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Cucurbit[10]uril

Simin Liu, Peter Y. Zavalij, and Lyle Isaacs*

Department of Chemistry and Biochemistry, University of Maryland, College Park, Maryland 20742

Abstract

Melamine diamine **1** is able to displace CB[5] from the CB[10]•CB[5] complex resulting in CB[10] •**1**₂ and precipitated CB[5]•**1**. We were able to isolate free CB[10] by treatment of CB[10]•**1** with acetic anhydride followed by washing with MeOH, DMSO, and water. The spacious cavity of CB [10] is able to complex large guests including a cationic calix[4]arene derivative in its 1,3-alternate form (CB[10]•1,3-*alt*-**3**). The addition of adamantane carboxylic acid (**4**) to CB[10]•**3** triggers a conformational change during the formation of termolecular complex CB[10]•*cone*-**3**•**4**.

In 1981, Mock disclosed the structure of cucurbit[6]uril (CB[6]) and subsequently delineated its outstanding binding properties toward ammonium ions in a series of elegant papers.¹ Nearly 20 years later, the groups of Kim and Day reported the preparation and isolation of the CB[n] homologues CB[5], CB[7], CB[8] and CB[10] as its CB[10]•CB[5] inclusion complex.² With their enhanced cavity size, the new members of the CB[n] family³ display a range of novel properties and applications including gas encapsulation, polarizability enhancement, and supramolecular dendrimer chemistry.⁴ Most notable, however, is the ability of CB[8] to simultaneously bind two aromatic guests which function as molecular machines in response to external stimuli.^{3b,5} In this paper we report the isolation of free CB[10] and disclose its unusual recognition properties. These results suggest that CB[10] will rival CB[8] for use as an advanced component for molecular machines and biomimetic systems.^{3,6}



We isolated CB[10]•CB[5] in good quantities using a modification of the procedure reported by Day.^{2b,2c} After much experimentation we discovered that treating a solution of CB[10] •CB[5] (Figure 1a) with a five equivalents of **1** results in the precipitation of the (CB[5]•**1**)_n exclusion complex and the formation of the CB[10]•**1**₂ inclusion complex (Figure 1b). ¹H NMR and x-ray crystallography indicates that **1** adopts a U-shape⁶ within the cavity of CB [10] (Figure 2); the two equivalents of **1** are arranged in a head-to-tail manner which results in a single set of resonances for H_b and H_c within CB[10]•**1**₂. The second equivalent of **1** is relatively weakly bound to CB[10] and can be removed by washing with MeOH to yield CB [10]•**1** (Figure 1c). Once again, **1** adopts a U-shape within the CB[10]•**1** complex; in this

E-mail: LIsaacs@umd.edu.

instance the top and bottom of CB[10] are differentiated and two sets of resonances are observed for H_b and H_c . Free CB[10] was obtained by heating CB[10]•1 in Ac₂O followed by washing with (CH₃)₂SO, MeOH, and H₂O (Figure 1d). CB[10] is quite stable in acidic solution (>1 month in 20% D₂O/DCl at room temperature) which enabled our investigations of its molecular recognition properties.

CB[10] is insoluble in D₂O (< 50 µM) but its inclusion complexes often are nicely soluble which allows their characterization by NMR. Alternatively, CB[10] can be dissolved in 20% DCl / D₂O for binding studies. An initial screen of many guests revealed that CB[10] – with its cavity volume of ≈ 870 Å³ – undergoes complexation with several chemically and biologically important substances (e.g. dyes, fluorophores, pharmaceuticals, and peptides) although some of these complexes occur as insoluble precipitates (Supporting Information). A soluble, kinetically stable complex was obtained with the more sizable and cationic guest (*R*)-2 which gave exclusively the termolecular complex CB[10]•(*R*)-2₂. Interestingly, when racemic (±)-2 was used, the racemic mixture of homochiral complexes (CB[10]•(*R*)-2₂ and CB [10]•(*S*)-2₂) was preferred relative to the heterochiral *meso*- complex (CB[10]•(*R*)-2•(*S*)-2) by a factor of three (Supporting Information). In combination, these results suggest that CB[10] may find application in drug delivery, for peptide sensing, and even to modulate the behavior of catalysts based on binaphthalene derived ligands.

Given the vast size of the CB[10] cavity we envisioned the encapsulation of smaller host molecules like cyclodextrins, calixarenes, or even CB[6] that would merge the advantageous features of these host families. In the event, only cationic calix[4]arene derivative **3** formed a soluble stable complex (CB[10]•**3** Figure 3a). Based on the number and multiplicity of resonances observed for CB[10]•**3**, we conclude that **3** adopts a mixture of the D_{2d} -symmetric 1,3-alternate conformation and a rapidly equilibrating mixture of cone, 1,2-alternate and partial cone conformers within the CB[10] host. Intrigued by the possibility of using allosteric effects to control the conformation of the macromolecular complex⁷ we studied the binding of small molecule guests to CB[10]•**3**. We found that substituted adamantanes (**4** – **8**) – which do not bind to **3** alone – induce a dramatic change in the conformer distribution during the formation of CB[10]•*cone*-**3**•adamantane complexes (Figure 3b).⁸ Scheme 1 shows an MMFF minimized model of the CB[10]•*cone*-**3**•4 complex.⁹ One of the hallmarks of biological allostery is the reversible response of the system to activator concentration. For this purpose we added stoichiometric amounts of CB[7] which sequesters **4** as its CB[7]•**4** complex^{3b,6d} and resets the system to its original CB[10]•**3** state (Figure 3c).

Just like the smaller CB[n] homologs, CB[10] retains the ability to bind a variety of chemically and biologically important cationic substances within its cavity. We have further demonstrated that CB[10] readily forms termolecular complexes (e.g. CB[10]•2₂ and CB[10]•*cone*-3•4); the vast cavity volume of CB[10] ($\approx 870 \text{ Å}^3$) suggests the potential formation of even higher molecularity complexes. The termolecular complexes already display a range of intriguing behavior including chiral recognition and efficient allosteric control of macromolecular geometry in response to a small molecule (e.g. 4). Overall, these results suggest that CB[10] will find broad application as an advanced component of molecular machines and biomimetic systems.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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- 8. Addition of 1 equiv. 4 to a solution of CB[10]•3 (500 μM) results in ≈ 90% formation of CB[10] *cone-3*•4 reflecting the strong binding of 4 to CB[10] •3. A larger number of equivalents of 5 8 are required to complete the conformation change presumably because of weaker binding interactions of tetracationic CB[10] •3 with these cationic guests.
- 9. The ¹H NMR spectrum of CB[10]•cone-3•4 does not show doubling of the H_g and H_j resonances as expected for the geometry shown in Scheme 1. We attribute this result to a dynamic process in which 4 reorients its CO₂H group between the two portals rapidly on the chemical shift timescale. The bulkier adamantanes 7 and 8 display two sets of resonances as expected.





Figure 1.

¹H NMR spectra (400 MHz, D₂O, 298 K) for: a) CB[10]•CB[5], b) CB[10]•**1**₂, c) CB[10]•**1**, d) CB[10] (20% D₂O/DCl).





Cross-eyed stereoview of the structure of $CB[10] \cdot \mathbf{1}_2$ in the crystal. Solvating water has been removed for clarity.

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Figure 3.

¹H NMR spectra recorded (400 MHz, D_2O / DCl , RT) for: a) CB[10]•3 (1,3-*alt* and dynamic equilibrium beween cone, 1,2-*alt* and partial cone), b) CB[10]•*cone*-3•4 with excess 4 (0.8 equiv.), and c) CB[10]•3 and CB[7]•4. Subscripts: 1,3 = 1,3-*alt*-3; dyn = dynamic equilibrium of 3.



Scheme 1.

Allosteric control of the conformations of CB[10]•3 (MMFF minimized) with 4 (purple) and CB[7].