:1067–1077.
 33. Grange, J. M., P. D. O. Davies, R. C. Brown, J. S. Woodhead, and T. Kardjito. 1985. A study of vitamin D levels in Indonesian patients with untreated pulmonary tuberculosis. *Tubercle* **66**:187–191.
 34. Grant, W. B. 2002. An estimate of premature cancer mortality in the U.S. due to inadequate doses of solar ultraviolet-B radiation. *Cancer* **94**:1867–1875.
 35. Hancock, R. E. W., and H. G. Sahl. 2006. Antimicrobial and host-defense peptides as new anti-infective therapeutic strategies. *Nat. Biotechnol.* **24**:1551–1557.
 36. Heilborn, J. D., M. F. Nilsson, G. Kratz, G. Weber, O. Sorensen, N. Borregaard, and M. Stahle-Backdahl. 2003. The cathelicidin anti-microbial peptide LL-37 is involved in re-epithelialization of human skin wounds and is lacking in chronic ulcer epithelium. *J. Invest. Dermatol.* **120**:379–389.
 37. Holick, M. F., E. S. Siris, N. Binkley, M. K. Beard, A. Khan, J. T. Katzner, R. A. Petruschke, E. L. Chen, and A. E. de Papp. 2005. Prevalence of vitamin D inadequacy among postmenopausal North American women receiving osteoporosis therapy. *J. Clin. Endocrinol. Metab.* **90**:3215–3224.
 38. Holick, M. F. 2006. Resurrection of vitamin D deficiency and rickets. *J. Clin. Invest.* **116**:2062–2072.
 39. Holick, M. F. 2006. High prevalence of vitamin D inadequacy and implications for health. *Mayo Clin. Proc.* **81**:353–373.
 40. Holick, M. F. 2007. Vitamin D deficiency. *N. Engl. J. Med.* **357**:266–281.
 41. Iannuzzi, M. C., B. A. Rybicki, and A. S. Teirstein. 2007. Medical progress: sarcoidosis. *N. Engl. J. Med.* **357**:2153–2165.
 42. Jenssen, H., P. Hamill, and R. E. W. Hancock. 2006. Peptide antimicrobial agents. *Clin. Microbiol. Rev.* **19**:491–511.
 43. Jones, G., S. A. Strugnell, and H. F. DeLuca. 1998. Current understanding of the molecular actions of vitamin D. *Physiological Rev.* **78**:1193–1231.
 44. Kawarau, A., E. Takeda, N. Tanida, K. Nakagawa, H. Yamamoto, K. Sawada, and T. Okano. 2006. Inhibitory effect of long term 1-alpha-hydroxyvitamin D3 administration on *Helicobacter pylori* infection. *J. Clin. Biochem. Nutr.* **38**:103–106.
 45. Kim, S., M. Yamazaki, L. A. Zella, N. K. Shevde, and J. W. Pike. 2006. Activation of receptor activator of NF-kappa B ligand gene expression by 1,25-dihydroxyvitamin D-3 is mediated through multiple long-range enhancers. *Mol. Cell. Biol.* **26**:6469–6486.
 46. Laaksi, I., J.-P. Ruohola, P. Tuohimaa, A. Auvinen, R. Haataja, H. Pihlajamäki, and T. Ylikomi. 2007. An association of serum vitamin D concentrations <40 nmol/L with acute respiratory tract infection in young Finnish men. *Am. J. Clin. Nutr.* **86**:714–717.
 47. Lim, W.-C., S. B. Hanauer, and Y. C. Li. 2005. Mechanisms of disease: vitamin D and inflammatory bowel disease. *Nat. Clin. Pract. Gastroenterol. Hepatol.* **2**:308–315.
 48. Lin, R., Y. Nagai, R. Sladek, Y. Bastien, J. Ho, K. Petrecca, G. Sotiropoulou, E. P. Diamandis, T. Hudson, and J. H. White. 2002. Expression profiling in squamous carcinoma cells reveals pleiotropic effects of vitamin D3 signaling on cell proliferation, differentiation and immune system regulation. *Mol. Endocrinol.* **16**:1243–1256.
 49. Lin, R., and J. H. White. 2004. The pleiotropic actions of vitamin D. *BioEssays* **26**:21–28.
 50. Liu, P. T., S. Stenger, H. Y. Li, L. Wenzel, B. H. Tan, S. R. Krutzik, M. T. Ochoa, J. Schaubert, K. Wu, C. Meinken, D. L. Kamen, M. Wagner, R. Bals, A. Steinmeyer, U. Zugel, R. L. Gallo, D. Eisenberg, M. Hewison, B. W. Hollis, J. S. Adams, B. R. Bloom, and R. L. Modlin. 2006. Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science* **311**:1770–1773.
 51. Liu, P. T., S. R. Krutzik, and R. L. Modlin. 2007. Therapeutic implications of the TLR and VDR partnership. *Trends Mol. Med.* **13**:117–124.
 52. Liu, P. T., S. Stenger, D. H. Tang, and R. L. Modlin. 2007. Cutting edge: vitamin D-mediated human antimicrobial activity against *Mycobacterium tuberculosis* is dependent on the induction of cathelicidin. *J. Immunol.* **179**:2060–2063.
 53. Malabanan, A., I. E. Veronikis, and M. F. Holick. 1998. Redefining vitamin D insufficiency. *Lancet* **351**:805–806.
 54. Martineau, A. R., R. J. Wilkinson, K. A. Wilkinson, S. M. Newton, B. Kampmann, B. M. Hall, G. E. Packe, R. N. Davidson, S. M. Eldridge, Z. J. Maunsell, S. J. Rainbow, J. L. Berry, and C. J. Griffiths. 2007. A single dose of vitamin D enhances immunity to mycobacteria. *Am. J. Respir. Crit. Care Med.* **176**:208–213.
 55. Matsuoka, L. Y., J. Wortsman, J. G. Haddad, P. Kolm, and B. W. Hollis. 1991. Racial pigmentation and the cutaneous synthesis of vitamin D. *Arch. Dermatol.* **127**:536–538.
 56. McKenna, N. J., and B. W. O'Malley. 2002. Combinatorial control of gene expression by nuclear receptors and coregulators. *Cell* **108**:465–474.
 57. Montone, K. T. 2007. Infectious diseases of the head and neck: a review. *Am. J. Clin. Pathol.* **128**:35–67.
 58. Muhe, L., S. Lulseged, K. E. Mason, and E. A. F. Simoes. 1997. Case-control study of the role of nutritional rickets in the risk of developing pneumonia in Ethiopian children. *Lancet* **349**:1801–1804.
 59. Munger, K. L., L. I. Levin, B. H. Hollis, N. S. Howard, and A. Ascherio. 2006. Serum 25-hydroxyvitamin D levels and risk of multiple sclerosis. *JAMA* **296**:2832–2838.
 60. Nesby-O'Dell, S., K. S. Scanlon, M. E. Cogswell, C. Gillespie, B. W. Hollis, A. C. Looker, C. Allen, C. Dougherty, E. W. Gunter, and E. A. Bowman. 2002. Hypovitaminosis D prevalence and determinants among African American and white women of reproductive age: Third National Health and Nutrition Examination Survey, 1988–1994. *Am. J. Clin. Nutr.* **76**:187–192.
 61. Nevado, J., S. P. Tenbaum, A. L. Castillo, A. Sanchez-Pacheco, and A. Aranda. 2007. Activation of the human immunodeficiency virus type 1 long terminal repeat by 1-alpha,25-dihydroxyvitamin D3. *J. Mol. Endocrinol.* **38**:587–601.
 62. Norman, A. W. 2006. Minireview. Vitamin D receptor: new assignments for an already busy receptor. *Endocrinology* **147**:5542–5548.
 63. Oberg, F., J. Botlin, and K. Nilsson. 1993. Functional antagonisms between vitamin-D3 and retinoic acid in the regulation of CD14 and CD23 expression during monocytic differentiation of U-937 cells. *J. Immunol.* **150**:3487–3495.
 64. Overbergh, L., B. Decallonne, D. Valckx, A. Verstuyf, J. Depovere, J. Laureys, O. Rutgeerts, R. Saint-Arnaud, R. Bouillon, and C. Mathieu. 2000. Identification and immune regulation of 25-hydroxyvitamin D-1-alpha-hydroxylase in murine macrophages. *Clin. Exp. Immunol.* **120**:139–146.
 65. Overbergh, L., K. Stoffels, M. Waer, A. Verstuyf, R. Bouillon, and C. Mathieu. 2006. Immune regulation of 25-hydroxyvitamin D-1 alpha-hydroxylase in human monocytic THP1 cells: mechanisms of interferon-gamma-mediated induction. *J. Clin. Endocrinol. Metab.* **91**:3566–3574.
 66. Patil, A., A. L. Hughes, and G. Zhang. 2004. Rapid evolution and diversification of mammalian α -defensins as revealed by comparative analysis of rodent and primate genomes. *Physiol. Genomics* **20**:1–11.
 67. Prosser, D. E., and G. Jones. 2004. Enzymes involved in the activation and inactivation of vitamin D. *Trends Biochem. Sci.* **29**:664–673.
 68. Proud, D., S. P. Sanders, and S. Wiehler. 2004. Human rhinovirus infection induces airway epithelial cell production of human beta-defensin 2 both in vitro and in vivo. *J. Immunol.* **172**:4637–4645.

69. **Provvedini, D. M., C. D. Tsoukas, L. J. Deftos, and S. C. Manolagas.** 1983. 1,25-Dihydroxyvitamin D3 receptors in human leukocytes. *Science* **221**: 1181–1183.
70. **Rachez, C., and L. P. Freedman.** 2000. Mechanisms of gene regulation by vitamin D-3 receptor: a network of coactivator interactions. *Gene* **246**:9–21.
71. **Ren, S., L. Nguyen, S. Wu, C. Encinas, J. S. Adams, and M. Hewison.** 2005. Alternative splicing of vitamin D-24-hydroxylase: a novel mechanism for the regulation of extrarenal 1,25-dihydroxyvitamin D synthesis. *J. Biol. Chem.* **280**:20604–20611.
72. **Rochel, N., J. M. Wurtz, A. Mitschler, B. Klaholz, and D. Moras.** 2000. The crystal structure of the nuclear receptor for vitamin D bound to its natural ligand. *Mol. Cell* **5**:173–179.
73. **Rook, G. A., J. Steele, L. Fraher, S. Barker, R. Karmali, J. O’Riordan, and J. Stanford.** 1986. Vitamin D3, interferon, and control of proliferation of *Mycobacterium tuberculosis* by human monocytes. *Immunology* **57**:159–163.
74. **Rosenfeld, M. G., and C. K. Glass.** 2001. Coregulator codes of transcriptional regulation by nuclear receptors. *J. Biol. Chem.* **276**:36865–36868.
75. **Roth, D. E., G. Soto, F. Arenas, C. T. Bautista, J. Ortiz, R. Rodriguez, L. Cabrera, and R. H. Gilman.** 2004. Association between vitamin D receptor gene polymorphisms and response to treatment of pulmonary tuberculosis. *J. Infect. Dis.* **190**:920–927.
76. **Roth, D. E., A. B. Jones, C. Prosser, J. L. Robinson, and S. Vohra.** 2008. Vitamin D receptor polymorphisms and the risk of acute lower respiratory tract infection in early childhood. *J. Infect. Dis.* **197**:676–680.
77. **Roy, S., A. Frodsham, B. Saha, S. K. Hazra, C. G. Mascie-Taylor, and A. V. S. Hill.** 1999. Association of vitamin D receptor genotype with leprosy type. *J. Infect. Dis.* **179**:187–191.
78. **Sadeghi, K., B. Wessner, U. Laggner, M. Ploder, D. Tamandl, J. Friedl, U. Zügel, A. Steinmeyer, A. Pollak, E. Roth, G. Boltz-Nitulescu, and A. Spitler.** 2006. Vitamin D3 down-regulates monocyte TLR expression and triggers hyporesponsiveness to pathogen-associated molecular patterns. *Eur. J. Immunol.* **36**:361–370.
79. **Schauber J., R. A. Dorschner, A. B. Coda, A. S. Büchau, P. T. Liu, D. Kiken, Y. R. Helfrich, S. Kang, H. Z. Elalich, A. Steinmeyer, U. Zügel, D. D. Bikle, R. L. Modlin, and R. L. Gallo.** 2007. Injury enhances TLR2 function and antimicrobial peptide expression through a vitamin D-dependent mechanism. *J. Clin. Investig.* **117**:803–811.
80. **Schwalfenberg, G.** 2007. Not enough vitamin D for Canadians. *Can. Fam. Physician* **53**:841–854.
81. **Shinkyo, R., T. Sakaki, M. Kamakura, M. Ohta, and K. Inouye.** 2004. Metabolism of vitamin D by human microsomal CYP2R1. *Biochem. Biophys. Res. Commun.* **324**:451–457.
82. **Stead, W. W., J. W. Senner, W. T. Reddick, and J. P. Lofgren.** 1990. Racial differences in sensitivity to infection by *Mycobacterium tuberculosis*. *N. Engl. J. Med.* **322**:422–427.
83. **Steinstraesser, L., B. Tippler, L. Mertens, E. Lamme, H. H. Homann, M. Lehnhardt, O. Wildner, H. U. Steinau, and K. Überla.** 2005. Inhibition of early steps in the lentiviral replication cycle by cathelicidin host defense peptides. *Retrovirology* **2**:Art 2.
84. **Stoffels, K., L. Overbergh, A. Giullietti, L. Verlinden, R. Bouillon, and C. Mathieu.** 2006. Immune regulation of 25-hydroxyvitamin-D-3-1 alpha-hydroxylase in human monocytes. *J. Bone Miner. Res.* **21**:37–47.
85. **Takeuchi, A., G. Reddy, T. Kobayashi, T. Okano, J. Park, and S. Sharma.** 1998. Nuclear factor of activated T cells (NFAT) as a molecular target for 1,25-dihydroxyvitamin D3-mediated effects. *J. Immunol.* **160**:209–218.
86. **Tavera-Mendoza, L., T. T. Wang, B. Lallemand, R. Zhang, Y. Nagai, V. Bourdeau, M. Ramiro Calderon, J. Desbarats, S. Mader, and J. H. White.** 2006. Convergence of vitamin D and retinoic acid signaling at a common hormone response element. *EMBO Rep.* **7**:180–185.
87. **Thomas, M. K., D. M. Lloyd-Jones, R. I. Thadhani, A. C. Shaw, D. J. Deraska, B. T. Kitch, E. C. Vamvakas, I. M. Dick, R. L. Prince, and J. S. Finkelstein.** 1998. Hypovitaminosis D in medical inpatients. *N. Engl. J. Med.* **338**:777–783.
88. **Thoma-Uszynski, S., S. Stenger, O. Takeuchi, M. T. Ochoa, M. Engele, P. A. Sieling, P. F. Barnes, M. Rollinghoff, P. L. Bolcskei, M. Wagner, S. Akira, M. V. Norgard, J. T. Belisle, J. T. Godowski, B. R. Bloom, and R. L. Modlin.** 2001. Induction of direct antimicrobial activity through mammalian Toll-like receptors. *Science* **291**:1544–1547.
89. **Thompson, P. D., P. W. Jurutka, G. K. Whitfield, S. M. Myskowski, K. R. Eichhorst, C. E. Dominguez, C. A. Haussler, and M. R. Haussler.** 2002. Liganded VDR induces CYP3A4 in small intestinal and colon cancer cells via DR3 and ER6 vitamin D responsive elements. *Biochem. Biophys. Res. Commun.* **299**:730–738.
90. **Thummel, K. E., C. Brimer, K. Yasuda, J. Thottassery, T. Senn, Y. Lin, H. Ishizuka, E. Kharasch, J. Schuetz, and E. Schuet.** 2001. Transcriptional control of intestinal cytochrome P-4503A by 1 α ,25-dihydroxyvitamin D3. *Mol. Pharmacol.* **60**:1399–1406.
91. **Uitterlinden, A. G., Y. Fang, J. B. J. van Meurs, H. A. P. Pols, and J. P. T. M. van Leeuwen.** 2004. Genetics and biology of vitamin D receptor polymorphisms. *Gene* **338**:143–156.
92. **Van Etten, E., and C. Mathieu.** 2005. Immunoregulation by 1,25-dihydroxyvitamin D3: basic concepts. *J. Ster. Biochem. Mol. Biol.* **97**:93–101.
93. **Vieth, R.** 2004. Why the optimal requirement for vitamin D3 is probably much higher than what is officially recommended for adults. *J. Steroid Biochem. Mol. Biol.* **89–90**:575–579.
94. **Villamor, E.** 2006. A potential role for vitamin D on HIV infection? *Nutr. Rev.* **64**:226–233.
95. **Wang, T. T., F. Nestel, V. Bourdeau, Y. Nagai, Q. Wang, J. Wu, L. Tavera-Mendoza, R. Lin, J. W. Hanrahan, S. Mader, and J. H. White.** 2004. Cutting edge: 1,25-dihydroxyvitamin D3 is a direct inducer of antimicrobial peptide gene expression. *J. Immunol.* **173**:2909–2912.
96. **Wang, T. T., L. Tavera-Mendoza, D. Laperriere, Y. Nagai, N. Burton MacLeod, E. Libby, R. Zhang, V. Bourdeau, A. Konstorum, B. Lallemand, S. Mader, and J. H. White.** 2005. Large-scale in silico and microarray-based genomic screening of 1,25-dihydroxyvitamin D₃ target genes. *Mol. Endocrinol.* **19**:2685–2695.
97. **Wayse, V., A. Yousafzai, K. Mogale, and S. Filteau.** 2004. Association of subclinical vitamin D deficiency with severe acute lower respiratory infection in Indian children under 5 y. *Eur. J. Clin. Nutr.* **58**:563–567.
98. **Weber, G., J. D. Heilborn, C. I. Chamorro Jimenez, A. Hammarsjö, H. Törmä, and M. Stähle.** 2005. Vitamin D induces the antimicrobial protein hCAP18 in human skin. *J. Investig. Dermatol.* **124**:1080–1082.
99. **Wehkamp, J., J. Schaubert, and E. F. Stange.** 2007. Defensins and cathelicidins in gastrointestinal infections. *Curr. Opin. Gastroenterol.* **23**:32–38.
100. **White, J. H.** 2004. Profiling 1,25-dihydroxyvitamin D3-regulated gene expression by microarray analysis. *J. Steroid Biochem. Mol. Biol.* **89–90**:239–244.
101. **Wilkinson, R. J., M. Llewelyn, Z. Toossi, P. Patel, G. Pasvol, A. Lalvani, D. Wright, M. Latif, and R. N. Davidson.** 2000. Influence of vitamin D deficiency and vitamin D receptor polymorphisms on tuberculosis amongst Gujarati Asians in West London: a case-control study. *Lancet* **355**:618–621.
102. **Wolf, J., and A. J. Daley.** 2007. Microbiological aspects of bacterial lower respiratory tract illness in children: atypical pathogens. *Paediatr. Respir. Rev.* **8**:212–220.
103. **Zanetti, M.** 2004. Cathelicidins: multifunctional peptides of the innate immunity. *J. Leukoc. Biol.* **75**:39–48.