

DNA Sequence of the Lymphotropic Variant of Minute Virus of Mice, MVM(i), and Comparison with the DNA Sequence of the Fibrotropic Prototype Strain

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The sequence of molecular clones of the genome of MVM(i), a lymphotropic variant of minute virus of mice, was determined and compared with that of MVM(p), the fibrotropic prototype strain. At the nucleotide level there are 163 base changes: 129 transitions and 34 transversions. Most nucleotide changes are silent, with only 27 amino acids changes predicted, of which 22 are conservative. Notable differences between the MVM(i) and MVM(p) genomes which may account for the cell specificities of these viruses occur within the 3' nontranslated regions. The differences discussed include the absence of a 65-base-pair direct repeat in MVM(i), the presence of only two polyadenylation sites in MVM(i) compared with four in MVM(p), and sequences that bear a resemblance to enhancer sequences. Also included in this paper is an important correction to the MVM(p) sequence (C. R. Astell, M. Thomson, M. Merchlinsky, and D. C. Ward, *Nucleic Acids Res.* 11:999-1018, 1983).

Autonomous parvoviruses are small, icosahedral animal viruses that contain a linear, single-stranded DNA genome (57). Replication of these viruses requires a host factor expressed during the S phase of the cell cycle (43, 50, 53, 58). In addition, these viruses have a requirement for a host-cell differentiated state (32, 33, 54). For several parvoviruses, individual strains of the same serotype can show both different pathological spectra *in vivo* (25, 27) and different host-cell specificities *in vitro* (11, 19, 29, 54). While studies of the strain-specific tissue tropisms exhibited by viruses of several other groups have indicated that this specificity is often dictated by the presence or absence of a receptor on the host cell (20, 24, 35, 36, 51, 60, 61), this is not the case for at least two serologically identical variants of minute virus of mice, MVM(p), the prototype strain, and MVM(i), an immunosuppressive variant (52).

MVM(i) was first isolated from a murine T-cell lymphoma (8) and shown to inhibit several T-cell-mediated functions *in vitro* (17). This immunosuppressive property of MVM(i) is not associated with the prototype virus MVM(p), which grows productively in cells of fibroblast origin. *In vitro* studies on the interaction of both strains of MVM with differentiated cells revealed that while these viruses show a tropism for cells of lymphoid [MVM(i)] or fibroblast [MVM(p)] origin, both viruses bind to receptors on productive and restrictive cell types. Furthermore, competition experiments showed that both viruses recognize the same receptor. Because the genomes of MVM(i) and MVM(p) are converted to viral replicative forms (RFs) in either the productive or restrictive cell type, it is presumed that penetration and uncoating of the viruses occur regardless of the final outcome of the virus-host cell interaction (52). Development of a plaque assay for both MVM(p) and MVM(i) in 324K cells, a strain of simian virus 40-transformed human kidney cells which is permissive for both

MVM strains, allowed a study of the interactions of both strains with host cells of lymphocyte or fibroblast origin (55). These results showed that the host range of each virus variant is specified by a viral genetic determinant which is stably inherited. In addition, the susceptible host-cell phenotype is dominant in T lymphocyte × fibroblast fusion hybrids, implying that different cell types express different developmentally regulated virus helper functions which can only be exploited by the virus variant which carries the appropriate strain-specific determinant (55).

The construction and characterization of an infectious clone of the prototype strain of MVM has been reported previously (30). As a first step in the process of mapping the lymphotropic and fibrotropic determinants of these viruses by constructing recombinant genomes, we decided to molecularly clone the MVM(i) genome and determine its complete sequence. This paper describes the construction of molecular clones of MVM(i) which were used to determine the sequences of the 3' and 5' ends of the viral genome. The sequence of most of the coding region was determined by direct cloning of restriction fragments from MVM(i) RF into M13 vectors. In addition, we determined two important corrections in our earlier sequence for MVM(p) (4). A preliminary report of some of these data has been presented previously (M. Blundell, M. Thomson, E. Gardiner, S. Cotmore, J. Bratton, P. Tattersall, and C. R. Astell, *Can. Fed. Biol. Soc.* 27:128, 1984).

While this manuscript was in preparation, a similar report on the sequence of MVM(i) has appeared (48), and we have included in our discussion a comparison of these results with those presented here.

MATERIALS AND METHODS

Material. Enzymes were purchased from Bethesda Research Laboratories, Inc. (Gaithersburg, Md.) and New England BioLabs, Inc. (Beverly, Mass.). Linkers were purchased from New England BioLabs. [α -³²P]dATP (2,000

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to 3,000 Ci/mmol) was purchased from Amersham Corp. (Arlington Heights, Ill.). Genetically engineered DNA polymerase (Klenow fragment) was a gift from C. Joyce.

Virus stocks and cell lines. The cloned stock of the prototype strain of MVM(p) was grown in A9 cells (53, 55) and used as the source of DNA for the sequence studies and the construction of molecular clones of this genome (4, 30). Derivation of the clonal isolate of the immunosuppressive strain, MVM(i), has been described previously (52) and is the same virus as that used in the studies of Sahli et al. (48). Stocks of MVM(i) are routinely grown in S49.1 TB2 cells, a T-cell-lymphoma line, as described previously (55). The viral DNA from which viral termini were cloned was prepared from MVM(i) particles grown in 324K cells, a simian virus 40-transformed newborn human kidney cell line.

Plasmids of MVM(p) genome. Molecular clones of MVM(p), pMM984 (30) and pABO (7), have been described previously.

Molecular cloning of MVM(i) genome. The 5' end of the MVM(i) genome was cloned in a manner similar to that of the MVM(p) genome (30). Single-stranded DNA purified from MVM(i) particles was extended with DNA polymerase (Klenow fragment) plus all four deoxyribonucleotide triphosphates. *Bam*HI linkers were ligated to the right-hand end, and the *Eco*RI-*Bam*HI (*Eco*RI-B) fragment (map units 70 to 100) was cloned between the *Eco*RI and *Bam*HI sites of pAT153. This clone was designated pEG222. The left-hand end of the viral genome was cloned in a similar way, taking advantage of the fact that 1% of the packaged viral strands of MVM(p) are of the plus orientation (9). The *Eco*RI-*Bam*HI (*Eco*RI-C) fragment (map units 1 to 21) was cloned between the *Eco*RI and *Bam*HI sites of pAT153. This clone was designated pEG618.

Cloning of MVM(i) RF restriction fragments in M13 vectors. MVM(i) RF DNA was grown in S49.1 TB2 cells, and RF DNA was isolated as described previously (55). Restriction fragments from partial and complete digests with *Hae*III, *Alu*I, and *Sau*3A were cloned into appropriate sites of M13 mp8 and mp9 by using the M13 shotgun cloning approach (31). To obtain clones in specific regions of the genome, we hybridized clones containing the appropriate complementary region from MVM(p) DNA (4). One small region could not be read on any of our clones (~ map unit 92) and was sequenced on a longer clone with a synthetic oligonucleotide primer, 5'-TAAGACTAATAAAGA-3'. It should be noted that using the M13 vectors we had difficulty obtaining large clones of the MVM(i) genome (particularly the right half). We have no explanation for this other than that from the sequence it is clear there are many *Eco*PI sites in the DNA, and our host cell at the time, JM103, carries a P1 lysogen which encodes the restriction and modification system (62).

Determination of nucleotide sequence. Sequence of the M13 clones was determined by the dideoxy terminator method (49). Sequences were aligned with that of MVM(p) on a computer, using the DNA SEQNCE program of A. Delaney (14). This approach gave virtually all the sequences between map units 8 and 94 with the exception of ~ map unit 92. The synthetic 15-mer described above was used to sequence this region from map units 91 to 94. The sequence in both strands was obtained for at least 80% of the genome.

Sequence of 3' and 5' ends of the genome. Because clones of the 3' and 5' palindromes were not expected (and indeed were never obtained) with single-stranded vectors, we sequenced the ends of the genome by using plasmid clones and the chemical sequencing method (28). The sequence of the

left-hand end (map units 0 to 10) was derived from clone pEG618. The palindromic sequence was resolved completely by running several different percent acrylamide gels (20, 12, and 8%) at 40 W per gel to ensure complete denaturation of the hairpin. The region between map units 87 and 100 including the 5' palindrome was sequenced by the chemical method with plasmid pEG222 DNA. The full-length 5' palindrome was propagated in *Escherichia coli* JC8111 (7).

RESULTS

Correction of MVM(p) sequence. From a comparison of the sequence of MVM(i) presented here with that of MVM(p) (4), we are now aware of two errors in the sequence of MVM(p). First, the region about the *Eco*RI site at map unit 70 was incorrect owing to an error at the computer level in joining the sequence of the *Eco*RI A fragment to the *Eco*RI B fragment from clone pABO. (NB: This clone contains both the A and B fragments with the A fragment inverted relative to B [7].) We have now confirmed this region on pMM984, the full-length infectious clone (30), and the correct sequence is given in Fig. 1. In comparing the sequence of MVM(i) with that of MVM(p), we were initially surprised to find that the sequences at ~ map unit 94 were identical (except for a few single base changes). However, in agreement with McMaster et al. (29), we observed a mobility difference of about 60 base pairs (bp) in the *Xba*I-*Msp*I fragment (map units ~86 to 99) of MVM(p) and MVM(i). We resequenced this region of the MVM(p) genome and discovered that there is a 65-bp direct repeat in this region (nucleotides 4720 to 4784 and 4785 to 4849; Fig. 1). This region in the MVM(p) genome was originally sequenced by the chemical method (28) with clone pABO (4). Because *Sau*3AI fragments were used to read this region, the 65-bp *Sau*3AI fragment contained within the direct repeat was missed. Sequencing leftward from the *Sau*96I site at nucleotide 4872 to get the overlap of the *Sau*3AI site should have detected the missing fragment. Unfortunately, at the time when these sequence experiments were done, commercial preparations of *Sau*96I were often not sufficiently pure to give sequences which could be read for the usual 150 to 200 nucleotides. Indeed, although this experiment was done, the overlap extended only ~25 to 30 nucleotides, a length which was considered adequate at the time. Clearly it was not, since a 65-bp repeat was missed. We subsequently sequenced this region with current preparations of *Sau*96I and read 198 nucleotides leftward from the *Sau*96I site, through both copies of the direct repeat. The possible significance of this direct repeat is discussed below.

Nucleotide sequence of MVM(i) genome. The sequence of MVM(i) is given in Fig. 1 (top line). This sequence is presented in comparison with that of the corrected sequence of MVM(p), using the DNA SEQNCE program (14). Nucleotide differences between the two sequences are displaced above [MVM(i)] and below [MVM(p)] the respective lines. The 65-bp repeat (nucleotides 4785 to 4849) in MVM(p) is further displaced to facilitate maintaining the homology lineup between the two genomes beyond this point. It should be noted here that the clone pEG222 is 21 nucleotides shorter than MVM(p) viral-strand DNA at the right-hand end of the genome owing to the DNA polymerase I (Klenow fragment) extension of viral DNA which stopped short of replicating the entire hairpin. The last 22 nucleotides of the MVM(i) right-end sequence shown here are inferred from the sequence of the clone between nucleotides 4880 and 4902.

It is clear from the sequence that with the exception of the

10 20 30 40 50 60 70 80 90 100
 ATTTTGAAGTACCAACCATGTTACGTAAGTGACGTGATGACGCGCGCT CGCGCGC GCCTTCGGACGTACACAGTCACTTACGTTTCACATGGTT
 ATTTTGAAGTACCAACCATGTTACGTAAGTGACGTGATGACGCGCGCT CGCGCGC GCCTTCGGACGTACACAGTCACTTACGTTTCACATGGTT
 10 20 30 40 50 60 69 79 89 99
 110 120 130 140 150 160 170 180 190 200
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 GGTCAGTTCTAAAAATGATAAGCGGTTTCAG GAGTTTA ACCAAGGCGCGAAAAAGGAAGTGGGCGTGGTTTAAAGTATATAAGCAA T CTGAAGTCAGT
 109 119 129 139 149 159 169 179 189 199
 210 220 230 240 250 260 270 280 290 300
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 TACTTATC TTTCTTTCATTCTGTGAGTCGAGACGC CAGAAAGAGAGTAACCAACTAACCATGGCTGGAAATGCTTACTCTGATGAAGTTTTGGGA CA
 209 219 229 239 249 259 269 279 289 299
 310 320 330 340 350 360 370 380 390 400
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 410 420 430 440 450 460 470 480 490 500
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 AAAAAA GAGCTGCAGGAGGACGAGCTGAAATCTTTACAACGAGGAGCGGAAACTAC TGGGACCAAAGCGAGGACATGGAATGGGAA C ACAGTGGAA
 409 419 429 439 449 459 469 479 489 499
 510 520 530 540 550 560 570 580 590 600
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 610 620 630 640 650 660 670 680 690 700
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 609 619 629 639 649 659 669 679 689 699
 710 720 730 740 750 760 770 780 790 800
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 GGCA CTAATGTTTACTGGAGCAGATGGTTGGTAAACAGCCTGTAATGTGCA CTAACACCAGCTGAAAGAATTAACCTAAGAGAAATAGCAGAAGACA
 709 719 729 739 749 759 769 779 789 799
 810 820 830 840 850 860 870 880 890 900
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 1009 1019 1029 1039 1049 1059 1069 1079 1089 1099

1110 1120 1130 1140 1150 1160 1170 1180 1190 1200
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 A AGA GTTTCATTAAAAC ACACCTTAAAGAGCT GTGCATAAAAAGAGTAACCTCACCAGA GACTGGATGATGATGCAGCCAGACAGTTACATTGAAATG
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 1210 1220 1230 1240 1250 1260 1270 1280 1290 1300
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 1209 1219 1229 1239 1249 1259 1269 1279 1289 1299
 1310 1320 1330 1340 1350 AG 1360 1370 1380 1390 1400
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 A AGCTGAAACCAGCAAACCTAACCAACTTTTCACTGCCTGACACAAGAACCTGCA ATTTTGTCTTTTCATGGCTGGAACATGTTAAAGTTTGCCATGC
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 1510 1520 1530 1540 1550 1560 1570 1580 1590 1600
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 A 1509 1519 1529 1539 1549 1559 1569 1579 1589 1599
 1610 1620 1630 1640 1650 1660 1670 1680 1690 1700
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 1910 1920 1930 1940 1950 1960 1970 1980 1990 2000
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 2010 2020 2030 2040 2050 2060 2070 2080 2090 2100
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 3210 3220 3230 3240 3250 3260 3270 3280 3290 3300
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3310 C 3320 3330 3340 3350 C 3360 3370 3380 3390 3400 T
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TGAAACCAAC ATAGCATC CCATACAGGTACTATTT TCGTTGACAGAGATCTTTTCAGT ACCTA GAAAATCAAGAAGGCACA TTGA CATAATG
C 3409 A 3419 T 3429 T 3439 C 3449 G 3459 C 3469 G 3479 A 3489 A 3499
3510 3520 3530 3540 3550 3560 3570 3580 3590 3600
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T ATGGGAACACCAAAGGAATGAATTCTCAATTTTTTACCATTGAGAACACACAACAAAATCACATTGCTCAGAAC GG GA GA TTTGC AC GG AC
G 3509 3519 3529 3539 3549 3559 3569 3579 3589 3599
3610 3620 3630 3640 3650 3660 3670 3680 3690 3700
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TACTACTTTGACACAAA CAGTTAAACT ACACACAC TGGCAAAC AACCGTCAACTTGGACAGCCTCCACTGCTGTCAACCTTTCTGAAGCTGAC
T 3609 TT 3619 C 3629 G 3639 C 3649 3659 3669 3679 3689 3699
3710 3720 3730 3740 G 3750 G A 3760 T 3770 3780 3790 3800
ACTGATGCAGGTACACTTACTGCTCAAGGGAGCAGACATGGA CAACACA ATGG GGTTAACTGGGTGAGTGAAGCAAT AGAACCAGACCTGCTCAAG
ACTGATGCAGGTACACTTACTGCTCAAGGGAGCAGACATGGA CAACACA ATGG GGTTAACTGGGTGAGTGAAGCAAT AGAACCAGACCTGCTCAAG
A 3709 3719 3729 3739 A 3749 A G 3759 C 3769 3779 3789 3799
3810 3820 3830 3840 3850 3860 3870 3880 3890 3900
G TAGGATTTTGTCA CCACACAATGACTTTGAAGCCAGCAGAGCTGGACCATTGCTGC CAAAAGTTCCAGCAGAT TTAAGGAGT GACA AGA
TAGGATTTTGTCA CCACACAATGACTTTGAAGCCAGCAGAGCTGGACCATTGCTGC CAAAAGTTCCAGCAGAT TTAAGGAGT GACA AGA
A 3809 3819 3829 3839 3849 3859 3869 3879 3889 3899
3910 3920 3930 3940 3950 3960 3970 3980 3990 4000
G AGCCAATGGCAGTGTAGATACAGTTATGGCAAACAGCATGGTGAATAATGGGCT CACA GGACCAGCACCAGAGCGCTACACATGGGATGAAACAA C
AGCCAATGGCAGTGTAGATACAGTTATGGCAAACAGCATGGTGAATAATGGGCT CACA GGACCAGCACCAGAGCGCTACACATGGGATGAAACAA C
T 3909 3919 3929 3939 3949 3959 3969 3979 3989 3999
4010 A 4020 G 4030 T 4040 T 4050 G 4060 4070 G 4080 4090 4100
TTTGGTTCAGG AGAGACACCA AGATGGTTTTATTCAATCAGCACC CTAGTTGTTCCACCACCCTAAATGG ATTCTTACAAATGCAAACCCTATTG
TTTGGTTCAGG AGAGACACCA AGATGGTTTTATTCAATCAGCACC CTAGTTGTTCCACCACCCTAAATGG ATTCTTACAAATGCAAACCCTATTG
T 4009 A 4019 4029 4039 A 4049 4059 4069 C 4079 4089 4099
4110 4120 4130 4140 4150 4160 4170 4180 4190 4200
A G ACTAAAAATGACATTCATTTTTCAAATGTTTTTAACAGCTATGGTCCACTAACTGCATTTTTACACCCAAGTCTGTATACCCTCAAGGACAAAATATG
G ACTAAAAATGACATTCATTTTTCAAATGTTTTTAACAGCTATGGTCCACTAACTGCATTTTTACACCCAAGTCTGTATACCCTCAAGGACAAAATATG
G 4109 4119 4129 4139 4149 4159 4169 4179 4189 4199
4210 4220 4230 4240 4250 4260 4270 4280 4290 4300
C GGACAAAGAAGTCTGATCTTGAACACAAAACCTAGACTTCACATAAAGTCTCCATTGT TGTAAAAAACAATGCACCTGGACAAAATGTTGGTTAGATTAGGA
GGACAAAGAAGTCTGATCTTGAACACAAAACCTAGACTTCACATAAAGTCTCCATTGT TGTAAAAAACAATGCACCTGGACAAAATGTTGGTTAGATTAGGA
T 4209 4219 4229 4239 4249 4259 4269 4279 4289 4299
4310 T 4320 G 4330 4340 4350 G 4360 T 4370 4380 4390 4400
CCAAA TAACTGACCA TATGATCCAAACGGAGCCACACTTTCTAGAATTGT AC TA GGTACATTTTTCTGAAAAGGAAAACTAACCATGAGAGCAA
CCAAA TAACTGACCA TATGATCCAAACGGAGCCACACTTTCTAGAATTGT AC TA GGTACATTTTTCTGAAAAGGAAAACTAACCATGAGAGCAA
C 4309 A 4319 4329 4339 4349 T 4359 C 4369 4379 4389 4399



FIG. 1. Homology between MVM(i) (upper line) and MVM(p) (lower line). The two sequences were aligned by using the homology routine of the DNA sequence program (14). Mismatch nucleotides are displaced above and below the lines. The 65-bp tandem repeat in MVM(p) (nucleotides 4785 to 4849) is further displaced to maintain the lineup of the two sequences.

65-bp repeat, the sequences of MVM(p) and MVM(i) are very similar. In the MVM(i) genome three differences are an insertion of a T at position 61, a deletion of a C at position 4828, and a compensatory insertion of a T at position 4864 relative to MVM(p). None of these changes occurs within the coding region of MVM(p) (41) or, presumably, of MVM(i). Since the 5' hairpin region of our sequence is derived from a clone, pEG222, it has only one orientation, in this case the flip orientation. It is interesting that the end of the loop in this flip orientation is 5'-TTT-3' on the C strand, whereas in MVM(p) this sequence is the complementary 5'-AAA-3'. Of the 163 bp changes between MVM(i) and MVM(p), 129 are transitions and 34 are transversions. Distribution of these changes is nonrandom throughout the genome. In a genome size of ~5,000 nucleotides, 163 random base changes would average 3 changes per 100 nucleotides. The region between map units 40 and 60 is highly conserved (there are only six base changes in the sequence

between nucleotides 2000 and 3000), whereas the regions between map units 8 to 20 and 70 to 80 have a much higher frequency of base changes (up to 8 to 10 changes per 100 nucleotides). This high degree of sequence conservation supports our previous suggestion that there may be multiple splice sites near map units 45 to 46 of MVM(p), with implied overlapping reading frames to generate different viral proteins (4). Another region (~ nucleotide 3580, map unit 70) seems to be a hot spot for base changes. There are eight changes within a distance of 25 nucleotides (Fig. 1). This region is within the coding region for the viral structural proteins; however, because all of these base changes occur in the third position of eight codons, the amino acid sequences between MVM(i) and MVM(p) in this region are identical (see below and Fig. 3).

Left-hand coding region. The sequence of MVM(i) is very similar to that of MVM(p) (4), with two major open reading frames which span almost the entire genome. The left-hand

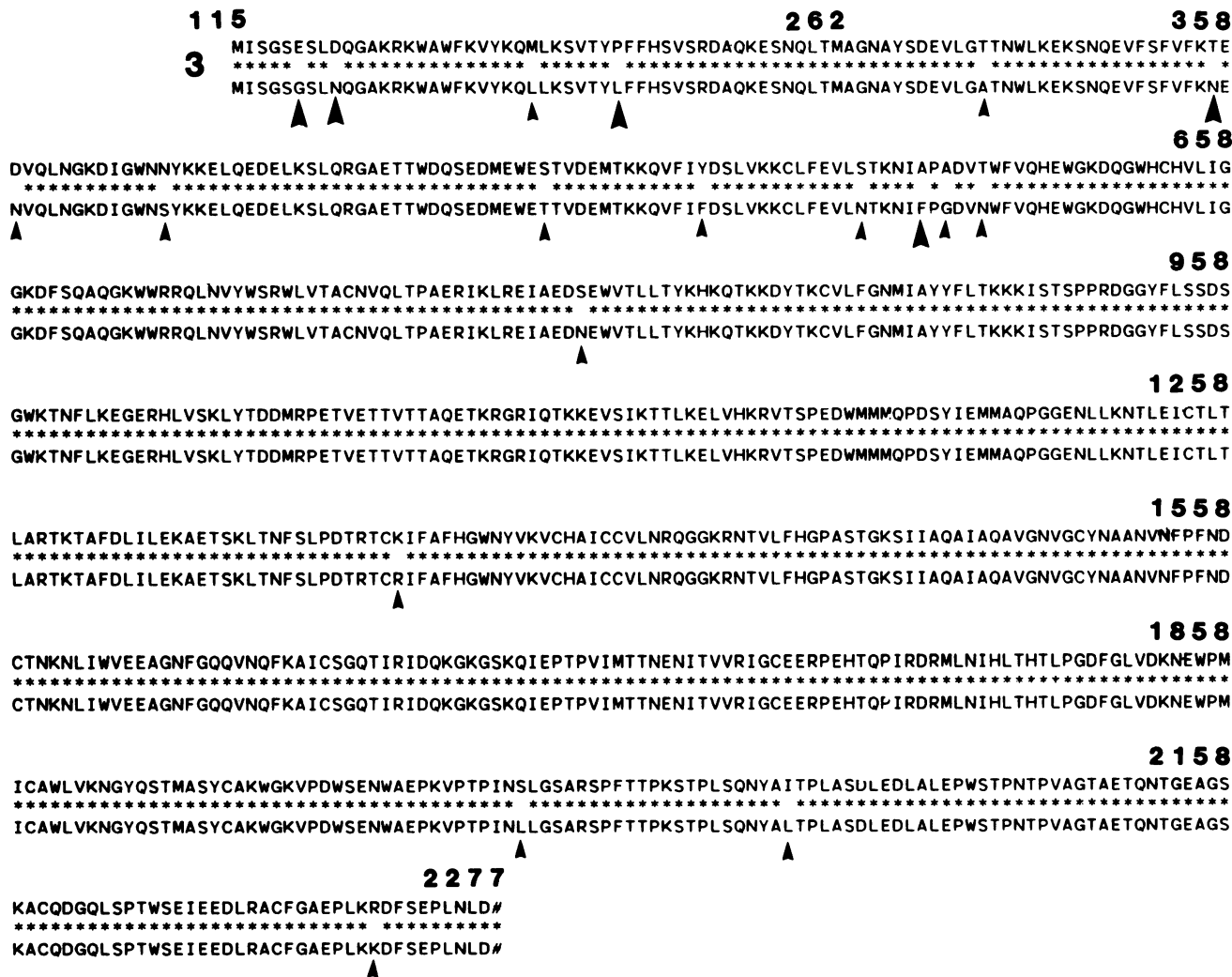


FIG. 2. Lineup of the left-hand open reading frame of MVM(i) (upper line) and MVM(p) (lower line). Identical amino acids are indicated by an asterisk. Conservative changes are noted by a small arrowhead below the amino acids. Radical changes are noted by a larger arrowhead. To facilitate comparison with Fig. 1, the numbers correspond to the nucleotide in the first position of the codon.

open reading frame (frame 3) extends from map units 2 to 45. The sequence of MVM(p) (4), transcription data (41), and subsequent studies on the mapping of the viral genome (13) indicate that NS-1, a nonstructural protein (83,000 daltons) of unknown function, maps to the left-hand coding region of the genome (13). A second nonstructural protein, NS-2 (24,000 daltons), has not yet been mapped precisely but is encoded in an mRNA transcribed from the left-hand (P4) promoter (13). Assuming that the protein products of the two strains, i and p, are similar, it is reasonable to compare the coding regions of the viruses to learn if there are significant differences at the amino acid level.

In Fig. 2, a lineup (14) of amino acids encoded within the potential left-hand open reading frames of MVM(i) (upper line) and MVM(p) (lower line) is given. Transcript R1 (4.8 kilobases) is a minor RNA component which encodes NS-1 (13). This transcript begins around nucleotide 201 (6), with the first methionine codon starting at nucleotide 262 of MVM(i) [261 of MVM(p)]. The protein (translated from reading frame 3) would begin with Met-Ala-Gly-Asn-Ala (MAGNA in Fig. 2). The first termination codon (TAA) in reading frame 3 occurs at position 2278. Beginning with the

methionine at 262, there are a total of 15 amino acid changes between the MVM(i) and MVM(p) coding regions. Thirteen of these changes are conservative: Thr→Ala, Thr→Asn, Asn→Ser, Ser→Thr, Tyr→Phe, Ser→Asn, Ala→Gly, Thr→Asn, Ser→Asn, Lys→Arg, Ser→Leu, Ile→Leu, and Arg→Lys. There are two radical changes: Gly→Glu at position 358, and Ala→Phe at position 583. We noted earlier that for MVM(p), reading frame 3 is open upstream of the promoter region and the mapped mRNA start site. From the most upstream methionine codon, at position 115, in this extra open reading frame, the protein should begin with the amino acid sequence Met-Ile-Ser-Gly-Ser (MISGS) if this ATG is a functional start site. This region is also open in the MVM(i) sequence, with four additional amino acid differences from the polypeptide encoded between nucleotides 115 and 261 of MVM(p). Three of these changes are radical: Glu→Gly, Asp→Asn, and Pro→Leu. The fourth change, Met→Leu, is conservative. The possible significance of this region of frame 3 will be discussed below.

Right-hand coding region. The right-hand open reading frame of MVM (frame 1) extends from map units 46 to 89. Mapping studies have shown this region to encode the

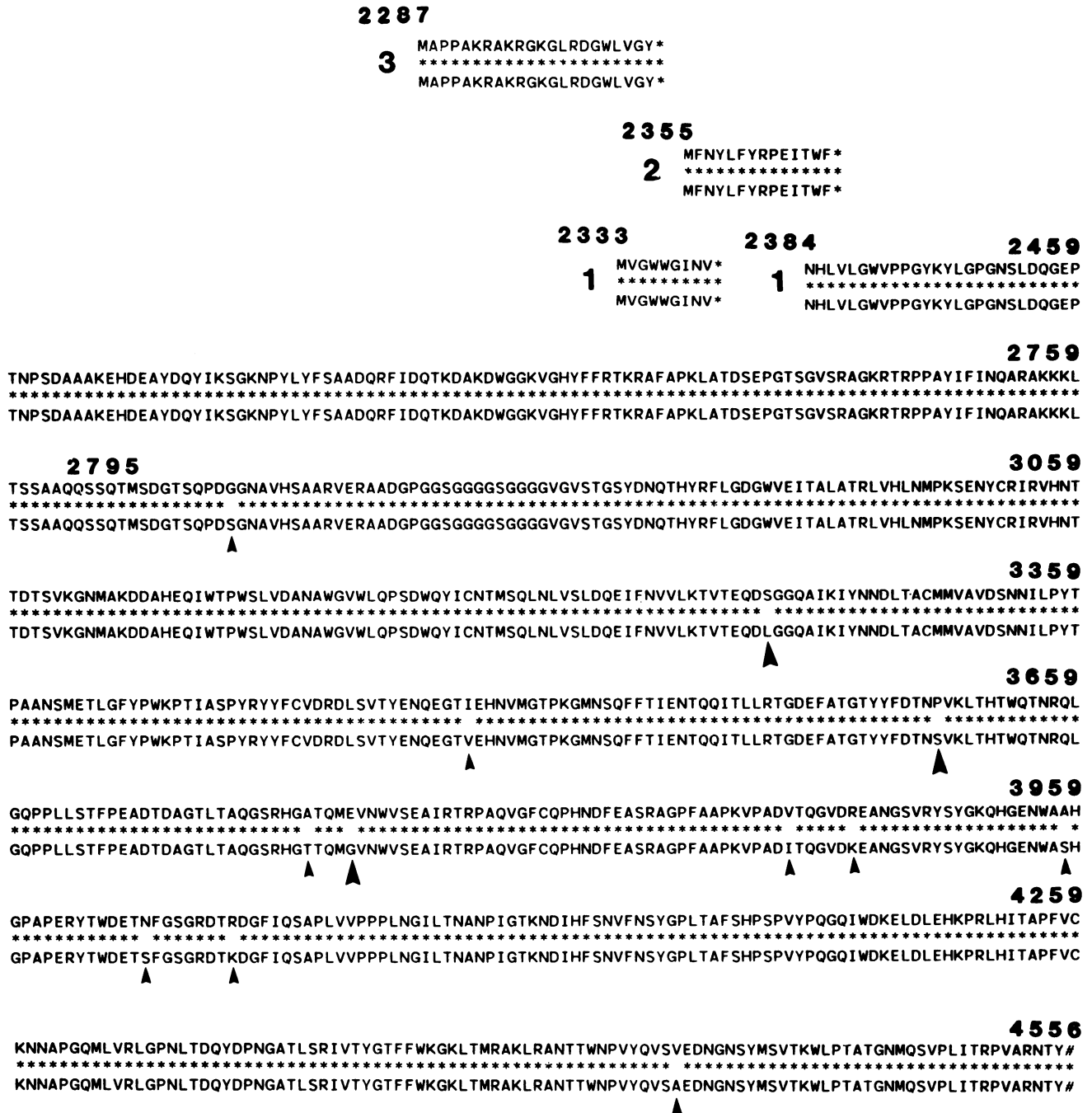


FIG. 3. Lineup of the right-hand open reading frame of MVM(i) (upper line) and MVM(p) (lower line). Identical amino acids are indicated by an asterisk. Conservative changes are emphasized by a small arrowhead below the amino acids. Radical changes are noted by a larger arrowhead. As discussed in the text and illustrated in Fig. 4, VP1 likely begins at the methionine at position 2287, and splicing of the transcript to frame 1 allows continued translation to the stop codon at nucleotide 4556. VP2 likely begins at 2795 and also terminates at the stop codon at nucleotide 4556 (see text for additional details). Numbers correspond to the first nucleotide of the codon.

overlapping structural polypeptides VP1 and VP2, which are both primary products of in vitro translation of viral mRNA (13, 46). The first methionine codon in this large right-hand open reading frame occurs at position 2795 (Fig. 3). The start of the mRNA(s) encoding these proteins maps to position 2005 ± 5 (6). We mapped this transcription start against the DNA sequence to position 2003, with two minor starts at 2006 and 2010 (C. Astell, unpublished data). Earlier, we

reasoned that the AUG at 2795 was unlikely to be the functional start signal for VP2 (4). It would require ribosomes to traverse some 790 nucleotides of mRNA and to bypass at least three upstream AUGs (in closed reading frames) to initiate here the translation of the most abundant viral gene product. However, structural studies on the N terminus of VP2 in H1 have indicated (40) that this is indeed the likely start codon for VP2 translation and that the amino

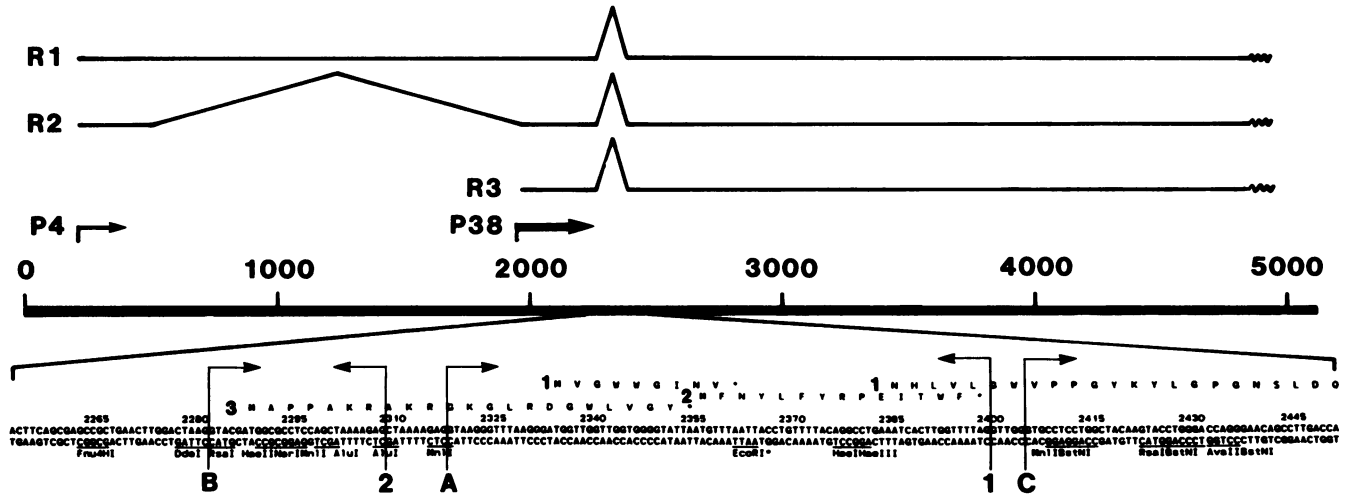


FIG. 4. Potential splice junctions at map unit ~45. The upper portion of the figure illustrates the RNA transcripts and splicing pattern for MVM(p) (41). As discussed in the text, a comparison of splice junctions at map unit ~45 for MVM(p), MVM(i), H1, and canine parvovirus indicate that a splice from A (5' donor site) to 1 (3' acceptor site) will generate the mRNA encoding VP1. Alternative splicing from B (5' donor site) to 1 (3' acceptor site) produces the probable mRNA encoding VP2 (see text for details). R1, R2, and R3 are the major spliced transcripts, and P4 and P38 are the two promoters (41).

acid sequence of VP2 is identical to the C-terminal three-quarters of VP1.

Transcription studies on MVM (41) have identified a minor splice, common to all transcripts, lying between nucleotides 2250 and 2450. It is significant that through this region, and for several hundred nucleotides on either side of it, MVM(p) and MVM(i) are more strongly conserved than in all other regions of the genome, as though additional constraints on random genetic drift apply to this region and not to others. The reason for this may be the potential use of a second open reading frame in this region, perhaps encoding part of NS-2, since the regions showing diminished drift between MVM(p) and MVM(i) delineate extensive dual open reading frames on either side of the minor splice. These dual open reading frames are conserved between MVM(p) (4), MVM(i) (this paper), H1 (46), and to some extent canine parvovirus (44).

We have noted previously (4) that the DNA sequence of MVM(p) around the minor splice region offers several close matches to consensus splice donor and acceptor splice sites (38), and we used both the conservation and divergence of sequence between MVM(p), MVM(i), H1, and canine parvovirus in this region to explore which splicing options are most likely used. We matched sequences from nucleotides 2251 through 2450 in MVM(i) with equivalent regions from MVM(p), H1, and canine parvovirus, using the University of Wisconsin Genetics Computer Group GAP program. This region from each of the four genomes was then searched individually for the 10 best fits to both donor and acceptor consensus sequences (38), using the University of Wisconsin Genetics Computer Group FITCONSENSUS program. Potential splice sites common to all four were then ranked (A, B, C, etc. for donors and 1, 2, etc. for acceptors) by their average quality of fit and are displayed on the MVM(p) sequence in Fig. 4. Three potential donor splice sites and two potential acceptor sequences conserved between the four genomes were identified in this way. (Note that acceptor site 1 was not recognized earlier [4] because our search at that time was restricted to TNCAG.) We can discount donor site C since it is downstream of both potential acceptor sequences. The DNA sequence downstream of acceptor site 2 has translation terminators in all three reading frames and

so would not allow translation of the VP1-specific region of the long right-hand open reading frame. While acceptor 1 is a 12-of-16-base fit to the consensus sequence, acceptor 2 is only a 9-of-16-base fit and has an AG dinucleotide at positions -4, -3, and -11, -10, from the A6 of the potential splice site which is never seen within 14 nucleotides of a functional acceptor site (38). On the basis of this analysis, we would predict that VP1 is synthesized starting with the methionine at nucleotide 2287 in MVM(i), with the sequence MAPPAKRA, and that this transcript is spliced from donor A to acceptor 1, joining a 10-amino acid N-terminal segment onto the right-hand long open reading frame through a fused glycine codon. An alternatively spliced transcript in which donor B is joined to acceptor 1 would circumvent the initiator methionine at nucleotide 2287 and allow the synthesis of VP2 to occur by delaying translation initiation until the ribosome had reached the methionine at nucleotide 2795 in MVM(i). For completeness, in Fig. 3 a lineup of the right-hand open reading frames (frame 1) of MVM(i) (upper line) and MVM(p) (lower line) includes potential leader regions (frames 1, 2, and 3) discussed earlier (4). However, assuming that VP1 begins in frame 3 at position 2287 as discussed above and is spliced to frame 1 with the 5' donor site A and 3' acceptor site 1 (Fig. 4), there are in the VP-1 coding region 12 amino acid changes between MVM(i) and MVM(p). Again, conservative changes predominate: Gly→Ser, Ile→Val, Ala→Thr, Val→Ile, Arg→Lys, Ala→Ser, Asn→Ser, Arg→Lys, and Val→Ala. There are three radical changes: Ser→Leu at position 3272, Pro→Ser at position 3619, and Glu→Gly at position 3755. It is noteworthy that the amino-terminal region unique to VP1 (i.e., not translated in VP2 if VP2 does begin at 2795) is invariant between MVM(i) and MVM(p).

Comparison of transcription signals between the two genomes. Two promoters with overlapping transcription units have been characterized for MVM(p) (41). The TATAAG of the left promoter (at position 177) is identical in i and p, as is the TATAAA of the second, much stronger promoter at position 1977. Whether the nucleotide changes in the vicinities of these consensus sequences are functionally significant is not known.

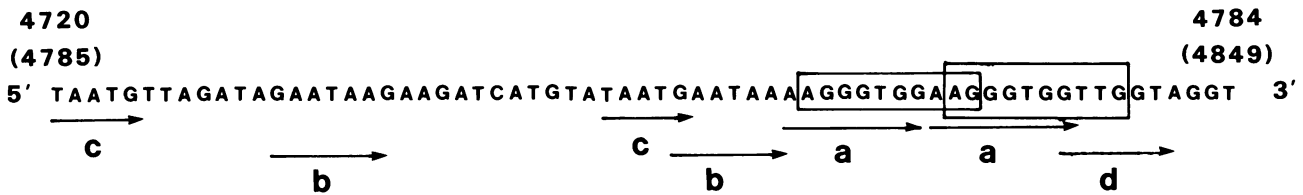
MVM(p) C-strand**H-1 C-strand**

FIG. 5. Nucleotide sequences of the 65- and 55-bp repeats found in MVM(p) and H1. Arrows below the sequence correspond to short repeated sequences found within and between these tandem repeats from the MVM(p) and H1 genomes. The boxed regions are regions which share some homology with the consensus sequence for enhancers.

The most obvious difference between i and p in transcription signals involves potential polyadenylation sites, AATAAA. The second polyadenylation signal in MVM(p) at nucleotide 4755 (repeated at 4820) is mutated to GGTAAG in MVM(i) (nucleotide 4756). Because this is within the 65-bp repeat in MVM(p), there are four canonical polyadenylation signals in MVM(p), at nucleotides 4602, 4755, 4820, and 4885, compared with only two in MVM(i) at nucleotides 4603 and 4821.

DISCUSSION

We constructed clones of the genome of MVM(i), the immunosuppressive strain of MVM, and determined the complete nucleotide sequence of these clones. The sequence is virtually identical to that of MVM(p) (4) with the exception that the sequence of MVM(p) was found to contain a duplication of 65 bp which was missed in the original sequence (4). The presence of the 65-bp duplication in MVM(p) relative to MVM(i) explains the size difference between the *XbaI-MspI* fragment (map units 86 to 99) observed by McMaster et al. (29) and confirmed by ourselves. It is noteworthy that a similar tandem repeat (of 55 bp) has been observed in another autonomous parvovirus genome (H1) as well as in defective interfering particles of H1 (45), though it is not present in the genome of a closely related virus, H3 (45). Both tandem repeats [MVM(p) and H1] (Fig. 5) contain a polyadenylation signal. The repeated sequence in H1 is 84% A+T in contrast with the repeat in MVM(p) which is only 66% A+T. When one compares the sequences of the 65-bp MVM(p) and 55-bp H1 repeats one of the most striking features is that on the complementary strand, the 65-bp repeat contains only one C residue while the 55-bp repeat does not contain a C. Within the MVM(p) 65-bp repeat there are several short direct repeats: AAGGGTGG (a), GAATAA (b), and TAATG (c). In H1 there is one short direct repeat, AATATA (e). Sequences b, c, and d (GTTGGT) are common to both the 65- and 55-bp repeat (Fig. 5).

The 65-bp direct repeats in the MVM(p) genome are of particular interest for three reasons. First, there is a polyadenylation signal within the repeat (hence two

polyadenylation sites at 4755 and 4820; Fig. 1). In MVM(i), these sites are lacking due to a mutation of AATAAA → GGTAAG (nucleotide 4756; Fig. 1). Hence the MVM(i) genome has two fewer potential polyadenylation sites than does the MVM(p) genome. It is of possible interest that many genes possess multiple polyadenylation sites and that alternative processing of the 3' ends of transcripts has been documented for several cellular genes as well as viral genes (1, 16, 23, 39, 47). While the factors that determine selection of a particular polyadenylation site are not well understood, it has been suggested that they probably reflect the growth state of the cell or the differentiated cell type (34). Second, Faust et al. (18) have recognized that the MVM genome contains 26 hexanucleotides, referred to as ψ , which conform to the consensus sequence $C_2A_{1-2}(C_{2-3}/T_2)$, and presented evidence that these sequences may be initiation sites for purified mouse DNA polymerase α primase in vitro. One of these sequences in the MVM(p) genome in the region of nucleotides 4762 to 4779 (3'-C₃AC₂T₂C₃AC₂A₂C₂-5') is repeated at nucleotides 4827 to 4844. As yet, there is no evidence that ψ sequences are used by the DNA polymerase α primase in vivo (18). The third reason why the 65-bp repeats of MVM(p) are of interest is that they contain sequences related to transcriptional enhancer elements which have been identified in viral and cellular genes. Some of these enhancers show tissue- or cell-specificity (5, 12, 15, 21, 26, 37, 42, 62), although it has been reported recently that the simian virus 40 72-bp repeats do not alter the species specificity of polyomavirus (10). Hen et al. (22) have compared enhancer sequences from 10 genomes and have suggested a consensus sequence GXTGTGG^{TTT}_{AAA}. Weiher et al. (59) have suggested a core sequence of (G)TGG^{TTT}_{AAA}(G) based on point mutations of the SV40 enhancer and comparison with other viral enhancers. Overlapping related sequences which occur within the MVM(p) 65-bp repeat are boxed in Fig. 5, as is a related sequence in the 55-bp repeat of H1. These MVM(p) sequences agree in 7 of 10 nucleotides with the former enhancer consensus sequence and in 7 of 8 nucleotides with the latter (viral) enhancer consensus sequence.

The structures of the 3' and 5' palindrome regions of the

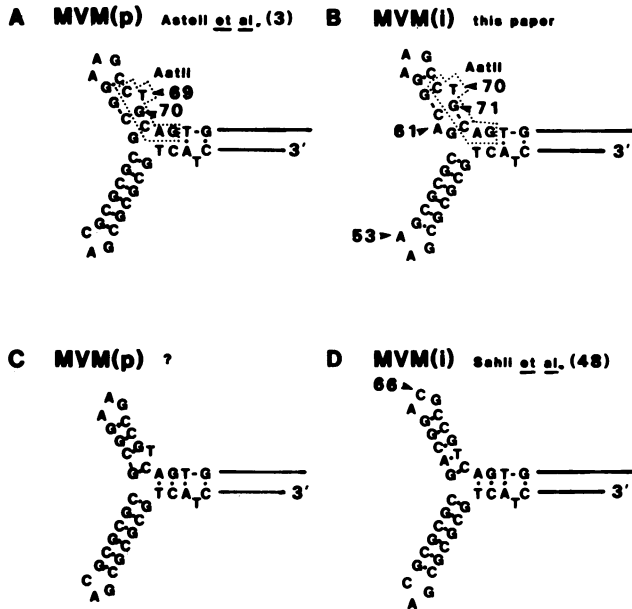


FIG. 6. Structure of the 3'-terminal hairpins from MVM(p) and MVM(i). The sequence determined for MVM(p) (3) is illustrated in Fig. 6A and that determined for MVM(i) (this paper) is given in Fig. 6B. Nucleotide differences between p and i are indicated by the arrowheads at positions 53 and 61. The sequence for MVM(i) determined by Sahli et al. (48) is illustrated in Fig. 6D and has an additional nucleotide change at position 66. Because we were not able to detect an *AatII* site at nucleotide 69 in MVM(i) or 68 in MVM(p), we believe that the sequence of Sahli et al. (48) is more probably correct. The correct sequence for the 3' hairpin of MVM(p) is illustrated in Fig. 6C.

MVM(i) genome indicate that they are very similar to those of MVM(p). The 3' palindrome is identical, except for an additional T at position 61 and a G→T transversion at position 53 in MVM(i). This additional base (an A in viral-strand DNA) occurs in the short arm of the Y-shaped hairpin in viral-strand DNA (3) and thus may alter the stability of this arm. The G→T base change at position 53 (C→A change in viral-strand DNA) lies within the 3-nucleotide loop of the long arm of the Y-shaped hairpin and would not be expected to alter the stability of this structure (Fig. 6A and B). The 5' palindrome is also highly conserved. Assuming that the inferred sequence of the last 21 nucleotides is correct, there are six single base changes. Each is matched by a compensatory base change such that in the hairpin configuration of viral-strand DNA, the number of potential base pairs and hence stability of the hairpin would not be expected to be altered. The loop end of the hairpin is AAA (flip orientation) and TTT (flop orientation), which is opposite to that of the MVM(p) genome. It is interesting to speculate on how such a change might have occurred. Possible explanations are three single point mutations, a gene conversion event, a double crossover, a single crossover event at the level of viral-strand DNA, or a single crossover event between flip and flop turnaround forms of RF DNA, distal to the asymmetric unpaired region of the hairpin. Of the five we consider the last to be most likely.

Comparison of our sequence for MVM(i) with that of Sahli et al. (48). We compared our sequence for MVM(i) with that of Sahli et al. (48) and found 11 differences in base changes (Table 1). The AC versus CA at position 70 is most likely to be due to the problem of resolving the hairpin on gels, noted

TABLE 1. Differences in sequence between MVM(i) of Sahli et al. (48) and MVM(i) of this paper

Nucleotide position	MVM(i) of Astell et al.	MVM(i) of Sahli et al.	Coding changes
66	T	G	
70	A	C	
71	C	A	
90	T	C	
1566	C	G	
1572	G	A	
1962	A	G	
2050	A	C	Ile→Leu
2832	C	G	Ala→Gly
4390	C	A	
4698	A	G	

earlier for MVM(p) (3). At this point we are not sure of the correct order for these nucleotides. We observed both orders for MVM(p) viral-strand DNA (C. Astell and M. B. Chow, unpublished data). More frequently the order was that reported earlier (3) and illustrated here in Fig. 6A. Also, sequencing studies of the complementary strand of MVM(p) RF DNA agreed with this order (2). However, we consider the order reported by Sahli et al. (48) to be more probably correct, for the following reason. Repeated attempts to restrict both MVM(p) and MVM(i) clones at a predicted *AatII* site at position 70, in this laboratory and in several other laboratories, have failed. Although the secondary structure in this region is a possible explanation for this problem, we must also consider the likelihood that the site simply does not exist, as is predicted by the Sahli sequence. This nucleotide reversal does not change the net number of base pairs in the short arm of the 3' hairpin of MVM(p), although it could possibly increase the stability of the structure. It increases by one the number of paired bases in the MVM(i) 3' hairpin, thereby significantly increasing its stability (Fig. 6B through D).

In addition to this difference, there are nine base change differences between the two sequences of MVM(i). Of the seven changes within the coding sequence, none would lead to radical amino acid differences. We found two changes which were not reported by Sahli et al. (48), and they found seven changes which we did not see.

A possible reason for the differences between our sequence of MVM(i) and that of Sahli et al. (48) is that we cloned from different MVM(i) stock preparations. Although both laboratories use stocks derived from the same genetically purified MVM(i) stock (52), our virus is routinely grown in S49.1 TB2 cells; in the case of the preparation from which the MVM(i) terminal plasmid clones were derived, the virus was grown in 324K cells. Sahli et al. (48) grew their preparation of MVM(i) in EL-4 lymphoma cells. Perhaps growth in these different host-cell types has allowed for some degree of genetic drift.

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ADDENDUM

After submission of this report, Rhode (J. Virol. 55:886-889, 1985) reported that sequences in the H1 genome

including the 55-bp repeat do not enhance expression of chloramphenicol acetyltransferase activity.

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