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Synthesis, Characterization and Application of Novel Chiral Ionic Liquids and their Polymers in Micellar Electrokinetic Chromatography

Syed Asad Ali Rizvi and Shahab A. Shamsi*

Department of Chemistry Center of Biotechnology and Drug Design Georgia State University Atlanta, GA 30303

Abstract

Two amino acid derived (leucinol and N-methyl pyrrolidinol) chiral ionic liquids are synthesized and characterized both in monomeric and polymeric forms. Leucinol based chiral cationic surfactant is room a temperature ionic liquid (RTIL), and pyrrolidinol based chiral cationic surfactant melts at 30-35 °C to form ionic liquid (IL). The monomeric and polymeric ILs are thoroughly characterized to determine critical micelle concentration, aggregation number, polarity, optical rotation and partial specific volume. Here in, we present the first enantioseparation using chiral IL as pseudostationary phase in capillary electrophoresis. Chiral separation of two acidic analytes, (±)-alpha-bromophenylacetic acid (\pm) - $(\alpha$ -BP-AA) and (\pm) -2-(2-chlorophenoxy)propanoic acid (\pm) -(2-PPA) can be achieved with both monomers and polymers of undecenoxy carbonyl-L-pryrrolidinol bromide (L-UCPB) and undecenoxy carbonyl-L-leucinol bromide (L-UCLB) at 25 mM surfactant concentration using phosphate buffer at pH 7.50. The chiral recognition seems to be facilitated by the extent of interaction of the acidic analytes with the cationic head group of chiral selectors. Polysodium Nundecenoxy carbonyl-L-leucine sulfate (poly-L-SUCLS) and polysodium N-undecenoxy carbonyl-L-leucinate (poly-L-SUCL) were compared at high and low pH for the enantioseparation of (±)-(2-PPA). At pH 7.5, poly-L-SUCLS, poly-L-SUCL and ±-2-PPA are negatively charged resulting in no enantioseparation. However, chiral separation was observed for (±)-(2-PPA) using poly-L-SUCLS at low pH (pH 2.00) at which analyte is neutral. The comparison of chiral separation of anionic and cationic surfactants demonstrates that the electrostatic interaction between the acidic analyte and cationic micelle plays a profound role in enantioseparation.

The separation of chiral compounds is currently the center of great interest. This interest can be attributed largely to a heightened awareness that enantiomers of a racemic drug usually display markedly different pharmacological activities. The human body metabolizes individual enantiomers by separate pathways to produce different pharmacological effects. Presently, a majority of commercially available drugs are synthetic and chiral. Most of these chiral drugs are obtained as a mixture of two enantiomers during synthesis. In order to avoid the possible undesirable effects of enantiomeric impurity in chiral drug, it is inevitable that only therapeutically active form be marketed. Hence there is a continuous need to develop technologies that have the ability to separate enantiomers.

Very recently, ionic liquids (ILs) have found great applications in efficient and environmentally benign chemical processing and chemical analysis. ^{5,6} By definition the ionic liquids (ILs) are organic salts with melting points (MP) below 100 °C or more often even lower than room

^{*}Corresponding author: Professor Shahab A. Shamsi, Department of Chemistry, Center for Biotechnology and Drug Design, Georgia state University, Atlanta, GA 30303, USA Phone: 404-651-1297; Fax: 404-651-2751 Email: chesas@langate.gsu.edu

temperature.⁷⁻¹¹ These compounds posses dual capability of dissolving both polar and nonpolar species and the most useful feature is that they do not evaporate even at high temperatures. ¹²⁻¹⁵ Most commonly, ILs are based on nitrogen-rich alkyl substituted heterocyclic cations, with a variety of anions (e.g., 1-ethyl-3-methylimidazolium tetrafluoroborate). Although, the reasons for low melting point of ILs are not clear, it is stated that, ILs consist of bulky inorganic anions with delocalized charged organic cations, which prevents the formation of a stable crystal lattice or random molecular packing resulting in lower melting points. ¹⁶ Due to these remarkable characteristics, ionic liquids have been used as, medium for liquid-liquid extractions, ¹⁷⁻¹⁹ mobile phase additives in high performance liquid chromatography (HPLC), ^{20,21} electrolytes in capillary electrophoresis (CE), ²²⁻²⁶ matrixes for matrix assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF MS), ^{27,28} stationary phases for gas chromatography (GC)²⁹⁻³² and as modifiers in micellar electrokinetic chromatography (MEKC). ^{33,34} However, there is no report in the literature about the use of ILs as chiral selector in CE.

Cationic surfactants are referred to as compounds containing at least one long hydrophobic chain attached to a positively charged nitrogen. These quaternary ammonium group containing surfactants are well known for displaying emulsifying properties, antimicrobial activity, components in cosmetic formulations, anti corrosive effects, phase transfer catalyst and as chiral induction medium (if chiral cationic surfactant) in organic reactions. ³⁵⁻⁴¹ As with the case of chiral anionic surfactants, amino acid based (both monomeric and polymeric) and ephedrine based (monomeric) chiral cationic surfactants have been used as chiral selectors in MEKC. ^{42,43} However, unlike chiral anionic polymeric surfactants, chiral cationic polymeric surfactants have not found great application so far, and only one report of chiral cationic polymeric surfactants as pseudostationary phase (PSP) in MEKC is reported. ⁴²

In this study, we report the synthesis, characterization and application of novel IL type surfactants and their polymers for chiral separation of acidic analytes in MEKC. Acidic analytes due to inherent negative charge poorly interact with most commonly employed chiral anionic surfactants at basic pH. As a result, still a large number of acidic analytes could not be resolved by MEKC. The cationic surfactant, undecenoxy carbonyl-L-leucinol bromide (L-UCLB) is an ionic liquid at room temperature, while undecenoxy carbonyl-L-pyrrolidinol bromide (L-UCPB) is a greasy solid that melts to form ionic liquid at 30-35 °C. In our case, quarternized nitrogen (chiral head group) is surrounded by hydrophobic tail and leucinol or pyrrolidinol side chain, which presumably prevent the proper packing of the cations and anions in regular three-dimensional patterns to form ionic liquids.

Current report is the first demonstration of MEKC chiral separation of several anionic compounds such as phenoxypropionic acid herbicide (\pm)-(2-PPA) and a very useful synthetic intermediate \pm -alpha-bromophenylacetic acid (\pm)-(α -BP-AA)^{44,45} using two synthetic chiral ionic liquids L-UCLB and L-UCPB as well their polymers. Chiral separation of acidic analyte is compared using polymeric anionic surfactants containing similar head group under both acidic and basic pH conditions.

MATERIALS AND METHODS

Standards and Chemicals

The analytes (\pm)-alpha-bromophenylacetic acid (\pm)-(α -BP-AA) and (\pm)-2-(2-chlorophenoxy) propanoic acid (\pm)-(2-PPA) were obtained as racemic mixture from Sigma Chemical Co (St. Louis, MO) and Aldrich (Milwaukee, WI), respectively. Chemicals used for the synthesis of surfactants included ω -undecylenyl alcohol, triphosgene, pyridine, dichloromethane, 2-bromoethylamine hydrobromide, L-leucinol, *N*-methylpryrrolidinol, 96% formic acid, 37%

formaldehyde, and isopropanol (HPLC grade) were also obtained from Aldrich (Milwaukee, WI) and were used as received.

Synthesis and Characterization of Monomeric Surfactants and Micelle Polymers

Choloroformate has been synthesized as reported earlier 46 by reacting triphosgene with unsaturated alcohol (step 1, Fig 1). The carbamate functionalized alkenyl bromide (step 2, Fig 1) was synthesized by dropwise addition of (10 mmoles) choloroformate over equimolar aqueous solution of 2-bromoethylamine hydrobromide and Na₂CO₃ and were stirred for 2 hrs. The resulting solution was extracted twice with dichloromethane, which then was washed three times with H₂O, dried over Na₂SO₄ and concentrated by evaporating solvent to yield product 1(89-93%). The N,N-dimethyl leucinol (product 2, step B, Fig 1) was synthesized by reductive alkylation of primary amine of leucinol using the well-known Eschweiler-Clark reaction (Yield 55-70%).⁴⁷⁻⁴⁹ The chiral ionic liquids were synthesized by refluxing the carbamate functionalized alkenyl bromide (product 1) with N,N-dimethyl leucinol or Nmethylpryrrolidinol for 48 hrs in isopropanol (IPA). After 48 hrs, the reaction mixture was concentrated by evaporating IPA, and the resulting fluid was dissolved in water and extracted with ethyl acetate. The aqueous solution of ionic liquids (products 3 and 4, Figure 2) were lyophilized (Yield 40-55%) at -50 °C collector temperature and 0.05 mBar pressure for 14 days (to ensure complete removal of water from both products). ¹H-NMR spectra of L-UCPB, L-UCLB and their polymers were recorded on a Varian Unity+ 300 MHz spectrometer using D₂O as the solvent. The surfactants were characterized and checked for purity by MALDI-TOF-MS (Fig 3A-B), ¹H-NMR and elemental analysis. **L-UCPB**, ¹H-NMR: δ 0.759-0.893 (b, 6H), 1.170 (m, 12H), 1.471 (m, 2H), 1.767 (m, 2H), 1.883 (b, 1H), 2.085 (m, 2H), 3.06 (b, 2H), 3.239-3.613 (b, 8H), 3.777-3.844 (m, 1H), 4.052 (d, *J*= 14.7, 2H), 4.379 (b, 2H), 4.789 (m, 2H), 5.626 (m, 1H). Anal. Calcd. for $C_{20}H_{39}N_2O_3Br + 2H_2O$: C, 50.95; H, 9.19; N, 5.94; Found: C, 51.56; H, 10.07; N, 5.88. **L-UCLB**, ¹H-NMR: δ 1.170 (b, 12H), 1.468 (b, 2H), 1.766-1.992 (b, 4H), 1.992-2.164 (b, 2H), 3.032 (b, 2H), 3.147 (b, 2H), 3.472 (b, 3H), 3.506-3.619 (b, 3H), 3.830 (b, 2H), 4.376 (b, 2H), 4.804 (m, 2H), 5.658 (m, 1H). Anal. Calcd. for C₂₂H₄₅N₂O₃Br + 2H₂O: C, 52.68; H, 9.85; N, 5.59; Found: C, 52.14; H, 9.00; N, 6.87.

The critical micelle concentration (CMC) was determined using a sigma 7⁰³ Digital Tensiometer (KVS Instruments USA, Monroe, Connecticut), by the Du NoÜy ring method at room temperature. Polymerization of the synthesized ionic liquids was achieved by ⁶⁰Co γirradiation (8 Mrad/h) of 100 mM aqueous solution for 30 hrs. To ensure complete polymerization, 30 hrs of continuous γ-irradiation was applied. The ¹H-NMR indicated the disappearance of double bond protons signal in the region of 4.8-5.0 and 5.7-5.9 ppm. After irradiation, the polymeric surfactant solutions were filtered and dialyzed against triply deionized water using regenerated cellulose (RC) dialysis membrane (Spectrum Laboratories, Inc, Rancho Dominguez, CA, USA) with a 1000 Da molecular mass cutoff for 24 hrs. Finally, the dialyzed solutions were lyophilized to obtain the dried polymeric surfactants. Further characterization, such as aggregation number and polarity of the amphiphilic ionic liquids (monomers and polymers) were determined by using pyrene emission vibronic fine structure method. 46 The partial specific volume of both monomer and polymer was determined using previously reported procedure. 46 The optical rotation of monomeric and the polymeric surfactants was obtained by an AUTOPOL III automatic polarimeter (Rudolph Research Analytical, Flanders, New Jersey) by measuring the optical rotation at 589 nm of a 10 mg/mL solution of each monomer and polymer in triply deionized water at 25 °C.

MEKC Instrumentation

All experiments were performed using an Agilent CE system (Agilent Technologies, Palo Alto, California) equipped with 0-30 kV high-voltage power supply, a diode array detector for UV detection and Chemstation software (V 9.0) for system control and data acquisition. The fused-

silica capillary was obtained from Polymicro Technologies (Phoenix, AZ). The total length of the capillary used with an Agilent CE system was $64.5~\rm cm$ ($56.0~\rm cm$ from inlet to detector, $50~\rm \mu m$ ID, $350~\rm \mu m$ OD), prepared by burning about 3 mm polyimide coating to create a detection window.

Capillary Electrophoresis Procedures and Calculations

The capillaries for all MEKC experiments were prepared by flushing with 1M NH₄OH for 60 min at 50 °C followed by 30 min rinse with triply deionized water at 20 °C. Between each injection, the capillary was flushed with 0.1 M NH₄OH and H₂O for 3 min each. Separations began after a 2 min rinse with the running buffer, followed by a 5 min flush with the running buffer containing ionic liquids. All separations were performed at -20 kV and at 20 °C. All surfactants (both monomers and polymers) were run with a new capillary (cut to the same length from the same capillary bundle) and was preconditioned using the identical flushing procedure as mentioned above. Chiral resolution (R_s) of acidic analytes (\pm)-(2-PPA) and (\pm)- α -BP-AA were calculated by Chemstation software using the peak width at half height method:

$$R_s = \frac{(2.35/2)(t_{r2} - t_{r1})}{W_{50(1)} + W_{50(2)}}$$

 $W_{50\;(1)}$ and $W_{50\;(2)}$ are the widths at 50% height for peak 1 and 2, respectively. The selectivity (α) is defined as t_2/t_1 , where t_1 and t_2 are the migration times of the first and second eluting enantiomers. Methanol was used as the t_0 marker and was measured from the time of injection to the first deviation from the baseline. Dodecanophenone was used as tracer for t_{mc} at 100 mM surfactants concentration of each monomer and polymer. The effective electrophoretic mobility of the monomers and polymers of ionic liquids was calculated by following equation:

$$\mu_{ep} = -\mu_{app} - \left(-\mu_{eof}\right)$$

Where, μ_{ep} , μ_{eof} and μ_{app} are effective electrophoretic mobility, electroosmotic mobility and apparent electrophoretic mobility respectively. The negative sign of μ_{app} and μ_{eof} is due to the fact that monomeric and polymeric ionic liquids coat the capillary wall and result in anodic electroosmotic flow, therefore negative voltage (-20 kV) has to be applied for separation.

Preparation of MEKC Buffers and Analyte Solutions

For all MEKC experiments, the final background electrolyte (BGE) consisted of a 25 mM each of Na₂HPO₄/NaH₂PO₄ buffered at pH 7.5. The desired pH value was obtained by using 1 M NaOH. The pH of BGE was adjusted before the addition of ILs (monomers and polymers). This BGE solution is finally filtered through a 0.45 µm Nalgene syringe filter (Rochester, NY). The running MEKC buffer solution was prepared by addition of 25 mM IL type surfactants to the BGE, followed by ultrasonication for about 25-30 min. The analytes prepared in 50/50 (v/v) of MeOH:H₂O at various concentrations were injected at a pressure of 50 mbar for 1-5s. The dodecanophenone was prepared in 100% MeOH at 3 mg/mL (stock solution), diluted to 1.8 mg/mL in 60:40 MeOH:H₂O and injected at a pressure of 50 mbar for 10s

RESULTS AND DISCUSSION

Physicochemical Properties

Table 1 represents the physicochemical properties of the synthesized enantiomerically pure chiral surfactants L-UCLB [room temperature ionic liquid (RTIL)] and L-UCPB [ionic liquid (IL), MP 30-35 °C] as well as their polymers, poly-L-UCLB and poly-L-UCPB. The L-UCLB exhibited higher polarity, lower CMC and partial specific volume (\bar{V}), significantly higher optical rotation but similar aggregation number (A) compared to L-UCPB. Similar trend was also observed for the poly-L-UCLB and poly-L-UCPB, except that the A value was higher for

the former polymer. Comparing physicochemical properties of monomeric and polymeric cationic surfactants, it can be noticed that A is always lower for the polymers than monomers, while polarity and V is always higher for polymeric surfactants.

Fig 3. shows the MALDI-TOF MS of both L-UCLB (A) and L-UCPB (B) in positive mode. Both L-UCLB and L-UCPB surfactants showed the molecular ion peak (base peak) at mass-to-charge ratios (m/z) of 385.3 and 355.3, respectively along with a fragment generated by the loss $C_5H_{11}O$. For L-UCLB the masses at 386.2 and 299.2 m/z, for L-UCPB the masses at 356.3 and 268.2 m/z are generated due to the ^{13}C isotope related to the molecular ion and the generated fragment ion, respectively. The generation of the cationic fragments (Z₂) for both ionic liquids as shown in Fig 3 is in accord with the previous observations that most of the fragments generated from cationic surfactants bear preformed cations (contain quaternary nitrogen of the cationic surfactants). 50,51

The electrophoretic parameters of monomeric and polymeric ionic liquids were also examined (Table 1) at 100 mM surfactant concentrations (at lower surfactant concentration t_{mc} marker was not observed even after 3 hrs). The reversed electroosmotic flow ($-\mu_{eof}$) and effective electrophoretic (μ_{ep}) mobilities of both poly-L-UCPB and poly-L-UCLB were slightly lower, while migration time window (t_{mc}/t_0) was greater compared to their respective monomers. In addition, the monomer and polymer of L-UCLB compared to the monomer and polymer of L-UCPB have lower μ_{ep} and provided larger t_{mc}/t_0 .

Enantioseparation of Acidic Analytes

The optimization of chiral resolution of (\pm) - $(\alpha$ -BP-AA) and (\pm) -(2-PPA) was performed by studying pH of the background electrolyte (BGE), type and concentration of BGE, organic modifiers and surfactant concentration. After optimizing the chiral MEKC conditions, chiral separation of (\pm) - $(\alpha$ -BP-AA) and (\pm) -(2-PPA) were compared using L-UCPB, L-UCLB and their respective polymers (poly-L-UCPB and poly-L-UCLB) to get insight on the factors affecting analyte-micelle interactions and ultimately chiral separation.

Enantioseparation of ±-alpha-bromophenylacetic acid

Figure 4(A) and 4(C) show the chiral separation of (\pm) - $(\alpha$ -BP-AA) at optimum separation conditions with L-UCPB and L-UCLB, respectively. Since (±)-(α-BP-AA) has dissociable carboxylic acid group with pKa = 2.40 ± 0.10) therefore, the effect of pH on enantioseparation was evaluated from pH 2.00-8.50 (data not shown). Although chiral resolution (R_s) at lower pH range (4.00-6.00) do not differ drastically, maximum R_s was obtained at pH 7.5 but no R_s at pH 2.00 and at pH > 7.5, R_s deteriorates (data not shown). One plausible explanation of R_s deterioration at pH > 7.5 could be the excess hydroxide ions (originated from the use of NaOH to adjust the BGE pH), which competes with the anionic chiral analyte for the positively charged ionic liquid at basic pH. The absence of any R_s at pH 2.00 and lower R_s at intermediate pH suggests that electrostatic interaction indeed contributes significantly to chiral recognition. It has been reported in the literature that in the presence of certain organized media (e.g., micelles), the pKa of the organic acid is altered up to more than 4 pH units. 43,52 Therefore, it is reasonable to believe that the amphiphilic ionic liquids might have increased the pKa of (\pm) - $(\alpha$ -BP-AA) such that maximum ionization occurs around pH 7.50. Hence, greater electrostatic interaction with the positively charged ionic liquids provided maximum chiral R_sat pH 7.50. On the other hand, L-UCPB and L-UCLB concentrations, as well as the use of organic modifiers (e.g., methanol, acetonitrile) did not show any significant variations in R_s

As depicted, L-UCPB provided almost twice as high chiral R_s for (\pm) - $(\alpha$ -BP-AA) compared to L-UCLB (Figure 4A vs. $\underline{4C}$). One possible explanation for enhanced chiral resolution provided by L-UCPB over L-UCLB could be due to the rigid ring system of L-UCPB, which

apparently allows maximum interaction via three-point interaction with \pm - α -Br-Ph-AA. The R_s trend is consistent to the findings of Thiobodeaux *et al.*, who observed that surfactants derived from L-proline (a rigid amino acid) provided better chiral separation for rigid chiral molecules (e.g., BNP). The analyte (\pm)-(α -BP-AA) has a chiral center, which is adjacent to a bromo group and a carboxylate group. Thus, it appears that the chiral recognition was greatly facilitated not only by electrostatic interactions between carboxylate group of the analyte and the positively charge nitrogen, but the presence of bromo group adjacent to the chiral center might also hydrogen bond with the –OH group of the ionic liquids (Figure 2).

Comparing the electropherograms in 4(A) vs. 4(C) and 4(B) vs. 4(D), it is obvious that monomers of both L-UCPB and L-UCLB provided better chiral resolution, selectivity and efficiency compared to the corresponding polymers. The probable reason behind this observation could be the polydispersity of the polymers, ⁵⁵ which usually is the case when surfactants are polymerized at concentration higher than the CMC. ^{55,56}

Enantioseparation of ±-2-(2-Chlorophenoxy)propanoic acid

As discussed above, in the case of (\pm)-(α -BP-AA) maximum chiral R_s was obtained at pH 7.50 and no R_s at pH 2.00. O'Keeffe *et al.*⁵⁷ and Haynes *et al.*⁵⁸ have reported the separation of (\pm)-(2-PPA) at pH 5.00-6.00 with a cationic substituted β -cyclodextrin. Similar to the case of (\pm)-(α -BP-AA) separation, the variation in surfactant concentration and addition of organic modifier showed no significant effects on chiral R_s of (\pm)-(2-PPA).

Figure 5(A) and 5(C) show the chiral separation of (\pm) -(2-PPA) at optimum MEKC parameters using L-UCPB and L-UCLB, respectively. The non-rigid leucine based (L-UCLB) chiral selector (Fig 5C) provided significantly higher chiral R_s of (\pm) -(2-PPA) than L-UCPB. This resolution trend is opposite to the separation of (\pm) -(α -BP-AA) (Fig 4A, 4C). As stated, the proximity of bromo and carboxylate group to the chiral center of (\pm) -(α -BP-AA) as well as the rigidity of the chiral selector was thought to be the key factors ensuring maximum enantioselectivity. However, in case of (\pm) -(2-PPA), the chloro group on the benzene ring is farther away from the chiral center. Furthermore, the non-rigidity of L-UCLB might have resulted in favorable hydrogen bonding interactions between the chloro group on the benzene ring and the primary alcohol of the L-leucinol. Comparing 5(A) vs. 5(B) and 5(C) vs. 5(D), it is clear that monomers and polymers of L-UCPB and L-UCLB show very similar stereoselective interactions with (\pm) -(2-PPA) as evident from the R_s and α values.

It is interesting to compare the enantioseparation capability between two polymeric chiral anionic surfactants [polysodium N-undecenoxy carbonyl-L-leucine sulfate (poly-L-SUCLS) and polysodium N-undecenoxy carbonyl-L-leucinate (poly-L-SUCL)] with the chiral cationic surfactants discussed earlier for racemic anionic analyte. The chiral separation of (\pm) -(2-PPA) with both sulfated and carboxylated head group polymeric surfactants were investigated at basic pH (Fig 6A-B). As we have mentioned earlier that anionic compounds are usually difficult to separate with anionic surfactant due to the electrostatic repulsion between similar charges. Hence, as expected no chiral resolution was obtained for (\pm) -(2-PPA) at pH 8.00. Since poly-L-SUCLS has sulfated head group, it can be used at any pH without any solubility problem. Therefore, we performed MEKC at pH 2.00 (Fig 6C) in order to minimize dissociation of carboxylic acid group of (\pm) -(2-PPA) (pKa 3.11 \pm 0.10). As can be seen in Fig 6C, partial chiral separation of (±)-(2-PPA) was achieved at pH 2.00. However, we could not improve this chiral R_s any further even after fine-tuning of the MEKC parameters (data not shown). Hence, comparing chiral separation of (±)-(2-PPA) with poly-L-UCLB (Fig 5D), poly-L-SUCLS (Fig 6A,C) and poly-L-SUCL (Fig 6B), it is clear that indeed electrostatic attraction interaction plays a dominant role in chiral recognition. In addition, structural features (e.g., rigidity and charges) of both analyte and chiral selector also seem to affect the chiral recognition.

CONCLUSIONS

This paper is the first demonstration of successful chiral separation of acidic analytes with synthetic chiral ILs in CE. Both L-UCLB and L-UCPB ionic liquid type surfactants were thoroughly characterized before and after the polymerization. It was found that chiral separation of the acidic analytes with the chiral ILs and their polymers is strongly dependent on the presence of opposite charge as well as the structural compatibility between chiral selector and the analyte. Even though we did not demonstrate the enantioseparation of large number of acidic analytes, we still believe that our findings will guide the future research in MEKC separation of acidic analytes with intelligently designed synthetic chiral ionic liquids.

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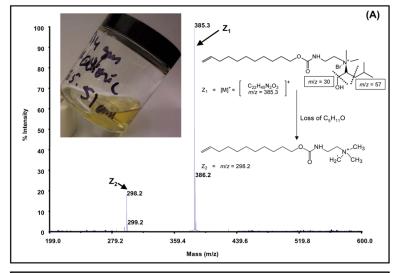
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Figure 1. Synthesis of the carbamate functionalized (A) alkenyl bromide and (B) *N*, *N*-dimethyl-L-leucinol.

Figure 2. Synthesis and polymerization of leucinol and pryrrolidinol derived ionic liquid and their polymers.



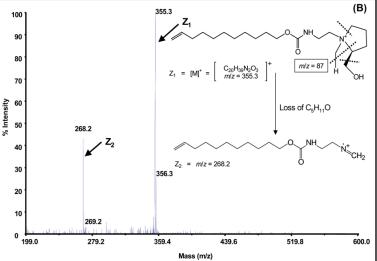


Figure 3. MALDI-TOF mass spectra (positive mode) with proposed cleavages and the corresponding fragment masses for IL type surfactants (A) undecenoxy carbonyl-L-leucinol bromide (L-UCLB) and (B) undecenoxy carbonyl-L-pryrrolidinol bromide (L-UCPB) after freeze drying on a lyophilizer at -50 °C collector temperature and 0.05 mbar pressure for 14 days. The mass spectra were obtained using α -cyano-4-hydroxycinnamic acid as MALDI-matrix.

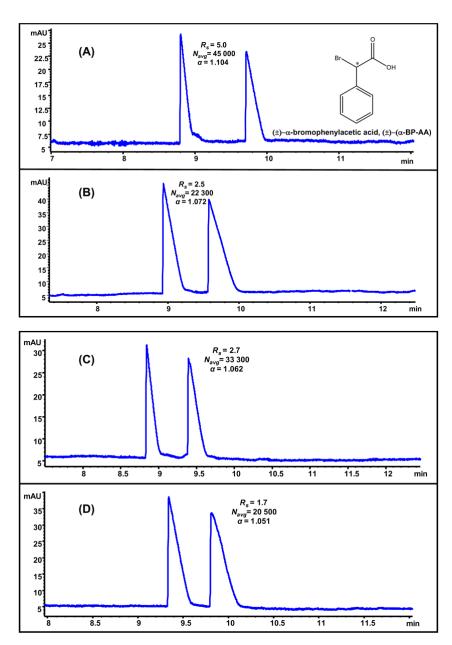


Figure 4. Comparison of 25 mM L-UCPB (A), poly-L-UCPB (B), 25 mM L-UCLB (C) and poly-L-UCLB (D) for enantioseparation of (\pm)-alpha-bromophenylacetic acid [(\pm)-(α -BP-AA), 2.5 mg/mL in MeOH/H₂O]. MEKC conditions: 50 mM NaH₂PO₄/ Na₂HPO₄, pH 7.5, pressure injection 50 mbar 5s, -20 kV, 20 °C, UV detection at 214 nm.

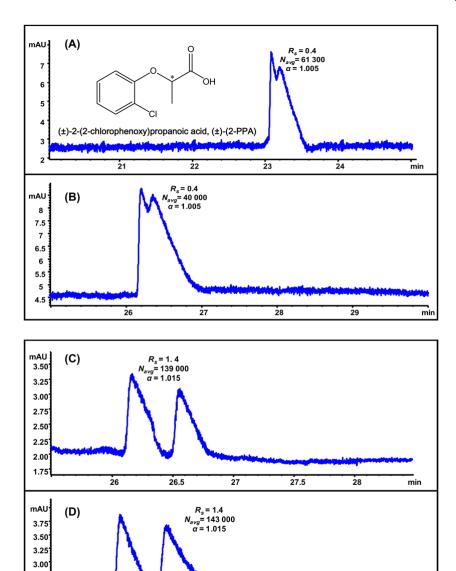


Figure 5. Comparison of 25 mM L-UCPB (A), poly-L-UCPB (B), 25 mM L-UCLB (C) and poly-L-UCLB (D) for enantioseparation of 2-(2-chlorophenoxy)propanoic acid[(\pm)-(2-PPA), 0.5 mg/mL in MeOH/H₂O]. MEKC conditions are same as in Fig. 4.

27

27.5

28

min

2.75

2.00

26

26.5

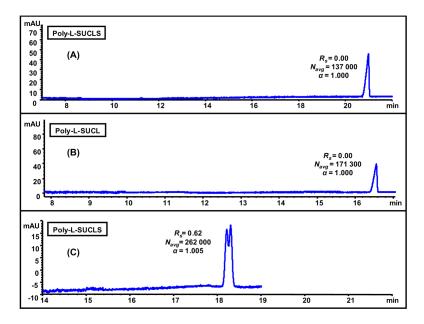


Figure 6. Comparison of 25 mM poly-L-SUCLS (A), 25 mM poly-L-SUCL (B), and 50 mM poly-L-SUCLS (C) for enantioseparation of chiral phenoxypropionic acid [(\pm)-(2-PPA), 0.5 mg/mL in MeOH/H₂O]. MEKC conditions (A, B): pH 8.00, 25 mM NH₄OAc/ 25 mM TEA, 15 °C, pressure injection 50 mbar s, +20 kV applied for separations, UV detection at 200 nm. (C) MEKC conditions same as 5(A) except pH 2.00, 25 mM NaH₂PO₄/ 25 mM CH₃COONa and -20 kV applied for separations.

Table 1

Physicochemical properties of the monomers and polymers of chiral amino acid derived cationic surfactants undecenoxy carbonyl-L-leucinol bromide (L-UCLB) and undecenoxy carbonyl-L-pryrrolidinol bromide (L-UCPB).

| Characteristic of the ionic liquid type monomeric surfactants | L-UCPB | L-UCLB |
|---|-------------------------------|-------------------------------|
| Critical micelle concentration | | |
| $(CMC)^{a)}[mM]$ | $1.15 \pm (0.01)^*$ | $0.84 \pm (0.05)^*$ |
| Aggregation number ^{b)} | $95 \pm (0.09)^*$ | 97 ± 0.04 |
| Polarity (I_1/I_3) ratio $^{c)}$ | $1.095 \pm (0.001)^*$ | 1.180 ± 0.040 |
| Optical rotation $^{d)}$ | $-2.35 + (0.02)^*$ | $+21.67 \pm 0.03$ |
| Partial specific volume ^{e)} | $0.8281 \pm (0.0036)^*$ | 0.7185 ± 0.00 |
| Electroosmotic mobility | -2.83×10^{-4} | -2.42×10^{-4} |
| $\mu_{\text{eof}} (\text{cm}^2 \text{V}^{-1} \text{S}^{-1})^f)$ | $(\pm 1.56 \times 10^{-5})^*$ | $(\pm 5.31 \times 10^{-6})^*$ |
| Effective electrophoretic mobility | 2.08×10^{-4} | 1.94×10^{-4} |
| $\mu_{\rm ep} ({\rm cm}^2 {\rm V}^{-1} {\rm S}^{-1})^{f)}$ | $(\pm 1.54 \times 10^{-5})^*$ | $(\pm 7.49 \times 10^{-7})^*$ |
| Migration-time window | 3.79 | 5.09 |
| $(t_{mc}/t_o)^{f}$ | (± 0.20)* | (± 0.38)* |
| Characteristic of the polymeric | (3, 3, | (333 3) |
| surfactants | poly-L-UCPB | poly-L-UCLB |
| Aggregation number b) | $34 \pm (0.780)^*$ | $25 \pm (0.034)^*$ |
| Polarity (I_1/I_3) ratio ^{C)} | $1.219 \pm (0.001)^*$ | $1.22 \pm (0.007)^*_{*}$ |
| Optical rotation ^{d)} | $-7.84 \pm (0.04)^*$ | $+17.45 \pm (0.64)^*$ |
| Partial specific volume ^{e)} | $0.8408 \pm (0.0075)^*$ | $0.7634 \pm (0.0008)^*$ |
| Electroosmotic mobility | -2.54×10^{-4} | -2.34×10^{-4} |
| $\mu_{\text{eof}} (\text{cm}^2 \text{V}^{-1} \text{S}^{-1})^{f})$ | $(\pm 3.67 \times 10^{-6})^*$ | $(\pm 3.12 \times 10^{-6})^*$ |
| Effective electrophoretic mobility | 2.02×10^{-4} | 1.91×10^{-4} |
| $\mu_{\rm ep} ({\rm cm}^2 {\rm V}^{-1} {\rm S}^{-1})^f)$ | $(\pm 2.96 \times 10^{-6})^*$ | $(\pm 3.52 \times 10^{-6})^*$ |
| Migration-time window | 4 87 | 5 38 |
| $(t_{mc}/t_o)^{f}$ | (± 0.16)* | (± 0.53)* |

a) Critical micelle concentration is determined by the surface tension measurements.

b) Aggregation number is determined by the florescence quenching experiment using pyrene as a probe and cetyl pyridinium chloride as a quencher.

 $^{^{(}C)}$ Polarities of the surfactants are determined using ratio of the fluorescence intensity (I_{1}/I_{3}) of pyrene.

 $^{^{}d)}$ Optical rotation of 10 mg/mL of monomer and micelle polymers were determined in triply deionized water were obtained at 589nm [sodium D line].

e)Partial specific volumes were determined by the density measurements at different surfactant concentrations.

f) The μ_{ep} values for all monomer and polymeric ionic liquids were determined using methanol as t_0 marker and dodecanophenone as t_{mc} tracer. Experimental conditions: 64.5 cm (56 cm effective length) \times 50 μ m ID capillary with an applied voltage of -20 kV at 25 °C using a running buffer of 25 mM each of NaH2PO4 and Na2HPO4, 100 mM monomer and polymeric ionic liquids; dodecanophenone introduction, 50 mbar for 10 s (1.5 mg/mL in 50:50 MeOH/H2O).

Standard deviations are given in parentheses.